



Table S1. Modulation of 5-LOX activity and comparison with COX-2 activity by (poly)phenols in cellular models.

1

Cellular model	Metabolite and assay conditions	Biological activity on 5-LOX and(or) COX-2 and molecular associated events	Effect on 5-LOX enzyme	Effect on COX-2 enzyme	References
Guinea pig isolated leukocytes	NDGA (0.5 – 30 μ M)	\downarrow LTB ₄ , 5-HETE and di-hydroxy acids formation (IC ₅₀ ~ 2 μ M) in A23187-stimulated cells in a dose-dependent manner	Not evaluated	Not evaluated	[1]
Isolated peritoneal mouse macrophages (prelabelled with [¹⁴ C]-arachidonic acid) and rat neutrophils. Cell-free assays with soybean 15-LOX were also carried out.	Quercetin and NDGA (0.01-100 μ M).	Effects in A23187-stimulated rat neutrophils: \downarrow LTB ₄ and 5-HETE formation in cells treated with quercetin (IC ₅₀ = 6.2 μ M) and NDGA (IC ₅₀ = 1 μ M); \downarrow PGE ₂ and TxB ₂ synthesis in cells treated with quercetin (IC ₅₀ = 6.2 and 18.0 μ M, respectively) and NDGA (IC ₅₀ = 42.0 and 17.6 μ M, respectively). Effects in zymosan-stimulated mouse macrophages: \downarrow PGE ₂ and 6-Keto-PGF _{1α} formation in cells treated with quercetin (IC ₅₀ = 8.2 and 8.5 μ M, respectively) and NDGA (IC ₅₀ = 2.4 and 4.2 μ M, respectively). Quercetin and NDGA inhibited soybean 15-LOX activity (IC ₅₀ = 3.2 and 3.6 μ M, respectively) in a cell-free assay.	Not evaluated.	Not evaluated.	[2]
Mouse mast tumor cells and rat-isolated platelets. Enzymatic assays with 5-LOX and	Caffeic acid and its phenethyl ester (CAPE) isolated from	\downarrow LTC ₄ and LTD ₄ synthesis (only caffeic acid at 10 and 100 μ M); \downarrow AA-	\downarrow 5-LOX activity by caffeic acid (IC ₅₀ = 3.7) and	\uparrow COX-2 activity by caffeic acid	[3]

COX-2 obtained from cloned mastocytoma P-815,2-E-6 cells were also carried out.	<i>Artemisia rubripes</i> Nakai (0.1-100 μ M).	induced platelet aggregation; \downarrow 12-HETE formation (IC_{50} = 30 μ M); no effect on HHT and TxB_2 formation at <10 μ M, but at higher doses (100 μ M) exerted an inhibitory effect (40% HHT and 80% 12-HETE); no effect on ADP-induced platelet aggregation.	CAPE (IC_{50} = 0.48 μ M)	(64%) and CAPE (188%)	
Human and porcine leukocytes and platelets. COX activity was tested in microsomes from ram vesicular glands.	6,7,4'-trihydroxyisoflavan (1 μ M – 1 mM).	\downarrow 5-HETE, 12-HETE, and LTB_4 formation, while sparing the production of HHT (15 μ M) in activated (human and porcine) leukocytes; \downarrow 12-HETE formation without inhibiting the synthesis of TxB_2 and HHT in platelets (at 100 μ M); IC_{50} values for 12-LOX (human platelets) and 5-LOX (leukocytes) were 22.0 and 1.6 μ M, respectively.	Not evaluated.	\downarrow HHT, $PGF_{2\alpha}$, PGD_2 , and PGE_2 synthesis in ram seminal vesicle microsomes (IC_{50} = 200.0 μ M). Total inhibition at 1 mM.	[4]
Human isolated platelets and leukocytes	NDGA	\downarrow 5-HETE (IC_{50} = 0.2 – 0.3 μ M), LTB_4 (IC_{50} = 0.5 μ M), LTC_4 (IC_{50} = 0.2 – 0.5 μ M), Δ^6 -trans- LTB_4 (IC_{50} = 0.2 μ M), ω -OH- LTB_4 (IC_{50} = 2 μ M), 12-HETE (IC_{50} = 5 – 30 μ M), 5S,12S-diHETE (IC_{50} = 0.2 μ M), 15-HETE (IC_{50} = 30 μ M) and HHT (IC_{50} = 5 – 100 μ M)	Not evaluated	Not evaluated	[5]
Human isolated platelets and leukocytes	NDGA (0.1 – 100 μ M)	\downarrow 5-HETE, LTB_4 , LTC_4 and 12-HETE	Not evaluated	Not evaluated	[6]
Human-isolated healthy donors	Caffeic acid (1 μ M) and esculetin (10 μ M)	\downarrow 5-HPETE and 5-HETE release in Streptolysin 0-stimulated PMNLs.	Not evaluated.	Not evaluated.	[7]

<i>Leishmania donovani</i> -infected macrophages	NDGA (3 μ M)	\downarrow 5-HETE and 12- or 15-HETE in infected cells	Not evaluated	Not evaluated	[8]
Human isolated leukocytes	NDGA	\downarrow 5-HETE (IC_{50} = 0.6 μ M) and weak effect on PGE ₂ formation (IC_{50} = 119 μ M) in A23187-stimulated cells	Not evaluated	Not evaluated	[9]
Human colonic mucosa	NDGA (10 μ g/mL)	\downarrow sulfidopeptide-LTs in A23187-stimulated human colonic mucosa; no significant effect on PGE ₂ formation	Not evaluated	Not evaluated	[10]
Human isolated neutrophils	NDGA (10 μ M)	\downarrow LTB ₄ and 5-HETE formation in A23187-stimulated neutrophils; no effect on HHT synthesis	Not evaluated	Not evaluated	[11]
Human isolated leukocytes.	Gingerdione, [6]-gingerol, and curcumin.	\downarrow 5-HETE release by [6]-gingerol (14% at 20 μ M) as well as by shogaol, capsaicin, gingerdione and curcumin (IC_{50} = 23, 100, 15.0 and 8.0 μ M, respectively); \downarrow PGE ₂ release (IC_{50} = 67, 73, 18.0, 68.0 and 52.0 μ M for shogaol, capsaicin, gingerdione, [6]-gingerol, and curcumin, respectively) in A23187-activated leukocytes.	Not evaluated.	Not evaluated.	[12]
Human isolated PMNL	3,5-, 4,5-, and 3,4-di-O-caffeoylquinic acid, caffeoylmalic acid, caffeoyltartaric acid, rosmarinic acid, caffeic acid, and chlorogenic acid (1 μ M – 1 mM).	\downarrow LTB ₄ and 5-HETE formation in A23187-stimulated PMNLs (IC_{50} = 55.6 – 92.5 μ M and 214 – 918 μ M for all compounds, respectively); \downarrow 15-HETE formation only in the presence of rosmarinic acid (IC_{50} = 455 μ M); \uparrow PGE ₂ in the presence of 3,4-di-O-caffeoylquinic acid, caffeic acid, and caffeoylmalic acid; \downarrow HHT in the presence of the compounds tested (IC_{50} = 80.0 – 90.2 μ M); chlorogenic acid was inactive.	Not evaluated.	Not evaluated.	[13]

Elicited rat peritoneal leukocytes.	Hypolaetin-8-glucoside, hypolaetin, and 14 other flavonoids (1 μ M – 1 mM).	<p>\downarrowLTB₄ and TxB₂ production in A23187-stimulated leukocytes by various flavonoids. Glycosides were less active than their aglycones (i.e., hypolaetin-8-glucoside/hypolaetin; gossypin/gossypetin; rutin/quercetin; naringin/naringenin), with the exception of naringin/naringenin.</p> <p>The flavonoids that stimulated DNA degradation also showed 5-LOX inhibitory activity, whereas those that lacked capacity to degrade DNA were related to COX inhibition.</p>	Not evaluated.	Not evaluated.	[14,15]
Rabbit peritoneal PMNLs	Isoliquiritigenin (10 and 30 μ M).	<p>\downarrow[¹⁴C]5-HETE production in a concentration-dependent manner in A23187-stimulated PMNLs; no inhibitory effect on PLA₂ activity obtained from rabbit platelet sonicates; \downarrow[¹⁴C]12-HETE formation in rabbit platelet sonicates.</p>	Not evaluated.	Not evaluated.	[16]
Porcine leukocytes	Isoflavonoids such as isoflavans and isoflavones (0.5 – 200 μ M)	<p>\downarrow5-LOX products (5-HETE, 5-HPETE, LTB₄ and stereoisomers of LTB₄); structure-activity investigation showed more effective inhibition in the presence of isoflavans</p>	Not evaluated	Not evaluated	[17]
Rat peritoneal neutrophils and human platelets	Curcumin (1–50 μ M).	<p>\downarrowLTB₄ synthesis in stimulated (calcium/A23187) rat peritoneal neutrophils (IC₅₀ = 30 μM; NDGA = 0.5 μM). \downarrowCOX-1 (IC₅₀ = 2 μM) and 12-LOX activity (30 μM) in platelets.</p>	Not evaluated.	Not evaluated.	[18]
Rat resident peritoneal macrophages.	Baicalein (1, 10 and 100 μ M).	<p>\downarrowLTC₄ synthesis in A23187-stimulated macrophages (IC₅₀ = 9.5 μM)</p>	Not evaluated.	Not evaluated.	[19]

Murine resident peritoneal macrophages.	Genistein (1–100 μ M).	<p>↓PGE₂ production (IC₅₀ = 20.0 μM) in response to zymosan, calcium ionophore A23187, and phorbol myristate acetate (PMA) stimulation; ↓LTC₄ production in response to zymosan and calcium ionophore A23187 (IC₅₀ = 15.0 and 10.0 μM, respectively). These effects were not exerted via inhibition of PLA₂, COX or LOX enzymes.</p>	Not evaluated.	Not evaluated.	[20]
Rat peritoneal leukocytes	Tanetin, fisetin, 3-hydroxy-flavone (1 – 100 μ M)	<p>↓LTB₄, TxB₂ and PGE₂ in A23187-stimulated cells in the presence of tanetin (IC₅₀ = 11 and 6–11 μM for 5-LOX and COX-2, respectively), fisetin (IC₅₀ = 11 and 50 μM for 5-LOX and COX-2, respectively) and 3-hydroxyflavone (IC₅₀ = 80 and 6–8 μM for 5-LOX and COX-2, respectively)</p>	Not evaluated	Not evaluated	[21]
Human isolated PMNL	RSV, piceid, 2,3,4',5-tetrahydroxystilbene-2-O-D-glucoside, α -(3,4-dihydroxy-phenyl)-cinnamic acid, α -(3,4-dihydroxy-phenyl)-3-hydroxycinnamic acid, α -(3,4-dihydroxy-phenyl)-4-hydroxycinnamic acid, α -(3,4-dihydroxy-phenyl)-3,4-dihydroxycinnamic acid, 3,3',4-trihydroxystilbene	<p>↓5-HETE, 5,12-diHETE, 15-HETE, HHT and PGE₂ in A23187-stimulated cells in the presence of RSV (IC₅₀ = 8.9, 6.7, 275, 4.4, 9.6 μM, respectively) and 3,3',4-trihydroxystilbene (IC₅₀ = 5.9, 0.63, 6.77, 19, 49 μM, respectively). Piceid only affected 5-HETE (IC₅₀ = 267 μM) and 5,12-diHETE (IC₅₀ = 201 μM) biosynthesis; no effects observed in the presence of the rest of compounds ↓LTC₄ in A23187-stimulated cells in the presence of RSV (IC₅₀ = 1.37 μM) and 3,3',4-trihydroxystilbene (IC₅₀ = 0.88 μM); ↓β-glucuronidase release</p>	Not evaluated	Not evaluated	[22]

		and ↑cAMP concentration in the presence of RSV and 3,3',4-trihydroxystilbene			
RBL-1 and RBL-2H3 cells	Honokiol	↓LTB ₄ and LTC ₄ formation in A23187-stimulated RBL-1 and IgE-treated RBL-2H3 cells; no effect on PLA ₂ , LTC ₄ synthase and LTA ₄ hydrolase activity	↓LTB ₄ and LTC ₄ in cell-free assays	Not evaluated	[23]
Isolated rat Kupffer cells and human phagocytic liver cells.	Silibinin (10, 25, 100 and 200 μM).	↓LTB ₄ formation (IC ₅₀ = 15 μM) and no effect on PGE ₂ formation in A23187-stimulated Kupffer cells. ↓Dose-dependent of O ₂ ⁻ and NO production (IC ₅₀ = 80 μM). No effect on TNF-α formation. ↓LTB ₄ formation in freshly isolated human phagocytic liver cells (10 and 25 μM).	Not evaluated.	Not evaluated.	[24]
Human platelets, white blood and endothelial cells.	Silibinin (1–25 μM to determine LOX products and 1–100 μM to determine COX products).	↓LTB ₄ formation (IC ₅₀ = 15 μM) and Cys-LT formation (IC ₅₀ = 14.5 μM) in A23187-stimulated human granulocytes, but no effect on PGE ₂ formation. ↓PGE ₂ formation in LPS-activated human monocytes (IC ₅₀ = 45 μM). ↓TxB ₂ and 6-keto-PGF _{1α} formation (IC ₅₀ = 69 and 52 μM, respectively), in human platelets and A23187-stimulated endothelial cells, respectively. ↓Hypochlorite production (IC ₅₀ = 7 μM), but not of O ₂ ⁻ production in PMA-activated human granulocytes.	Not evaluated.	Not evaluated.	[25]

