



Review

Melatonin/Sericin Wound Healing Patches: Implications for Melanoma Therapy

Katarzyna Adamiak ¹, Vivian A. Gaida ², Jasmin Schäfer ², Lina Bosse ², Clara Diemer ², Russel J. Reiter ³, Andrzej T. Slominski ^{4,5}, Kerstin Steinbrink ², Alina Sionkowska ¹ and Konrad Kleszczyński ^{2,*}

¹ Department of Biomaterials and Cosmetic Chemistry, Faculty of Chemistry, Nicolaus Copernicus University, Gagarin 7, 87-100 Toruń, Poland; kadamiak@doktorant.umk.pl (K.A.); alinas@umk.pl (A.S.)

² Department of Dermatology, University of Münster, Von-Esmarch-Str. 58, 48149 Münster, Germany; vgaida@uni-muenster.de (V.A.G.); jschae1@uni-muenster.de (J.S.); lbosse1@uni-muenster.de (L.B.); cdiemer@uni-muenster.de (C.D.); kerstin.steinbrink@ukmuenster.de (K.S.)

³ Department of Cell Systems and Anatomy, Long School of Medicine, UT Health, San Antonio, TX 78229, USA; reiter@uthscsa.edu

⁴ Department of Dermatology, Comprehensive Cancer Center, University of Alabama at Birmingham, Birmingham, AL 35294, USA; aslominski@uabmc.edu

⁵ Pathology and Laboratory Medicine Service, VA Medical Center, Birmingham, AL 35294, USA

* Correspondence: konrad.kleszczyński@ukmuenster.de; Tel.: +49-251-83-56523; Fax: +49-251-83-58646

Abstract: Melatonin and sericin exhibit antioxidant properties and may be useful in topical wound healing patches by maintaining redox balance, cell integrity, and regulating the inflammatory response. In human skin, melatonin suppresses damage caused by ultraviolet radiation (UVR) which involves numerous mechanisms associated with reactive oxygen species/reactive nitrogen species (ROS/RNS) generation and enhancing apoptosis. Sericin is a protein mainly composed of glycine, serine, aspartic acid, and threonine amino acids removed from the silkworm cocoon (particularly *Bombyx mori* and other species). It is of interest because of its biodegradability, anti-oxidative, and anti-bacterial properties. Sericin inhibits tyrosinase activity and promotes cell proliferation that can be supportive and useful in melanoma treatment. In recent years, wound healing patches containing sericin and melatonin individually have attracted significant attention by the scientific community. In this review, we summarize the state of innovation of such patches during 2021–2023. To date, melatonin/sericin-polymer patches for application in post-operational wound healing treatment has been only sparingly investigated and it is an imperative to consider these materials as a promising approach targeting for skin tissue engineering or regenerative dermatology.

Keywords: melatonin; sericin; wound healing; biomaterials; scaffolds; hydrogels; skin regeneration



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1. Introduction

The skin with subcutis constitutes the largest organ in the human body, exposed to external and internal aging factors. The stochastic process of skin aging implies functional and phenotypic variability in cutaneous and immune cells, which occurs along with functional and structural changes in extracellular matrix constituents, including collagen and elastin. Wrinkling, roughness, skin laxity, and decrease in elasticity are the main clinical features of the skin aging process [1–3].

The crucial external factor of skin aging is the excessive exposure to ultraviolet radiation (UVR) which is connected with hyperpigmentary changes [4] and wrinkling [5], and also induces common types of skin cancer such as basal cell carcinoma [6,7], squamous cell carcinoma [8–10], and malignant melanoma [11,12]. The UVR spectra are: UVC (200–280 nm), UVB (280–315 nm) and UVA (315–400 nm), and radiation [13–15]. The UVC and 280–290 wavelengths of UVB are absorbed by the ozone layer of the atmosphere; however, the *stratum corneum* is able to absorb the UVC radiation after the exposition to

non-natural light sources [14]. The UVA can penetrate the reticular dermis and induce biological effects, yet not as efficiently as the UVB radiation [16], which is absorbed by the upper layers of the epidermis and can penetrate the papillary dermis [13,15]. The UVR has an influence on many complex processes in the human body and can also trigger systemic reactions [17,18]. It can upregulate local and systemic neuroendocrine systems [18]. The locally induced cytokines, urocortins, enkephalins, and melanocortins can impose systemic effects while released into the circulation, like the agitation of the central hypothalamic–pituitary–adrenal axis, opioidogenic effects, and immunosuppression, as well as vitamin D synthesis and activation [17–22].

The UV lights have a tremendous impact on biological organisms and the origin of life on Earth [23–26]. The energy of UVR is similar to the energy of covalent bonds, which means that any molecule electron excitation by UV light can disrupt or create a covalent bond. Organic molecules exposed to the energy of UV light commute to chemical bonds with high-energy levels, achieving molecular complexity [18,26]. The cells can use their energy increasing the enthalpy of the system [26]. UVB is crucial in photosynthesis, providing various forms of vitamin D which can be enzymatically activated at the local and systemic levels in different organisms [21,22,27].

In human skin, reactive oxygen species/reactive nitrogen species (ROS/RNS) production can be induced by UVR, thereby elevating the secretion of pro-inflammatory cytokines. Increased cell proliferation and oxidation processes can disrupt cell membrane integrity and induce DNA fragmentation [7]. The topical delivery of antioxidants helps to maintain redox balance in epidermal cells, regulates pro-inflammatory cytokines release, and prevents oxidative damage and DNA fragmentation in these cells [28–35].

Melatonin (*N*-acetyl-5-methoxyindolamine) and sericin exert antioxidant properties and could be useful in topical wound healing patches by maintaining redox balance, cell integrity, and regulating the inflammatory response [36–39]. In human skin, melatonin suppresses the damage caused by UVR through numerous mechanisms associated with ROS/RNS production and programmed cell death (apoptosis) [40–43]. Melatonin derived from tryptophan constitutes an active pleiotropic molecule in the human organism, synthesized by cells of the pineal gland and also peripheral organs such as the skin, gastrointestinal tract, and lymphocytes [44–46]. Melatonin influences biorhythms, scavenges free radicals, enhances DNA repair, influences the gene expression of anti-oxidative enzymes, and stimulates wound healing properties [47–51]. Sericin as an effective antioxidant exhibits skin protective activity against UVB and UVA radiation-induced damage [52]. Chromophores' UVB absorption is predominant, whereas UVA is weakly absorbed by DNA and cellular chromophores with greater ROS/RNS generation, which leads to oxidative changes in the cells [13,53]. Sericin is produced by silkworm's glands (e.g., *Bombyx mori*, *Bombyx mandarins*, and other species) and has been explored in biomaterial applications because of its biodegradability, anti-oxidative, and antibacterial properties [54–59]. Furthermore, sericin inhibits tyrosinase activity and promotes cell proliferation, which can be supportive and useful in melanoma treatment [60–64]. After surgery, the potential bacterial infections could exacerbate epidermal/dermal damage and delay wound healing. Thus, developing biomaterials for melanoma therapy that reduces the risk of infection, maintaining redox balance in skin cells and preventing cancer recurrence is essential for patient recovery [65–70]. Sericin as a biomaterial has been used in the preparation of a variety of tissue engineering materials such as composites, hydrogels, membranes, nanofibers, and nanoparticles [71–75]. However, poor mechanical strength and high production costs of sericin-based scaffolds make it difficult to introduce this molecule for medical use. Using a biopolymer with improved mechanical properties and with the incorporation sericin and melatonin as active compounds may be highly useful for wound healing in melanoma-affected patients [76–80].

2. Melatonin and Sericin against the Skin Aging Process

The process of aging has a prominent impact on the skin's healing function, mainly by prolonging the inflammatory phase and increasing ROS/RNS production [81,82]. The skin constitutes a protective barrier between external and internal environments and has the sensory capacity to maintain body homeostasis in response to deleterious factors. Skin aging is a natural process with progressive functional and morphological changes, determined by the overall exposure to both intrinsic and extrinsic factors, which may vary depending on skin regions within diverse ethnicities. The clinical signs of skin aging are visible as wrinkles, a rough-textured appearance, a loss of elasticity, and laxity [83,84].

The physiological process of skin aging is characterized primarily by fluctuation changes in endocrine circadian rhythmicity, gene expression, and hormonal descent, which are the reasons for appearance of morphological and functional alterations [85–88]. With increasing age, the occurrence of changes in the endocrine glands, the steroidogenic system, skin cholesterol synthesis, proopiomelanocortin (POMC) expression, and POMC-derived peptides production with the focus on the melanocortin receptor 1 (MC1R) and MC2R agonists, are more frequent and may lead to skin alterations and lesions [89–92]. Vitamin D production, essential to maintain proper skin functions and immunity, also decreases with age [92–95].

On the molecular level, the process of skin aging comprises ROS/RNS generation, diminished antioxidant protection, changes in gene expression, and defects in cellular DNA mechanisms. Along with the senescence of the organism, the mitochondrial DNA content and number decreases [96–100], but there is also enhanced ROS/RNS generation with reduced oxidative phosphorylation and adenosine triphosphate production which leads to mitochondria-mediated apoptosis [101–103]. Melatonin has an antioxidant capacity which relies on the indirect receptor-mediated stimulation of antioxidant enzymes to resist the oxidative stress [104–112]. Melatonin and its metabolites are also known for their anti-inflammatory and mitochondrial protective capability [32,113–120], which help to maintain proper skin functions [121–123]. Melatonin and its metabolites have a major role in human epidermal keratinocytes protection against UVB radiation, in particular *N*¹-acetyl-*N*²-formyl-5-methoxykynuramine (AFMK), *N*¹-acetyl-5-methoxykynuramine (AMK), 5-methoxytryptamine (5-MT), and 6-hydroxymelatonin (6(OH)MEL), which ameliorate the disruptive effects of UVR. Studies have shown that melatonin also protects the dermal fibroblasts from the deleterious action of UVA and UVB [124–137]. Thus, melatonin and its metabolites counteract photodamage and premature skin aging [138–140] (Figure 1). Finally, due to stimulated expression of involucrin, keratin-10 and keratin-14, topically applied melatonin enhances the epidermal barrier function of the skin and increases the activity of keratinocytes *ex vivo* [141,142] (Figure 1). The mechanism of action of melatonin and its metabolites would include the activation of the membrane-bound MT1 and T2 receptors [5] or the recently identified aryl hydrocarbon (AhR) and peroxisome proliferator-activated receptor gamma (PPAR- γ) [143], or the receptor independent mechanisms mentioned above.

Studies using dermal fibroblasts have shown that silk sericin stimulates collagen synthesis, which also indicates its anti-aging properties [144,145]. Results have revealed that silk sericin activates collagen type I synthesis and suppresses oxidative stress, maintaining unaltered fibroblast growth kinetics and cellular structure [146]. Next to ROS/RNS-scavenging activity, sericin also exhibits anti-tyrosinase and anti-elastase properties. Recent studies have shown that particular sericin strains have an anti-proliferative activity on peripheral blood mononuclear cells; *in vitro* IFN- γ secretion was decreased, without affecting TNF and IL-10 release. Thus, sericin may be useful for dermatological use [62,147].

The recent inventions using sericin in anti-aging treatments include extract loaded-sericin hydrogel as a topical agent [148], naringenin microemulsion-loaded sericin gel [149], and gold silk sericin/niacinamide/signaline complex [150]. Extract-loaded sericin hydrogels in six formulations were examined for anti-melanogenesis on the B16F10 melanoma cell line, UVR-preventive properties of human keratinocytes (HaCaT), and anti-aging ef-

fectiveness on normal human dermal fibroblasts. The study showed that the hydrogel increased the anthocyanin penetration through the skin. The most promising formulation using the purple waxy corn cob (*Zea mays* L.) extract demonstrated the highest tyrosinase activity inhibition, melanin pigment reduction, collagenase/elastase inhibition, collagen type I production, and elevation of cell viability.

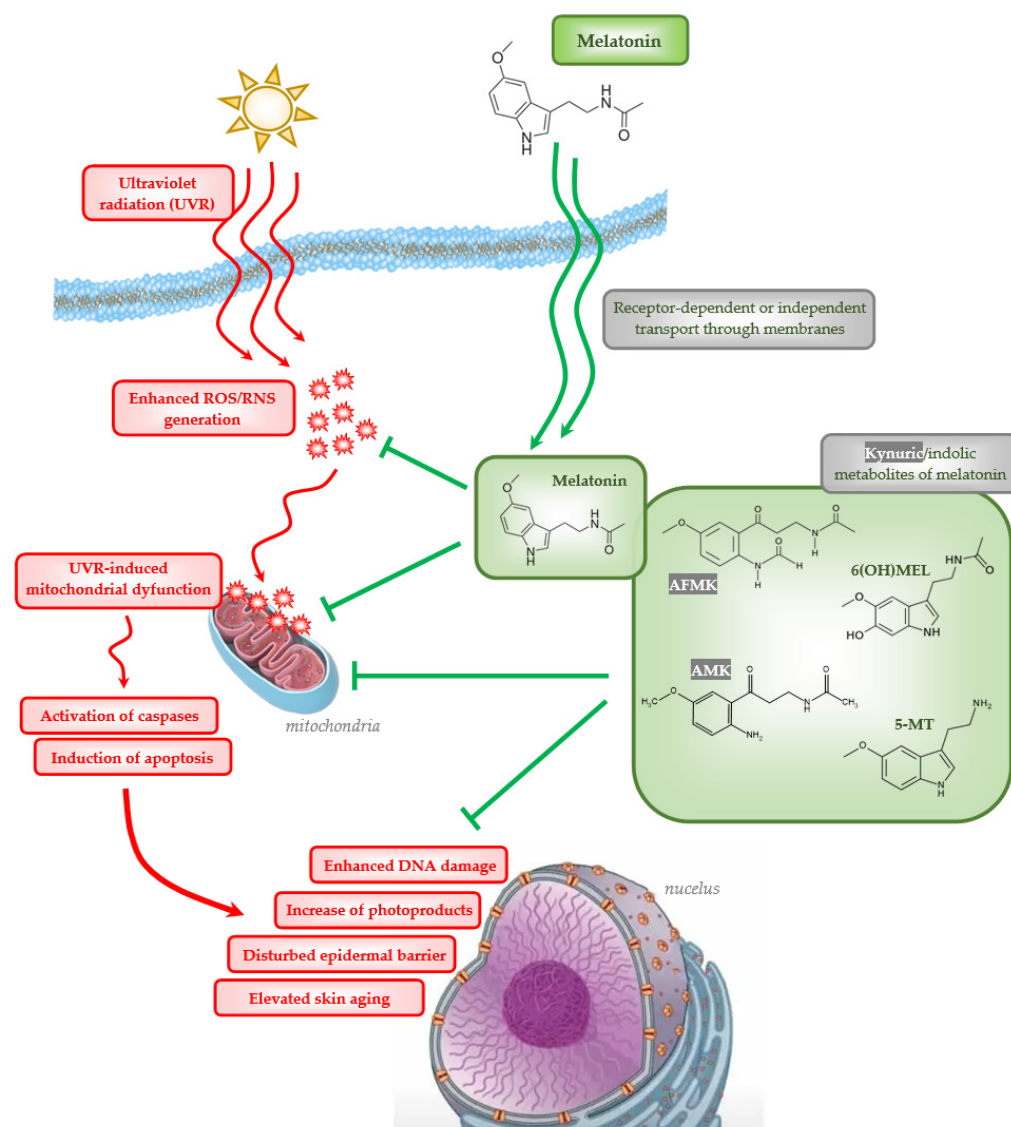


Figure 1. Cellular changes induced by ultraviolet radiation (UVR) including enhanced oxidative stress (ROS/RNS generation), mitochondrial dysfunction, DNA damage and protective action of melatonin as well as its kynuric (AFMK, AMK) and indolic (6(OH)MEL, 5-MT) metabolites.

Thus, the purple waxy corn cob extract-loaded sericin hydrogel as a topical agent indicates a great potential in anti-aging products [148]. Namely, naringenin microemulsion-loaded sericin gel showed an inhibition of UVR-induced photoaging and increased free radical scavenging. The in vitro cytotoxicity study on skin cancer cells enhanced anti-proliferative activity by increasing ROS/RNS in cancer cells with caspase-3 (Casp-3) activation [149]. The randomized study with the gold silk sericin/niacinamide/signaline complex have shown the efficacy of daily application in improving the condition of the skin with an antiaging effect [150].

UV light constitutes an important factor in skin aging and disturbances in skin proliferation. UV light up-regulates the nuclear factor kappa B (NF-κB) and releases pro-

inflammatory cytokines, with a simultaneous increased generation of ROS/RNS. Next, free radicals affecting DNA decrease protein tyrosine phosphates and up-regulate matrix metalloproteinase generation, which leads to collagen decomposition [151,152]. Although, in healthy conditions urocanic acid, produced in the upper layers in the human skin, constitutes a natural skin UV absorber, excessive exposure to UV light should be avoided due to the harmful effects induced by UV light [153].

