

Figure S1. Transcriptional analysis of HF and i-HF. A) Gene Ontology (GO) enrichment analysis based on up- or downregulation of biological processes terms from differentially expressed genes (DEG) found in i-HF compared to HF of the three different donors. B-C) Gene set enrichment analysis (GSEA) were conducted on the pool of the three donors associated with the indicated biological pathways in i-HF compared to HF are shown. P-adj=Adjusted p-value, NES=Normalized enrichment score. D) Heatmap illustrating the transcriptional landscape associated with the regulation of TP53 activity gene set for each cell line. Color scale reflects the standard deviation of gene expression level with upregulation in red and downregulation in blue. Hierarchical clustering indicates level of similarities among samples.

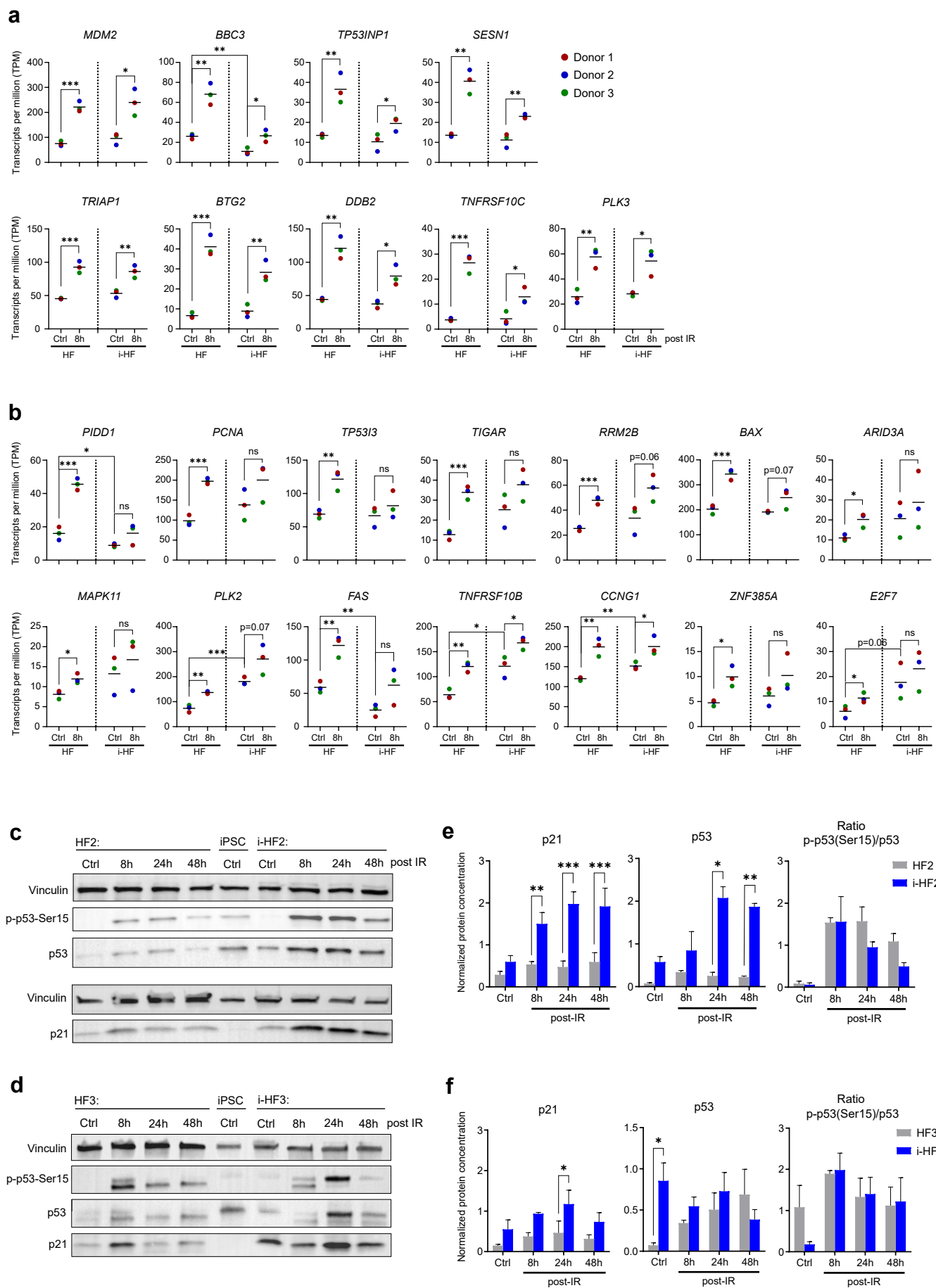


Figure S2. p53-related effectors gene and protein expression in HF and i-HF following IR. A-B) Number of mRNA transcripts per million (TPM) mapped reads from RNA-seq data of each of the three donors of HF and i-HF at basal level (Ctrl) and 8h post exposure to 15 Gy IR. Shown are the DEG common for both cell line or only in HF as shown in the Venn diagram from figure 2B. Lines represent the mean value. Data was analyzed using multiple unpaired t tests between HF and i-HF for each group. C-D) Representative western blot membranes showing protein expression of p21, p53 and phosphorylated p53 at serine 15 (p-p53(Ser15)) from clone 1 of HF and i-HF donor 2 and 3 after 8, 24 and 48h exposure to 15 Gy IR. Vinculin was used as a loading control. E-F) Quantification of p21 and p53 protein expression normalized on vinculin signal with the quantified ratio of p-p53(Ser15) on total p53 protein signal of donor 2 and 3 respectively. Data analysis was performed using a two-way ANOVA followed by a Fisher's LSD post-hoc test. Shown is mean \pm SEM of n=3 individual western blots.