Rat platelets and rat PMNL	2-(3,4-Dihydroxyphenyl) ethanol (DPE), 2-(4-Hydroxyphenyl) ethanol, 2-(3,4-dihydroxyphenyl) acetic acid, caffeic acid, protocatechuic acid, <i>p</i> -coumaric acid, <i>o</i> -coumaric acid, vanillic acid, and syringic acid (0.1 μ M – 1 mM)	\downarrow LTB ₄ production in A23187-stimulated PMNLs and 12-HETE production in intact platelets (IC ₅₀ of DPE = 26.0 and 50.0 μ M, respectively). DPE showed the highest inhibitory (compared with the rest of the compounds) activity against 5- and 12-LOX, especially in intact cells.	\downarrow 5-HETE synthesis (IC ₅₀ of DPE = 13.0) (5-LOX isolated from PMNLs)	\downarrow 12-HETE synthesis (IC ₅₀ of DPE = 4.2 μ M), and no effect on TxB ₂ synthesis (COX-1 and 12-LOX isolated from platelets)	[26]
PMNL from healthy donors.	Hydroxytyrosol (0.1 μ M – 1 mM).	\downarrow LTB ₄ production (IC ₅₀ = 1.2 μ M) and its ω -oxidized metabolites (20-hydroxy and 20-carboxy-LTB ₄) in A23187-stimulated PMNLs	Not evaluated.	Not evaluated.	[27]
RBL-1 cells	YPE-01 (derived from yakuchinones), yakuchinone B, dimethyl-yakuchinone B; NDGA was used a control of 5-LOX inhibition	\downarrow LTB ₄ and LTC ₄ biosynthesis in A23187-treated RBL-1 cells in the presence of YPE-01 (IC ₅₀ = 0.035 and 0.046 μ M, respectively), yakuchinone B (IC ₅₀ = 0.49 and 0.61 μ M, respectively), dimethyl-yakuchinone B (IC ₅₀ = 1.14 and 2.11 μ M, respectively) and NDGA (IC ₅₀ = 0.27 and 0.5 μ M, respectively)	\downarrow 5-LOX activity by YPE-01 (IC ₅₀ = 0.28 μ M), yakuchinone B (IC ₅₀ = 0.37 μ M), dimethyl-yakuchinone B (IC ₅₀ = 0.22 μ M) and NDGA (IC ₅₀ = 0.49 μ M) in cell-free assay	\downarrow COX-1 and COX-2 activity by yakuchinone B and dimethyl-yakuchinone B in cell-free assays	[28]
Human PMNL from healthy donors.	<i>trans</i> -RSV (0.44 – 440 μ M).	\downarrow LTB ₄ , 6- <i>trans</i> - and 12- <i>trans</i> -epi-LTB ₄ production in A23187-stimulated PMNLs (IC ₅₀ = 48.2 \pm 7.0 μ M); formation of 5-LOX-derived metabolites was virtually abolished at 220 μ M.	Not evaluated.	Not evaluated.	[29]
Bovine PMNL	Hyperoside, rutoside, and chlorogenic acid (25 – 100 μ M).	No effect on LTB ₄ or 12(S)-HETE biosynthesis at concentrations \leq 100 μ M in A23187-stimulated PMNLs.	Not evaluated.	Not evaluated.	[30]

Human isolated PMNLs and rat basophilic leukemia cells RBL-1	Myr-3-glur and NDGA	<p>↓LTB₄ formation in A23187-stimulated PMNLs treated with Myr-3-glur for 2 h (IC₅₀ = 2.2 μM) or with NDGA for 5 min (IC₅₀ = 0.5 μM); no effect of Myr-3-glur in cells incubated for 5 min.</p> <p>Myr-3-glur inhibited AA-induced platelet aggregation and ↓PGE₂, PGD₂, and PGI₂ biosynthesis ex vivo (to a similar extent as indomethacin)</p> <p>↓COX-1 activity obtained from platelets (IC₅₀ = 0.5 μM) and ram seminal vesicle (IC₅₀ = 10 μM)</p>	<p>↓5-LOX activity in a crude enzyme preparation from RBL-1 (IC₅₀ = 0.1 μM for Myr-3-glur and IC₅₀ = 0.26 μM for NDGA)</p>	<p>↓COX-2 activity (IC₅₀ = 0.26 μM) obtained from sheep placenta</p>	[31]
Rat peritoneal leukocytes.	Gnaphalin, quercetin and galangin (20 – 160 μM).	<p>Gnaphalin effects: ↓LTB₄ production in A23187-stimulated rat peritoneal leukocytes (IC₅₀ = 81.8±12.9 μM; ↓TxB₂ production in rat peritoneal leukocytes stimulated with A23187 (IC₅₀ = 39.9±3.9 μM), chemotactic peptide fMLP (IC₅₀ = 12.0 μM) and AA (IC₅₀ = 57.7±5.1 μM); no effect on the secretion of lysozyme, myeloperoxidase and β-glucuronidase pro-inflammatory enzymes from neutrophil secretory granules; no scavenging effect against hydrogen peroxide or hypochlorous acid.</p> <p>Quercetin effects: inhibition of LTB₄ (100%) and TxB₂ (55%) at 160 μM.</p> <p>Galangin effects: inhibition of LTB₄ (11%) and TxB₂ (84%) at 40 μM.</p>	Not evaluated.	Not evaluated.	[32]
Rat peritoneal leukocytes.	Oleuropein, tyrosol, hydroxytyrosol, and	<p>↓LTB₄ production in A23187-stimulated rat peritoneal leukocytes treated</p>	Not evaluated.	Not evaluated.	[33]

	caffeic acid (40, 100 and 200 μ M).	with hydroxytyrosol, oleuropein, caffeic acid, and tyrosol (IC_{50} = 15, 80, 200, and 500 μ M, respectively); \downarrow ROS production in PMA-stimulated rat leukocytes; no substantial inhibition on TxB_2 exerted by the compounds tested			
Human isolated PMNLs	3-methylcatechol, 4-methylcatechol, 4-nitrocatechol, guaiacol, pyrogallol, propylgallate, 1,2,4-trihydroxybenzene, 1,3,5-trihydroxybenzene (1.8 nM – 1800 μ M)	\downarrow LTB ₄ (IC_{50} = 5 – 900 μ M) and PGE ₂ (IC_{50} = 45 – 900 μ M) synthesis in A23187-stimulated cells (at concentrations higher than 18 μ M); 1,3,5-trihydroxybenzene exerted no inhibition on LTB ₄ formation (IC_{50} >> 1800 μ M)	Not evaluated	Not evaluated	[34]
Human erythroleukemia K562 cells.	RSV (30 μ M).	\downarrow LTB ₄ and PGE ₂ synthesis in H ₂ O ₂ -treated K562 cells; \downarrow purified 15-LOX (IC_{50} = 25.0 \pm 3.0 μ M) activity.	\downarrow purified 5-LOX (IC_{50} = 2.5 \pm 0.3 μ M) activity.	\downarrow PGH synthase (IC_{50} = 20.0 \pm 2.0 μ M) activity	[35]
Rat isolated peritoneal leukocytes	6-hydroxy-kaempferol 3,6-dimethyl ether; 6-hydroxy-kaempferol 3,6,4'-trimethyl ether; quercetagenin 3,6,3'-trimethyl ether; 6-hydroxy-luteolin 6-methyl ether; 6-hydroxy-luteolin 6,3'-dimethyl ether	\downarrow TxB ₂ (IC_{50} = 22 – 182 μ M) and LTB ₄ (IC_{50} = 58 – 182 μ M) biosynthesis in A23187-stimulated cells	Not evaluated	Not evaluated	[36]
RBL-2H3	Magnolol (0.1 – 20 μ M)	\downarrow LTB ₄ and LTC ₄ formation (at 10 and 20 μ M); no effect on IgE-induced β -hexosaminidase release; \downarrow AA release at 10 μ M; \downarrow [Ca ⁺²] release in IgE-treated cells	\downarrow LTB ₄ and LTC ₄ (using AA or LTA ₄ as substrates) formation at 10 and	Not evaluated	[37]

				25 μ M in cell-free assays		
Monocytic leukemia cells MM6, human platelets and PMNLs isolated from healthy donors.	Hyperforin (0.1 – 100 μ M).	\downarrow LTB ₄ and 15-HETE formation in A23187-stimulated PMNLs (IC ₅₀ = 1.0–2.0 and >10 μ M, respectively); \downarrow HHT production in thrombin- or A23187- and exogenous AA-stimulated human platelets (IC ₅₀ = 0.3, 3.0 and 3.0 μ M, respectively) as well as in platelet lysates (IC ₅₀ = 3 μ M). Hyperforin could not interfere with COX-2 product formation (6-keto-PGF _{1α}) in MM6 cells; and did not significantly inhibit 12-LOX or 15-LOX in intact platelets or leukocytes, respectively.	\downarrow purified 5-LOX (IC ₅₀ = 90 nM) activity acting via uncompetitive mechanisms.	Not evaluated.	[38]	
Rat peritoneal leukocytes	Erycristagallin (\leq 100 μ M)	\downarrow LTB ₄ formation in A23187-stimulated leukocytes (IC ₅₀ = 23.4 μ M) without cytotoxic effects; no effect on COX-1 from platelets at 100 μ M	Not evaluated.	Not evaluated.	[39]	
RAW264.7 macrophages and HT-29 human colon cancer cells.	Curcumin and THC (10, 20 and 50 μ M).	\downarrow LTB ₄ and PGE ₂ formation as well as AA release in LPS-stimulated RAW264.7 cells at 10 μ M; \downarrow AA release in A23187-stimulated HT-29 cells at 10 μ M; \downarrow cPLA ₂ activity from microsomal fraction of HT-29 cells; \downarrow cPLA ₂ level and cPLA ₂ phosphorylation in LPS-treated RAW264.7 cells at 50 μ M; \downarrow PGF _{2α} , PGE ₂ , PGD ₂ , HHT, 15-HETE and 11-HETE in LPS-stimulated RAW264.7 cell lysates treated with 0THC at 50 μ M; at the same concentration curcumin reduced the biosynthesis of PGD ₂ , 15-HETE and 11-	\downarrow Human recombinant 5-LOX expression in LPS-treated RAW264.7 cells (IC ₅₀ = 0.7 and 3 μ M for curcumin and THC, respectively).	\downarrow COX-2 expression in LPS-treated RAW264.7 cells (curcumin at 20 μ M), but increase COX-2 levels without LPS stimulation.	[40]	

		HETE; higher inhibitory effect on the peroxidase activity of COX-1 than that of COX-2			
Rat peritoneal leukocytes.	Sigmoidin A and sigmoidin B (5 – 100 μ M).	\downarrow LTB ₄ in calcium/A23187- stimulated rat peritoneal leukocytes treated with sigmoidin A (IC ₅₀ = 31 μ M), sigmoidin B (44% inhibition at the same concentration); apigenin used as reference compound also showed inhibitory activity (IC ₅₀ = 14 μ M). No inhibition of COX-1 activity; no reduction of 12-HHT synthesis at 100 μ M of both compounds.	Not evaluated.	Not evaluated.	[41]
PMNLs from healthy donors.	Curcumin, eugenol, cinnamaldehyde and quercetin (1 – 150 μ M).	\downarrow 5-HETE production in PMNLs (IC ₅₀ = 30.0, 26.0, 35.0 and 25.0 μ M for curcumin, eugenol, cinnamaldehyde and quercetin, respectively); NDGA (IC ₅₀ = 28.0 μ M).	Not evaluated.	Not evaluated.	[42]
Neutrophils isolated from rats	Magnolol (0.1 – 10 μ M)	\downarrow COX products formation, but low effect on 5-LOX products biosynthesis at 3 μ M; Inhibition of COX and 5-LOX products production at 10 μ M; \downarrow TxB ₂ formation (IC ₅₀ = 0.5 μ M); \uparrow LTB ₄ biosynthesis at concentrations <3 μ M, but inhibition at higher concentrations; Magnolol alone modified 5-LOX distribution in the cells, \uparrow AA release (PLA ₂ activity), \uparrow cPLA ₂ translocation and phosphorylation (effects dose-dependent) and had no effect on MAPK phosphorylation. These effects were absent in the presence of A23187.	\downarrow COX-1 and COX-2 activity (IC ₅₀ = 26.0 \pm 1.9 and 31.2 \pm 2.8 μ M, respectively) in cell-free assays	Weak inhibition of 5-LOX activity (IC ₅₀ ~ 90 μ M) in cell-free assay	[43]