3. Melanoma: A Tumor of Melanocyte Origin

Melanoma, a tumor of melanocyte origin, constitutes one of the most formidable types of skin malignancy [154–156] described by local invasiveness, recurrence, early metastasis, and high mortality risk [157–161]. The standard treatment is surgical resection [162–164], while alternative therapies like chemotherapy, radiotherapy, or photodynamic therapy are focused on the elimination of the melanoma cells [165,166]. Nevertheless, the limitations of the treatment can cause prolonged stress for patients. Insufficient light penetration depth using photodynamic therapy may be a barrier for reaching pigmented lesions [167]. Furthermore, the complete tumor resection with residual tumor tissues may lead to major cutaneous defects [168,169]. The risk of the wound infection in post-surgical treatment is an emerging issue in the wound healing process. Therefore, it is essential to promote skin regeneration during melanoma treatment [170]. Throughout the years, various methods like autologous/allogenic skin grafts or tissue-engineered scaffolds were developed [171–173]. However, wound healing patches have drawn attention; understanding the requirements of melanoma treatment are indispensable to develop a strategy with integrated wound healing and therapeutic effects.

To better understand the process of melanoma wound healing, it is important to focus on the molecular bases of tumor development and possible ways to impede them [174–176]. Namely, the risk factors of melanoma induction are: UVR, burns, melanocytic nevi, fair skin or gene mutations (BRAF, NRAS, KIT) [177–180]. Also in the focus of attention is a tumor suppressor gene cyclin-dependent kinase inhibitor 2A (CDKN2A), and the encoding of p16INK4A and p14ARF proteins as a major genetic risk factor [181,182]. In the cell cycle of p16INK4 α , the transition from G1 to S phase is regulated and the cycle also acts as a CDK inhibitor, blocking the phosphorylation and inactivation of the Rb protein. However, p14ARF shows anti-proliferation activity, inhibiting the disintegration of the p53 tumor suppressor. During the G2/M phase in normal conditions, the expression of p53 stops the cell cycle or induces apoptosis, which can be a response, for example, to UVR-induced DNA damage [183].

This process can activate the heat shock proteins (HSPs) which protect the cells from physical or environmental stressors [184]. In vitro studies have shown that after the exposure to UV light, melanoma cells release the HSP70 which activates anti-melanoma T cells [185–188]. Thus, HSPs play an essential role as molecular chaperones by assisting the correct folding of nascent and stress-accumulated misfolded proteins, and by preventing their aggregation [189]. Additionally, HSPs have a protective function; they allow the cells to survive in otherwise lethal conditions. Various mechanisms have been proposed to account for the cytoprotective functions of HSPs. Namely, several of these proteins have been demonstrated to directly interact with components of the cell signaling pathways, for instance those of the tightly regulated caspase-dependent programmed cell death machinery, upstream, downstream and at the mitochondrial level. HSPs can also affect caspase-independent apoptosis-like processes by interacting with apoptogenic factors, such as the apoptosis-inducing factor (AIF) or by acting at the lysosome level [189]. Melatonin is known to downregulate the expression of HSP40, HSP70, and HSP90 [32,190,191] to reduce oxidative stress. Thus, melatonin may constitute the additional defense in melanoma [32,192–195]. The anti-inflammatory properties of melatonin could also be an answer to the NF- κ B activation, which is known to be the regulator in oncogenesis. NF- κ B activation promotes cell proliferation and inhibits apoptosis and p53 in cancer development.

The anti-inflammatory mechanism of melatonin's purpose is to regulate cyclooxygenase-2 (COX-2), inducible nitric oxide synthase (iNOS), and cytokines [196].

Recent studies have confirmed that melatonin and its indolic and kynurenic derivatives downstream the pathway of melanogenesis, causing a drop in the cyclic adenosine monophosphate (cAMP) level and the microphthalmia-associated transcription factor (MITF) and causing the resultant collapse in tyrosinase (TYR) activity and melanin content. These findings can be a breakthrough for the future studies on pigmentation in melanoma therapies [122,142,196–199]. Since the inhibition of melanogenesis in advanced melanomas can serve as an adjuvant strategy in the systemic therapy of melanoma [200], melatonin as a mitochondrial protector with anti-inflammatory properties and direct antioxidants should be explored in topical and transepidermal delivery, especially in skin damage after UVR exposition. It is also known to be very effective in alopecia and atopic dermatitis treatment. It may be also considered as a skin barrier protectant in chronopharmacology [42,45,130]. In topical delivery, melanoma treatment using a combination of melatonin/sericin also indicates antioxidant and mitochondrial protection activity which may constitute a potential therapeutic strategy, including in the antibacterial properties of sericin that promote wound healing processes [201,202]. We also acknowledge the potential limitation of the accumulation of melatonin in tumor cells, which may increase their resistance to pharmacotherapy or radiotherapy, as discussed recently [203].

4. State of Innovation in Melatonin-Polymer Wound Healing Patches

Melatonin, a derivative of tryptophan, was initially characterized and isolated by Lerner et al. [204]. Melatonin is an indoleamine, which is defined as a indole heterocycle that contains two chains, i.e., 5-methoxy and 3-amide group. The indole structure is rich in electrons and has high resonance stability and reactivity which is the reason for melatonin's free radical scavenging capacity [204]. To date, it has been reported that melatonin neutralizes a high number of ROS including hydrogenperoxide (H_2O_2), peroxy radicals ($ROO\bullet$), hydroxyl radical ($\bullet OH$), singlet oxygen (1O_2), and RNS, such as peroxynitrite ($ONOO^-$) or nitric oxide radicals ($NO\bullet$) [205–211]. The high antioxidant potential of melatonin enables the protection of the cells against oxidative stress more efficiently than other antioxidants. For instance, Figure 2 summarizes the cascade interaction between ROS/RNS and melatonin as well as its metabolites. Thus, melatonin has a capacity to detoxify numerous toxic ROS or RNS, whereas the scavenging ability of other antioxidants is significantly lower [212].

Furthermore, Table 1 presents melatonin as an active substance that is found in wound healing patches in combination with polymers: silk fibroin/methacrylate [213], polycaprolactone [214], carboxymethyl cellulose [215], methacrylated gelatin/thiolated pectin hydrogel [216], chitosan-sulfonated ethylene–propylene–diene terpolymer, sulfonated ethylene–propylene–diene terpolymer [217], nanoclay [218], polycaprolactone/sodium alginate [219], polycaprolactone/gelatin [220], collagen/chitosan cross-linked by glyoxal [221], gelatin [222], carboxymethyl chitosan [223], chitosan/collagen [224], chitosan–polycaprolactone/polyvinyl alcohol [225], chitosan–polycaprolactone [226], and collagen with aminated xanthan gum [227]. The main applications of melatonin-polymer patches are wound healing [214,223,224] and wound dressings [222,225], but they are also used in diabetic wound repair [215], skin delivery [218], and skin tissue engineering [221]. Furthermore, melatonin-loaded polymer patches were also used in cartilage repair [213], vital pump regeneration [216], tendon regeneration [219], membranes [217], nerve tissue engineering [220], skin tissue regeneration [227], and osteosarcoma treatment [226]. Polymers used in inventions were mainly synthetic but modified biopolymers including polycaprolactone [214], carboxymethyl cellulose [215], methacrylated gelatin/thiolated pectin hydrogel [216], chitosan-sulfonated ethylene–propylene–diene terpolymer [217], sulfonated ethylene–propylene–diene terpolymer [217], carboxymethyl chitosan [223], and chitosan–polycaprolactone [225]. A combination of natural and synthetic polymer were also observed, e.g., silk fibroin/methacrylate [213], polycaprolactone/sodium alginate [219], polycaprolactone/gelatin [220], collagen/chitosan cross-linked

by glyoxal [221], chitosan/collagen [224], and collagen with aminated xanthan gum [227], although natural polymer matrix nanoclay was rare [218], as was gelatin alone [222]. Other active substances in wound healing patches found in recent years are γ -cyclodextrin [215], tideglusib [216], and silver nanoparticles [227].

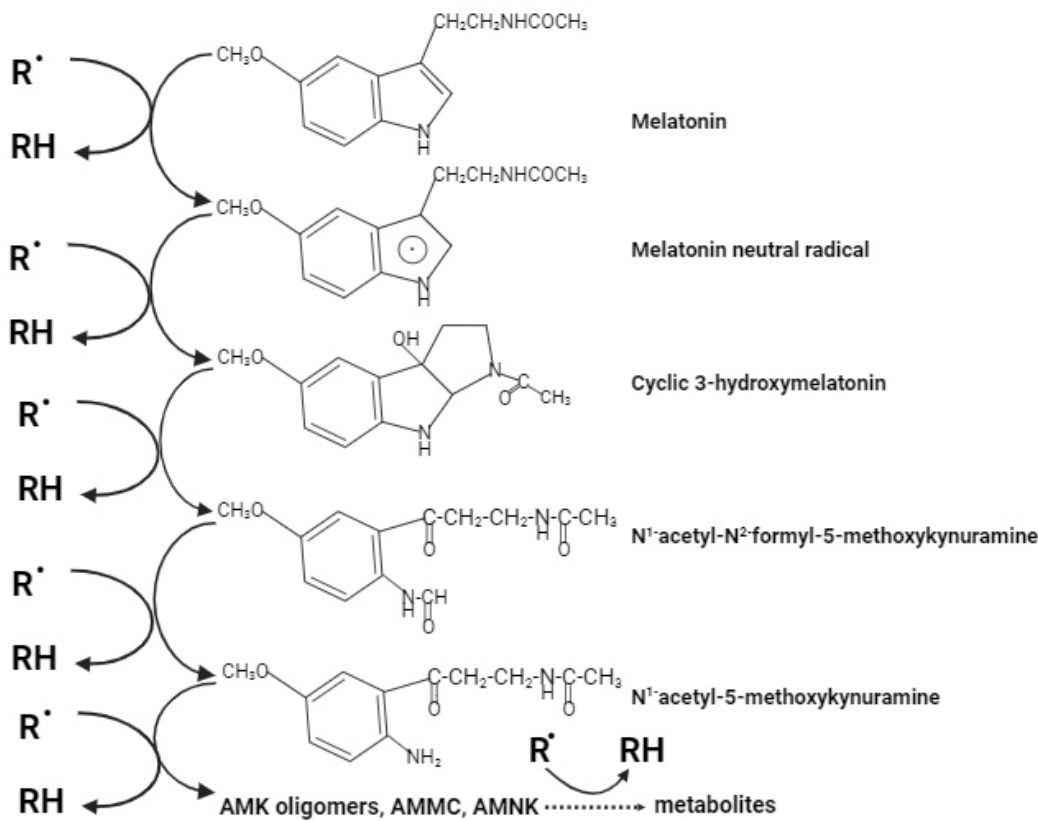


Figure 2. The cascade reaction with ROS/RNS products (or metabolites) of melatonin which are involved in terms of attenuation of UVR-induced changes as described above and presented in Figure 1.

Table 1. State of innovation in melatonin-polymer patches.

Invention	Matrix Polymer	Active Substance	Application	Origin	Year	Literature
Biomimetic melatonin-loaded silk fibroin/GelMA scaffolds	Silk fibroin/gel methacrylate	Melatonin	Cartilage repair	Natural/synthetic	2023	[213]
Melatonin-loaded polycaprolactone fiber mats	Polycaprolactone	Melatonin	Wound healing	Synthetic	2023	[214]
Carboxymethyl cellulose-based injectable hydrogel loaded with a composite of melatonin and γ -cyclodextrin	Carboxymethyl cellulose	Melatonin and γ -cyclodextrin	Diabetic wound repair	Synthetic	2023	[215]
Injectable methacrylated gelatin/thiolated pectin hydrogels carrying melatonin/tideglusib-loaded core/shell PMMA/silk fibroin electrospun fibers	Methacrylated gelatin/thiolated pectin hydrogels	Melatonin/tideglusib	Vital pulp regeneration	Synthetic	2023	[216]

Table 1. Cont.

Invention	Matrix Polymer	Active Substance	Application	Origin	Year	Literature
Chitosan–sEDMP and melatonin–chitosan–sEDMP composite membranes	Chitosan-sulfonated ethylene–propylene–diene terpolymer (Chi-sEPDM) membrane Sulfonated ethylene–propylene–diene terpolymer (sEPDM) membrane	Melatonin	Membranes	Synthetic	2023	[217]
Melatonin/nanoclay hybrids	Nanoclay	Melatonin	Skin delivery	Natural	2022	[218]
Biomimetic multilayer polycaprolactone/sodium alginate hydrogel scaffolds loaded with melatonin	Polycaprolactone/sodium alginate	Melatonin	Tendon regeneration	Natural/synthetic	2022	[219]
Melatonin-polycaprolactone/gelatin electrospun fibrous scaffolds	Polycaprolactone/gelatin	Melatonin	Nerve tissue engineering	Natural/synthetic	2022	[220]
Melatonin-cultured collagen/chitosan scaffolds cross-linked by a glyoxal solution	Collagen/chitosan 3D scaffolds cross-linked by glyoxal	Melatonin	Skin tissue engineering	Natural/synthetic	2022	[221]
Melatonin-loaded gelatin sponge	Gelatin sponge	Melatonin	Wound dressing	Natural	2022	[222]
Melatonin-loaded carboxymethyl chitosan (CMCS)-based hydrogel	Carboxymethyl chitosan (CMCS)-based hydrogel	Melatonin	Wound healing	Synthetic	2021	[223]
Polymeric matrix loaded with melatonin	Chitosan/collagen (CTS/Coll)-contained biomaterials	Melatonin	Wound healing	Natural/synthetic	2021	[224]
Nanofiber wound dressing loaded with melatonin	Chitosan–polycaprolactone (PCL)/polyvinylalcohol (PVA)	Melatonin	Wound dressing	Synthetic	2021	[225]
3D-printing magnesium–polycaprolactone loaded with melatonin	Polycaprolactone	Melatonin	Osteosarcoma treatment	Synthetic	2021	[226]
Bio-hybrid hydrogel comprising collagen-capped silver nanoparticles and melatonin	Bio-hybrid hydrogel system comprising collagen and aminated xanthan gum	Silver nanoparticles and melatonin	Tissue regeneration in skin defects	Natural/synthetic	2021	[227]

5. State of Innovation in Sericin-Polymer Wound Healing Patches

Sericin is a “glue-like” protein coiled around the protein core which keeps the fibroin filaments together. This macromolecule constitutes a globular protein built from coil and β -sheets [228]. The coil structure for β -sheet can change in response to temperature, mechanical stretching properties, and moisture absorption. In 50–60 °C water solution, the protein acquires its soluble form. At lower temperatures, the coil structure converts into β -sheets resulting in the gel formation [229]. Sericin exhibits a hydrophilic character and is composed of 18 amino acids with polar groups such as hydroxyl, carboxyl, and amino groups. The group formation is capable of forming crosslinks, co-polymerizations, and compositions with other polymers [230]. The significant structure of sericin defines its biological properties that includes anti-bacterial activity, antioxidant, and biocompatibility [231].

The antioxidant and photoprotective potential of sericin against UVB in human epidermal keratinocytes was confirmed using the flow cytometry assessment [232]. It was revealed that treatment with sericin significantly attenuated apoptosis by inhibiting the

expression of pro-apoptotic proteins and upregulation of the anti-apoptotic Bcl-2 proteins family, as well as preventing Casp-3 activation [232]. The role of sericin in preventing mitochondrial damage was also confirmed by the inhibition of hydrogen peroxide formation. As a consequence, the intracellular ROS/RNS and activation of poly-ADP-ribose polymerase enzyme (PARP) were distinctly reduced. Studies have shown that sericin is a potent antioxidant and anti-apoptotic agent [232,233]. Additionally, the antioxidant properties of sericin are related to high serine and threonine content, whereas hydroxyl groups act as a chelator of trace elements such as copper and iron [234,235].

In recent years, wound healing patches containing sericin have been the center of attention (Table 2). Sericin as a component of the matrix appeared next to polymers: poly (vinyl alcohol) [236], sodium carboxy-methyl-cellulose and polyvinyl alcohol [237], placenta-derived extracellular matrix [238], silk fiber [239], PVA/chitosan [240], PVA [241], robust alcohol, polyurethane/chitosan [242], carboxymethyl cellulose [243], poly (*N*-isopropylacrylamide) [244], cellulose/silk nonwoven fabric [245], collagen-fibrin [246], poly(2-hydroxyethyl methacrylate) [247], cellulose [248], carboxymethyl cellulose as hydrogels [249], cellulose/poly(vinyl alcohol) [250], poly lactide-co-glycolic acid [251], PVA/collagen [252], PVA/chitosan [253], polycaprolactone/cellulose acetate/fibroin [254], poly(ethyleneterephthalate)-*g*-poly (hydroxyethylmethacrylate, PET-*g*-HEMA) nanofibers [255], sericin/chitosan/polyvinyl alcohol [256], gelatin [257], fibroin [258], poly(Σ -caprolactone)/poly(ethylene oxide) [259]. There are also sericin-based patches with only sericin or sericin treated with HRP/H₂O₂ as a matrix, but in significantly smaller amounts [260,261].

Table 2. State of innovation in sericin-polymer patches.

