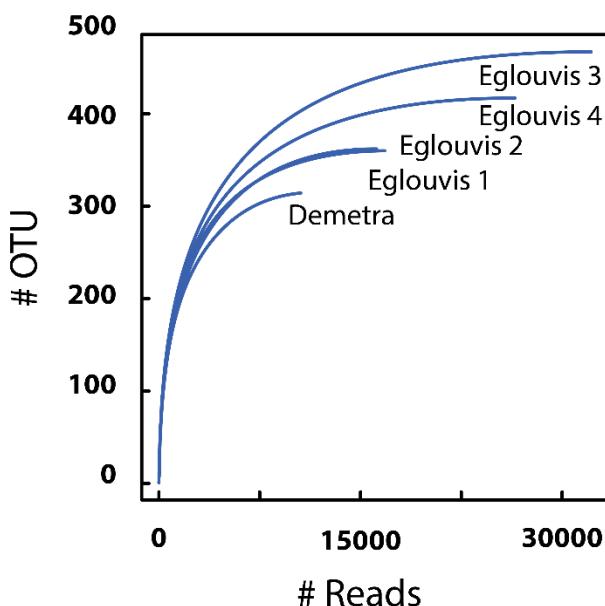
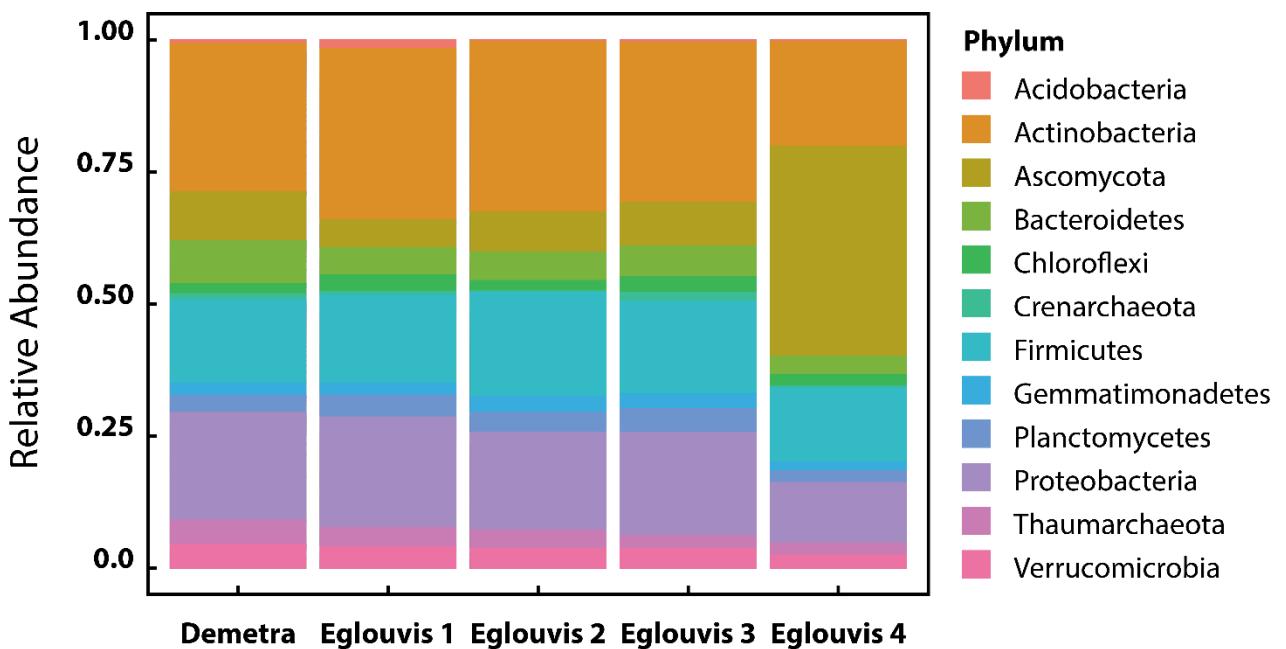