BMMC cells	Ginkgetin (1 – 50 μ M).	\downarrow LTC ₄ in BMMC activated with KL (100 ng/ml) (IC ₅₀ = 0.33 μ M); \downarrow PGD ₂ in BMMC activated with KL (100 ng/ml), IL-10 (100 U/ml) and LPS (100 ng/ml) (IC ₅₀ = 0.75 μ M); \downarrow degranulation reaction in a dose dependent manner (IC ₅₀ = 6.52 μ M)	Not evaluated.	\downarrow COX-2 protein level.	[44]
Rat peritoneal neutrophils and RAW 264.7 macrophages	5-O-demethylnobiletin (0.01 – 10 μ M)	\downarrow LTB ₄ formation in rat neutrophils (IC ₅₀ = 0.35 μ M)	\downarrow 5-LOX activity (cell lysate) by 57% at 0.25 μ M	\downarrow COX-2 expression RAW cells (5-20% inhibition)	[45]
Human monocyte line U937.	Procyanidin B ₂ (50 and 100 μ M).	\downarrow LOX-1 protein and mRNA levels in ox-LDL treated macrophages.	\downarrow 5-LOX protein and mRNA levels in ox-LDL treated macrophages.	Not evaluated.	[46]
BMMC	Five coumarins (psoralen, xanthotoxin, scopoletin, umbelliferone, and (+)-marmesin) and three flavonoids (apigenin, luteolin and cynaroside) (12.5 and 25.0 μ g/mL).	\downarrow LTC ₄ production in BMMC activated with KL (100 ng/ml) at 25.0 μ g/mL; \downarrow PGD ₂ formation in BMMC activated with KL (100 ng/ml), IL-10 (100 U/ml) and LPS (100 ng/ml) at 12.5 μ g/mL. All compounds showed COX-2/5-LOX dual inhibitory activity.	Not evaluated.	Not evaluated.	[47]
BMMC cells	Methyl gallate (5 – 160 μ M).	\downarrow LTC ₄ (IC ₅₀ = 5.3 μ M) in BMMC activated with KL (100 ng/ml). \downarrow PGD ₂ (IC ₅₀ = 17.0 μ M) in BMMC activated with KL (100 ng/ml), IL-10 (100 U/ml) and LPS (100 ng/ml).	Not evaluated.	\downarrow COX-2 activity (colorimetric COX inhibitor screening assay kit) (IC ₅₀ = 19.0 μ M); no effect on COX-2 protein levels at \leq 80 μ M.	[48]
BMMC cells	Ochnaflavone (1.5 – 50 μ M).	\downarrow LTC ₄ (IC ₅₀ = 6.5 μ M) in BMMC activated with KL (100 ng/ml).	Not evaluated.	\downarrow COX-2 protein levels.	[49]

		↓PGD ₂ (IC ₅₀ = 0.6 μM) in BMMC activated with KL (100 ng/ml), IL-10 (100 U/ml) and LPS (100 ng/ml); ↓Degranulation reaction in a dose dependent manner (IC ₅₀ = 3.0 μM).			
PMNLs from healthy donors.	Eugenol (5 – 60 μM).	↓LTC ₄ and 5-HETE in A23187- and AA-stimulated PMNLs (IC ₅₀ = 30.0 and 26.0 μM, respectively).	Not evaluated.	Not evaluated.	[50]
Human peripheral blood eosinophils	Genistein (1nM – 10 μM)	↓LTC ₄ in A23187-stimulated cells (IC ₅₀ = 80 nM); ↓p38 and MK2 phosphorylation	↓5-LOX translocation	Not evaluated	[51]
Rat peritoneal leukocytes.	Sakuranetin, 7-O-methylaromadendrin and 3-acetyl-7-O-methylaromadendrin (5 – 100 μM).	↓LTB ₄ production A23187-stimulated rat peritoneal leukocytes treated with 7-O-methylaromadendrin, sakuranetin, and 3-acetyl-7-O-methylaromadendrin (IC ₅₀ = 62, 9 and 15 μM, respectively); as a reference compound, apigenin also showed inhibitory effects (IC ₅₀ = 14 μM); ↓PLA ₂ activity (only 7-O-methylaromadendrin); ↓Elastase release (only sakuranetin at 100 μM); none of the flavanones tested inhibited PGE ₂ biosynthesis in RAW 264.7 mouse macrophages.	↓LTB ₄ production in homogenized rat peritoneal polymorphonuclear leukocytes (only sakuranetin at 20 μM).	Not evaluated.	[52]
Human gastric epithelial cells (AGS) and murine macrophage RAW264.7	NDGA and geraniin (10 μM).	↓5-HETE production in AGS cells; ↓IL-8 and TNF-α in AGS cells and macrophages; attenuation of cPLA ₂ induction by NDGA, but not geraniin.	↓5-LOX protein levels.	No effect on COX-2 protein levels.	[53]
BMMC cells	Meso-dihydroguaiaic acid (0.8 – 25 μM).	↓LTC ₄ (IC ₅₀ = 1.3 μM) in BMMC activated with KL (100 ng/ml); ↓PGD ₂ (IC ₅₀ = 9.8 μM) in BMMC activated with KL (100 ng/ml), IL-10 (100 U/ml) and LPS (100 ng/ml); ↓Degranulation	Not evaluated.	No effect on COX-2 protein levels.	[54]

		reaction in a dose dependent manner (IC ₅₀ = 11.4 μM).			
PMNLs from healthy donors.	Carnosic acid and carnosol (0.03 – 30 μM).	↓LTB ₄ , 5(S),12(S)-diHETE, and 5-H(p)ETE in A23187-stimulated PMNL treated with carnosol and carnosic acid (IC ₅₀ = 7.0 and 15-20 μM, respectively); ↓12-HETE and 12-HPETE synthesis in A23187-stimulated PMNL treated with carnosol (IC ₅₀ = 13 μM), but no effect on cells treated with carnosic acid; Antagonise intracellular Ca ²⁺ mobilisation induced by a chemotactic stimulus, and attenuate formation of ROS and the secretion of human leukocyte elastase.	↓5-LOX activity (IC ₅₀ = 0.1 and 1.0 μM for carnosic acid and carnosol, respectively)	Not evaluated.	[55]
PBMC isolated from whole human blood.	Quercetin, 3'-O-methylquercetin (MQ), quercetin-3'-O-sulfate (QS), quercetin-3-O-glucuronide (QG) and 3'-O-methylquercetin-3-O-glucuronide (MQG) (1 – 10 μM).	↓LTB ₄ synthesis in A23187-stimulated PBMCs treated with quercetin (IC ₅₀ = 2.0 μM) and MQ at 2.0 μM; ↓PGE ₂ formation in LPS-activated PBMCs treated with quercetin and MQ (IC ₅₀ = 2.0 μM); lack of effect for the rest of compounds tested	Not evaluated.	Not evaluated.	[56]
Rat adherent macrophages from granuloma	NDGA (10 μM)	↓LTB ₄ synthesis in stimulated cells	↓5-LOX expression (mRNA and protein)	Not evaluated	[57,58]
BBMCs isolated from mice	Isolated phenolic compounds from <i>Ailanthus altissima</i> leaves	Scopoletin, quercetin and luteolin inhibited the biosynthesis of PGD ₂ (IC ₅₀ = 39.2, 7.3 and 2.5 μM, respectively) and LTC ₄ (IC ₅₀ = 59.9, 5.1 and 1.8 μM, respectively); no effect of astragaloside and scopolin at the concentrations tested.	Not evaluated	Not evaluated	[59]

		The methanolic extract and solvent fractions of <i>Ailanthus altissima</i> inhibited PGD ₂ (49.0 – 57.4%) and LTC ₄ (96.7 – 99.8%) at 50 µg/mL		
Human neutrophils	Hyperforin (0.3 – 30 µM)	↓5-LOX product biosynthesis (IC ₅₀ = 1.9 µM) in intact neutrophils via inhibition of 5-LOX translocation to the nuclear membrane. This effect was reduced in the presence of OAG (IC ₅₀ ≥ 10 µM)	↓5-LOX activity in cell homogenates (IC ₅₀ ~ 30 µM), in partially purified (IC ₅₀ ~ 7 µM) and purified enzyme (IC ₅₀ = 1.6 µM). This inhibition occurs via interaction with the Trp residues within the 5-LOX C2-like domain and blocking the binding of 5-LOX with CLP. Besides, this inhibitory effect was independent of the redox tone and blocked in the presence of OAG (IC ₅₀ ≥ 10 µM) and PC.	Not evaluated [60]
RBL-1 cells and RAW 264.7 cells.	(-)-Nyasol (2–100 µM).	↓Cys-LT production in A23187-treated RBL-1 cells at 2 – 10 µM. As a reference compound, NDGA also exerted inhibitory effects at 5 µM;	Not evaluated.	No effect on COX-2 protein levels. [61]

			↓PGD ₂ and NO production in LPS-treated RAW 264.7 cells at 1 – 100 μM.			
RBL-1 and RAW 264.7 cells	<i>Schisandra fructus</i> methanolic extract and the individual compounds such as vanillic acid; NDGA as control of 5-LOX inhibition		↓5-LOX products formation (Cys-LTs) y RBL-1 cells treated with the extract (IC ₅₀ = 71.1 μg/mL) and NDGA (IC ₅₀ = 0.2 – 1 μM); vanillic acid had no effect; the extract and vanillic acid exerted less than 50% inhibition against PGE ₂ and NO formation at the concentrations tested.	Not evaluated	Not evaluated	[62]
Rat endothelial cells (YPEN-1).	Morin (1 and 5 μM).		↓ROS generation; ↓NF-κB activation (via reduced DNA binding activity, IκBα phosphorylation and p65/p50 nuclear translocation); modulation of ERK and p38 signal transduction; ↓iNOS protein levels in <i>t</i> -BHP-induced YPEN-1 cells.	↓5-LOX protein levels in <i>t</i> -BHP-induced YPEN-1 cells.	↓COX-2 protein levels in <i>t</i> -BHP-induced YPEN-1 cells.	[63]
Murine macrophage RAW264.7 cells.	Curcumin (0.63 – 80 μM).		↓PGE ₂ formation in LPS-stimulated RAW264.7 cells; ↑HO-1 mRNA expression and protein level; no effect on iNOS (mRNA expression and protein level), cPLA ₂ (mRNA expression)	No effect on 5-LOX mRNA expression.	No effect on COX-2 mRNA expression and protein level.	[64]
BMCMC cells	NDGA (3 μM).		↑PGE ₂ and ↓LTB ₄ /LTC ₄ production in BMCMCs stimulated with ConA (mimics IgE cross-linking), substance P (following culture of cells with IL-4) or A23187; ↓degranulation of BMCMCs stimulated with ConA and A23187.	Not evaluated.	Not evaluated.	[65]
RBL-1	13 phenolic compounds isolated from <i>Lonicera japonica</i> (20		The compounds that exerted the highest inhibition on 5-LOX products (Cys-LTs) formation were protocatechuic acid (35.8%), 3,5-	Not evaluated	Not evaluated	[66]

	μM); NDGA as control of 5-LOX inhibition	dicaFFEoylquinic acid methylester (15.1%), luteolin (97.0%), quercetin 3-O-β-D-glucopyranoside (11.1%) and NDGA (100%)			
Human isolated PBMC	Quercetin (25 μM)	↓TLR-2 and TLR-4 mRNA expression; ↓NF-κB activation; ↓IL-6 biosynthesis; ↓iNOS mRNA expression; ↓PGE ₂ biosynthesis	↓5-LOX production;	↓COX-2 mRNA expression; ↓COX production;	[67]
RBL-1 cells	Scopoletin, scopolin, scoparone, esculetin chlorogenic acid, quercetin, isorhamnetin-3-O-galactoside, isorhamnetin-3-O-robinobioside, capillarisin, and NDGA (0.1 – 50 μM)	↓LTB ₄ biosynthesis in A23187-stimulated cells. The most effective compounds were NDGA (95.7% inhibition at 1 μM), as well as esculetin, quercetin, and capillarisin (IC ₅₀ = 6.6, 0.7 and 17.7 μM, respectively).	Not evaluated	Not evaluated	[68]
Rat peritoneal macrophages	<i>Platyclusus orientalis</i> chloroform extract (12.5 – 100 μg/mL) and the individual compounds hinokiol and acacetin (12.5 – 100 μM); 12.5 μM NDGA was used as control of 5-LOX inhibition	↓5-HETE and LTB ₄ ; ↓12-HHT; the extract showed no inhibitory effect on LTA ₄ hydrolase	↓5-LOX activity (43.2% for honokiol and 48.8% for acacetin at 100 μM; IC ₅₀ = 7.1 μM for NDGA) in a cell-free assay	Moderate or weak COX-2 inhibition in cell-free assays	[69]
RBL-2H3 cells	Extracts of <i>Foeniculum vulgare</i> (12.5 – 200 mg/mL), its individual compounds (50 μM) and 1 μM NDGA as positive control	↓5-LOX products (Cys-LTs) formation in cells treated with the extracts (only at 200 μg/mL); the individual phenolics (scopoletin and umbelliferone) had no effect on 5-LOX activity; 1 μM NDGA exerted a strong inhibition (92.4%)	Not evaluated	Not evaluated	[70]