Invention	Matrix Polymer	Active Substance	Application	Origin	Year	Literature
Silk sericin/poly (vinyl alcohol) hydrogel	Poly (vinyl alcohol) hydrogel	Sericin	Infected large burn wound healing	Natural/synthetic	2023	[236]
Silk sericin-based hydrogel	Sodium carboxy-methyl-cellulose and polyvinylalcohol	Sericin	Acute wounds	Natural/synthetic	2023	[237]
Sericin/human placenta-derived extracellular matrix scaffolds	Placenta-derived extracellular matrix	Sericin/human placenta	Wound treatment	Natural	2023	[238]
Resveratrol loaded native silk fiber-sericin hydrogel	Hydrogel	Resveratrol	Wound healing	Natural	2023	[239]
PVA/sericin/chitosan nanofibrous matrix	PVA/sericin/chitosan	Sericin	Wound dressing	Natural/synthetic	2023	[240]
Sericin/PVA hydrogels	Sericin/PVA hydrogels	Sericin	Wound dressing	Natural/synthetic	2023	[241]
Robust alcohol soluble polyurethane/chitosan/silk sericin (APU/CS/SS) nanofiber artificial skin	Robust alcohol soluble polyurethane/chitosan/silk sericin	N/A	Artificial skin	Natural/synthetic	2023	[242]
Turmeric-loaded carboxymethyl cellulose/silk sericin dressings	Carboxymethyl cellulose/silk sericin	Turmeric	Wound healing	Natural/synthetic	2023	[243]

Table 2. Cont.

Invention	Matrix Polymer	Active Substance	Application	Origin	Year	Literature
Silver-loaded anti-bacterial sericin/poly (N-isopropylacrylamide) hydrogel	Sericin/poly (N-isopropylacrylamide) hydrogel	Silver/sericin	Wound healing	Natural/synthetic	2023	[244]
Cellulose/silk nonwoven fabric/silk sericin sandwich membrane	Cellulose/silk nonwoven fabric/silk sericin	Sericin	Wound healing	Natural/synthetic	2023	[245]
Silk sericin-functionalized dense collagen/fibrin hybrid hydrogels	Collagen/fibrin	Sericin/collagen	Regenerative scaffolds	Natural	2023	[246]
Sericin poly(2-hydroxyethyl methacrylate) hydrogel scaffolds	Poly(2-hydroxyethyl methacrylate)	Sericin	Tissue engineering	Natural/synthetic	2023	[247]
Microstructured bacterial cellulose-silk sericin	Cellulose-silk sericin	Sericin	Wound dressing	Natural/synthetic	2022	[248]
Carboxymethyl cellulose/sericin-based hydrogels	Carboxymethyl cellulose/sericin	Sericin	Wound healing	Natural/synthetic	2022	[249]
Porous bacterial cellulose/poly(vinyl alcohol)-based silk sericin and azithromycin release system	Cellulose/poly(vinyl alcohol)-based silk sericin	Sericin/azithromycin	Wound healing	Natural/synthetic	2022	[250]
Silk sericin/PLGA electrospun scaffolds	Silk sericin/poly lactide-co-glycolic acid	Sericin	Periodontal tissue engineering	Natural/synthetic	2022	[251]
Polyvinyl alcohol (PVA) hydrogel with collagen hydrolysate and silk sericin	PVA/collagen/sericin	Collagen/sericin	Wound healing	Natural/synthetic	2022	[252]
PVA/sericin/chitosan nanofibrous wound dressing matrix	PVA/sericin/chitosan	Sericin	Wound dressing matrix	Natural/synthetic	2022	[253]
Polycaprolactone/cellulose acetate blended nanofiber mats containing sericin and fibroin for biomedical application	Polycaprolactone/cellulose acetate/sericin/fibroin	Sericin	Biomedical application	Natural/synthetic	2022	[254]
PET-based nanofiber dressing material coated with silk sericin capped-silver nanoparticles	Poly (ethylene terephthalate)-g-poly (hydroxyethylmethacrylate)nanoparticles (PET-g-HEMA) nanofibers	Silver	Wound dressing	Natural/synthetic	2021	[255]

Table 2. Cont.

Invention	Matrix Polymer	Active Substance	Application	Origin	Year	Literature
Silver nanoparticles@organic frameworks/graphene oxide (Ag@MOF-GO) in sericin/chitosan/polyvinyl alcohol hydrogel	Sericin/chitosan/polyvinyl alcohol	Silver nanoparticles	Wound healing	Natural/synthetic	2021	[256]
Poly(lactic-co-glycolic acid) (PLGA) nanoparticles incorporated into sericin/gelatin nanofibers	Sericin/gelatin	Poly(lactic-co-glycolic acid) nanoparticles	Wound healing	Natural/synthetic	2021	[257]
Silk sericin/fibroin electrospinning dressings	Silk sericin/fibroin	Sericin	Wound dressing	Natural/synthetic	2021	[258]
Poly(Σ -caprolactone) poly(ethylene oxide) sandwich type nanofibers containing sericin-capped silver nanoparticles	Poly(Σ -caprolactone)/poly(ethylene oxide)	Silver nanoparticles	Wound healing	Natural/synthetic	2021	[259]
Sericin scaffolds with ethanol post-treatments	Sericin	Sericin	Tissue engineering	Natural/synthetic	2023	[260]
Horseradish peroxidase-mediated cross-linked sericin hydrogels	Sericin treated HRP/H ₂ O ₂	Sericin	Wound healing	Natural/synthetic	2021	[261]

The main active substances in those inventions included resveratrol [239], turmeric [243], silver nanoparticles [255], poly(lactic-co-glycolic acid) nanoparticles [257], and sericin as a sole ingredient [260,261] or in combinations with human placenta [238], silver [244], collagen [246] or azithromycin [250]. The applications of sericin-polymer patches in recent years involved wound treatment [238,239,243–245,249,250,252,254,257,259,261], including acute [237] and infected large burn wounds [236], artificial skin [242], wound dressings [240,241,248,253,255,258], tissue engineering [247], regenerative scaffolds [246], and periodontal tissue engineering [251]. The composition of the wound healing patches in the years 2021–2023 was mostly a combination of natural and synthetic components, where sericin was the natural ingredient. There were also fully natural patches designed from sericin and human placenta [238], as well as the combination of sericin and collagen [246].

The studies have shown that nanofiber (NF) wound dressing loaded with 20% melatonin (NF + 20% MEL) indicated the highest mRNA expression of collagen type 1 (COL1A1) on the 14th day of treatment compared to Comfeel Plus, NF + 10% MEL, nanofiber dressing, and the non-treated group. Thus, NF + 20% MEL electrospun wound dressing could be used as an effective matrix for accelerating the wound healing process reducing the costs of wound repair [225].

Another study has shown that the melatonin-loaded hydrogel significantly increased the percent of effectiveness of wound closure, promoted tissue granulation and re-epithelialization, leading to accelerated collagen deposition, when compared to the control group and the hydrogel with no melatonin groups. Therefore, it is essential to conduct further research using melatonin in wound healing patches [223].

6. Discussion: Melatonin/Sericin Collagen Scaffolds as Possible Wound Healing Patches

Recent advances in melatonin-polymer wound healing patches showed that melatonin is the main active ingredient [213–227]. Melatonin was also combined with other active substances to enhance regenerative, healing or anti-bacterial properties of dressings such as γ -cyclodextrin [215], tideglusib [216], and silver nanoparticles [227]. In comparison, sericin was used most often as a polymer in combinations with polyvinyl alcohol [237], carboxymethyl cellulose [243], sodium carboxymethylcellulose and polyvinyl alcohol [237], poly lactide-co-glycolic acid [251], PVA/chitosan [253], PVA/collagen [252], poly (*N*-isopropylacrylamide) [244], polyurethane/chitosan [242], poly(2-hydroxyethyl methacrylate) [247], polycaprolactone/cellulose acetate/fibroin [254], poly (ethylene terephthalate)-*g*-poly(hydroxyethylmethacrylate) (PET-*g*-HEMA) nanofibers [255], and poly(Σ -caprolactone)/poly(ethylene oxide) [259]. Less often occurring polymers used in combinations with sericin were gelatin [257], fibroin [258], cellulose [248], collagen/fibrin [246], placenta-derived extracellular matrix [238], silk fiber [239], and cellulose/silk nonwoven fabric [245]. Sericin used separately as a matrix was relatively rare, probably because of its mechanical properties [260,261]. Sericin used in the mentioned inventions were treated with HRP/H₂O₂ to enhance the mechanical properties [261]. When applied as a polymer of the matrix, it was used in combination with resveratrol [239], turmeric [243], silver nanoparticles [255], poly(lactic-co-glycolic acid) nanoparticles [257] as the active ingredients. Although sericin also constituted the main substance in the wound healing patches over the years 2021–2023 [236–261], it was effectively used in combinations with human placenta [238], silver [244], collagen [246] or azithromycin [250] to increase the efficacy of wound-healing dressings (Figure 3).

Polymers including melatonin were similar to those that were used in sericin wound healing patches such as polycaprolactone [214,259] and carboxymethyl cellulose [215,249], whereas similar polymers used in different combinations included silk fibroin/methacrylate, methacrylated gelatin/thiolated pectin hydrogel, poly(2-hydroxyethyl methacrylate) [213,216,247], chitosan-sulfonated ethylene-propylene-diene terpolymer, sulfonated ethylene-propylene-diene terpolymer [217], collagen/chitosan cross-linked by glyoxal [221], carboxymethyl chitosan [223], chitosan/collagen [224], PVA/sericin/chitosan [240,253,256], polycaprolactone/gelatin [220], gelatin [257], collagen with aminated xanthan gum [227] and collagen-fibrin [246]. Sericin-polymer patches in recent years have had a variety of applications as follows: wound treatment [238,239,243–245,249,250,252,254,256,257,259,261], including acute [237] and infected large burn wounds [236], artificial skin [242], wound dressings [240,241,248,253,255,258], tissue engineering [247], regenerative scaffolds [246], and periodontal tissue engineering [251]. The main applications of melatonin-polymer patches in the years 2021–2023 were wound healing [214,223,224] and wound dressings [222,225], also used in diabetic wound repair [215], skin delivery [218], and skin tissue engineering [221]. During this period, melatonin-polymer patches were employed in cartilage repair [213], vital pump regeneration [216], tendon regeneration [216], membranes [217], nerve tissue engineering [220], skin tissue regeneration [227], and osteosarcoma treatment [226]. Active compounds combined with resveratrol and turmeric exhibit antioxidant and anti-inflammatory properties [239,243], whereas silver nanoparticles or poly(lactic-co-glycolic acid) nanoparticles reveal anti-bacterial effect [255,257]. Thus, for wound healing patches it was essential to include them as proof of the regeneration processes of the skin. Finally, numerous reports have shown that both melatonin and sericin but also resveratrol and turmeric exhibit high antioxidant potential [262–265].

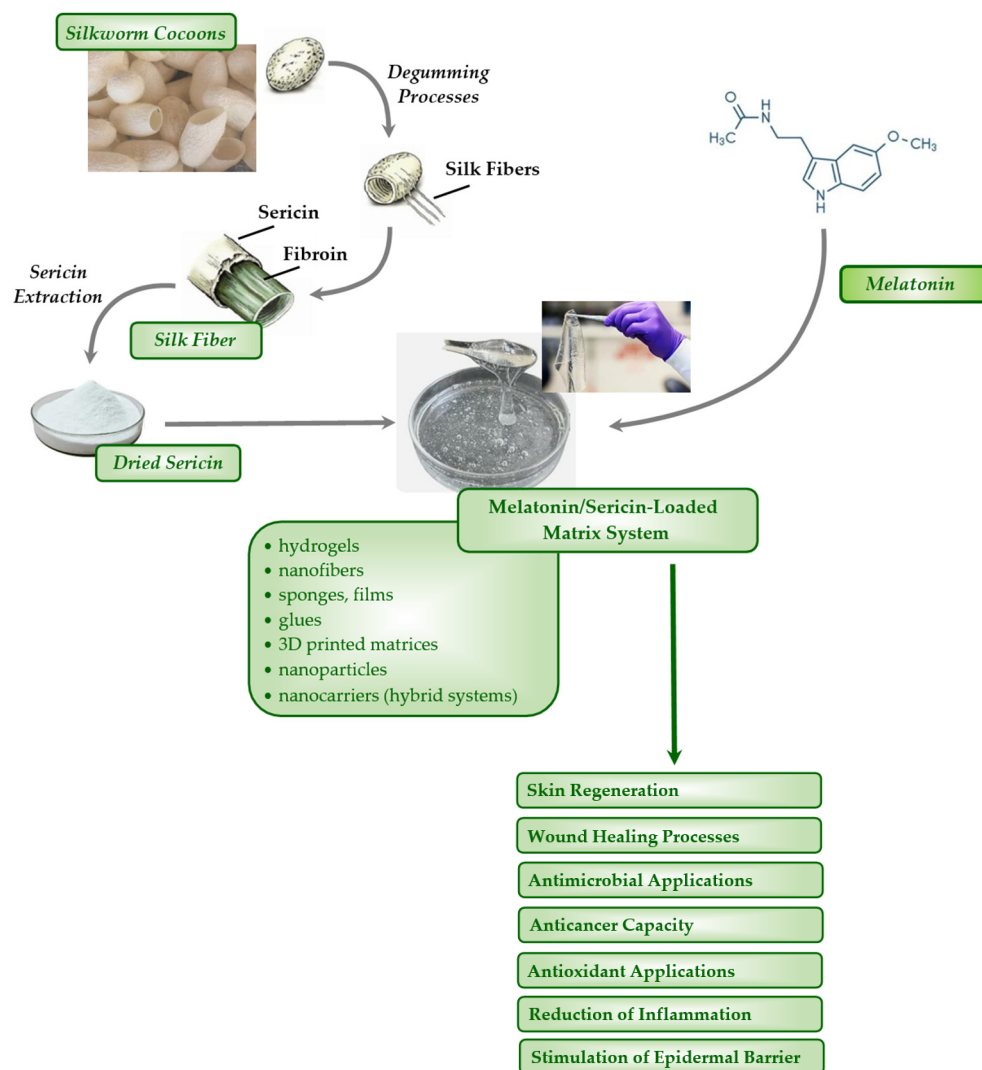


Figure 3. Melatonin/sericin-loaded matrix systems and their potential impact on skin-related applications.

The elevating secretion of pro-inflammatory cytokines, increased cell proliferation and oxidation processes can damage cell membrane integrity and lead to DNA fragmentation [7,27]. Although UVR is one of the major melanoma risk factors it is also essential for initiating vitamin D synthesis, while active forms of vitamin D also have photoprotective properties [266–268]. The topical treatment of large or non-resectable melanoma lesions including lentigo maligna using a combination of melatonin and sericin also shows that mitochondrial and antioxidant protection activity may lead to the development of a valuable additional strategy targeting melanoma, regarding the antibacterial properties of sericin that accelerate wound healing [269,270].

7. Conclusions, Challenges and Future Perspectives

Sericin-polymer patches have a variety of applications such as wound treatment [237,238], including acute [237] and infected large burn wounds [236], artificial skin [242], wound dressings [240,241], tissue engineering [247], regenerative scaffolds [246], and periodontal tissue engineering [251], whereas the main applications of melatonin-polymer patches in years 2021–2023 were wound healing and wound dressings [222–225]; they were also used in diabetic wound repair [215], skin delivery [218], and skin tissue engineering [221]. During this period, melatonin-polymer patches were also used in cartilage repair, vital pump regeneration, tendon regeneration, membranes, nerve tissue engineering, skin tissue regeneration, and osteosarcoma treatment [213,216,217,220,226,227]. Research has shown

that both melatonin and sericin exhibit high antioxidant potential that is beneficial in melanoma treatment [271]. Melatonin/sericin-polymer patches have not been exploited for their application in post-melanoma wound healing treatment; this is the field that would likely benefit from further investigation.