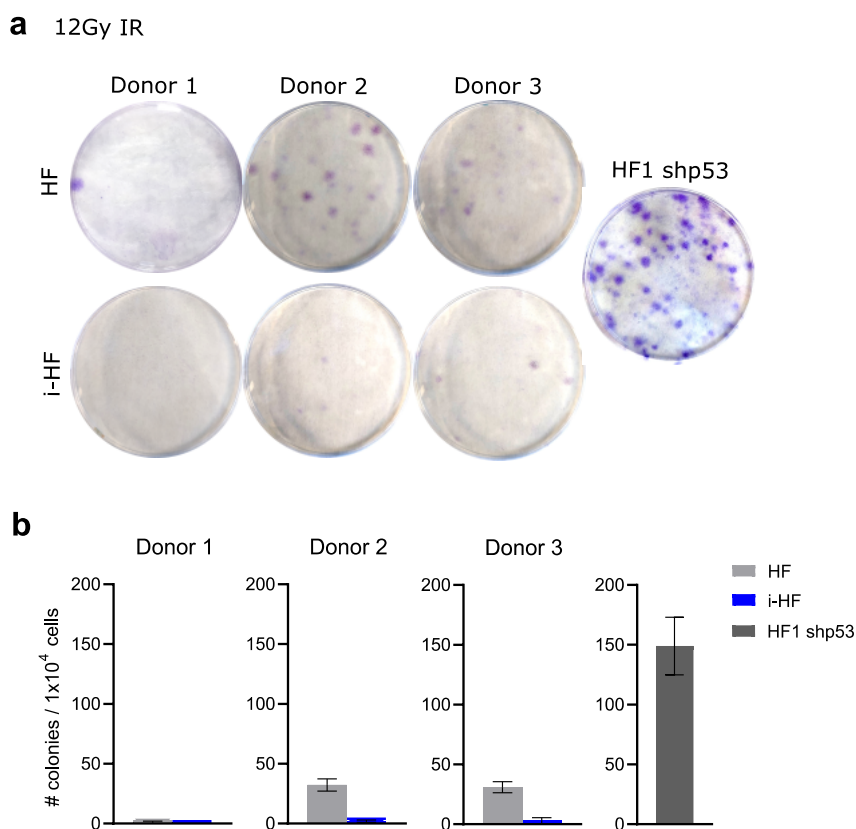


Figure S3. I-HF cell lines are more sensitive to IR. A) Representative images showing the results of a colony formation assay on cells exposed or not to 12 Gy IR. B) Quantification of the number of colonies counted following senescence induction on cell lines described in panel A. Shown is mean \pm SD from 3 replicates of one experiment.