RBL-2H3 cells	Macelignan (5, 10 and 20 μ M)	No cytotoxic effect; $\downarrow\beta$ -hexosaminidase (8 – 32% inhibition) and histamine release (8 – 27% inhibition); \downarrow Ca ²⁺ intracellular concentration; \downarrow PGE ₂ and LTC ₄ biosynthesis; \downarrow IL-4, IL-13, and TNF- α formation; \downarrow Akt and MAPK phosphorylation	\downarrow 5-LOX mRNA expression	\downarrow COX-2 mRNA expression	[71]
PMNL from healthy donors, whole human blood and HEK293 cells	Caffeic acid and CAPE (0.1-30 μ M).	CAPE exerted a \downarrow LT biosynthesis (LTB ₄ , 20-COOH- and 20-OH-LTB ₄ as well as 5-HETE) in thapsigargin-stimulated PMNLs (53-85% inhibition at 1 μ M; IC ₅₀ = 0.52 μ M) as well as in zymosan-stimulated whole blood treated (32% reduction at 1 μ M; IC ₅₀ = 1.79 μ M); CAPE also exerted a \downarrow AA release in thapsigargin-stimulated PMNLs (32% reduction at 1 μ M); Caffeic acid exerted no effects.	CAPE exerted \downarrow 5-LOX activity (IC ₅₀ = 0.13 μ M) whereas caffeic acid had no effect on the enzyme isolated from 5-LOX-transfected HEK293	Not evaluated.	[72]
Rat basophilic leukemia (RBL-2H3) cells.	Panduratin A (1 – 20 μ M).	The effects observed in A23187- and PMA-stimulated RBL-2H3 cells were: \downarrow LTB ₄ and PGE ₂ formation as well as \downarrow AA release; \downarrow Ca ²⁺ influx and β -hexosaminidase and histamine secretion at 20 μ M; \downarrow IL-4, IL-13, and TNF- α mRNA expression; \downarrow Akt, ERK, p38, and JNK phosphorylation	\downarrow 5-LOX mRNA expression.	\downarrow COX-2 mRNA expression.	[73]
Primary avian polymorphonuclear leukocytes.	Caffeic acid (5 μ M).	\downarrow LTB ₄ production in leukocytes stimulated with TLR agonists (FLG, PAM or CpG).	Not evaluated.	Not evaluated.	[74]
Feline esophageal epithelial cells.	Eupatilin (25-200 μ M).	\downarrow H ₂ O ₂ -induced cytotoxicity (dose-dependent effect), \downarrow p38 and JNK activation, and LTB ₄ production at 150 μ M (12 h pretreatment).	\downarrow 5-LOX expression in H ₂ O ₂ -induced cells at 150 μ M (12 h pre-treatment)	Not evaluated.	[75]

Human neutrophils isolated from peripheral blood of healthy donors.	3,3',4'-Tri-hydroxy-flavone, 3,5,7-tri-hydroxy-flavone, 3,7,3'-tri-hydroxy-flavone and 3,7,4'-tri-hydroxy-flavone (10 and 25 μ M), NDGA and quercetin (10 μ M)	\downarrow LTB ₄ production in A23187- and AA-stimulated neutrophils (except for 3,7,3'-tri-hydroxy-flavone at 10 μ M); \downarrow oxidation of luminol, amplex red and APF by reactive species generated by PMA-stimulated neutrophils; \downarrow COX-1 activity in a cell-free system for 3,5,7-tri-hydroxy-flavone and 3,7,3'-tri-hydroxy-flavone in a concentration-dependent manner.	Not evaluated.	\downarrow COX-2 activity in a cell-free system for 3,5,7-tri-hydroxyflavone in a concentration-dependent manner. [76]
RAW264.7 cells and rat peritoneal macrophages.	Baicalein and its glycoside, baicalin (1.5 – 100 μ M).	\downarrow LTB ₄ production in rat peritoneal macrophages in the presence of exogenous AA (IC ₅₀ = 35.6 μ M); \downarrow PGE ₂ in LPS-stimulated RAW264.7 cells by baicalein (IC ₅₀ = 28.6 μ M).	No effect on 5-LOX expression (mRNA and protein level).	\downarrow COX-2 mRNA expression and protein level in LPS-stimulated RAW264.7 cells. [77]
Human isolated PMNL or monocytes	Cannaflavin A and B (0.1 – 10 μ M)	\downarrow 5-LOX products (LTB ₄ , tr-LTB ₄ , 5-HPETE) with IC ₅₀ = 1.6 – 2.4 μ M for cannaflavin A; \downarrow PGE ₂ biosynthesis (IC ₅₀ = 8.8 μ M for cannaflavin A) in LPS stimulated monocytes; \downarrow 12-HHT in platelets (IC ₅₀ > 10 μ M for cannaflavin A)	\downarrow 5-LOX (IC ₅₀ = 0.8 and 0.9 μ M for cannaflavin A and B, respectively) activity	\downarrow mPGES-1 activity by both compounds (cannaflavin A showed IC ₅₀ = 1.8 μ M); \downarrow COX-1 and COX-2 activity (cannaflavin A showed IC ₅₀ > 10 μ M) [78]
Human isolated neutrophils	Luteolin, NDGA, quercetin, 3',4'-dihydroxyflavone, apigenin, acacetin, chrysoeriol, diosmetin, taxifolin, chrysin, (\pm)-eriodictyol (\pm)-naringenin and other related flavonoids (10 and 40 μ M)	The highest inhibition of LTB ₄ synthesis was exerted by luteolin (IC ₅₀ = 1.6 \pm 0.3 μ M), 3',4'-dihydroxyflavone (IC ₅₀ = 1.7 \pm 0.1 μ M), quercetin (IC ₅₀ = 4.0 \pm 1.2 μ M), NDGA (56.5 \pm 2.5% inhibition at 1 μ M) and eriodictyol (53.4 \pm 4.8% inhibition at 40 μ M).	Not evaluated	Not evaluated [79]

BMMC	Homoisoflavanone (5,7-di-hydroxy-3-(3-hydroxy-4-methoxy-benzyl)-chroman-4-one) (1.4 – 14.4 μ M).	\downarrow LTB ₄ , LTC ₄ and PGD ₂ in DNP-IgE and DNP-HSA-activated BMCMCs; \downarrow mRNA level and release of IL-6 and TNF- α ; \downarrow degranulation of BMCMCs; no effect on COX-1 protein levels	\downarrow 5-LOX protein levels in DNP-IgE and DNP-HSA-activated BMCMCs.	\downarrow COX-2 protein levels in DNP-IgE and DNP-HSA-activated BMCMCs.	[80]
BMMCs-isolated from bone marrow of C57BL/6	Curcumin (1 – 10 μ M)	\downarrow LTC ₄ and Ca ⁺² influx in IgE/Ag-induced BMCMCs; \downarrow COX-2-dependent PGD ₂ synthesis; \downarrow cPLA ₂ phosphorylation and translocation; inhibition of NF- κ B, MAPK, and Syk pathways (all the effects were dose-dependent).	\downarrow 5-LOX translocation	Not evaluated	[81]
PMNL from healthy donors.	Pinostilbene, rhapontigenin, isorhapontigenin, RSV, oxyresveratrol, pterostilbene	\downarrow LTB ₄ (IC ₅₀ = 18.5 \pm 7.8 and 9.32 \pm 3.3 μ M for oxyresveratrol and pterostilbene, respectively; IC ₅₀ > 50 μ M for the rest of compounds) Pinostilbene exerted the highest COX-1 and COX-2 inhibition (IC ₅₀ = 1.9 \pm 1.6 and 0.35 \pm 0.23, respectively), whereas rhapontigenin was the weakest compound (IC ₅₀ = 24.55 \pm 10.38 and 36.12 \pm 10.43, respectively)	Molecular docking analysis showed that pterostilbene formed hydrogen bonds with His372 and Thr364	Molecular docking analysis showed that pinostilbene formed hydrogen bonds with Arg129, Phe518 and Gln192;	[82]
PMNL from healthy donors.	Xanthohumol, xanthohumol C, 8-prenylnaringenin, 4-hydroxy-colupulone, humulone, cascadone and humudifucol (up to 10 μ M).	\downarrow LTB ₄ and 5-HPETE in A23187- and AA-stimulated PMNLs treated with xanthohumol (IC ₅₀ = 2.9 μ M); no effects observed with the rest of compounds.	Not evaluated.	Not evaluated.	[83]
Human peripheral neutrophils from human whole blood.	Daidzein, dihydrodaidzein, and equol (0.01 – 1 μ M).	\downarrow LTB ₄ A23187- and AA- stimulated neutrophils (IC ₅₀ = 0.2 μ M for equol; 20% of inhibition for daidzein and dihydrodaidzein 1 μ M); no effect on the enzymatic hydrolysis of LTA ₄ to LTB ₄ ; \downarrow free radical peroxidation of AA (IC ₅₀	Not evaluated.	Not evaluated.	[84]

		= 0.6 μ M for daidzein and >1.0 μ M for equol and dihydrodaidzein); \downarrow MPO activity (IC_{50} = 0.45, 0.7 and >1.0 μ M for equol, dihydrodaidzein and daidzein, respectively)			
Isolated human PMNL	CAPE (1 μ M)	\downarrow 5-LOX products	Molecular docking analysis showed affinity = -8.5 Kcal/mol and interaction with His372	Not evaluated.	[85]
RAW 264.7 macrophages	Isoorientin (1 – 25 μ M; 16 h)	\downarrow TNF- α , IL-1 β , iNOS expression and NF- κ B pathway;	\downarrow 5-LOX protein expression	\downarrow COX-2 protein expression	[86]
Mouse mast cell line (PB-3c).	Daidzein, genistein and their glycosides (daidzin and genistin), equol, quercetin and kaempferol (50 μ M; 48 h).	\downarrow LTB ₄ production A23187- and AA-stimulated PB-3c cells (100% inhibition exerted by equol, quercetin and kaempferol, 72.5% by genistein, and a weak but significant suppression by daidzin and daidzein).	Not evaluated.	Not evaluated.	[87]
Isolated human PMNL and platelets	Baicalein, apigenin, quercetin, CAPE, myricetin (0.1 – 20 μ M)	\downarrow 5-LOX products (including LTB ₄ , 6- <i>trans</i> -LTB ₄ , 6- <i>trans</i> -12- <i>epi</i> -LTB ₄ , 20-OH-LTB ₄ , 20-COOH-LTB ₄ , and 5-HETE); IC_{50} = 11.0 and 5.97 μ M in PMNL for baicalein and quercetin, respectively). \downarrow 12-HETE (IC_{50} = 1.78, 1.34 and 1.28 μ M, respectively) and 12-HHT (IC_{50} = 9.92, 2.91 and 3.79 μ M, respectively) in platelets treated with baicalein, quercetin and CAPE	Molecular docking analysis using soybean 15-LOX; (i) CAPE's affinity = -5.4 kcal/mol and interaction with Arg426, Ser510, His518 and Phe576 (ii) Baicalein's affinity = -1.4 kcal/mol and interaction with	Not evaluated.	[88]