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References

- Slominski, A.; Wortsman, J. Neuroendocrinology of the skin. *Endocr. Rev.* **2000**, *21*, 457–487. [\[CrossRef\]](#) [\[PubMed\]](#)
- Slominski, A.; Zmijewski, M.; Skobowiat, C.; Zbytek, B.; Slominski, R.; Steketee, J. *Sensing the Environment: Regulation of Local and Global Homeostasis by the Skin's Neuroendocrine System*; Advances in Anatomy, Embryology and Cell Biology; Springer Science and Business Media: Dordrecht, The Netherlands, 2012; Volume 212, pp. 1–115.
- Bocheva, G.; Slominski, R.; Slominski, A. Neuroendocrine aspects of skin aging. *Int. J. Mol. Sci.* **2019**, *20*, 2798. [\[CrossRef\]](#) [\[PubMed\]](#)
- Choi, W.; Miyamura, Y.; Wolber, R.; Smuda, C.; Reinhold, W.; Liu, H.; Kolbe, L.; Hearing, V. Regulation of human skin pigmentation In Situ by repetitive UV exposure: Molecular characterization of responses to UVA and/or UVB. *J. Investig. Dermatol.* **2010**, *130*, 1685–1696. [\[CrossRef\]](#) [\[PubMed\]](#)
- Edwards, C.; Pearse, A.; Marks, R.; Nishimori, Y.; Matsumoto, K.; Kawai, M. Degenerative Alterations of Dermal Collagen Fiber Bundles in Photodamaged Human Skin and UV-Irradiated Hairless Mouse Skin: Possible Effect on Decreasing Skin Mechanical Properties and Appearance of Wrinkles. *J. Investig. Dermatol.* **2001**, *117*, 1458–1463. [\[CrossRef\]](#) [\[PubMed\]](#)
- Moan, J.; Grigalavicius, M.; Baturaite, Z.; Dahlback, A.; Juzeniene, A. The relationship between UV exposure and incidence of skin cancer. *Photodermatol. Photoimmunol. Photomed.* **2015**, *31*, 26–35. [\[CrossRef\]](#) [\[PubMed\]](#)
- Bajgar, R.; Moukova, A.; Chalupnikova, N.; Kolarova, H. Differences in the effects of broad-band UVA and narrow-band UVB on epidermal keratinocytes. *Int. J. Environ. Res. Public Health* **2021**, *18*, 12480. [\[CrossRef\]](#) [\[PubMed\]](#)
- Berman, B. Basal cell carcinoma and actinic keratoses: Patients' perceptions of their disease and current treatments. *Int. J. Dermatol.* **2001**, *40*, 573–576. [\[CrossRef\]](#) [\[PubMed\]](#)
- Brash, D.; Rudolph, J.; Simon, J.; Lin, A.; McKenna, G.; Baden, H.; Halperin, A.; Ponten, J. A role for sunlight in skin cancer: UV-induced p53 mutations in squamous cell carcinoma. *Proc. Natl. Acad. Sci. USA* **1991**, *88*, 10124–10128. [\[CrossRef\]](#) [\[PubMed\]](#)
- Collins, G.; Nickoonahand, N.; Morgan, M. Changing demographics and pathology of nonmelanoma skin cancer in the last 30 years. *Semin. Cutan. Med. Surg.* **2004**, *23*, 80–83. [\[CrossRef\]](#)
- Ley, R.D. Ultraviolet radiation A-induced precursors of cutaneous melanoma in *Monodelphis domestica*. *Cancer Res.* **1997**, *57*, 3682–3684.
- Gidanian, S.; Mentelle, M.; Meyskens, F.; Farmer, P. Melanosomal damage in normal human melanocytes induced by UVB and metal uptake—A basis for the pro-oxidant state of melanoma. *Photochem. Photobiol.* **2008**, *84*, 556–564. [\[CrossRef\]](#) [\[PubMed\]](#)
- Bjorn, L. *Photobiology: The Science of Life and Light*, 2nd ed.; Springer: New York, NY, USA, 2008.
- Morison, W. *Phototherapy and Photochemotherapy for Skin Disease*, 3rd ed.; CRC Press: Boca Raton, FL, USA, 2005.
- Gilchrest, B. *Photodamage*, 1st ed.; Blackwell Science: Cambridge, MA, USA, 1995.
- Brenner, M.; Hearing, V.J. The protective role of melanin against UV damage in human skin. *Photochem. Photobiol.* **2008**, *84*, 539–549. [\[CrossRef\]](#) [\[PubMed\]](#)
- Skobowiat, C.; Dowdy, J.; Sayre, R.; Tuckey, R.; Slominski, A. Cutaneous hypothalamic-pituitary-adrenal axis homolog: Regulation by ultraviolet radiation. *Am. J. Physiol. Endocrinol. Metab.* **2011**, *301*, 484–493. [\[CrossRef\]](#)
- Slominski, R.M.; Chen, J.Y.; Raman, C.; Slominski, A.T. Photo-neuro-immuno-endocrinology: How the ultraviolet radiation regulates the body, brain, and immune system. *Proc. Natl. Acad. Sci. USA* **2024**, *121*, e2308374121. [\[CrossRef\]](#) [\[PubMed\]](#)

19. Slominski, A. Ultraviolet radiation (UVR) activates central neuro-endocrine-immune system. *Photodermatol. Photoimmunol. Photomed.* **2015**, *31*, 121–123. [[CrossRef](#)] [[PubMed](#)]
20. Skobowiat, C.; Slominski, A. UVB Activates Hypothalamic-Pituitary-Adrenal Axis in C57BL/6 Mice. *J. Investig. Dermatol.* **2015**, *135*, 1638–1648. [[CrossRef](#)] [[PubMed](#)]
21. Slominski, A.T.; Tuckey, R.C.; Jetten, A.M.; Holick, M.F. Recent Advances in Vitamin D Biology: Something New under the Sun. *J. Investig. Dermatol.* **2023**, *143*, 2340–2342. [[CrossRef](#)] [[PubMed](#)]
22. Holick, M.F.; Slominski, A.T. Photobiology of vitamin D. In *Feldman and Pike's Vitamin D*; Hewison, M., Ed.; Academic Press: Oxford, UK, 2024; pp. 27–45.
23. Cockell, C.; Gerda, H. The History of the UV Radiation Climate of the Earth—Theoretical and Space-based Observations. *Photochem. Photobiol.* **2001**, *73*, 447–451. [[CrossRef](#)] [[PubMed](#)]
24. Cockell, C. Ultraviolet radiation and the photobiology of earth's early oceans. *Orig. Life Evol. Biosph.* **2000**, *30*, 467–500. [[CrossRef](#)]
25. Dworkin, J.; Deamer, D.; Sandford, S.; Allamandola, L. Self-assembling amphiphilic molecules: Synthesis in simulated interstellar/precometary ices. *Proc. Natl. Acad. Sci. USA* **2001**, *98*, 815–819. [[CrossRef](#)]
26. Slominski, A.; Zmijewski, M.; Plonka, P.; Szaflarski, J.; Paus, R. How UV Light Touches the Brain and Endocrine System through Skin, and Why. *Endocrinology* **2018**, *159*, 1992–2007. [[CrossRef](#)] [[PubMed](#)]
27. Kim, T.K.; Slominski, R.; Pyza, E.; Kleszczyński, K.; Tuckey, R.; Reiter, R.J.; Holick, M.; Slominski, A.T. Evolutionary formation of melatonin and vitamin D in early life forms: Insects take centre stage. *Biol. Rev.* **2024**, *in press*. [[CrossRef](#)]
28. Ho, Y.; Wu, J.; Chang, C. A new natural antioxidant biomaterial from *Cinnamomum osmophloeum* Kanehira leaves represses melanogenesis and protects against DNA damage. *Antioxidants* **2019**, *8*, 474. [[CrossRef](#)] [[PubMed](#)]
29. Bose, V.; Balaganesan, V.; Govindaraj, G.; Veerichetty, V. Cellular antioxidant and cytotoxic activity of astaxanthin and ellagic acid on UV irradiated skin melanoma cells and gel formulation. *Mater. Today Proc.* **2023**, *in press*.
30. Galano, A.; Tan, D.; Reiter, R. Melatonin: A versatile protector against oxidative DNA damage. *Molecules* **2018**, *23*, 530. [[CrossRef](#)] [[PubMed](#)]
31. Salucci, S.; Burattini, S.; Buontempo, F.; Martelli, A.; Falcieri, E.; Battistelli, M. Protective effect of different antioxidant agents in UVB-irradiated keratinocytes. *Eur. J. Histochem.* **2017**, *61*, 215–221. [[CrossRef](#)] [[PubMed](#)]
32. Kleszczyński, K.; Zwicker, S.; Tukaj, S.; Kasperkiewicz, M.; Zillikens, D.; Wolf, R.; Fischer, T.W. Melatonin compensates silencing of heat shock protein 70 and suppresses ultraviolet radiation-induced inflammation in human skin ex vivo and cultured keratinocytes. *J. Pineal Res.* **2015**, *58*, 117–126. [[CrossRef](#)] [[PubMed](#)]
33. López-Burillo, S.; Tan, D.; Rodríguez-Gallego, V.; Manchester, L.; Mayo, J.; Sainz, R.; Reiter, R. Melatonin and its derivatives cyclic 3-hydroxymelatonin, N¹-acetyl-N²-formyl-5-methoxykynuramine and 6-methoxymelatonin reduce oxidative DNA damage induced by Fenton reagents. *J. Pineal Res.* **2003**, *34*, 178–184. [[CrossRef](#)]
34. Sliwinski, T.; Rozej, W.; Morawiec-Bajda, A.; Morawiec, Z.; Reiter, R.; Blasiak, J. Protective action of melatonin against oxidative DNA damage—Chemical inactivation *versus* base-excision repair. *Mutat. Res. Genet. Toxicol. Environ. Mutagen.* **2007**, *634*, 220–227. [[CrossRef](#)] [[PubMed](#)]
35. Fischer, T.; Kleszczyński, K.; Hardkop, L.; Kruse, N.; Zillikens, D. Melatonin enhances antioxidative enzyme gene expression (CAT, GPx, SOD), prevents their UVR-induced depletion, and protects against the formation of DNA damage (8-hydroxy-2'-deoxyguanosine) in Ex Vivo human skin. *J. Pineal Res.* **2013**, *54*, 303–312. [[CrossRef](#)]
36. Hu, X.; Tian, X.; Yang, C.; Ling, F.; Liu, H.; Zhu, X.; Pei, M.; Yang, H.; Liu, T.; Xu, Y.; et al. Melatonin-loaded self-healing hydrogel targets mitochondrial energy metabolism and promotes annulus fibrosus regeneration. *Mater. Today Bio* **2023**, *23*, 100811. [[CrossRef](#)]
37. Wei, L.; Yu, M.; Xie, D.; Wang, L.; Ye, C.; Zhu, Q.; Liu, F.; Yang, L. Melatonin-stimulated MSC-derived exosomes improve diabetic wound healing through regulating macrophage M1 and M2 polarization by targeting the PTEN/AKT pathway. *Stem Cell Res. Ther.* **2020**, *11*, 259.
38. Rupesh, D.; Acharya, C.; Bindu, P.; Kundu, S. Antioxidant potential of silk protein sericin against hydrogen peroxide-induced oxidative stress in skin fibroblasts. *BMB Rep.* **2008**, *41*, 236–241.
39. Boni, B.; Lamboni, L.; Bakadia, B.; Hussein, S.; Yang, G. Combining silk sericin and surface micropatterns in bacterial cellulose dressings to control fibrosis and enhance wound healing. *Eng. Sci.* **2020**, *10*, 68–77. [[CrossRef](#)]
40. Slominski, A.; Zmijewski, M.; Semak, I.; Kim, T.; Janjetovic, Z.; Slominski, R.; Zmijewski, J. Melatonin, mitochondria, and the skin. *Cell. Mol. Life Sci.* **2017**, *74*, 3913–3925. [[CrossRef](#)] [[PubMed](#)]
41. Holtkamp, C.; Warmus, D.; Bonowicz, K.; Gagat, M.; Linowiecka, K.; Wolnicka-Glubisz, A.; Reiter, R.; Böhm, M.; Slominski, A.; Steinbrink, K.; et al. Ultraviolet Radiation-Induced Mitochondrial Disturbances Are Attenuated by Metabolites of Melatonin in Human Epidermal Keratinocytes. *Metabolites* **2023**, *13*, 861. [[CrossRef](#)] [[PubMed](#)]
42. Slominski, A.; Kleszczyński, K.; Semak, I.; Janjetovic, Z.; Zmijewski, M.; Kim, T.; Slominski, R.; Reiter, R.; Fischer, T. Local melatonergic system as the protector of skin integrity. *Int. J. Mol. Sci.* **2014**, *15*, 17705–17732. [[CrossRef](#)] [[PubMed](#)]
43. Kleszczyński, K.; Tukaj, S.; Kruse, N.; Zillikens, D.; Fischer, T. Melatonin prevents ultraviolet radiation-induced alterations in plasma membrane potential and intracellular pH in human keratinocytes. *J. Pineal Res.* **2013**, *54*, 89–99. [[CrossRef](#)] [[PubMed](#)]
44. Acuña-Castroviejo, D.; Escames, G.; Venegas, C.; Díaz-Casado, M.; Lima-Cabello, E.; López, L.; Sergio Rosales-Corral, D.; Reiter, R. Extrapineal melatonin: Sources, regulation, and potential functions. *Cell. Mol. Life Sci.* **2014**, *71*, 2997–3025. [[CrossRef](#)] [[PubMed](#)]

45. Slominski, A.; Wortsman, J.; Tobin, D. The cutaneous serotonergic/melatonergic system: Securing a place under the sun. *FASEB J.* **2015**, *19*, 176–194. [[CrossRef](#)] [[PubMed](#)]
46. Slominski, A.; Hardeland, R.; Zmijewski, M.; Slominski, R.; Reiter, R.; Paus, R. Melatonin: A Cutaneous Perspective on its Production, Metabolism, and Functions. *J. Investig. Dermatol.* **2018**, *138*, 490–499. [[CrossRef](#)]
47. Vasey, C.; McBride, J.; Penta, K. Circadian rhythm dysregulation and restoration: The role of melatonin. *Nutrients* **2021**, *13*, 3480. [[CrossRef](#)]
48. Du Plessis, S.; Hagenaar, K.; Lampiao, F. The In Vitro effects of melatonin on human sperm function and its scavenging activities on NO and ROS. *Andrologia* **2010**, *42*, 112–116. [[CrossRef](#)] [[PubMed](#)]
49. Liu, R.; Fu, A.; Hoffman, A.; Zheng, T.; Zhu, Y. Melatonin enhances DNA repair capacity possibly by affecting genes involved in DNA damage responsive pathways. *BMC Cell Biol.* **2013**, *14*, 1. [[CrossRef](#)] [[PubMed](#)]
50. Ackermann, K.; Plomp, R.; Lao, O.; Middleton, B.; Revell, V.; Skene, D.; Kayser, M. Effect of sleep deprivation on rhythms of clock gene expression and melatonin in humans. *Chronobiol. Int.* **2013**, *30*, 901–909. [[CrossRef](#)] [[PubMed](#)]
51. Pugazhenth, K.; Kapoor, M.; Clarkson, A.; Hall, I.; Appleton, I. Melatonin accelerates the process of wound repair in full-thickness incisional wounds. *J. Pineal Res.* **2008**, *44*, 387–396. [[CrossRef](#)] [[PubMed](#)]
52. Kumar, J.; Alam, S.; Jain, A.; Ansari, K.; Mandal, B. Protective activity of silk sericin against UV radiation-induced skin damage by downregulating oxidative stress. *ACS Appl. Bio Mater.* **2018**, *1*, 2120–2132. [[CrossRef](#)] [[PubMed](#)]
53. Young, A. Chromophores in human skin. *Phys. Med. Biol.* **1997**, *42*, 789–802. [[CrossRef](#)] [[PubMed](#)]
54. Akturk, O.; Tezcaner, A.; Bilgili, H.; Deveci, M.; Gecit, M.; Keskin, D. Evaluation of sericin/collagen membranes as prospective wound dressing biomaterial. *J. Biosci. Bioeng.* **2011**, *112*, 279–288. [[CrossRef](#)] [[PubMed](#)]
55. Tao, G.; Cai, R.; Wang, Y.; Zuo, H.; He, H. Fabrication of antibacterial sericin based hydrogel as an injectable and mouldable wound dressing. *Mater. Sci. Eng. C* **2021**, *119*, 111597. [[CrossRef](#)] [[PubMed](#)]
56. Chirila, T.; Suzuki, S.; McKirdy, N. Further development of silk sericin as a biomaterial: Comparative investigation of the procedures for its isolation from *Bombyx mori* silk cocoons. *Prog. Biomater.* **2016**, *5*, 135–145. [[CrossRef](#)]
57. Chirila, T.; Suzuki, S.; Bray, L.; Barnett, N.; Harkin, D. Evaluation of silk sericin as a biomaterial: In Vitro growth of human corneal limbal epithelial cells on *Bombyx mori* sericin membranes. *Prog. Biomater.* **2013**, *2*, 14. [[CrossRef](#)]
58. Kundu, B.; Kundu, S. Silk sericin/polyacrylamide In Situ forming hydrogels for dermal reconstruction. *Biomaterials* **2012**, *33*, 7456–7467. [[CrossRef](#)] [[PubMed](#)]
59. Mandal, B.; Priya, A.; Kundu, S. Novel silk sericin/gelatin 3-D scaffolds and 2-D films: Fabrication and characterization for potential tissue engineering applications. *Acta Biomater.* **2009**, *5*, 3007–3020. [[CrossRef](#)] [[PubMed](#)]
60. Aramwit, P.; Damrongsakkul, S.; Kanokpanont, S.; Srichana, T. Properties and antityrosinase activity of sericin from various extraction methods. *Biotechnol. Appl. Biochem.* **2010**, *55*, 91–98. [[CrossRef](#)] [[PubMed](#)]
61. Jena, K.; Pandey, J.; Kumari, R.; Sinha, A.; Gupta, V.; Singh, G. Tasar silk fiber waste sericin: New source for anti-elastase, anti-tyrosinase and anti-oxidant compounds. *Int. J. Biol. Macromol.* **2018**, *114*, 1102–1108. [[CrossRef](#)] [[PubMed](#)]
62. Chlapanidas, T.; Faragò, S.; Lucconi, G.; Perteghella, S.; Galuzzi, M.; Mantelli, M.; Avanzini, M.; Tosca, M.; Marazzi, M.; Vigo, D.; et al. Sericins exhibit ROS-scavenging, anti-tyrosinase, anti-elastase, and In Vitro immunomodulatory activities. *Int. J. Biol. Macromol.* **2013**, *58*, 47–56. [[CrossRef](#)] [[PubMed](#)]
63. Liu, J.; Qi, C.; Tao, K.; Zhang, J.; Zhang, J.; Xu, L.; Jiang, X.; Zhang, Y.; Huang, L.; Xie, H.; et al. Sericin/dextran injectable hydrogel as an optically trackable drug delivery system for malignant melanoma treatment. *ACS Appl. Mater. Interfaces* **2016**, *8*, 6411–6422. [[CrossRef](#)] [[PubMed](#)]
64. Villani, A.; Scalvenzi, M.; Micali, G.; Lacarrubba, F.; Fornaro, L.; Martora, F.; Potestio, L. Management of Advanced Invasive Melanoma: New Strategies. *Adv. Ther.* **2023**, *40*, 3381–3394. [[CrossRef](#)] [[PubMed](#)]
65. Wang, S.; Zheng, H.; Zhou, L.; Cheng, F.; Liu, Z.; Zhang, H.; Zhang, Q. Injectable redox and light responsive MnO₂ hybrid hydrogel for simultaneous melanoma therapy and multidrug-resistant bacteria-infected wound healing. *Biomaterials* **2020**, *260*, 120314. [[CrossRef](#)] [[PubMed](#)]
66. Yuan, C.; Zhang, D.; Tang, Y.; Guo, Z.; Lin, K.; Yu, Y.; Li, J.; Cai, Q. Fibrous dressing containing bioactive glass with combined chemotherapy and wound healing promotion for post-surgical treatment of melanoma. *Biomater. Adv.* **2023**, *149*, 213387. [[CrossRef](#)]
67. Dhall, S.; Do, D.; Garcia, M.; Wijesinghe, D.; Brandon, A.; Kim, J.; Sanchez, A.; Lyubovitsky, J.; Gallagher, S.; Nothnagel, E.; et al. A novel model of chronic wounds: Importance of redox imbalance and biofilm-forming bacteria for establishment of chronicity. *PLoS ONE* **2014**, *9*, e109848. [[CrossRef](#)]
68. Dryden, M. Complicated skin and soft tissue infection. *J. Antimicrob. Chemother.* **2010**, *65*, 35–44. [[CrossRef](#)] [[PubMed](#)]
69. Widgerow, A.; King, K.; Tocco-Tussardi, I.; Banyard, D.; Chiang, R.; Awad, A.; Afzel, H.; Bhatnagel, S.; Melkumyan, S.; Wirth, G.; et al. The burn wound exudate—An under-utilized resource. *Burns* **2015**, *41*, 11–17. [[CrossRef](#)] [[PubMed](#)]
70. Song, J.; Razaq, A.; Khan, N.; Iqbal, H.; Ni, J. Chitosan/poly (3-hydroxy butyric acid-co-3-hydroxy valeric acid) electrospun nanofibers with cephradine for superficial incisional skin wound infection management. *Int. J. Biol. Macromol.* **2023**, *250*, 126229. [[CrossRef](#)] [[PubMed](#)]
71. Nayak, S.; Talukdar, S.; Kundu, S. Potential of 2D crosslinked sericin membranes with improved biostability for skin tissue engineering. *Cell Tissue Res.* **2012**, *347*, 783–794. [[CrossRef](#)] [[PubMed](#)]