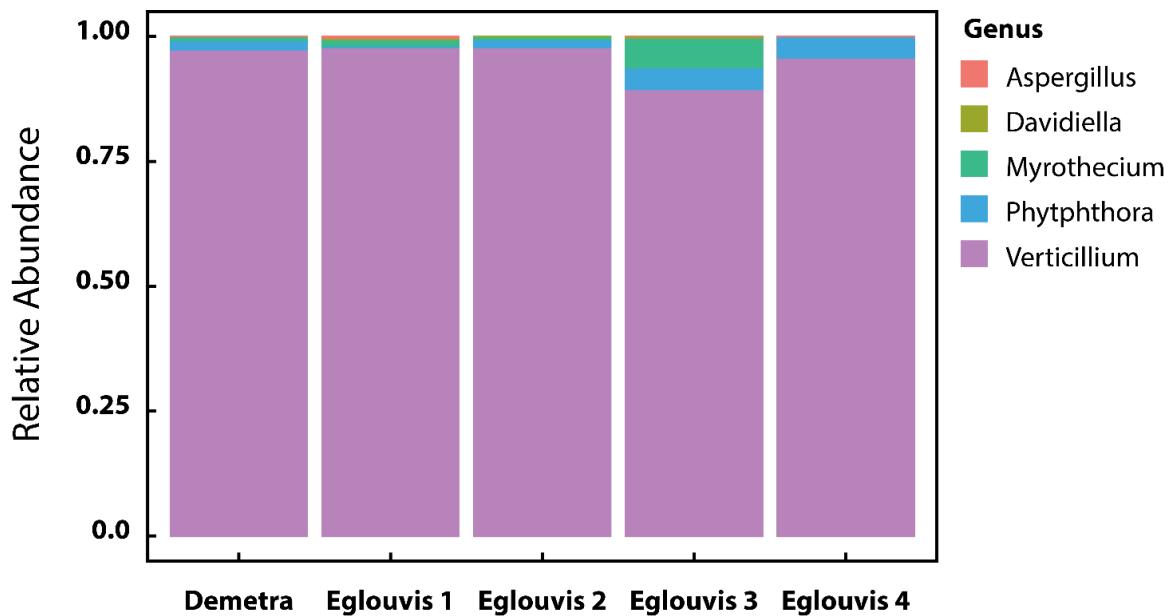
## Supplementary Figures and Tables



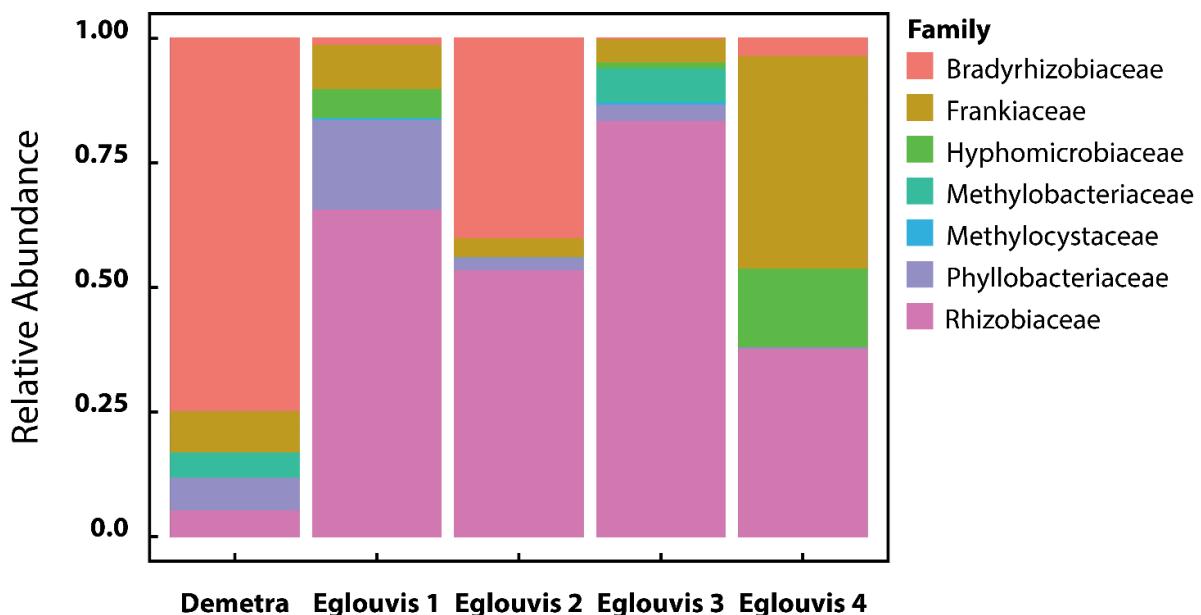
**Figure S1.** Rarefaction curves of lentil samples.