			Fe, His518 and Phe576 (iii) Quercetin's affinity = 0.8 kcal/mol and interaction with Asp766, His518 and Phe576		
Isolated human PMNL and whole blood	CAPE (0.1 – 10 μ M)	\downarrow 5-LOX products (including LTB ₄ , tr-LTB ₄ , 20-OH-LTB ₄ , 20-COOH-LTB ₄ , and 5-HETE); IC ₅₀ = 0.97 and 3.58 μ M in PMNL and whole blood, respectively)	Molecular docking analysis into the active site of 5-LOX (crystal structure) showed affinity = -8.8 Kcal/mol and interaction with Leu420 and His372	Not evaluated	[89]
Caco-2	Hydroxytyrosol (1 μ M)	\downarrow PGE ₂ , LTB ₄ , 5-HETE, 12-HETE, 15-HETE, 13-HODE	Not evaluated.	Not evaluated.	[90]
RAW 264.7 macrophages	Ellagic acid-3,3',4-trimethoxy-4'-O- α -L-rhamnopyranoside (1 – 100 μ M for toxicity assays; 1 – 5 μ M for anti-inflammatory assays)	\downarrow NO, TNF- α , IL-6 and NF- κ B expression; \uparrow IL-10	\downarrow 5-LOX level in culture medium	\downarrow COX-2 level in culture medium	[91]
Human monocytes, neutrophils and platelets.	Ginkgolic acid (0.1 – 10 μ M).	\downarrow Tr-LTB ₄ isomers and 5-HPETE in A23187-stimulated intact human neutrophils in the presence or absence of exogenous AA (IC ₅₀ = 3.8 and 2.9 μ M, respectively); \downarrow 12-HHT formation by isolated COX-1 (IC ₅₀ = 8.1 μ M); \downarrow 12-HHT and TxB ₂ formation in AA-stimulated platelets (IC ₅₀ = 2.1 and 2.2 μ M,	\downarrow Tr-LTB ₄ isomers and 5-HPETE synthesized by human recombinant 5-LOX (IC ₅₀ = 0.2 μ M)	No effect on COX-2 activity (human recombinant isolated enzyme)	[92]

		respectively); ↓PGE ₂ , PGD ₂ , TxB ₂ , PGF _{2α} , 11-HETE, 12-HHT, PGE ₁ and TxB ₁ in LPS-stimulated monocytes.			
PBMCs from healthy donors.	Apigenin, EGCG, genistein, naringenin, nobiletin, wogonin), RSV and its dimer (ε-viniferin), tetramer (hopeaphenol), and imine analogues (IRA) (1 – 10 μM)	↓LTB ₄ production by A23187- stimulated neutrophils treated with genistein, resveratrol, IRA (1 and 10 μM) and ε-viniferin (10 μM); LC-MS based targeted oxylipin metabolomics showed that: RSV inhibited the COX-1/2 pathway and exerted a weak attenuation of the 12/15-LOX activity; ε-viniferin caused a clear substrate shunt towards the remaining AA cascade enzymes (15-LOX, COX-1/2, cytochrome P450); IRA had no impact on 15-LOX activity, but elevated the formation of COX-derived PGs, having no inhibitory effects on COX-1/2 activity.	Not evaluated.	Not evaluated.	[93]
Murine mast cell line (MC/9).	Kuwanon G and Morusin (0.5–10 μM).	↓LTB ₄ production in PMA- and A23187-stimulated MC/9 mast cells treated with morusin (2 and 4 μM); ↓histamine production by kuwanon G (2.5–10 μM) and morusin (4 μM).	↓5-LOX nuclear protein level, but not cytoplasmic level in PMA- and A23187- stimulated MC/9 mast cells treated with kuwanon G (2.5 – 10 μM) and morusin (4 μM)	Not evaluated.	[94]
BMMC	Taxifolin (dihydroquercetin) (1, 2 and 5 μM).	↓LTC ₄ in IgE/Ag-stimulated and DNP-HSA-activated BMMCs.	↓5-LOX nuclear protein levels in	↓COX-2 protein levels in IgE/Ag-stimulated and	[95]

		<p>↓IL-6 and MAPKs protein levels, translocation of cPLA₂ and Akt/IKK/NF-κB pathway in IgE/Ag-stimulated and DNP-HSA-activated BMMCs.</p> <p>↓degranulation, histamine release, phosphorylation of PLCγ, and Ca²⁺ mobilization in IgE/Ag-stimulated and DNP-HSA-activated BMMCs.</p>	<p>IgE/Ag-stimulated and DNP-HSA-activated BMMCs.</p>	<p>DNP-HSA-activated BMMCs.</p>	
PMNLs from healthy donors.	<p>Uro-A, Uro-B, Uro-C and IsoUro-A, and their conjugates Uro-A glur, Uro-B glur, IsoUro-A-glur and Uro-A sulfate (1, 5 and 15 μM).</p>	<p>↓LTB₄ and 5-HETE (dose-dependent) synthesis in A23187/LPS-stimulated PMNLs treated with Uro-C (15 μM); ↓PGE₂ formation A23187/LPS-stimulated PMNLs treated with Uro-A or IsoUro-A (15 μM); ↓HKE₂ and HKD₂ production (5-LOX/COX-2 pathway) in A23187/LPS-stimulated PMNLs treated with Uro-A, IsoUro-A, and Uro-C at 15 μM; ↓HKD₂ in the cells treated with Uro-B at 15 μM.</p>	<p>↑No effect on 5-LOX protein level in A23187/LPS-stimulated PMNLs treated with Uro-C at 15 μM.</p>	<p>↓COX-2 protein level in A23187/LPS-stimulated PMNLs treated with Uro-A and IsoUro-A at 15 μM; no effect of these compounds on COX-2 activity</p>	[96]
Caco-2	<p>5,7 di-hydroxy-3,3',4'-trimethoxyflavone and 3,5,6,7,4'-pentamethoxy-flavone</p>	<p>↓TNF-α and IL-1β as well as NF-κB activation compared to LPS-treated cells.</p>	<p>No effect on 5-LOX expression</p>	<p>↓COX-2 expression in LPS-treated cells</p>	[97]
Human neutrophils	<p>NDGA (1 μM)</p>	<p>↓TLB₄, tr-LTB₄, 20-OH-LTB₄, 5-HETE, 12-HETE and 15-HETE</p>	<p>No effect on 5-LOX expression</p>	<p>No effect on COX-2 expression</p>	[98]
RAW 264.7 macrophages	<p>Celery ethanol extract, and karafsin (apigenin-7-O-β-(5'-E-p-coumaroyl)-β-apiofuranoside)</p>	<p>No cytotoxic effects; ↓NO release (0.0625 – 1 mg/mL).</p>	<p>↓5-LOX activity (25 – 500 mg/mL)</p>	<p>↓COX-2 activity (25 – 500 mg/mL)</p>	[99]

Rat basophilic leukemia cell line (RBL-2H3) cells.	Tricin (30.2 μ M – 1.5 mM).	\downarrow LTB ₄ , LTC ₄ and PGE ₂ formation in anti-DNP-IgE plus DNP-HAS-stimulated RBL-2H3 cells; \downarrow hexosaminidase release; \downarrow IL-4 and TNF- α level; \downarrow cPLA2, Akt, ERK, p38, JNK, protein kinase C, phospholipase C1, Lyn and Syk phosphorylation; minimal effect on Fyn.	\downarrow 5-LOX protein levels in anti-DNP-IgE plus DNP-HAS-stimulated RBL-2H3 cells.	\downarrow COX-2 protein levels in anti-DNP-IgE plus DNP-HAS-stimulated RBL-2H3 cells.	[100]
Rat basophilic leukemia RBL-2H3 cells and human non-small-cell lung carcinoma A549 cells.	Red-kerneled rice proanthocyanidin -RRP (catechin octamer) and catechin monomer (1 – 10 μ M).	\downarrow LTB ₄ and PGE ₂ formation in A23187-stimulated RBL-2H3 and A549 cells treated with RRP at 1 μ M; RRP exerted inhibition of 12-LOX (IC ₅₀ = 39.0 μ M) and p12-LOX (IC ₅₀ = 47.5 μ M) activity; weak inhibitory effect on 15-LOX-2 (IC ₅₀ >100 μ M); catechin exerted \downarrow 15-LOX-2 (IC ₅₀ = 20.4 μ M) and COX-1 (IC ₅₀ = 30 μ M)	RRP and catechin exerted non-competitive inhibition against human 5-LOX (IC ₅₀ = 15.1 and 2.2 μ M, respectively) and rat 5-LOX (IC ₅₀ = 7.0 and 9.0 μ M, respectively)	RRP exerted no effect on COX-1 or COX-2 activity up to 500 μ M	[101]

Abbreviations: **AA**, arachidonic acid; **ADP**, adenosine diphosphate; **APF**, 2-[6-(4'-amino)phenoxy-3H-xanthen-3-on-9-yl]benzoic acid; **BMCMCs**, canine bone marrow derived cultured mast cells; **BMMC**, mice bone marrow-derived mast cells; **cAMP**, cyclic adenosine monophosphate; **CAPE**, caffeic acid phenethyl ester; **CLP**, coactosin-like protein; **ConA**, concanavalin A; **COX**, cyclooxygenase; **CpG**, unmethylated CpG motifs of bacteria DNA; **cPLA2**, cytoplasmic phospholipase-2; **Cys-LTs**, cysteinyl leukotrienes; **DNP-HAS**, dinitrophenyl human serum albumin; **DNP-IgE**, dinitrophenyl immunoglobulin E; **DPE**, 2-(3,4-dihydroxy-phenyl)-ethanol; **EGCG**, epigallocatechin gallate; **FLG**, flagellin; **fMLP**, formyl methionyl leucyl phenylalanine; **HETE**, hydroxyeicosatetraenoic acid; **HHT**, hydroxy-5,8,10-heptadecatetraenoic acid; **HKE₂**, hemiketal E₂; **HKD₂**, hemiketal D₂; **HODE**, hydroxyoctadecadienoic acid; **HO-1**, hemoxygenase-1; **HPETE**, hydroxyperoxyeicosatetraenoic acid; **iNOS**, inducible nitric oxide synthase; **KL**, c-kit ligand; **LOX**, lipoxygenase; **LOX-1**, lectin-like oxidized low-density lipoprotein receptor-1; **LPS**, lipopolysaccharide; **LT**, leukotrienes; **MAPK**, mitogen activated protein kinase; **MK2**, mitogen-activated protein kinase-activated protein kinase2; **mPGES**, microsomal prostaglandin E synthase; **MPO**, myeloperoxidase; **MQ**, 3'-O-methylquercetin; **MQG**, 3'-O-methylquercetin-3-O-glucuronide; **Myr-3-glur**, myricetin-3-O- β -D-glucuronide; **NDGA**, nordihydroguaiaretic acid; **NF- κ B**, nuclear factor kappa-light-chain-enhancer of activated B cells; **NO**, nitric oxide; **OAG**, 1-oleoyl-2-acetyl glycerol; **ox-LDL**, oxidized low density lipoprotein; **p12-LOX**, porcine leukocyte-like 12 lipoxygenase; **PAM**, bacterial lipoprotein mimic palmitoyl-3-cysteine-serine-lysine-4; **PBMCs**, peripheral blood mononuclear cells; **PC**, phosphatidil coline; **PG**, prostaglandins; **PLA₂**, phospholipase-2; **PMA**, phorbol 12-myristate 13-acetate; **PMNLs**, polymorphonuclear leukocytes; **QG**, quercetin-3-O-glucuronide; **QS**, quercetin-3'-O-sulfate; **ROS**, reactive oxygen species; **RRP**, red-kerneled rice proanthocyanidin; **RSV**, resveratrol; **THC**, tetrahydrocurcumin; **TLR**, toll-like receptor; **TNF- α** , tumor necrosis factor alpha; **Tr-LTB₄**, trans-LTB₄; **Trp**, tryptophan; **TxB₂**, thromboxane B₂; **Uro**, urolithins;