72. Yuan, L.; Jiang, X.; Jiang, M.; Guo, Y.; Liu, Y.; Ming, P.; Li, S.; Zhou, P.; Cai, R.; Yu, K.; et al. Biocompatible gellan gum/sericin hydrogels containing halloysite@ polydopamine nanotubes with hemostasis and photothermal antibacterial properties for promoting infectious wound repair. *Mater. Des.* **2023**, *227*, 111744. [\[CrossRef\]](#)
73. Oviedo, M.; Montoya, Y.; Alvarez, C.; Bustamante, J. Influence of Electrospinning Parameters on the Physicochemical Properties of Polycaprolactone, Chitosan, and Sericin Membranes. *Mater. Proc.* **2023**, *11*, 5. [\[CrossRef\]](#)
74. Rui, Z.; Li, X.; Sun, B.; Zhang, Y.; Zhang, D.; Tang, Z.; Chen, X.; Wang, C. Electrospun chitosan/sericin composite nanofibers with antibacterial property as potential wound dressings. *Int. J. Biol. Macromol.* **2014**, *68*, 92–97.
75. Sood, A.; Bhaskar, R.; Won, S.; Seok, Y.; Kumar, A.; Han, S. Disulfide bond-driven hyaluronic acid/sericin nanoparticles for wound-healing application. *J. Nanostruct. Chem.* **2023**, *13*, 463–480. [\[CrossRef\]](#)
76. Lee, H.; Jang, M.; Park, B.; Um, I. Structural Characteristics and Properties of Redissolved Silk Sericin. *Polymers* **2023**, *15*, 3405. [\[CrossRef\]](#)
77. Teh, T.; Toh, S.; Goh, J. Optimization of the silk scaffold sericin removal process for retention of silk fibroin protein structure and mechanical properties. *Biomed. Mater.* **2010**, *5*, 35008. [\[CrossRef\]](#)
78. Turbiani, F.; Tomadon, J.; Seixas, F.; Gimenes, M. Properties and structure of sericin films: Effect of the crosslinking degree. *Chem. Eng. Trans.* **2011**, *24*, 1489–1494.
79. Wang, J.; Shang, J.; Ren, F.; Leng, X. Study of the physical properties of whey protein: Sericin protein-blended edible films. *Eur. Food Res. Technol.* **2010**, *231*, 109–116. [\[CrossRef\]](#)
80. Sothornvit, R.; Chollakup, R. Properties of sericin–glucomannan composite films. *Int. J. Food Sci. Technol.* **2009**, *44*, 1395–1400. [\[CrossRef\]](#)
81. Roubenoff, R.; Harris, T.; Abad, L.; Wilson, P.; Dallal, G.; Dinarello, C. Monocyte cytokine production in an elderly population: Effect of age and inflammation. *J. Gerontol. A Biol. Sci. Med. Sci.* **1998**, *53*, 20–26. [\[CrossRef\]](#) [\[PubMed\]](#)
82. Gorni, D.; Finco, A. Oxidative stress in elderly population: A prevention screening study. *Aging Med.* **2020**, *3*, 205–213. [\[CrossRef\]](#) [\[PubMed\]](#)
83. Makrantonaki, E.; Bekou, V.; Zouboulis, C. Genetics and skin aging. *Derm.-Endocrinol.* **2012**, *4*, 280–284. [\[CrossRef\]](#) [\[PubMed\]](#)
84. Venkatesh, S.; Maymone, M.; Vashi, N. Aging in skin of color. *Clin. Dermatol.* **2019**, *37*, 351–357. [\[CrossRef\]](#) [\[PubMed\]](#)
85. Mazzocchi, G.; De Cata, A.; Greco, A.; Damato, M.; Marzulli, N.; Dagostino, M.; Carughi, S.; Perfetto, F.; Tarquini, R. Aging related changes of circadian rhythmicity of cytotoxic lymphocyte subpopulations. *J. Circadian Rhythm.* **2010**, *8*, 6. [\[CrossRef\]](#) [\[PubMed\]](#)
86. Barth, E.; Srivastava, A.; Wengerodt, D.; Stojiljkovic, M.; Axer, H.; Witte, O.; Kretz, A.; Marz, M. Age-dependent expression changes of circadian system-related genes reveal a potentially conserved link to aging. *Aging* **2021**, *13*, 25694–25716. [\[CrossRef\]](#)
87. Gorelik, S.; Belousova, O.; Treneva, E.; Bulgakova, S.; Zakharova, N.; Nesterenko, S. Effect of daily rhythms of cortisol secretion on the rate of aging in men. *Arch. Razi Inst.* **2022**, *77*, 1233–1239.
88. Martinez-Nicolas, A.; Madrid, J.; García, F.; Campos, M.; Moreno-Casbas, M.; Almada-Pagán, P.; Lucas-Sanchez, A.; Rol, M. Circadian monitoring as an aging predictor. *Sci. Rep.* **2018**, *8*, 15027. [\[CrossRef\]](#) [\[PubMed\]](#)
89. Cirillo, N.; Prime, S. Keratinocytes synthesize and activate cortisol. *J. Cell. Biochem.* **2011**, *112*, 1499–1505. [\[CrossRef\]](#) [\[PubMed\]](#)
90. Jackson, E.; Heidl, M.; Imfeld, D.; Meeus, L.; Schuetz, R.; Campiche, R. Discovery of a highly selective MC1R agonists pentapeptide to be used as a skin pigmentation enhancer and with potential anti-aging properties. *Int. J. Mol. Sci.* **2019**, *20*, 6143. [\[CrossRef\]](#) [\[PubMed\]](#)
91. Slominski, A.T.; Slominski, R.M.; Raman, C.; Chen, J.Y.; Athar, M.; Elmets, C. Neuroendocrine signaling in the skin with a special focus on the epidermal neuropeptides. *Am. J. Physiol. Cell Physiol.* **2022**, *323*, C1757–C1776. [\[CrossRef\]](#) [\[PubMed\]](#)
92. Bocheva, G.; Slominski, R.; Slominski, A. The impact of vitamin D on skin aging. *Int. J. Mol. Sci.* **2021**, *22*, 9097. [\[CrossRef\]](#) [\[PubMed\]](#)
93. Slominski, A.; Brożyna, A.; Zmijewski, M.; Jóźwicki, W.; Jetten, A.; Mason, R.; Tuckey, R.; Elmets, C. Vitamin D signaling and melanoma: Role of vitamin D and its receptors in melanoma progression and management. *Lab. Investig.* **2017**, *97*, 706–724. [\[CrossRef\]](#) [\[PubMed\]](#)
94. Janjetovic, Z.; Slominski, A.T. Promising Functions of Novel Vitamin D Derivatives as Cosmetics: A New Fountain of Youth in Skin Aging and Skin Protection. *Cosmetics* **2024**, *11*, 37. [\[CrossRef\]](#)
95. Slominski, A.; Brożyna, A.; Skobowiat, C.; Zmijewski, M.; Kim, T.; Janjetovic, Z.; Oak, A.; Jozwicki, W.; Jetten, A.; Mason, R.S.; et al. On the role of classical and novel forms of vitamin D in melanoma progression and management. *J. Steroid. Biochem. Mol. Biol.* **2018**, *177*, 159–170. [\[CrossRef\]](#) [\[PubMed\]](#)
96. Noh, E.; Park, J.; Song, H.; Kim, J.; Lee, M.; Song, H.; Hong, O.; Whang, P.; Han, M.; Kwon, K.; et al. Skin aging-dependent activation of the PI3K signaling pathway via downregulation of PTEN increases intracellular ROS in human dermal fibroblasts. *Oxid. Med. Cell. Longev.* **2016**, *2016*, 6354261. [\[CrossRef\]](#)
97. Bratic, A.; Larsson, N.G. The role of mitochondria in aging. *J. Clin. Investig.* **2013**, *123*, 951–957. [\[CrossRef\]](#)
98. Trifunovic, A.; Wredenberg, A.; Falkenberg, M.; Spelbrink, J.N.; Rovio, A.T.; Bruder, C.E.; Bohlooly, Y.M.; Gidlöf, S.; Oldfors, A.; Wibom, R.; et al. Premature ageing in mice expressing defective mitochondrial DNA polymerase. *Nature* **2004**, *429*, 417–423. [\[CrossRef\]](#) [\[PubMed\]](#)
99. McCullough, J.L.; Kelly, K.M. Prevention and treatment of skin aging. *Ann. N. Y. Acad. Sci.* **2006**, *1067*, 323–331. [\[CrossRef\]](#) [\[PubMed\]](#)

100. Zhang, R.; Wang, Y.; Ye, K.; Picard, M.; Gu, Z. Independent impacts of aging on mitochondrial DNA quantity and quality in humans. *BMC Genom.* **2017**, *18*, 890. [\[CrossRef\]](#)
101. Anderson, A.; Bowman, A.; Boulton, S.; Manning, P.; Birch-Machin, M. A role for human mitochondrial complex II in the production of reactive oxygen species in human skin. *Redox Biol.* **2014**, *2*, 1016–1022. [\[CrossRef\]](#) [\[PubMed\]](#)
102. Zungu, I.; Hawkins Evans, D.; Abrahamse, H. Mitochondrial responses of normal and injured human skin fibroblasts following low level laser irradiation—An In Vitro study. *Photochem. Photobiol.* **2009**, *85*, 987–996. [\[CrossRef\]](#)
103. Birch-Machin, M.; Bowman, A. Oxidative stress and ageing. *Br. J. Dermatol.* **2016**, *175*, 26–29. [\[CrossRef\]](#)
104. Reiter, R.J.; Mayo, J.C.; Tan, D.X.; Sainz, R.M.; Alatorre-Jimenez, M.; Qin, L. Melatonin as an antioxidant: Under promises but over delivers. *J. Pineal Res.* **2016**, *61*, 259–278. [\[CrossRef\]](#)
105. Tan, D.X.; Chen, L.D.; Poeggeler, B.; Manchester, L.C.; Reiter, R.J. Melatonin: A potent endogenous hydroxyl radical scavenger. *Endocrine* **1993**, *1*, 57–60.
106. Reiter, R.; Tan, D.; Rosales-Corral, S.; Galano, A.; Jou, M.; Acuna-Castroviejo, D. Melatonin mitigates mitochondrial meltdown: Interactions with SIRT3. *Int. J. Mol. Sci.* **2018**, *19*, 2439. [\[CrossRef\]](#)
107. Hardeland, R. Antioxidant protection by melatonin: Multiplicity of mechanisms from radical detoxification to radical avoidance. *Endocrine* **2005**, *27*, 119–130. [\[CrossRef\]](#)
108. Reiter, R.; Rosales-Corral, S.; Tan, D.; Jou, M.; Galano, A.; Xu, B. Melatonin as a mitochondria-targeted antioxidant: One of evolution's best ideas. *Cell. Mol. Life Sci.* **2017**, *74*, 3863–3881. [\[CrossRef\]](#)
109. Reiter, R.; Tan, D.; Manchester, L.; Qi, W. Biochemical reactivity of melatonin with reactive oxygen and nitrogen species: Are view of the evidence. *Cell Biochem. Biophys.* **2001**, *34*, 237–256. [\[CrossRef\]](#)
110. Bonnefont-Rousselot, D.; Collin, F. Melatonin: Action as antioxidant and potential applications in human disease and aging. *Toxicology* **2010**, *278*, 55–67. [\[CrossRef\]](#) [\[PubMed\]](#)
111. Yuksel Egrilmez, M.; Kocturk, S.; Aktan, S.; Oktay, G.; Resmi, H.; Simsek Keskin, H.; Akdogan, G.; Ozkan, S. Melatonin prevents UVB-induced skin photoaging by inhibiting oxidative damage and MMP expression through JNK/AP-1 signaling pathway in human dermal fibroblasts. *Life* **2022**, *12*, 950. [\[CrossRef\]](#)
112. Ayata, A.; Mollaoglu, H.; Yilmaz, H.; Akturk, O.; Ozguner, F.; Altuntas, I. Oxidative stress-mediated skin damage in an experimental mobile phone model can be prevented by melatonin. *J. Dermatol.* **2004**, *31*, 878–883. [\[CrossRef\]](#)
113. Ashrafizadeh, M.; Najafi, M.; Kavyiani, N.; Mohammadinejad, R.; Farkhondeh, T.; Samarghandian, S. Anti-inflammatory activity of melatonin: A focus on the role of NLRP3 inflammasome. *Inflammation* **2021**, *44*, 1207–1222. [\[CrossRef\]](#) [\[PubMed\]](#)
114. Mayo, J.; Sainz, R.; Tan, D.; Hardeland, R.; Leon, J.; Rodriguez, C.; Reiter, R.J. Anti-inflammatory actions of melatonin and its metabolites, N^1 -acetyl- N^2 -formyl-5-methoxykynuramine (AFMK) and N^1 -acetyl-5-methoxykynuramine (AMK), in macrophages. *J. Neuroimmunol.* **2005**, *165*, 139–149. [\[CrossRef\]](#) [\[PubMed\]](#)
115. Tan, D.; Manchester, L.; Qin, L.; Reiter, R. Melatonin: A mitochondrial targeting molecule involving mitochondrial protection and dynamics. *Int. J. Mol. Sci.* **2016**, *17*, 2124. [\[CrossRef\]](#)
116. Hardeland, R. Melatonin and inflammation-story of a double-edged blade. *J. Pineal Res.* **2018**, *65*, e12525. [\[CrossRef\]](#)
117. Galano, A.; Tan, D.X.; Reiter, R.J. On the free radical scavenging activities of melatonin's metabolites, AFMK and AMK. *J. Pineal Res.* **2013**, *54*, 245–257. [\[CrossRef\]](#)
118. Tan, D.; Manchester, L.; Burkhardt, S.; Sainz, R.; Mayo, J.; Kohen, R.; Shohami, E.; Huo, Y.; Hardeland, R.; Reiter, R. N^1 -acetyl- N^2 -formyl-5-methoxykynuramine, a biogenic amine and melatonin metabolite, functions as a potent antioxidant. *FASEB J.* **2001**, *15*, 2294–2296. [\[CrossRef\]](#) [\[PubMed\]](#)
119. Slominski, A.; Fischer, T.W.; Zmijewski, M.; Wortsman, J.; Semak, I.; Zbytek, B.; Slominski, R.; Tobin, D. On the role of melatonin in skin physiology and pathology. *Endocrine* **2005**, *27*, 137–148. [\[CrossRef\]](#) [\[PubMed\]](#)
120. Fischer, T.; Zmijewski, M.; Zbytek, B.; Sweatman, T.; Slominski, R.; Wortsman, J.; Slominski, A. Oncostatic effects of the indole melatonin and expression of its cytosolic and nuclear receptors in cultured human melanoma cell lines. *Int. J. Oncol.* **2006**, *29*, 665–672. [\[CrossRef\]](#) [\[PubMed\]](#)
121. Galano, A.; Tan, D.; Reiter, R. Melatonin as a naturally against oxidative stress: A physicochemical examination. *J. Pineal Res.* **2011**, *51*, 1–16. [\[CrossRef\]](#)
122. Kobayashi, H.; Kromminga, A.; Dunlop, T.; Tychsen, B.; Conrad, F.; Suzuki, N.; Memezawa, A.; Bettermann, A.; Aiba, S.; Carlberg, C.; et al. A role of melatonin in neuroectodermal-mesodermal interactions: The hair follicle synthesizes melatonin and expresses functional melatonin receptors. *FASEB J.* **2005**, *19*, 1710–1712. [\[CrossRef\]](#) [\[PubMed\]](#)
123. Li, W.; Wang, Z.; Cao, J.; Dong, Y.; Chen, Y. Melatonin improves skin barrier damage caused by sleep restriction through gut microbiota. *J. Pineal Res.* **2023**, *75*, e12874. [\[CrossRef\]](#)
124. Skobowiat, C.; Brożyna, A.; Janjetovic, Z.; Jeayeng, S.; Oak, A.; Kim, T.; Panich, U.; Reiter, R.; Slominski, A. Melatonin and its derivatives counteract the ultraviolet B radiation-induced damage in human and porcine skin ex vivo. *J. Pineal Res.* **2018**, *65*, e12501. [\[CrossRef\]](#)
125. Izykowska, I.; Cegielski, M.; Gebarowska, E.; Podhorska-Okolow, M.; Piotrowska, A.; Zabel, M.; Dziegiel, P. Effect of melatonin on human keratinocytes and fibroblasts subjected to UVA and UVB radiation In Vitro. *In Vivo* **2009**, *23*, 739–745.
126. Kleszczynski, K.; Hardkop, L.; Fischer, T. Differential effects of melatonin as a broad range UV-damage preventive dermato-endocrine regulator. *Derm.-Endocrinol.* **2011**, *3*, 27–31. [\[CrossRef\]](#)