**Figure S2:** Relative abundances of lentil samples for the 12 most abundant phyla including pathogen *Kabatiella microstictica*.



**Figure S3.** Relative abundances of lentil samples for the pathogens at genus level.



**Figure S4.** Relative abundances of lentil samples for N<sub>2</sub>-fixation bacteria at family level.

**Table S1.** Amplicon primers, PCR reagents, and PCR cycling program.

	PCR for 16S amplicons	PCR for ITS2 amplicons
<b>Amplicon primers</b>		
Forward primer	515F 5'-GTGCCAGCMGCCGCGTAA- 3'	ITS4 5'-TCCTCCGCTTATTGATATGC-3'
Reverse primer	806R 5'-GGACTACHVHHHTWTCTAAT-3'	ITS9 5'-AACGCAGCRAAIIGYGA- 3'
<b>PCR reagents' recipe</b>		
gDNA template (10 ng·μL)	3 μL	3 μL
Forward primer (10 μM)	1 μL	1 μL
Reverse primer (10 μM)	1 μL	1 μL
2x Platinum II		
Hot-Start Master Mix	25 μL	25 μL
Molecular water	10 μL	10 μL
1x Platinum™ GC Enhancer	10 μL	10 μL
Total volume	50 μL	50 μL
<b>PCR cycling conditions</b>		
Initial denaturation	94°C for 120 s	94°C for 120 s
Denaturation	94 °C for 15 s	94°C for 15 s
Annealing	52°C for 15 s	60°C for 15 s
Extension	68°C for 15 s	68°C for 15 s
Cycles	35	35
Final Hold	4 °C	4 °C

**Table S2.** Metrics of  $\alpha$  diversity among the studied genotypes; Eglouvis landrace (Eglouvis 1–4) and the commercial cultivar Demetra.

Sample	Observed	Chao1	ACE	Shannon	Simpson	InvSimpson	Fisher
Eglouvis 1	360	360	360	4.490	0.978	46.1	64.6
Eglouvis 2	362	362	362	4.453	0.977	43.5	65.7
Eglouvis 3	467	467	467	4.590	0.979	48.4	77.4
Eglouvis 4	417	417	417	4.492	0.976	41.6	70.3
Demetra	315	315	315	4.344	0.972	35.4	59.6

**Table S3.** Relative abundances of common taxa at phylum level for (a) Demetra, Eglouvis 1, and Eglouvis 2, and (b) Demetra, Eglouvis 3, and Eglouvis 4.

(a)

Sample	Demetra	Eglouvis 1	Eglouvis 2
<b>Acidobacteria</b>	0.109	0.729	0.162
<b>Actinobacteria</b>	0.238	0.387	0.375
<b>Ascomycota</b>	0.353	0.274	0.374
<b>Bacteroidetes</b>	0.358	0.330	0.312
<b>Crenarchaeota</b>	0.450	0.394	0.156
<b>Firmicutes</b>	0.212	0.380	0.407
<b>Gemmatimonadetes</b>	0.260	0.352	0.388
<b>Planctomycetes</b>	0.220	0.412	0.368
<b>Proteobacteria</b>	0.276	0.409	0.315
<b>Thaumarchaeota</b>	0.311	0.357	0.332
<b>Verrucomicrobia</b>	0.287	0.371	0.342

(b)

Sample	Demetra	Eglouvis 3	Eglouvis 4
<b>Acidobacteria</b>	0.217	0.408	0.375
<b>Actinobacteria</b>	0.167	0.469	0.364
<b>Ascomycota</b>	0.184	0.315	0.501
<b>Bacteroidetes</b>	0.233	0.451	0.316
<b>Chloroflexi</b>	0.123	0.444	0.433
<b>Crenarchaeota</b>	0.150	0.714	0.137
<b>Firmicutes</b>	0.146	0.457	0.397
<b>Gemmatimonadetes</b>	0.155	0.476	0.370