References

1. Bokoch, G.M.; Reed, P.W. Evidence for Inhibition of Leukotriene A4 Synthesis by 5,8,11,14-Eicosatetraenoic Acid in Guinea Pig Polymorphonuclear Leukocytes. *J. Biol. Chem.* **1981**, *256*, 4156–4159.
2. Chang, J.; Skowronek, M.D.; Cherney, M.L.; Lewis, A.J. Differential Effects of Putative Lipoxygenase Inhibitors on Arachidonic Acid Metabolism in Cell-Free and Intact Cell Preparations. *Inflammation* **1984**, *8*, 143–155, doi:10.1007/BF00916090.
3. Koshihara, Y.; Neichi, T.; Murota, S.; Lao, A.; Fujimoto, Y.; Tatsuno, T. Caffeic Acid Is a Selective Inhibitor for Leukotriene Biosynthesis. *Biochim. Biophys. Acta* **1984**, *792*, 92–97.
4. Kuhl, P.; Shiloh, P.; Jha, H.; Murawski, U.; Zilliken, F. 6,7,4'-Trihydroxyisoflavan: A Potent and Selective Inhibitor of 5-Lipoxygenase in Human and Porcine Peripheral Blood Leukocytes. *Prostaglandins* **1984**, *28*, 783–804, doi:10.1016/0090-6980(84)90035-2.
5. Salari, H.; Braquet, P.; Borgeat, P. Comparative Effects of Indomethacin, Acetylenic Acids, 15-HETE, Nordihydroguaiaretic Acid and BW755C on the Metabolism of Arachidonic Acid in Human Leukocytes and Platelets. *Prostaglandins. Leukot. Med.* **1984**, *13*, 53–60, doi:10.1016/0262-1746(84)90102-1.
6. Sirois, P.; Saura, C.; Salari, H.; Borgeat, P. Comparative Effects of Etodolac, Indomethacin, and Benoxaprofen on Icosanoid Biosynthesis. *Inflammation* **1984**, *8*, 353–364, doi:10.1007/BF00918212.
7. Bremm, K.D.; König, W.; Pfeiffer, P.; Rauschen, I.; Theobald, K.; Thelestam, M.; Alouf, J.E. Effect of Thiol-Activated Toxins (Streptolysin O, Alveolysin, and Theta Toxin) on the Generation of Leukotrienes and Leukotriene-Inducing and -Metabolizing Enzymes from Human Polymorphonuclear Granulocytes. *Infect. Immun.* **1985**, *50*, 844–851, doi:10.1128/iai.50.3.844-851.1985.
8. Reiner, N.E.; Malemud, C.J. Arachidonic Acid Metabolism by Murine Peritoneal Macrophages Infected with *Leishmania Donovanii*: In Vitro Evidence for Parasite-Induced Alterations in Cyclooxygenase and Lipoxygenase Pathways. *J. Immunol. Baltim. Md 1950* **1985**, *134*, 556–563.
9. Bockor, A.M.; Pugsley, T.A. Effect of CI-922, a Potential New Antiallergy Agent, on Arachidonic Acid Metabolism in Vitro. *Inflammation* **1986**, *10*, 435–441, doi:10.1007/BF00915827.
10. Dreyling, K.W.; Hoppe, U.; Peskar, B.A.; Morgenroth, K.; Kozushek, W.; Peskar, B.M. Leukotriene Synthesis by Human Gastrointestinal Tissues. *Biochim. Biophys. Acta* **1986**, *878*, 184–193, doi:10.1016/0005-2760(86)90145-1.
11. Nielsen, O.H.; Bukhave, K.; Ahnfelt-Rønne, I.; Elmgreen, J. Arachidonic Acid Metabolism in Human Neutrophils: Lack of Effect of Cyclosporine A. *Int. J. Immunopharmacol.* **1986**, *8*, 419–426, doi:10.1016/0192-0561(86)90126-8.
12. Flynn, D.L.; Rafferty, M.F.; Bockor, A.M. Inhibition of Human Neutrophil 5-Lipoxygenase Activity by Gingerdione, Shogaol, Capsaicin and Related Pungent Compounds. *Prostaglandins. Leukot. Med.* **1986**, *24*, 195–198, doi:10.1016/0262-1746(86)90126-5.

13. Kimura, Y.; Okuda, H.; Okuda, T.; Hatano, T.; Arichi, S. Studies on the Activities of Tannins and Related Compounds, X. Effects of Caffeetannins and Related Compounds on Arachidonate Metabolism in Human Polymorphonuclear Leukocytes. *J. Nat. Prod.* **1987**, *50*, 392–399, doi:10.1021/np50051a009. 29 30
14. Moroney, M.A.; Alcaraz, M.J.; Forder, R.A.; Carey, F.; Hoult, J.R. Selectivity of Neutrophil 5-Lipoxygenase and Cyclo-Oxygenase Inhibition by an Anti-Inflammatory Flavonoid Glycoside and Related Aglycone Flavonoids. *J. Pharm. Pharmacol.* **1988**, *40*, 787–792, doi:10.1111/j.2042-7158.1988.tb05173.x. 31 32
15. Laughton, M.J.; Evans, P.J.; Moroney, M.A.; Hoult, J.R.; Halliwell, B. Inhibition of Mammalian 5-Lipoxygenase and Cyclo-Oxygenase by Flavonoids and Phenolic Dietary Additives. Relationship to Antioxidant Activity and to Iron Ion-Reducing Ability. *Biochem. Pharmacol.* **1991**, *42*, 1673–1681, doi:10.1016/0006-2952(91)90501-u. 33 34 35
16. Yamamoto, S.; Aizu, E.; Jiang, H.; Nakadate, T.; Kiyoto, I.; Wang, J.C.; Kato, R. The Potent Anti-Tumor-Promoting Agent Isoliquiritigenin. *Carcinogenesis* **1991**, *12*, 317–323, doi:10.1093/carcin/12.2.317. 36 37
17. Voss, C.; Sepulveda-Boza, S.; Zilliken, F.W. New Isoflavonoids as Inhibitors of Porcine 5-Lipoxygenase. *Biochem. Pharmacol.* **1992**, *44*, 157–162, doi:10.1016/0006-2952(92)90049-o. 38 39
18. Ammon, H.P.; Safayhi, H.; Mack, T.; Sabieraj, J. Mechanism of Antiinflammatory Actions of Curcumine and Boswellic Acids. *J. Ethnopharmacol.* **1993**, *38*, 113–119, doi:10.1016/0378-8741(93)90005-p. 40 41
19. Butenko, I.G.; Gladtschenko, S.V.; Galushko, S.V. Anti-Inflammatory Properties and Inhibition of Leukotriene C4 Biosynthesis in Vitro by Flavonoid Baicalein from *Scutellaria Baicalensis* Georgy Roots. *Agents Actions* **1993**, *39 Spec No*, C49-51, doi:10.1007/BF01972717. 42 43
20. Glaser, K.B.; Sung, A.; Bauer, J.; Weichman, B.M. Regulation of Eicosanoid Biosynthesis in the Macrophage. Involvement of Protein Tyrosine Phosphorylation and Modulation by Selective Protein Tyrosine Kinase Inhibitors. *Biochem. Pharmacol.* **1993**, *45*, 711–721, doi:10.1016/0006-2952(93)90147-o. 44 45
21. HOULT, J.R.S.; PANG, L. -H.; BLAND-WARD, P.A.; FORDER, R.A.; WILLIAMS, C.A.; HARBORNE, J.B. Inhibition of Leucocyte 5-Lipoxygenase and Cyclo-oxygenase but Not Constitutive Nitric Oxide Synthase by Tanetin, a Novel Flavonol Derived from Feverfew, *Tanacetum Parthenium*. *Pharm. Pharmacol. Commun.* **1995**, *1*, 71–74, doi:10.1111/j.2042-7158.1995.tb00394.x. 46 47 48
22. Kimura, Y.; Okuda, H.; Kubo, M. Effects of Stilbenes Isolated from Medicinal Plants on Arachidonate Metabolism and Degranulation in Human Polymorphonuclear Leukocytes. *J. Ethnopharmacol.* **1995**, *45*, 131–139, doi:10.1016/0378-8741(94)01206-f. 49 50
23. Hamasaki, Y.; Muro, E.; Miyanji, S.; Yamamoto, S.; Kobayashi, I.; Sato, R.; Zaitu, M.; Matsuo, M.; Ichimaru, T.; Tasaki, H.; et al. Inhibition of Leukotriene Synthesis by Honokiol in Rat Basophilic Leukemia Cells. *Int. Arch. Allergy Immunol.* **1996**, *110*, 278–281, doi:10.1159/000237299. 51 52
24. Dehmlow, C.; Erhard, J.; de Groot, H. Inhibition of Kupffer Cell Functions as an Explanation for the Hepatoprotective Properties of Silibinin. *Hepatol. Baltim. Md* **1996**, *23*, 749–754, doi:10.1053/jhep.1996.v23.pm0008666328. 53 54
25. Dehmlow, C.; Murawski, N.; de Groot, H. Scavenging of Reactive Oxygen Species and Inhibition of Arachidonic Acid Metabolism by Silibinin in Human Cells. *Life Sci.* **1996**, *58*, 1591–1600, doi:10.1016/0024-3205(96)00134-8. 55 56

26. Kohyama, N.; Nagata, T.; Fujimoto, S.; Sekiya, K. Inhibition of Arachidonate Lipoxygenase Activities by 2-(3,4-Dihydroxyphenyl)Ethanol, a Phenolic Compound from Olives. *Biosci. Biotechnol. Biochem.* **1997**, *61*, 347–350, doi:10.1271/bbb.61.347. 57
27. Petroni, A.; Blasevich, M.; Papini, N.; Salami, M.; Sala, A.; Galli, C. Inhibition of Leukocyte Leukotriene B4 Production by an Olive Oil-Derived Phenol Identified by Mass-Spectrometry. *Thromb. Res.* **1997**, *87*, 315–322, doi:10.1016/s0049-3848(97)00133-3. 58
28. Yamazaki, R.; Aiyama, R.; Matsuzaki, T.; Hashimoto, S.; Yokokura, T. Anti-Inflammatory Effect of YPE-01, a Novel Diarylheptanoid Derivative, on Dermal Inflammation in Mice. *Inflamm. Res. Off. J. Eur. Histamine Res. Soc. Al* **1998**, *47*, 182–186, doi:10.1007/s000110050315. 59
29. Rotondo, S.; Rajtar, G.; Manarini, S.; Celardo, A.; Rotillo, D.; de Gaetano, G.; Evangelista, V.; Cerletti, C. Effect of Trans-Resveratrol, a Natural Polyphenolic Compound, on Human Polymorphonuclear Leukocyte Function. *Br. J. Pharmacol.* **1998**, *123*, 1691–1699, doi:10.1038/sj.bjp.0701784. 60
30. Müller, K.; Ziereis, K.; Paper, D.H. Ilex Aquifolium: Protection against Enzymatic and Non-Enzymatic Lipid Peroxidation. *Planta Med.* **1998**, *64*, 536–540, doi:10.1055/s-2006-957509. 61
31. Hiermann, A.; Schramm, H.W.; Laufer, S. Anti-Inflammatory Activity of Myricetin-3-O-b-D-Glucuronide and Related Compounds. **1998**, *47*, 7. 62
32. de la Puerta, R.; Forder, R.A.; Hoult, J.R. Inhibition of Leukocyte Eicosanoid Generation and Radical Scavenging Activity by Gnaphalin, a Lipophilic Flavonol Isolated from Helichrysum Picardii. *Planta Med.* **1999**, *65*, 507–511, doi:10.1055/s-1999-14005. 63
33. de la Puerta, R.; Ruiz Gutierrez, V.; Hoult, J.R. Inhibition of Leukocyte 5-Lipoxygenase by Phenolics from Virgin Olive Oil. *Biochem. Pharmacol.* **1999**, *57*, 445–449, doi:10.1016/s0006-2952(98)00320-7. 64
34. Alanko, J.; Riutta, A.; Holm, P.; Mucha, I.; Vapaatalo, H.; Metsä-Ketelä, T. Modulation of Arachidonic Acid Metabolism by Phenols: Relation to Their Structure and Antioxidant/Prooxidant Properties. *Free Radic. Biol. Med.* **1999**, *26*, 193–201, doi:10.1016/s0891-5849(98)00179-8. 65
35. MacCarrone, M.; Lorenzon, T.; Guerrieri, P.; Agrò, A.F. Resveratrol Prevents Apoptosis in K562 Cells by Inhibiting Lipoxygenase and Cyclooxygenase Activity. *Eur. J. Biochem.* **1999**, *265*, 27–34, doi:10.1046/j.1432-1327.1999.00630.x. 66
36. Williams, C.A.; Harborne, J.B.; Geiger, H.; Hoult, J.R. The Flavonoids of Tanacetum Parthenium and T. Vulgare and Their Anti-Inflammatory Properties. *Phytochemistry* **1999**, *51*, 417–423, doi:10.1016/s0031-9422(99)00021-7. 67
37. Hamasaki, Y.; Kobayashi, I.; Zaitu, M.; Tsuji, K.; Kita, M.; Hayasaki, R.; Muro, E.; Yamamoto, S.; Matsuo, M.; Ichimaru, T.; et al. Magnolol Inhibits Leukotriene Synthesis in Rat Basophilic Leukemia-2H3 Cells. *Planta Med.* **1999**, *65*, 222–226, doi:10.1055/s-1999-13984. 68
38. Albert, D.; Zündorf, I.; Dingermann, T.; Müller, W.E.; Steinhilber, D.; Werz, O. Hyperforin Is a Dual Inhibitor of Cyclooxygenase-1 and 5-Lipoxygenase. *Biochem. Pharmacol.* **2002**, *64*, 1767–1775, doi:10.1016/s0006-2952(02)01387-4. 69
39. Njamen, D. Anti-Inflammatory Activity of Erycristagallin, a Pterocarpene from Erythrina Mildbraedii. *Eur. J. Pharmacol.* **2003**, *468*, 67–74, doi:10.1016/S0014-2999(03)01664-9. 70
- 71 72 73 74 75 76 77 78 79 80 81 82 83