127. Cho, J.; Kim, C.; Lee, K. Modification of gene expression by melatonin in UVB-irradiated HaCaT keratinocyte cell lines using a cDNA microarray. *Oncol. Rep.* **2007**, *17*, 573–577. [\[CrossRef\]](#)
128. Kleszczynski, K.; Zillikens, D.; Fischer, T. Melatonin enhances mitochondrial ATP synthesis, reduces reactive oxygen species formation, and mediates translocation of the nuclear erythroid 2-related factor 2 resulting in activation of phase-2 antioxidant enzymes (γ -GCS, HO-1, NQO1) in ultraviolet radiation-treated normal human epidermal keratinocytes (NHEK). *J. Pineal Res.* **2016**, *61*, 187–197. [\[PubMed\]](#)
129. Fischer, T.; Zmijewski, M.; Wortsman, J.; Slominski, A. Melatonin maintains mitochondrial membrane potential and attenuates activation of initiator (casp-9) and effector caspases (casp-3/casp-7) and PARP in UVR-exposed HaCaT keratinocytes. *J. Pineal Res.* **2008**, *44*, 397–407. [\[CrossRef\]](#) [\[PubMed\]](#)
130. Fischer, T.; Sweatman, T.; Semak, I.; Sayre, R.; Wortsman, J.; Slominski, A. Constitutive and UV-induced metabolism of melatonin in keratinocytes and cell-free systems. *FASEB J.* **2006**, *20*, 897–908. [\[CrossRef\]](#) [\[PubMed\]](#)
131. Lee, K.; Lee, W.; Suh, S.; Kim, S.; Lee, S.; Ryoo, Y.; Kim, B. Melatonin reduces ultraviolet-B induced cell damage and polyamine levels in human skin fibroblasts in culture. *Exp. Mol. Med.* **2003**, *35*, 263–268. [\[CrossRef\]](#) [\[PubMed\]](#)
132. Rezzani, R.; Rodella, L.; Favero, G.; Damiani, G.; Paganelli, C.; Reiter, R. Attenuation of ultraviolet A-induced alterations in NIH3T3 dermal fibroblasts by melatonin. *Br. J. Dermatol.* **2014**, *170*, 382–391. [\[CrossRef\]](#) [\[PubMed\]](#)
133. Ryoo, Y.; Suh, S.; Mun, K.; Kim, B.; Lee, K. The effects of the melatonin on ultraviolet-B irradiated cultured dermal fibroblasts. *J. Dermatol. Sci.* **2001**, *27*, 162–169. [\[CrossRef\]](#) [\[PubMed\]](#)
134. Janjetovic, Z.; Jarrett, S.; Lee, E.; Duprey, C.; Reiter, R.; Slominski, A. Melatonin and its metabolites protect human melanocytes against UVB-induced damage: Involvement of NRF2-mediated pathways. *Sci. Rep.* **2017**, *7*, 1274. [\[CrossRef\]](#) [\[PubMed\]](#)
135. Lee, J.; Moon, J.; Nazim, U.; Lee, Y.; Seol, J.; Eo, S.; Lee, J.; Park, S. Melatonin protects skin keratinocyte from hydrogen peroxide-mediated cell death via the SIRT1 pathway. *Oncotarget* **2016**, *7*, 12075–12088. [\[CrossRef\]](#) [\[PubMed\]](#)
136. Ranieri, D.; Avitabile, D.; Shiota, M.; Yokomizo, A.; Naito, S.; Bizzarri, M.; Torrisi, M. Nuclear redox imbalance affects circadian oscillation in HaCaT keratinocytes. *Int. J. Biochem. Cell Biol.* **2015**, *65*, 113–124. [\[CrossRef\]](#)
137. Haslam, I.; Jadkauskaite, L.; Szabo, I.; Staeger, S.; Hesebeck-Brinckmann, J.; Jenkins, G.; Bhogal, R.; Lim, F.; Farjo, N.; Farjo, B.; et al. Oxidative damage control in a human (mini-) organ: Nrf2 activation protects against oxidative stress-induced hair growth inhibition. *J. Investig. Dermatol.* **2017**, *137*, 295–304. [\[CrossRef\]](#)
138. Scheuer, C. Melatonin for prevention of erythema and oxidative stress in response to ultraviolet radiation. *Dan. Med. J.* **2017**, *64*, B5358. [\[PubMed\]](#)
139. Dong, K.; Goyarts, E.; Rella, A.; Pelle, E.; Wong, Y.; Pernodet, N. Age associated decrease of MT-1 melatonin receptor in human dermal skin fibroblasts impairs protection against UV-induced DNA damage. *Int. J. Mol. Sci.* **2020**, *21*, 326. [\[CrossRef\]](#) [\[PubMed\]](#)
140. Scheuer, C.; Pommergaard, H.C.; Rosenberg, J.; Gogenur, I. Dose dependent sun protective effect of topical melatonin: A randomized, placebo-controlled, double-blind study. *J. Dermatol. Sci.* **2016**, *84*, 178–185. [\[CrossRef\]](#) [\[PubMed\]](#)
141. Kim, T.; Kleszczynski, K.; Janjetovic, Z.; Sweatman, T.; Lin, Z.; Li, W. Metabolism of melatonin and biological activity of intermediates of melatonergic pathway in human skin cells. *FASEB J.* **2013**, *27*, 2742–2755. [\[CrossRef\]](#) [\[PubMed\]](#)
142. Slominski, A.; Chassalevris, N.; Mazurkiewicz, J.; Maurer, M.; Paus, R. Murine skin as a target for melatonin bioregulation. *Exp. Dermatol.* **1994**, *3*, 44–50. [\[CrossRef\]](#) [\[PubMed\]](#)
143. Slominski, A.T.; Kim, T.K.; Slominski, R.M.; Song, Y.; Qayyum, S.; Placha, W.; Janjetovic, Z.; Kleszczynski, K.; Atigadda, V.; Song, Y.; et al. Melatonin and Its Metabolites Can Serve as Agonists on the Aryl Hydrocarbon Receptor and Peroxisome Proliferator-Activated Receptor Gamma. *Int. J. Mol. Sci.* **2023**, *24*, 15496. [\[CrossRef\]](#) [\[PubMed\]](#)
144. Aramwit, P.; Kanokpanont, S.; Nakpheng, T.; Srichana, T. The effect of sericin from various extraction methods on cell viability and collagen production. *Int. J. Mol. Sci.* **2010**, *11*, 2200–2211. [\[CrossRef\]](#)
145. Aramwit, P.; Kanokpanont, S.; De-Eknamkul, W.; Kamei, K.; Srichana, T. The effect of sericin with variable amino-acid content from different silk strains on the production of collagen and nitric oxide. *J. Biomater. Sci. Polym. Ed.* **2009**, *20*, 1295–1306. [\[CrossRef\]](#) [\[PubMed\]](#)
146. Kitisin, T.; Maneekan, P.; Luplertlop, N. In Vitro characterization of silk sericin as an anti-aging agent. *J. Agric. Sci.* **2013**, *5*, 54–62. [\[CrossRef\]](#)
147. Sangwong, G.; Sumida, M.; Sutthikhum, V. Antioxidant activity of chemically and enzymatically modified sericin extracted from cocoons of Bombyx mori. *Biocatal. Agric. Biotechnol.* **2016**, *5*, 155–161. [\[CrossRef\]](#)
148. Kanpipit, N.; Nualkaew, N.; Thapphasaraphong, S. The Potential of Purple Waxy Corn Cob (*Zea mays* L.) Extract Loaded-Sericin Hydrogel for Anti-Hyperpigmentation, UV Protection and Anti-Aging Properties as Topical Product Applications. *Pharmaceuticals* **2023**, *16*, 35. [\[CrossRef\]](#) [\[PubMed\]](#)
149. Parashar, P.; Pal, S.; Dwivedi, M.; Saraf, S. Augmented Therapeutic Efficacy of Naringenin through Microemulsion-Loaded Sericin Gel against UVB-Induced Photoaging. *AAPS PharmSciTech* **2020**, *21*, 215. [\[CrossRef\]](#) [\[PubMed\]](#)
150. Berardesca, E.; Ardigo, M.; Cameli, N.; Mariano, M.; Agozzino, M.; Matts, P. Randomized, double-blinded, vehicle-controlled, split-face study to evaluate the effects of topical application of a Gold Silk Sericin/Niacinamide/Signaline complex on biophysical parameters related to skin ageing. *Int. J. Cosmet. Sci.* **2015**, *37*, 606–612. [\[CrossRef\]](#) [\[PubMed\]](#)
151. Fisher, G.; Wang, Z.; Datta, S.; Varani, J.; Kang, S.; Voorhees, J. Pathophysiology of premature skin aging induced by ultraviolet light. *N. Engl. J. Med.* **1997**, *337*, 1419–1429. [\[CrossRef\]](#) [\[PubMed\]](#)

152. Hudson, L.; Rashdan, E.; Bonn, C.; Chavan, B.; Rawlings, D.; Birch-Machin, M. Individual and combined effects of the infrared, visible, and ultraviolet light components of solar radiation on damage biomarkers in human skin cells. *FASEB J.* **2020**, *34*, 3874–3883. [[CrossRef](#)] [[PubMed](#)]
153. Barresi, C.; Stremnitzer, C.; Mlitz, V.; Kezic, S.; Kammeyer, A.; Ghannadan, M.; Posa-Markaryan, K.; Selden, C.; Tschachler, E.; Eckhart, L. Increased sensitivity of histidinemic mice to UVB radiation suggests a crucial role of endogenous urocanic acid in photoprotection. *J. Investig. Dermatol.* **2011**, *131*, 188–194. [[CrossRef](#)]
154. Andor, N.; Graham, T.; Jansen, M.; Xia, L.; Aktipis, C.; Petritsch, C.; Ji, H.; Maley, C. Pan-cancer analysis of the extent and consequences of intratumor heterogeneity. *Nat. Med.* **2016**, *22*, 105–113. [[CrossRef](#)] [[PubMed](#)]
155. Dalle Carbonare, L.; Minoia, A.; Vareschi, A.; Piritore, F.; Zouari, S.; Gandini, A.; Meneghel, M.; Elia, R.; Lorenzi, P.; Antoniazzi, F.; et al. Exploring the Interplay of RUNX2 and CXCR4 in Melanoma Progression. *Cells* **2024**, *13*, 408. [[CrossRef](#)] [[PubMed](#)]
156. Colebatch, A.; Scolyer, R. Trajectories of premalignancy during the journey from melanocyte to melanoma. *Pathology* **2018**, *50*, 16–23. [[CrossRef](#)]
157. Elder, D.; Gimotty, P.; Guerry, D. Cutaneous melanoma: Estimating survival and recurrence risk based on histopathologic features. *Dermatol. Ther.* **2005**, *18*, 369–385. [[CrossRef](#)]
158. Baade, P.; Whiteman, D.; Janda, M.; Cust, A.; Neale, R.; Smithers, B.; Green, A.; Khosrotehrani, K.; Mar, V.; Soyer, P.; et al. Long-term deaths from melanoma according to tumor thickness at diagnosis. *Int. J. Cancer* **2020**, *147*, 1391–1396. [[CrossRef](#)]
159. Owen, C.; Shoushtari, A.; Chauhan, D.; Palmieri, D.J.; Lee, B.; Rohaan, M.; Mangana, J.; Atkinson, V.; Zaman, F.; Young, A.; et al. Management of early melanoma recurrence despite adjuvant anti-PD-1 antibody therapy. *Ann. Oncol.* **2020**, *31*, 1075–1082. [[CrossRef](#)] [[PubMed](#)]
160. Dedeilia, A.; Lwin, T.; Li, S.; Tarantino, G.; Tunsiricharoenkul, S.; Lawless, A.; Sharova, T.; Liu, D.; Boland, G.; Cohen, S. Factors Affecting Recurrence and Survival for Patients with High-Risk Stage II Melanoma. *Ann. Surg. Oncol.* **2024**, *31*, 2713–2726. [[CrossRef](#)]
161. Arnold, M.; Singh, D.; Laversanne, M.; Vignat, J.; Vaccarella, S.; Meheus, F.; Cust, A.; Vries, E.; Whiteman, D.; Bray, F. Global burden of cutaneous melanoma in 2020 and projections to 2040. *JAMA Dermatol.* **2022**, *158*, 495–503. [[CrossRef](#)] [[PubMed](#)]
162. Enomoto, L.; Levine, E.; Shen, P.; Votanopoulos, K. Role of surgery for metastatic melanoma. *Surg. Clin.* **2020**, *100*, 127–139. [[CrossRef](#)]
163. Joyce, D.; Skitzki, J. Surgical management of primary cutaneous melanoma. *Surg. Clin.* **2020**, *100*, 61–70. [[CrossRef](#)]
164. Koizumi, S.; Inozume, T.; Nakamura, Y. Current surgical management for melanoma. *J. Dermatol.* **2024**, *51*, 312–323. [[CrossRef](#)] [[PubMed](#)]
165. Patil, T.; Rohiwal, S.; Tiwari, A. Stem cells: Therapeutic implications in chemotherapy and radiotherapy resistance in cancer therapy. *Curr. Stem Cell Res. Ther.* **2023**, *18*, 750–765. [[CrossRef](#)]
166. Pires, L.; Demidov, V.; Wilson, B.; Salvio, A.; Moriyama, L.; Bagnato, V.; Vitkin, I.; Kurachi, C. Dual-agent photodynamic therapy with optical clearing eradicates pigmented melanoma in preclinical tumor models. *Cancers* **2020**, *12*, 1956. [[CrossRef](#)]
167. Mallidi, S.; Anbil, S.; Bulin, A.; Obaid, G.; Ichikawa, M.; Hasan, T. Beyond the barriers of light penetration: Strategies, perspectives and possibilities for photodynamic therapy. *Theranostics* **2016**, *6*, 2458–2487. [[CrossRef](#)]
168. Rio, A.; Mas, J.O.; Moreno, G.; Sanchez, D.; Castresana, I.; Cuxart, J. Reconstruction Using Perforator Propeller Flaps After Malignant Melanoma Resection of the Lower Extremity. *Plast. Surg.* **2022**, *32*, 276–282.
169. Wing, W. Reconstruction of head and neck melanoma defects. *Oral Maxillofac. Surg. Clin.* **2020**, *34*, 283–298.
170. Mohiuddin, J.; Chu, B.; Facciabene, A.; Poirier, K.; Wang, X.; Doucette, A.; Zheng, C.; Xu, W.; Anstadt, E.; Amaravadi, R.; et al. Association of antibiotic exposure with survival and toxicity in patients with melanoma receiving immunotherapy. *J. Nat. Cancer Inst.* **2021**, *113*, 162–170. [[CrossRef](#)] [[PubMed](#)]
171. Huynh, M.; Olaiya, O.; Kim, P.; Gallo, L.; Dunn, E.; Farrokhyar, F.; McRae, M.; Voinescos, S.; McRae, M. A comparison of skin grafts versus local flaps for facial skin cancer from the patient perspective: Protocol for a feasibility study. *Jpn. J. Clin. Oncol.* **2023**, *53*, 489–493. [[CrossRef](#)] [[PubMed](#)]
172. Mamsen, F.; Kiilerich, C.; Hesselfeldt-Nielsen, J.; Saltvig, I.; Remvig, C.; Trøstrup, H.; Schmidt, V. Risk stratification of local flaps and skin grafting in skin cancer-related facial reconstruction: A retrospective single-center study of 607 patients. *J. Pers. Med.* **2022**, *12*, 2067. [[CrossRef](#)] [[PubMed](#)]
173. Zhang, X.; Sun, Y.; Hou, Z.; Luo, B.; Li, C.; Jiang, K.; Liu, J.; Yao, G.; Tang, J. Application of dermal regenerative template in reconstructing skin defects after plantar malignant melanoma excision. *J. BUON* **2021**, *26*, 2146–2153.
174. Heo, J.; Jeon, E.; Joo, K.; Cha, H. Locoregional melanoma therapy by tissue adhesive microneedle patch-assisted trans-tumoral delivery of anticancer drug. *Biotechnol. Bioprocess Eng.* **2023**, *28*, 473–482. [[CrossRef](#)]
175. Xu, Q.; Wang, Y.; Chen, T.; Lao, C.; Gao, H.; Wei, R.; Feng, B.; Zhi, W.; Weng, J.; Wang, J. A distinctive nanocomposite hydrogel integrated platform for the healing of wound after the resection of melanoma. *Materialia* **2020**, *14*, 100931. [[CrossRef](#)]
176. Du, S.; Suo, H.; Xie, G.; Lyu, Q.; Mo, M.; Xie, Z.; Zhou, N.; Zhang, L.; Tao, J.; Zhu, J. Self-powered and photothermal electronic skin patches for accelerating wound healing. *Nano Energy* **2022**, *93*, 106906. [[CrossRef](#)]
177. Andreassi, L. UV exposure as a risk factor for skin cancer. *Expert Rev. Dermatol.* **2011**, *6*, 445–454. [[CrossRef](#)]
178. Cho, E.; Rosner, B.; Colditz, G. Risk factors for melanoma by body site. *Cancer Epidemiol. Biomarkers Prev.* **2005**, *14*, 1241–1244. [[CrossRef](#)]