<b>Planctomycetes</b>	0.140	0.546	0.315
<b>Proteobacteria</b>	0.188	0.469	0.343
<b>Thaumarchaeota</b>	0.239	0.358	0.403
<b>Verrucomicrobia</b>	0.192	0.440	0.368

Table S4. Top 10% of key species (31/158) after degree centrality analysis

Species	Degree Centrality (Links)
<i>Actinoplanes liguriensis</i>	20
<i>Arthrobacter sp. 'SMCC G965'</i>	20
<i>Arthrobacter sp. 'SMCC ZAT200'</i>	19
<i>Cellulomonas fimi</i>	20
<i>Cryobacterium psychrophilum</i>	18
<i>Glaciibacter superstes</i>	18
<i>Microbacterium oxydans</i>	18
<i>Micromonospora chaiyaphumensis</i>	19
<i>Pimelobacter simplex</i>	18
<i>Pseudonocardia zijingensis</i>	18
<i>Terracoccus luteus</i>	20
<i>Thermobispora bispora</i>	20
<i>Verticillium dahliae</i>	18
<i>Chitinophaga sancti</i>	18
<i>Cytophaga hutchinsonii</i>	20
<i>Hymenobacter sp. VUG-A141a</i>	18
<i>Pedobacter sp. MJ11</i>	18
<i>Salinibacter ruber</i>	18
<i>Bacillus halmapalus</i>	20
<i>Bacillus sp. TA2.A1</i>	20
<i>Coprothermobacter proteolyticus</i>	18
<i>Marvinbryantia formatexigens</i>	20
<i>Paenibacillus polymyxa</i>	18
<i>Ruminococcus flavefaciens</i>	20
<i>Symbiobacterium thermophilum</i>	19
<i>Gemmatimonas aurantiaca</i>	18
<i>Planctomycete Zi62</i>	18
<i>Angiococcus disciformis</i>	18
<i>Corallococcus coralloides</i>	18
<i>Shewanella frigidimarina</i>	20
<i>uncultured proteobacterium</i>	18

Table S5. Top 10% (16 /158) key species after closeness centrality analysis.

Species	Closeness Centrality
<i>Actinomadura pelletieri</i>	0.001892
<i>Actinoplanes missouriensis</i>	0.001769
<i>Amycolatopsis coloradensis</i>	0.001755
<i>Lechevalieria aerocolonigenes</i>	0.001892
<i>Microbispora rosea</i>	0.001901
<i>Streptosporangium roseum</i>	0.001755
<i>Hymenobacter aerophilus</i>	0.001901
<i>Hymenobacter antarcticus</i>	0.001901
<i>Prolixibacter bellariivorans</i>	0.001641
<i>Acetobacterium woodii</i>	0.001892
<i>Bacillus firmus</i>	0.001641
<i>Bacillus niaci</i>	0.00188
<i>Bacillus sp. S207</i>	0.00188
<i>Planctomyces limnophilus</i>	0.001901
uncultured <i>Geobacter</i> sp.	0.001901
uncultured Rhodocyclaceae bacterium	0.001892

Table S6. Top 10% of key species (15/158) after betweenness centrality.

Species	Betweenness
<i>Actinoplanes liguriensis</i>	427.067
<i>Clavibacter michiganensis</i>	530.2385
<i>Rubrobacter xylanophilus</i>	538.571
<i>Streptomyces maritimus</i>	543.7762
<i>Streptomyces virginiae</i>	631.2566
<i>Thermobispora bispora</i>	427.067
<i>Cytophaga hutchinsonii</i>	631.2399
<i>Thermomicrombium roseum</i>	461.6074
<i>Bacillus halmapalus</i>	427.067
<i>Bacillus litoralis</i>	424.3583
<i>Bacillus sp. TA2.A1</i>	631.2399
<i>Desulfitobacterium hafniense</i>	424.3583
<i>Desulfonispora thiosulfatigenes</i>	483.5359
<i>Buchnera aphidicola</i>	541.219
<i>Pseudoxanthomonas dokdonensis</i>	462.2815