40. Hong, J.; Bose, M.; Ju, J.; Ryu, J.-H.; Chen, X.; Sang, S.; Lee, M.-J.; Yang, C.S. Modulation of Arachidonic Acid Metabolism by Curcumin and Related Beta-Diketone Derivatives: Effects on Cytosolic Phospholipase A(2), Cyclooxygenases and 5-Lipoxygenase. *Carcinogenesis* **2004**, *25*, 1671–1679, doi:10.1093/carcin/bgh165. 84–86
41. Njamien, D.; Mbafor, J.T.; Fomum, Z.T.; Kamanyi, A.; Mbanya, J.-C.; Recio, M.C.; Giner, R.M.; Máñez, S.; Ríos, J.L. Anti-Inflammatory Activities of Two Flavanones, Sigmoidin A and Sigmoidin B, from *Erythrina Sigmoidea*. *Planta Med.* **2004**, *70*, 104–107, doi:10.1055/s-2004-815484. 87–88
42. Prasad, N.S.; Raghavendra, R.; Lokesh, B.R.; Naidu, K.A. Spice Phenolics Inhibit Human PMNL 5-Lipoxygenase. *Prostaglandins Leukot. Essent. Fatty Acids* **2004**, *70*, 521–528, doi:10.1016/j.plefa.2003.11.006. 89–90
43. Hsu, M.-F.; Lu, M.-C.; Tsao, L.-T.; Kuan, Y.-H.; Chen, C.-C.; Wang, J.-P. Mechanisms of the Influence of Magnolol on Eicosanoid Metabolism in Neutrophils. *Biochem. Pharmacol.* **2004**, *67*, 831–840, doi:10.1016/j.bcp.2003.09.040. 91–92
44. Son, J.K.; Son, M.J.; Lee, E.; Moon, T.C.; Son, K.H.; Kim, C.-H.; Kim, H.P.; Kang, S.S.; Chang, H.W. Ginkgetin, a Biflavone from Ginkgo Biloba Leaves, Inhibits Cyclooxygenases-2 and 5-Lipoxygenase in Mouse Bone Marrow-Derived Mast Cells. *Biol. Pharm. Bull.* **2005**, *28*, 2181–2184, doi:10.1248/bpb.28.2181. 93–94
45. Bas, E.; Recio, M.C.; Giner, R.M.; Máñez, S.; Cerdá-Nicolás, M.; Ríos, J.-L. Anti-Inflammatory Activity of 5- O -Demethylnobiletin, a Polymethoxyflavone Isolated from *Sideritis Tragoriganum*. *Planta Med.* **2006**, *72*, 136–142, doi:10.1055/s-2005-873191. 95–96
46. Chen, D.-M.; Cai, X.; Kwik-Urbe, C.L.; Zeng, R.; Zhu, X.-Z. Inhibitory Effects of Procyanidin B(2) Dimer on Lipid-Laden Macrophage Formation. *J. Cardiovasc. Pharmacol.* **2006**, *48*, 54–70, doi:10.1097/01.fjc.0000242052.60502.21. 97–98
47. Kim, J.S.; Kim, J.C.; Shim, S.H.; Lee, E.J.; Jin, W.; Bae, K.; Son, K.H.; Kim, H.P.; Kang, S.S.; Chang, H.W. Chemical Constituents of the Root of *Dystaenia Takeshimana* and Their Anti-Inflammatory Activity. *Arch. Pharm. Res.* **2006**, *29*, 617–623, doi:10.1007/BF02968244. 99–100
48. Kim, S.J.; Jin, M.; Lee, E.; Moon, T.C.; Quan, Z.; Yang, J.H.; Son, K.H.; Kim, K.-U.; Son, J.K.; Chang, H.W. Effects of Methyl Gallate on Arachidonic Acid Metabolizing Enzymes: Cyclooxygenase-2 and 5-Lipoxygenase in Mouse Bone Marrow-Derived Mast Cells. *Arch. Pharm. Res.* **2006**, *29*, 874–878, doi:10.1007/BF02973908. 101–103
49. Son, M.J.; Moon, T.C.; Lee, E.K.; Son, K.H.; Kim, H.P.; Kang, S.S.; Son, J.K.; Lee, S.H.; Chang, H.W. Naturally Occurring Biflavonoid, Ochnaflavone, Inhibits Cyclooxygenases-2 and 5-Lipoxygenase in Mouse Bone Marrow-Derived Mast Cells. *Arch. Pharm. Res.* **2006**, *29*, 282–286, doi:10.1007/BF02968571. 104–105
50. Raghavendra, H.; Diwakar, B.T.; Lokesh, B.R.; Naidu, K.A. Eugenol--the Active Principle from Cloves Inhibits 5-Lipoxygenase Activity and Leukotriene-C4 in Human PMNL Cells. *Prostaglandins Leukot. Essent. Fatty Acids* **2006**, *74*, 23–27, doi:10.1016/j.plefa.2005.08.006. 106–107
51. Kalhan, R.; Smith, L.J.; Nlend, M.C.; Nair, A.; Hixon, J.L.; Sporn, P.H.S. A Mechanism of Benefit of Soy Genistein in Asthma: Inhibition of Eosinophil P38-Dependent Leukotriene Synthesis. *Clin. Exp. Allergy* **2007**, *0*, 071104193151002-???, doi:10.1111/j.1365-2222.2007.02862.x. 108–109
52. Hernández, V.; Recio, M.C.; Máñez, S.; Giner, R.M.; Ríos, J.-L. Effects of Naturally Occurring Dihydroflavonols from *Inula Viscosa* on Inflammation and Enzymes Involved in the Arachidonic Acid Metabolism. *Life Sci.* **2007**, *81*, 480–488, doi:10.1016/j.lfs.2007.06.006. 110–111

53. Park, S.; Han, S.-U.; Lee, K.-M.; Park, K.H.; Cho, S.W.; Hahm, K.-B. 5-LOX Inhibitor Modulates the Inflammatory Responses Provoked by Helicobacter Pylori Infection. *Helicobacter* **2007**, *12*, 49–58, doi:10.1111/j.1523-5378.2007.00469.x. 112
113
54. Moon, T.C.; Seo, C.S.; Haa, K.; Kim, J.C.; Hwang, N.K.; Hong, T.G.; Kim, J.H.; Kim, D.H.; Son, J.K.; Chang, H.W. Meso-Dihydroguaiaretic Acid Isolated from Saururus Chinensis Inhibits Cyclooxygenase-2 and 5-Lipoxygenase in Mouse Bone Marrow-Derived Mast Cells. *Arch. Pharm. Res.* **2008**, *31*, 606–610, doi:10.1007/s12272-001-1200-y. 114
115
116
55. Poeckel, D.; Greiner, C.; Verhoff, M.; Rau, O.; Tausch, L.; Hörnig, C.; Steinhilber, D.; Schubert-Zsilavec, M.; Werz, O. Carnosic Acid and Carnosol Potently Inhibit Human 5-Lipoxygenase and Suppress pro-Inflammatory Responses of Stimulated Human Polymorphonuclear Leukocytes. *Biochem. Pharmacol.* **2008**, *76*, 91–97, doi:10.1016/j.bcp.2008.04.013. 117
118
119
56. Loke, W.M.; Proudfoot, J.M.; Stewart, S.; McKinley, A.J.; Needs, P.W.; Kroon, P.A.; Hodgson, J.M.; Croft, K.D. Metabolic Transformation Has a Profound Effect on Anti-Inflammatory Activity of Flavonoids Such as Quercetin: Lack of Association between Antioxidant and Lipoxygenase Inhibitory Activity. *Biochem. Pharmacol.* **2008**, *75*, 1045–1053, doi:10.1016/j.bcp.2007.11.002. 120
121
122
57. Anogianaki, A.; Castellani, M.L.; Madhappan, B.; Salini, V.; Vecchiet, J.; Tetè, S.; Frydas, S.; Perrella, A.; De Lutiis, M.A.; Neri, G.; et al. RANTES (CCL5) Potentiates Calcium Ionophore in the Production of LTB4 in Rat Adherent Macrophages from Granuloma Induced by KMnO4: Inhibition by NDGA. *Pharmacol. Res.* **2008**, *57*, 49–55, doi:10.1016/j.phrs.2007.11.002. 123
124
125
58. Castellani, M.L.; Conti, P.; Felaco, M.; Vecchiet, J.; Ciampoli, C.; Cerulli, G.; Boscolo, P.; Theoharides, T.C. Substance P Upregulates LTB4 in Rat Adherent Macrophages from Granuloma Induced by KMnO4. *Neurotox. Res.* **2009**, *15*, 49–56, doi:10.1007/s12640-009-9004-6. 126
127
59. Jin, M.-I.; Bae, K.-H.; Chang, H.-W.; Son, J.-K. Anti-Inflammatory Compounds from the Leaves of Ailanthus Altissima Meihua JIN. *Biomol. Ther.* **2009**, *17*, 86–91, doi:10.4062/biomolther.2009.17.1.86. 128
129
60. Feisst, C.; Pergola, C.; Rakonjac, M.; Rossi, A.; Koeberle, A.; Dodt, G.; Hoffmann, M.; Hoernig, C.; Fischer, L.; Steinhilber, D.; et al. Hyperforin Is a Novel Type of 5-Lipoxygenase Inhibitor with High Efficacy in Vivo. *Cell. Mol. Life Sci. CMLS* **2009**, *66*, 2759–2771, doi:10.1007/s00018-009-0078-3. 130
131
61. Lim, H.; Nam, J.W.; Seo, E.-K.; Kim, Y.S.; Kim, H.P. (-)-Nyasol (Cis-Hinokiresinol), a Norneolignan from the Rhizomes of Anemarrhena Asphodeloides, Is a Broad Spectrum Inhibitor of Eicosanoid and Nitric Oxide Production. *Arch. Pharm. Res.* **2009**, *32*, 1509–1514, doi:10.1007/s12272-009-2102-4. 132
133
62. Lim, H.; Son, K.H.; Bae, K.H.; Hung, T.M.; Kim, Y.S.; Kim, H.P. 5-Lipoxygenase-Inhibitory Constituents from Schizandra Fructus and Magnolia Flos. *Phytother. Res. PTR* **2009**, *23*, 1489–1492, doi:10.1002/ptr.2783. 134
135
63. Kim, J.M.; Lee, E.K.; Park, G.; Kim, M.K.; Yokozawa, T.; Yu, B.P.; Chung, H.Y. Morin Modulates the Oxidative Stress-Induced NF-KappaB Pathway through Its Anti-Oxidant Activity. *Free Radic. Res.* **2010**, *44*, 454–461, doi:10.3109/10715761003610737. 136
137
64. Saw, C.L.L.; Huang, Y.; Kong, A.-N. Synergistic Anti-Inflammatory Effects of Low Doses of Curcumin in Combination with Polyunsaturated Fatty Acids: Docosahexaenoic Acid or Eicosapentaenoic Acid. *Biochem. Pharmacol.* **2010**, *79*, 421–430, doi:10.1016/j.bcp.2009.08.030. 138
139