179. Bauer, J.; Garbe, C. Acquired melanocytic nevi as risk factor for melanoma development. A comprehensive review of epidemiological data. *Pigment Cell Res.* **2003**, *16*, 297–306. [\[CrossRef\]](#) [\[PubMed\]](#)
180. Gutiérrez-Castañeda, L.; Nova, J.; Tovar-Parra, J. Frequency of mutations in BRAF, NRAS, and KIT in different populations and histological subtypes of melanoma: A systemic review. *Melanoma Res.* **2020**, *30*, 62–70. [\[CrossRef\]](#) [\[PubMed\]](#)
181. Potrony, M.; Puig-Butillé, J.; Aguilera, P.; Badenas, C.; Carrera, C.; Malvey, J.; Puig, S. Increased prevalence of lung, breast, and pancreatic cancers in addition to melanoma risk in families bearing the cyclin-dependent kinase inhibitor 2A mutation: Implications for genetic counseling. *J. Am. Acad. Dermatol.* **2014**, *71*, 888–895. [\[CrossRef\]](#)
182. Abdel-Rahman, M.; Pilarski, R.; Massengill, J.; Christopher, B.; Noss, R.; Davidorf, F. Melanoma candidate genes CDKN2A/p16/INK4A, p14ARF, and CDK4 sequencing in patients with uveal melanoma with relative high-risk for hereditary cancer predisposition. *Melanoma Res.* **2011**, *21*, 175–179. [\[CrossRef\]](#)
183. Roesch, A.; Volkenandt, M. *Dermatology*, 3rd ed.; Springer: Berlin, Germany, 2009; pp. 1416–1432.
184. Li, B.; Smith, C.; Laing, J.; Gober, M.; Liu, L.; Aurelian, L. Overload of the heat-shock protein H11/HspB8 triggers melanoma cell apoptosis through activation of transforming growth factor- β -activated kinase 1. *Oncogene* **2007**, *26*, 3521–3531. [\[CrossRef\]](#)
185. Knudsen, S.; Schardt, A.; Buhl, T.; Boeckmann, L.; Schön, M.; Neumann, C.; Haenssle, H. Enhanced T-cell activation by immature dendritic cells loaded with HSP70-expressing heat-killed melanoma cells. *Exp. Dermatol.* **2010**, *19*, 108–116. [\[CrossRef\]](#)
186. Park, K.; Kim, D.; Choi, H.; Kim, K.; Chung, J.; Eun, H.; Lee, S.; Seo, J.S. Overexpression of HSP70 prevents ultraviolet B-induced apoptosis of a human melanoma cell line. *Arch. Dermatol. Res.* **2000**, *292*, 482–487. [\[CrossRef\]](#) [\[PubMed\]](#)
187. Russo, A.; Cardile, V.; Caggia, S.; Gunther, G.; Troncoso, N.; Garbarino, J. Boldo prevents UV light and nitric oxide-mediated plasmid DNA damage and reduces the expression of Hsp70 protein in melanoma cancer cells. *J. Pharm. Pharmacol.* **2011**, *63*, 1219–1229. [\[CrossRef\]](#)
188. Roh, B.; Kim, D.; Cho, M.; Park, Y.; Whang, K. Expression of heat shock protein 70 in human skin cells as a photoprotective function after UV exposure. *Ann. Dermatol.* **2008**, *20*, 184. [\[CrossRef\]](#)
189. Lanneau, D.; Brunet, M.; Frisan, E.; Solary, E.; Fontenay, M.; Garrido, C. Heat shock proteins: Essential proteins for apoptosis regulation. *J. Cell. Mol. Med.* **2008**, *12*, 743–761. [\[CrossRef\]](#) [\[PubMed\]](#)
190. Assefa, Z.; Van Laethem, A.; Garmyn, M.; Agostinis, P. Ultraviolet radiation-induced apoptosis in keratinocytes: On the role of cytosolic factors. *Biochim. Biophys. Acta Rev. Cancer* **2005**, *1755*, 90–106. [\[CrossRef\]](#) [\[PubMed\]](#)
191. Strozyk, E.; Kulms, D. The role of AKT/mTOR pathway in stress response to UV-irradiation: Implication in skin carcinogenesis by regulation of apoptosis, autophagy and senescence. *Int. J. Mol. Sci.* **2013**, *14*, 15260–15285. [\[CrossRef\]](#) [\[PubMed\]](#)
192. Cabrera, J.; Negrín, G.; Estévez, F.; Loro, J.; Reiter, R.; Quintana, J. Melatonin decreases cell proliferation and induces melanogenesis in human melanoma SK-MEL-1 cells. *J. Pineal Res.* **2010**, *49*, 45–54. [\[CrossRef\]](#) [\[PubMed\]](#)
193. Brożyna, A.; Jóźwicki, W.; Roszkowski, K.; Filipiak, J.; Slominski, A. Melanin content in melanoma metastases affects the outcome of radiotherapy. *Oncotarget* **2016**, *7*, 17844–17853. [\[CrossRef\]](#) [\[PubMed\]](#)
194. Roberts, J.; Wiechmann, A.; Hu, D. Melatonin receptors in human uveal melanocytes and melanoma cells. *J. Pineal Res.* **2000**, *28*, 165–171. [\[CrossRef\]](#) [\[PubMed\]](#)
195. Souza, A.; Visconti, M.; Castrucci, A. Melatonin biological activity and binding sites in human melanoma cells. *J. Pineal Res.* **2003**, *34*, 242–248. [\[CrossRef\]](#) [\[PubMed\]](#)
196. Pfeffer, M.; von Gall, C.; Wicht, H.; Korf, H.W. The Role of the Melatonergic System in Circadian and Seasonal Rhythms-Insights From Different Mouse Strains. *Front. Physiol.* **2022**, *13*, 883637. [\[CrossRef\]](#) [\[PubMed\]](#)
197. Gautier, C.; Theret, I.; Lizzo, G.; Ferry, G.; Guénin, S.P.; Boutin, J.A. Why Are We Still Cloning Melatonin Receptors? A Commentary. *Methods Mol. Biol.* **2022**, *2550*, 267–281.
198. Song, Y.; Wang, S. Melatonin synergistically enhances docetaxel induced endoplasmic reticulum stress to promote apoptosis by suppressing NF- κ B activation in cervical cancer. *Med. Oncol.* **2023**, *40*, 219. [\[CrossRef\]](#)
199. Xiong, Y.; Ma, C.; Li, Q.; Zhang, W.; Zhao, H.; Ren, P.; Zhang, K.; Lei, X. Melatonin ameliorates simulated-microgravity-induced mitochondrial dysfunction and lipid metabolism dysregulation in hepatocytes. *FASEB J.* **2023**, *37*, 14947. [\[CrossRef\]](#) [\[PubMed\]](#)
200. Slominski, R.M.; Sarna, T.; Płonka, P.M.; Raman, C.; Brożyna, A.A.; Slominski, A.T. Melanoma, Melanin, and Melanogenesis: The Yin and Yang Relationship. *Front. Oncol.* **2022**, *12*, 842496. [\[CrossRef\]](#) [\[PubMed\]](#)
201. Möller, J.; Linowiecka, K.; Gagat, M.; Brożyna, A.; Foksiński, M.; Wolnicka-Glubisz, A.; Pyza, E.; Reiter, R.; Tulic, M.; Slominski, A.; et al. Melanogenesis Is Directly Affected by Metabolites of Melatonin in Human Melanoma Cells. *Int. J. Mol. Sci.* **2023**, *24*, 14947. [\[CrossRef\]](#)
202. Stefan, J.; Kim, T.K.; Schedel, F.; Janjetovic, Z.; Crossman, D.K.; Steinbrink, K.; Slominski, R.M.; Zmijewski, J.; Tulic, M.; Reiter, R.; et al. Differential and overlapping effects of melatonin and its metabolites on keratinocyte function: Bioinformatics and metabolic analyses. *Antioxidants* **2021**, *10*, 618. [\[CrossRef\]](#) [\[PubMed\]](#)
203. Slominski, R.M.; Raman, C.; Chen, J.Y.; Slominski, A.T. How cancer hijacks the body's homeostasis through the neuroendocrine system. *Trends Neurosci.* **2023**, *46*, 263–275. [\[CrossRef\]](#) [\[PubMed\]](#)
204. Lerner, A.; Case, J.; Takahashi, Y.; Lee, T.; Mori, W. Isolation of melatonin, the pineal gland factor that lightens melanocytes. *J. Am. Chem. Soc.* **1958**, *80*, 2587. [\[CrossRef\]](#)
205. Tan, D.X.; Manchester, L.; Reiter, R.; Plummer, B.; Limson, J.; Weintraub, S.; Qi, W. Melatonin directly scavenges hydrogen peroxide: A potentially new metabolic pathway of melatonin biotransformation. *Free Radic. Biol. Med.* **2000**, *29*, 1177–1185. [\[CrossRef\]](#) [\[PubMed\]](#)

206. Galano, A. On the direct scavenging activity of melatonin towards hydroxyl and a series of peroxy radicals. *Phys. Chem. Chem. Phys.* **2011**, *13*, 7178–7188. [[CrossRef](#)] [[PubMed](#)]
207. Horstman, J.; Wrona, M.; Dryhurst, G. Further insights into the reaction of melatonin with hydroxyl radical. *Bioorg. Chem.* **2002**, *30*, 371–382. [[CrossRef](#)]
208. Schaefer, M.; Hardeland, R. The melatonin metabolite N^1 -acetyl-5-methoxykynuramine is a potent singlet oxygen scavenger. *J. Pineal Res.* **2009**, *46*, 49–52. [[CrossRef](#)]
209. Harasimowicz, J.; Marques, K.; Silva, A.; Costa, R.; Prior, J.; Rodrigues, S.; Santos, J. Chemiluminometric evaluation of melatonin and selected melatonin precursors' interaction with reactive oxygen and nitrogen species. *Anal. Biochem.* **2012**, *420*, 1–6. [[CrossRef](#)] [[PubMed](#)]
210. Zhang, H.; Squadrito, G.; Uppu, R.; Pryor, W. Reaction of peroxynitrite with melatonin: A mechanistic study. *Chem. Res. Toxicol.* **1999**, *12*, 526–534. [[CrossRef](#)]
211. Noda, Y.; Mori, A.; Liburdy, R.; Packer, L. Melatonin and its precursors scavenge nitric oxide. *J. Pineal Res.* **1999**, *27*, 159–163. [[CrossRef](#)]
212. Miguel, R.; Martínez, A. A new free radical scavenging cascade involving melatonin and three of its metabolites (3OHM, AFMK and AMK). *Comput. Theor. Chem.* **2018**, *1123*, 111–118.
213. Mingzhuang, H.; Zhang, Y.; Liu, Y.; Ge, X.; Hu, X.; Zhao, Z.; Tian, X.; Liu, T.; Yang, H.; Chen, X.; et al. Biomimetic melatonin-loaded silk fibroin/GelMA scaffold strengthens cartilage repair through retrieval of mitochondrial functions. *J. Mater. Sci. Technol.* **2023**, *146*, 102–112.
214. Azizoglu, G.; Azizoglu, E.; Barker, T.; Özer, Ö. Single and multi-dose drug loaded electrospun fiber mats for wound healing applications. *J. Drug Deliv. Sci. Technol.* **2023**, *81*, 104168. [[CrossRef](#)]
215. Yamei, W.; Xiao, D.; Tang, Y.; Xia, Y.; Zhong, Y.; Zhang, L.; Sui, X.; Wang, B.; Feng, X.; Xu, H.; et al. Carboxymethyl cellulose-based injectable hydrogel loaded with a composite of melatonin and γ -cyclodextrin with antioxidant property for diabetic wound repair. *Cellulose* **2023**, *30*, 1791–1810.
216. Atila, D.; Keskin, D.; Lee, Y.; Lin, F.; Hasirci, V.; Tezcaner, A. Injectable methacrylated gelatin/thiolated pectin hydrogels carrying melatonin/tideglusib-loaded core/shell PMMA/silk fibroin electrospun fibers for vital pulp regeneration. *Colloids Surf. B* **2023**, *222*, 113078. [[CrossRef](#)]
217. Păncescu, F.; Rikabi, A.; Oprea, O.; Grosu, A.; Nechifor, A.; Grosu, V.A.; Tanczos, S.Z.; Dumitru, F.; Nechifor, G.; Bungău, S. Chitosan–sEPDM and Melatonin–Chitosan–sEPDM Composite Membranes for Melatonin Transport and Release. *Membranes* **2023**, *13*, 282. [[CrossRef](#)]
218. Borrego-Sánchez, A.; Muñoz-Santiburcio, D.; Viseras, C.; Hernández-Laguna, A.; Sainz-Díaz, I. Melatonin/nanoclay hybrids for skin delivery. *Appl. Clay Sci.* **2022**, *218*, 106417. [[CrossRef](#)]
219. Yao, Z.; Qian, Y.; Jin, Y.; Wang, S.; Li, J.; Yuan, W.E.; Fan, C. Biomimetic multilayer polycaprolactone/sodium alginate hydrogel scaffolds loaded with melatonin facilitate tendon regeneration. *Carbohydr. Polym.* **2022**, *277*, 118865. [[CrossRef](#)]
220. Tingkuo, C.; Jiang, H.; Li, X.; Zhang, D.; Zhu, Y.; Chen, X.; Yang, H. Proliferation and differentiation study of melatonin functionalized polycaprolactone/gelatin electrospun fibrous scaffolds for nerve tissue engineering. *Int. J. Biol. Macromol.* **2022**, *197*, 103–110.
221. Kaczmarek-Szczepańska, B.; Pin, J.; Zasada, L.; Sonne, M.; Reiter, R.; Słomiński, A.; Steinbrink, K.; Kleszczyński, K. Assessment of melatonin-cultured collagen/chitosan scaffolds cross-linked by a glyoxal solution as biomaterials for wound healing. *Antioxidants* **2022**, *11*, 570. [[CrossRef](#)]
222. Nongmaithem, C.; Bhattacharya, K.; Marbaniang, D.; Pal, P.; Ray, S.; Mazumder, B. Evaluation of a novel melatonin-loaded gelatin sponge as a wound dressing. *J. Vasc. Nur.* **2022**, *40*, 2–10.
223. Chen, K.; Tong, C.; Yang, J.; Cong, P.; Liu, Y.; Shi, X.; Liu, X.; Zhang, J.; Zou, R.; Xiao, K.; et al. Injectable melatonin-loaded carboxymethyl chitosan (CMCS)-based hydrogel accelerates wound healing by reducing inflammation and promoting angiogenesis and collagen deposition. *J. Mater. Sci. Technol.* **2021**, *63*, 236–245. [[CrossRef](#)]
224. Kaczmarek-Szczepańska, B.; Ostrowska, J.; Kozłowska, J.; Szota, Z.; Brożyna, A.; Dreier, R.; Reiter, R.; Słomiński, A.; Steinbrink, K.; Kleszczyński, K. Evaluation of polymeric matrix loaded with melatonin for wound dressing. *Int. J. Mol. Sci.* **2021**, *22*, 5658. [[CrossRef](#)]
225. Mirmajidi, T.; Chogan, F.; Rezayan, A.; Sharifi, A. In Vitro and In Vivo evaluation of a nanofiber wound dressing loaded with melatonin. *Int. J. Pharm.* **2021**, *596*, 120213. [[CrossRef](#)]
226. Weilin, Z.; Zhao, W.; Li, Q.; Zhao, D.; Qu, J.; Yuan, Z.; Cheng, Z.; Zhu, X.; Zhuang, X.; Zhang, Z. 3D-printing magnesium-polycaprolactone loaded with melatonin inhibits the development of osteosarcoma by regulating cell-in-cell structures. *J. Nanobiotechnol.* **2021**, *19*, 263.
227. Ragothaman, M.; Villalan, A.; Dhanasekaran, A.; Palanisamy, T. Bio-hybrid hydrogel comprising collagen-capped silver nanoparticles and melatonin for accelerated tissue regeneration in skin defects. *Mater. Sci. Eng. C* **2021**, *128*, 112328. [[CrossRef](#)]
228. Sinohara, H. Glycopeptides isolated from sericin of the silkworm, *Bombyx mori*. *Comp. Biochem. Physiol. B Biochem. Comp. Biochem.* **1979**, *63*, 87–91. [[CrossRef](#)]
229. Hoyoung, L.; Ahn, D.; Jeon, E.; Fam, D.; Lee, J.; Lee, W. Macroscopic assembly of sericin toward self-healable silk. *Biomacromolecules* **2021**, *22*, 4337–4346.