65. Lin, T.-Y.; London, C.A. Characterization and Modulation of Canine Mast Cell Derived Eicosanoids. *Vet. Immunol. Immunopathol.* **2010**, *135*, 118–127, doi:10.1016/j.vetimm.2009.11.010. 140
141
66. Lee, E.J.; Kim, J.S.; Kim, H.P.; Lee, J.-H.; Kang, S.S. Phenolic Constituents from the Flower Buds of *Lonicera Japonica* and Their 5-Lipoxygenase Inhibitory Activities. *Food Chem.* **2010**, *120*, 134–139, doi:10.1016/j.foodchem.2009.09.088. 142
143
67. Bhaskar, S.; Shalini, V.; Helen, A. Quercetin Regulates Oxidized LDL Induced Inflammatory Changes in Human PBMCs by Modulating the TLR-NF-KB Signaling Pathway. *Immunobiology* **2011**, *216*, 367–373, doi:10.1016/j.imbio.2010.07.011. 144
145
68. Kwon, O.S.; Choi, J.S.; Islam, Md.N.; Kim, Y.S.; Kim, H.P. Inhibition of 5-Lipoxygenase and Skin Inflammation by the Aerial Parts of *Artemisia Capillaris* and Its Constituents. *Arch. Pharm. Res.* **2011**, *34*, 1561–1569, doi:10.1007/s12272-011-0919-0. 146
147
69. Fan, S.-Y.; Zeng, H.-W.; Pei, Y.-H.; Li, L.; Ye, J.; Pan, Y.-X.; Zhang, J.-G.; Yuan, X.; Zhang, W.-D. The Anti-Inflammatory Activities of an Extract and Compounds Isolated from *Platycladus Orientalis* (Linnaeus) Franco in Vitro and Ex Vivo. *J. Ethnopharmacol.* **2012**, *141*, 647–652, doi:10.1016/j.jep.2011.05.019. 148
149
70. Lee, J.H.; Lee, D.U.; Kim, Y.S.; Kim, H.P. 5-Lipoxygenase Inhibition of the Fructus of *Foeniculum Vulgare* and Its Constituents. *Biomol. Ther.* **2012**, *20*, 113–117, doi:10.4062/biomolther.2012.20.1.113. 150
151
71. Han, Y.S.; Kim, M.-S.; Hwang, J.-K. Macelignan Inhibits Histamine Release and Inflammatory Mediator Production in Activated Rat Basophilic Leukemia Mast Cells. *Inflammation* **2012**, *35*, 1723–1731, doi:10.1007/s10753-012-9490-1. 152
153
72. Boudreau, L.H.; Maillet, J.; LeBlanc, L.M.; Jean-François, J.; Touaibia, M.; Flamand, N.; Surette, M.E. Caffeic Acid Phenethyl Ester and Its Amide Analogue Are Potent Inhibitors of Leukotriene Biosynthesis in Human Polymorphonuclear Leukocytes. *PLoS One* **2012**, *7*, e31833, doi:10.1371/journal.pone.0031833. 154
155
73. Choi, Y.; Kim, M.S.; Hwang, J.-K. Inhibitory Effects of Panduratin A on Allergy-Related Mediator Production in Rat Basophilic Leukemia Mast Cells. *Inflammation* **2012**, *35*, 1904–1915, doi:10.1007/s10753-012-9513-y. 156
157
74. Kogut, M.H.; He, H.; Genovese, K.J. Bacterial Toll-like Receptor Agonists Induce Sequential NF-KB-Mediated Leukotriene B4 and Prostaglandin E2 Production in Chicken Heterophils. *Vet. Immunol. Immunopathol.* **2012**, *145*, 159–170, doi:10.1016/j.vetimm.2011.11.003. 158
159
75. Lim, J.C.; Park, S.Y.; Nam, Y.; Nguyen, T.T.; Sohn, U.D. The Protective Effect of Eupatilin against Hydrogen Peroxide-Induced Injury Involving 5-Lipoxygenase in Feline Esophageal Epithelial Cells. *Korean J. Physiol. Pharmacol. Off. J. Korean Physiol. Soc. Korean Soc. Pharmacol.* **2012**, *16*, 313–320, doi:10.4196/kjpp.2012.16.5.313. 160
162
76. Gomes, A.; Couto, D.; Alves, A.; Dias, I.; Freitas, M.; Porto, G.; Duarte, J.A.; Fernandes, E. Trihydroxyflavones with Antioxidant and Anti-Inflammatory Efficacy. *BioFactors Oxf. Engl.* **2012**, *38*, 378–386, doi:10.1002/biof.1033. 163
164
77. Li, L.; Zeng, H.; Shan, L.; Yuan, X.; Li, Y.; Liu, R.; Zhang, W. The Different Inhibitory Effects of Huang-Lian-Jie-Du-Tang on Cyclooxygenase 2 and 5-Lipoxygenase. *J. Ethnopharmacol.* **2012**, *143*, 732–739, doi:10.1016/j.jep.2012.07.037. 165
166

78. Werz, O.; Seegers, J.; Schaible, A.M.; Weinigel, C.; Barz, D.; Koeberle, A.; Allegrone, G.; Pollastro, F.; Zampieri, L.; Grassi, G.; et al. Cannflavins from Hemp Sprouts, a Novel Cannabinoid-Free Hemp Food Product, Target Microsomal Prostaglandin E2 Synthase-1 and 5-Lipoxygenase. *PharmaNutrition* **2014**, *2*, 53–60, doi:10.1016/j.phanu.2014.05.001. 167–169
79. Ribeiro, D.; Freitas, M.; Tomé, S.M.; Silva, A.M.S.; Porto, G.; Cabrita, E.J.; Marques, M.M.B.; Fernandes, E. Inhibition of LOX by Flavonoids: A Structure-Activity Relationship Study. *Eur. J. Med. Chem.* **2014**, *72*, 137–145, doi:10.1016/j.ejmech.2013.11.030. 170–171
80. Lee, Y.S.; Hur, S.; Kim, T.-Y. Homoisoflavanone Prevents Mast Cell Activation and Allergic Responses by Inhibition of Syk Signaling Pathway. *Allergy* **2014**, *69*, 453–462, doi:10.1111/all.12356. 172–173
81. Li, X.; Lu, Y.; Jin, Y.; Son, J.-K.; Lee, S.H.; Chang, H.W. Curcumin Inhibits the Activation of Immunoglobulin E-Mediated Mast Cells and Passive Systemic Anaphylaxis in Mice by Reducing Serum Eicosanoid and Histamine Levels. *Biomol. Ther.* **2014**, *22*, 27–34, doi:10.4062/biomolther.2013.092. 174–175
82. Kutil, Z.; Kvasnicova, M.; Temml, V.; Schuster, D.; Marsik, P.; Cusimamani, E.F.; Lou, J.-D.; Vanek, T.; Landa, P. Effect of Dietary Stilbenes on 5-Lipoxygenase and Cyclooxygenases Activities In Vitro. *Int. J. Food Prop.* **2015**, *18*, 1471–1477, doi:10.1080/10942912.2014.903416. 176–177
83. Forino, M.; Pace, S.; Chianese, G.; Santagostini, L.; Werner, M.; Weinigel, C.; Rummler, S.; Fico, G.; Werz, O.; Tagliatela-Scafati, O. Humudifucol and Bioactive Prenylated Polyphenols from Hops (*Humulus Lupulus* Cv. “Cascade”). *J. Nat. Prod.* **2016**, *79*, 590–597, doi:10.1021/acs.jnatprod.5b01052. 178–179
84. Tsen, S.Y.; Tan, X.Y.; Tan, Y.M.; Yan, B.Y.; Loke, W.M. Relative Inhibitions of 5-Lipoxygenase and Myeloperoxidase and Free-Radical Scavenging Activities of Daidzein, Dihydrodaidzein, and Equol. *J. Med. Food* **2016**, *19*, 543–548, doi:10.1089/jmf.2015.3557. 180–181
85. Roy, P.-P.; Faye, D.; Blanchard, S.; Cormier, M.; Doiron, J.A.; Surette, M.E.; Touaibia, M. New Caffeic Acid Phenylethyl Ester Analogs Bearing Substituted Triazole: Synthesis and Structure-Activity Relationship Study towards 5-Lipoxygenase Inhibition. *J. Chem.* **2017**, *2017*, e2380531, doi:10.1155/2017/2380531. 182–183
86. Anilkumar, K.; Reddy, G.V.; Azad, R.; Yarla, N.S.; Dharmapuri, G.; Srivastava, A.; Kamal, M.A.; Pallu, R. Evaluation of Anti-Inflammatory Properties of Isoorientin Isolated from Tubers of *Pueraria Tuberosa*. *Oxid. Med. Cell. Longev.* **2017**, *2017*, 1–10, doi:10.1155/2017/5498054. 184–185
87. Takasugi, M.; Muta, E.; Yamada, K.; Arai, H. A New Method to Evaluate Anti-Allergic Effect of Food Component by Measuring Leukotriene B4 from a Mouse Mast Cell Line. *Cytotechnology* **2018**, *70*, 177–184, doi:10.1007/s10616-017-0129-9. 186–187
88. Doucet, M.S.; Jougoux, J.-L.; Poirier, S.J.; Cormier, M.; Léger, J.L.; Surette, M.E.; Pichaud, N.; Touaibia, M.; Boudreau, L.H. Identification of Peracetylated Quercetin as a Selective 12-Lipoxygenase Pathway Inhibitor in Human Platelets. *Mol. Pharmacol.* **2019**, *95*, 139–150, doi:10.1124/mol.118.113480. 188–189
89. Mbarik, M.; Poirier, S.J.; Doiron, J.; Selka, A.; Barnett, D.A.; Cormier, M.; Touaibia, M.; Surette, M.E. Phenolic Acid Phenylesters and Their Corresponding Ketones: Inhibition of 5-Lipoxygenase and Stability in Human Blood and HepaRG Cells. *Pharmacol. Res. Perspect.* **2019**, *7*, e00524, doi:10.1002/prp2.524. 190–191
90. Storniolo, C.E.; Martínez-Hovelman, N.; Martínez-Huélamo, M.; Lamuela-Raventos, R.M.; Moreno, J.J. Extra Virgin Olive Oil Minor Compounds Modulate Mitogenic Action of Oleic Acid on Colon Cancer Cell Line. *J. Agric. Food Chem.* **2019**, *67*, 11420–11427, doi:10.1021/acs.jafc.9b04816. 192–193

91. Prabha, B.; Sini, S.; Priyadarshini, T.S.; Sasikumar, P.; Gopalan, G.; Joseph, J.P.; Jithin, M.M.; Sivan, V.V.; Jayamurthy, P.; Radhakrishnan, K.V. Anti-Inflammatory Effect and Mechanism of Action of Ellagic Acid-3,3',4-Trimethoxy-4'-O- α -L-Rhamnopyranoside Isolated from *Hopea Parviflora* in Lipopolysaccharide-Stimulated RAW 264.7 Macrophages. *Nat. Prod. Res.* **2019**, 1–5, doi:10.1080/14786419.2019.1690486.
92. Gerstmeier, J.; Seegers, J.; Witt, F.; Waltenberger, B.; Temml, V.; Rollinger, J.M.; Stuppner, H.; Koeberle, A.; Schuster, D.; Werz, O. Ginkgolic Acid Is a Multi-Target Inhibitor of Key Enzymes in Pro-Inflammatory Lipid Mediator Biosynthesis. *Front. Pharmacol.* **2019**, *10*, 797, doi:10.3389/fphar.2019.00797.
93. Hartung, N.M.; Fischer, J.; Ostermann, A.I.; Willenberg, I.; Rund, K.M.; Schebb, N.H.; Garscha, U. Impact of Food Polyphenols on Oxylipin Biosynthesis in Human Neutrophils. *Biochim. Biophys. Acta Mol. Cell Biol. Lipids* **2019**, *1864*, 1536–1544, doi:10.1016/j.bbalip.2019.05.002.
94. Jin, S.E.; Ha, H.; Shin, H.-K.; Seo, C.-S. Anti-Allergic and Anti-Inflammatory Effects of Kuwanon G and Morusin on MC/9 Mast Cells and HaCaT Keratinocytes. *Mol. Basel Switz.* **2019**, *24*, E265, doi:10.3390/molecules24020265.
95. Pan, S.; Zhao, X.; Ji, N.; Shao, C.; Fu, B.; Zhang, Z.; Wang, R.; Qiu, Y.; Jin, M.; Kong, D. Inhibitory Effect of Taxifolin on Mast Cell Activation and Mast Cell-Mediated Allergic Inflammatory Response. *Int. Immunopharmacol.* **2019**, *71*, 205–214, doi:10.1016/j.intimp.2019.03.038.
96. Giménez-Bastida, J.A.; González-Sarriás, A.; Espín, J.C.; Schneider, C. Inhibition of 5-Lipoxygenase-Derived Leukotrienes and Hemiketals as a Novel Anti-Inflammatory Mechanism of Urolithins. *Mol. Nutr. Food Res.* **2020**, *64*, e2000129, doi:10.1002/mnfr.202000129.
97. Habib, E.S.; El-Bsoumy, E.; Ibrahim, A.K.; Helal, M.A.; El-Magd, M.A.; Ahmed, S.A. Anti-Inflammatory Effect of Methoxyflavonoids from *Chiliadenus Montanus* (*Jasonia Montana*) Growing in Egypt. *Nat. Prod. Res.* **2020**, 1–5, doi:10.1080/14786419.2020.1802272.
98. Gilbert, N.C.; Gerstmeier, J.; Schexnaydre, E.E.; Börner, F.; Garscha, U.; Neau, D.B.; Werz, O.; Newcomer, M.E. Structural and Mechanistic Insights into 5-Lipoxygenase Inhibition by Natural Products. *Nat. Chem. Biol.* **2020**, *16*, 783–790, doi:10.1038/s41589-020-0544-7.
99. Mostafa, E.S.; Nawwar, M.A.M.; Mostafa, D.A.; Ragab, M.F.; Swilam, N. Karafsin, a Unique Mono-Acylated Flavonoid Apiofurnoside from the Leaves of *Apium Graveolens* Var. *Secalinum Alef*: In Vitro and in Vivo Anti-Inflammatory Assessment. *Ind. Crops Prod.* **2020**, *158*, 112901, doi:10.1016/j.indcrop.2020.112901.
100. Lee, J.-Y.; Park, S.-H.; Jhee, K.-H.; Yang, S.-A. Tricin Isolated from Enzyme-Treated *Zizania Latifolia* Extract Inhibits IgE-Mediated Allergic Reactions in RBL-2H3 Cells by Targeting the Lyn/Syk Pathway. *Mol. Basel Switz.* **2020**, *25*, E2084, doi:10.3390/molecules25092084.
101. Toda, K.; Tsukayama, I.; Nagasaki, Y.; Konoike, Y.; Tamenobu, A.; Ganeko, N.; Ito, H.; Kawakami, Y.; Takahashi, Y.; Miki, Y.; et al. Red-Kerneled Rice Proanthocyanidin Inhibits Arachidonate 5-Lipoxygenase and Decreases Psoriasis-like Skin Inflammation. *Arch. Biochem. Biophys.* **2020**, *689*, 108307, doi:10.1016/j.abb.2020.108307.