230. Pornanong, A.; Siritientong, T.; Kanokpanont, S.; Srichana, T. Formulation and characterization of silk sericin–PVA scaffold crosslinked with genipin. *Int. J. Biol. Macromol.* **2010**, *47*, 668–675.
231. Kunz, R.; Brancalhão, R.; Ribeiro, L.; Natali, M. Silkworm sericin: Properties and biomedical applications. *BioMed Res. Int.* **2016**, *2*, 1–19. [\[CrossRef\]](#)
232. Rupesh, D.; Mandal, M.; Ghosh, S.; Kundu, S. Silk sericin protein of tropical tasar silkworm inhibits UVB-induced apoptosis in human skin keratinocytes. *Mol. Cell. Biochem.* **2008**, *311*, 111–119.
233. Praveen, K.; Mandal, B. Silk sericin induced pro-oxidative stress leads to apoptosis in human cancer cells. *Food Chem. Toxicol.* **2019**, *123*, 275–287.
234. Dash, R.; Ghosh, S.; Kaplan, D.; Kundu, S. Purification and biochemical characterization of a 70 kDa sericin from tropical tasar silkworm, *Antheraea mylitta*. *Comp. Biochem. Physiol. B Biochem. Mol. Biol.* **2007**, *147*, 129–134. [\[CrossRef\]](#)
235. Kato, N.; Sato, S.; Yamanaka, A.; Yamada, H.; Fuwa, N.; Nomura, M. Silk Protein, Sericin, Inhibits Lipid Peroxidation and Tyrosinase Activity. *Biosci. Biotechnol. Biochem.* **1998**, *62*, 145–147. [\[CrossRef\]](#)
236. Bakadia, B.; Lamboni, L.; Ahmed, A.; Zheng, R.; Boni, B.; Shi, Z.; Song, S.; Souho, T.; Mukole, B.; Qi, F.; et al. Antibacterial silk sericin/poly (vinyl alcohol) hydrogel with antifungal property for potential infected large burn wound healing: Systemic evaluation. *Smart Mater. Med.* **2023**, *4*, 37–58. [\[CrossRef\]](#)
237. Fariha, M.; Tahir, H.; Ali, S.; Ali, A.; Tehreem, A.; Durr, S.; Zaidi, S.; Adnan, M.; Ijaz, F. Characterization and Evaluation of Silk Sericin-Based Hydrogel: A Promising Biomaterial for Efficient Healing of Acute Wounds. *ACS Omega* **2023**, *8*, 32090–32098.
238. Jayavardhini, B.; Dharmalingam, S.; Sathiyaraj, W.; Rajendran, S.; Rymbai, S.; Senthil, R.; Atchudan, R. Sericin/Human Placenta-Derived Extracellular Matrix Scaffolds for Cutaneous Wound Treatment—Preparation, Characterization, In Vitro and In Vivo Analyses. *Pharmaceutics* **2023**, *15*, 362. [\[CrossRef\]](#)
239. Li, Y.; Wang, S.; Li, Y.; Zhang, G.; Wu, T.; Wei, Y.; Cao, X.; Yan, H.; Liang, P.; Yan, Z.; et al. Resveratrol loaded native silk fiber-sericin hydrogel double interpenetrating bioactive wound dressing facilitates full-thickness skin wound healing. *Biomed. Mater.* **2023**, *18*, 045007. [\[CrossRef\]](#)
240. Bakhsheshi-Rad, H.; Ismail, A.; Aziz, M.; Akbari, M.; Hadisi, Z.; Omid, M.; Chen, X. Development of the PVA/CS nanofibers containing silk protein sericin as a wound dressing: In Vitro and In Vivo assessment. *Int. J. Biomol. Macromol.* **2020**, *149*, 513–521. [\[CrossRef\]](#)
241. Ekasurya, W.; Joses, S.; Dita, P.; Asri, P.; Asri, L. Synthesis and Degradation Properties of Sericin/PVA Hydrogels. *Gels* **2023**, *9*, 76. [\[CrossRef\]](#)
242. Jingwen, L.; Cui, T.; Xu, X.; Du, Y.; Wang, L.; Chen, S.; Pang, J. Robust Alcohol Soluble Polyurethane/Chitosan/Silk Sericin (APU/CS/SS) Nanofiber Scaffolds Toward Artificial Skin Extracellular Matrices via Microfluidic Blow-Spinning. *Adv. Fiber Mater.* **2023**, *5*, 349–361.
243. Piyachat, C.; Pengsuk, C.; Lirdprapamongkol, K.; Thanyacharoen, T.; Techasakul, S.; Svasti, J.; Nooeaid, P. Turmeric Herb Extract-Incorporated Biopolymer Dressings with Beneficial Antibacterial, Antioxidant and Anti-Inflammatory Properties for Wound Healing. *Polymers* **2023**, *15*, 1090. [\[CrossRef\]](#)
244. Yusu, W.; Li, H.; Xu, L.; Yan, J.; Wang, X. Preparation and properties of temperature-sensitive silver-loaded antibacterial sericin/poly (N-isopropylacrylamide) hydrogel. *J. Macromol. Sci. B* **2023**, 1–15.
245. Wang, K.; Hazra, R.; Ma, Q.; Khan, M.; Hoque, A.; Jiang, L.; Quadir, M.; Zhang, Y.; Wang, S.; Han, G. Robust biocompatible bacterial cellulose/silk nonwoven fabric/silk sericin sandwich membrane with strong UV-blocking and antioxidant properties. *Cellulose* **2023**, *30*, 3973–3993. [\[CrossRef\]](#)
246. Griffanti, G.; McKee, M.; Nazhat, S. Mineralization of Bone Extracellular Matrix-like Scaffolds Fabricated as Silk Sericin-Functionalized Dense Collagen–Fibrin Hybrid Hydrogels. *Pharmaceutics* **2023**, *15*, 1087. [\[CrossRef\]](#)
247. Nantaprapa, T.; Sonjan, S.; Promkrainit, S.; Daengmankhong, J.; Phimmuan, P.; Mahasaranon, S.; Jongjitwimol, J.; Charoensit, P.; Ross, G.; Viennet, C.; et al. Porous Poly (2-hydroxyethyl methacrylate) Hydrogel Scaffolds for Tissue Engineering: Influence of Crosslinking Systems and Silk Sericin Concentration on Scaffold Properties. *Polymers* **2023**, *15*, 4052. [\[CrossRef\]](#)
248. Boni, B.; Lamboni, L.; Mao, L.; Bakadia, B.; Shi, Z.; Yang, G. In Vivo performance of microstructured bacterial cellulose-silk sericin wound dressing: Effects on fibrosis and scar formation. *Eng. Sci.* **2022**, *19*, 175–185. [\[CrossRef\]](#)
249. El-Samad, L.; Hassan, M.; Basha, A.; El-Ashram, S.; Radwan, E.; Aziz, K.; Tamer, T.; Augustyniak, M.; El Wakil, A. Carboxymethyl cellulose/sericin-based hydrogels with intrinsic antibacterial, antioxidant, and anti-inflammatory properties promote re-epithelization of diabetic wounds in rats. *Int. J. Pharm.* **2022**, *629*, 122328. [\[CrossRef\]](#) [\[PubMed\]](#)
250. Moise, B.; Boni, B.; Ahmed, A.; Zheng, R.; Shi, Z.; Ullah, M.; Lamboni, L.; Yang, G. In Situ synthesized porous bacterial cellulose/poly (vinyl alcohol)-based silk sericin and azithromycin release system for treating chronic wound biofilm. *Macromol. Biosci.* **2022**, *22*, 2200201.
251. Konstantina, C.; Karavasili, C.; Adamoudi, E.; Bouropoulos, N.; Tzetzis, D.; Bakopoulou, A.; Fatouros, D. Silk sericin/PLGA electrospun scaffolds with anti-inflammatory drug-eluting properties for periodontal tissue engineering. *Biomater. Adv.* **2022**, *133*, 112723.
252. Apirujee, P.; Reddy, N.; Aramwit, P. Enhancing clinical applications of PVA hydrogel by blending with collagen hydrolysate and silk sericin. *J. Polym. Res.* **2022**, *29*, 110.
253. Karthick, S.; Manjari, K.; Devi, M. Biocompatible and bioactive PVA/Sericin/Chitosan nanofibrous wound dressing matrix. *Appl. Surf. Sci. Adv.* **2023**, *13*, 100362. [\[CrossRef\]](#)

254. Barnthip, N.; Teeka, J.; Kantha, P.; Teepoo, S.; Damjuti, W. Fabrication and characterization of polycaprolactone/cellulose acetate blended nanofiber mats containing sericin and fibroin for biomedical application. *Sci. Rep.* **2022**, *12*, 22370. [\[CrossRef\]](#)
255. Gök, Z.; Yiğitoğlu, M.; Vargel, İ.; Şahin, Y.; Alçıgır, M. Synthesis, characterization and wound healing ability of PET based nanofiber dressing material coated with silk sericin capped-silver nanoparticles. *Mater. Chem. Phys.* **2021**, *259*, 124043. [\[CrossRef\]](#)
256. Zhang, M.; Wang, D.; Ji, N.; Lee, S.; Wang, G.; Zheng, Y.; Zhang, X.; Yang, L.; Qin, Z.; Yang, Y. Bioinspired design of sericin/chitosan/Ag@ MOF/GO hydrogels for efficiently combating resistant bacteria, rapid hemostasis, and wound healing. *Polymers* **2021**, *13*, 2812. [\[CrossRef\]](#)
257. Akolpoğlu, B.; Gündüz, U.; Tezcaner, A.; Keskin, D. Topical delivery of heparin from PLGA nanoparticles entrapped in nanofibers of sericin/gelatin scaffolds for wound healing. *Int. J. Pharm.* **2021**, *597*, 120207. [\[CrossRef\]](#)
258. Lin, N.; Zuo, B. Silk sericin/fibroin electrospinning dressings: A method for preparing a dressing material with high moisture vapor transmission rate. *J. Biomater. Sci. Polym. Ed.* **2021**, *32*, 1983–1997. [\[CrossRef\]](#)
259. İnal, M.; Gün Gök, Z.; Kartal, E.; Verim, N.; Murat, S.; Apaydın, T.; Yiğitoğlu, M. The Fabrication of Poly (Σ -caprolactone)–Poly (ethylene oxide) Sandwich Type Nanofibers Containing Sericin-Capped Silver Nanoparticles as an Antibacterial Wound Dressing. *J. Nanosci. Nanotechnol.* **2021**, *21*, 3041–3049. [\[CrossRef\]](#) [\[PubMed\]](#)
260. Arango, M.; Osorio, Y.; Osorno, J.; Parra, S.; Alvarez-López, C. Effect of Ethanol Post-Treatments over Sericin Scaffolds for Tissue Engineering Applications. *J. Polym. Environ.* **2023**, *31*, 1800–1811. [\[CrossRef\]](#)
261. Baptista-Silva, S.; Borges, S.; Costa-Pinto, A.; Costa, R.; Amorim, M.; Dias, J.; Ramos, O.; Alves, P.; Granja, P.; Soares, R.; et al. In Situ forming silk sericin-based hydrogel: A novel wound healing biomaterial. *ACS Biomater. Sci. Eng.* **2021**, *7*, 1573–1586. [\[CrossRef\]](#) [\[PubMed\]](#)
262. Kukula-Koch, W.; Sz wajgier, D.; Gawel-Beben, K.; Strzepak-Gomółka, M.; Głowniak, K.; Meissner, H. Is Phytomelatonin Complex Better than Synthetic Melatonin? The Assessment of the Antiradical and Anti-Inflammatory Properties. *Molecules* **2021**, *26*, 6087. [\[CrossRef\]](#)
263. Joyjamras, K.; Chaotham, C.; Chanvorachote, P. Response surface optimization of enzymatic hydrolysis and ROS scavenging activity of silk sericin hydrolysates. *Pharm. Biol.* **2022**, *60*, 308–318. [\[CrossRef\]](#) [\[PubMed\]](#)
264. Intagliata, S.; Spadaro, A.; Lorenti, M.; Panico, A.; Siciliano, E.; Barbagallo, S.; Macaluso, B.; Kamble, S.; Modica, M.; Montenegro, L. In Vitro antioxidant and anti-glycation activity of resveratrol and its novel triester with trolox. *Antioxidants* **2020**, *10*, 12. [\[CrossRef\]](#) [\[PubMed\]](#)
265. Dushman, T.; Volpato de Oliveira, T.; Giacobbo de Marco, I.; Palioto, G.; Dushman, E. Bioactive compounds and antioxidant, antimicrobial and cytotoxic activities of extracts of *Curcuma longa*. *J. Food Meas. Charact.* **2021**, *15*, 3752–3760.
266. Slominski, A.; Chaiprasongsuk, A.; Janjetovic, Z.; Kim, T.; Stefan, J.; Slominski, R.; Hanumanthu, V.; Raman, C.; Qayyum, S.; Song, Y.; et al. Photoprotective Properties of Vitamin D and Lumisterol Hydroxyderivatives. *Cell Biochem. Biophys.* **2020**, *78*, 165–180. [\[CrossRef\]](#)
267. Chaiprasongsuk, A.; Janjetovic, Z.; Kim, T.; Tuckey, R.; Li, W.; Raman, C.; Panich, U.; Slominski, A. CYP11A1-derived vitamin D₃ products protect against UVB-induced inflammation and promote keratinocytes differentiation. *Free Radic. Biol. Med.* **2020**, *1*, 87–98. [\[CrossRef\]](#)
268. Chaiprasongsuk, A.; Janjetovic, Z.; Kim, T.; Jarrett, S.; D’Orazio, J.; Holick, M.; Tang, E.; Tuckey, R.; Panich, U.; Li, W.; et al. Protective effects of novel derivatives of vitamin D₃ and lumisterol against UVB-induced damage in human keratinocytes involve activation of Nrf2 and p53 defense mechanisms. *Redox Biol.* **2019**, *24*, 101206. [\[CrossRef\]](#)
269. Moreno, A.; Freitas Saito, R.; Tiago, M.; Massaro, R.; Pagni, R.; Pegoraro, R.; Cruz Souza, P.; Reiter, R.; Campa, A.; Soengas, M.; et al. Melatonin inhibits human melanoma cells proliferation and invasion via cell cycle arrest and cytoskeleton remodeling. *Melatonin Res.* **2020**, *3*, 194–209. [\[CrossRef\]](#)
270. Joyjamras, K.; Netcharoensirisuk, P.; Roytrakul, S.; Chanvorachote, P.; Chaotham, C. Recycled Sericin Hydrolysates Modified by Alcalase® Suppress Melanogenesis in Human Melanin-Producing Cells via Modulating MITF. *Int. J. Mol. Sci.* **2022**, *23*, 3925. [\[CrossRef\]](#) [\[PubMed\]](#)
271. Bisevac, J.; Djukic, M.; Stanojevic, I.; Stevanovic, I.; Mijuskovic, Z.; Djuric, A.; Gobeljic, B.; Banovic, T.; Vojvodic, D. Association between oxidative stress and melanoma progression. *J. Med. Biochem.* **2018**, *37*, 12–20. [\[CrossRef\]](#) [\[PubMed\]](#)

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