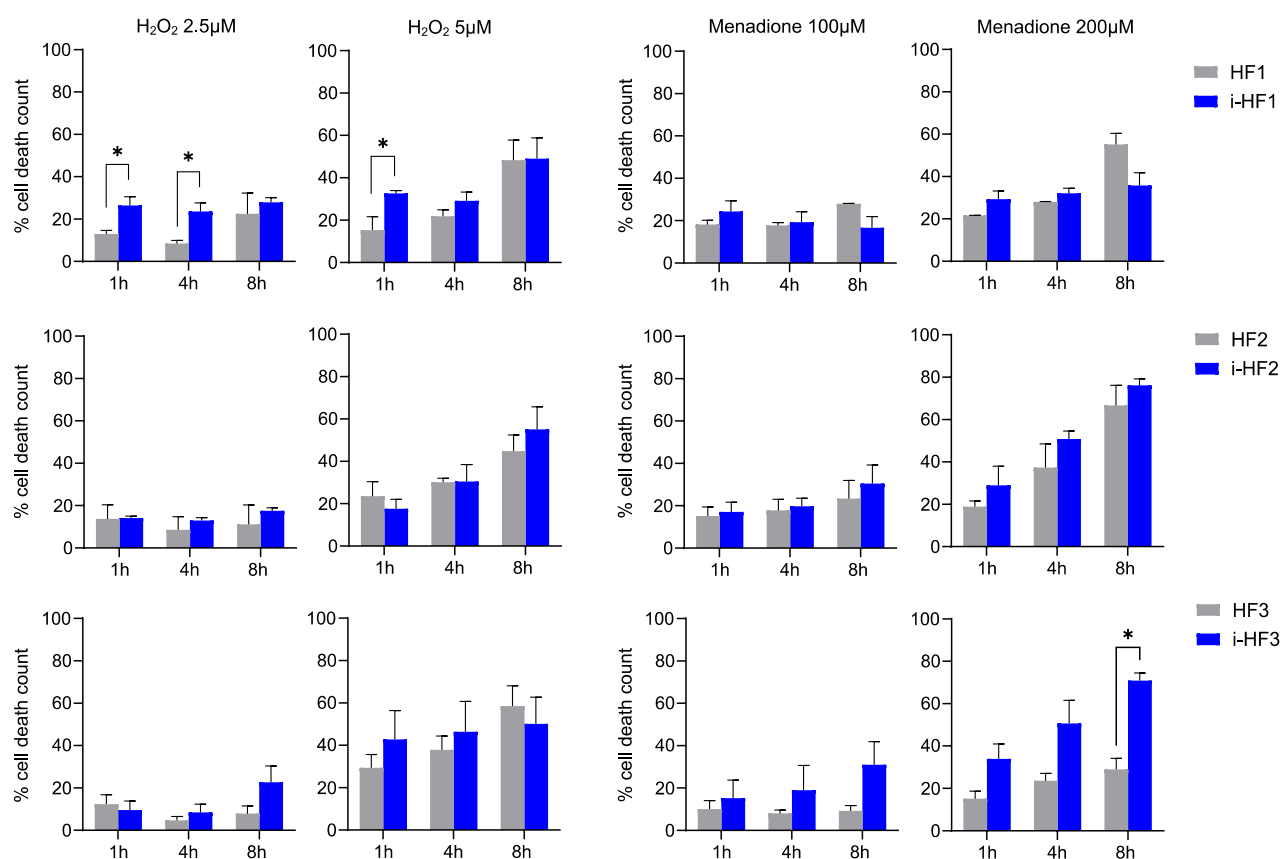


Figure S4. i-HF from donors 1 and 3 exhibit increased sensitivity to oxidative stress. Percentage of cell death caused by two different doses of H₂O₂ or menadione, two known inducers of oxidative stress. Cell death was assessed by propidium iodide staining using flow cytometry on the indicated HF and i-HF lines at different timepoints (1h, 4h and 8h) after cells were treated for 1h with the indicated reagent. Multiple unpaired t tests were applied for each timepoint. Shown is mean±SEM of n=3 independent experiments.

Supplementary Table S1: List of antibodies

| Antibodies | Supplier reference | Dilution | Application |
|--------------------------------|----------------------------------|-----------------|--------------------|
| TRA1-60 | Life technologies - A24881 | 1:500 | IF |
| SSEA4 | Life technologies - A24881 | 1:500 | IF |
| Vimentin | Invitrogen – MA5-16409 | 1:200 | IF |
| FSP-1 | Sigma – F4771 | 1:200 | IF |
| hTERT | Santa cruz – sc-393013 | 1:200 | IF |
| Iso PE Rat IgG2bk (RTK4530) | Biologend - 400608 | | FACS |
| Iso FITC IgG2ak (MOPC-173) | Biologend - 400208 | | FACS |
| Iso APC IgG1k (MOPC-21) | Biologend - 400122 | | FACS |
| CD44 (IM7) | Biologend - 103008 | | FACS |
| HLA-1 - FITC | Biologend – cl W6/32 311404 | | FACS |
| CD73 - APC | Biologend – cl AD2 344006 | | FACS |
| BrdU | BD – cl B44 347580 | 1:250 | IF |
| P21 | R&D – AF1047 | 1:1000 | WB |
| p53 | Santa Cruz - sc-126 | 1:1000 | WB |
| Phosphor-p53 (Ser15) | Cell signaling – 9284S | 1:1000 | WB |
| CDK4 | Cell signaling – cl D9G3E 12790S | 1:1000 | WB |
| p16 | BD – 51-1325GR | 1:500 | WB |
| Vinculin | Santa Cruz – sc-73614 | 1:5000 | WB |
| yH2Ax (Ser139) | EMD Millipore – cl JBW301 | 1:250 | IF |
| 53BP1 | Novus – NB100-304 | 1:1500 | IF |

Supplementary Table S2: List of primers

| | |
|---------------------|---------------------------|
| <i>18S Fw</i> | TCAACTTTTCGATGGTAGTCGCCGT |
| <i>18S Rev</i> | TCCTTGGATGTGGTAGCCGTTTCT |
| <i>CDKN1A Fw</i> | GGCAGACCAGCATGACAGATT |
| <i>CDKN1A Rev</i> | GCGGATTAGGGCTTCCTCTT |
| <i>GADD45A Fw</i> | CTGGAGGAAGTGCTCAGCAAA |
| <i>GADD45A Rev</i> | GCACAACACCACGTTATCGG |
| <i>Sce1 NHEJ Fw</i> | AGGTCCCTGAGGCTACCTCCAGTTC |
| <i>Sce1 NHEJ Rv</i> | CATAGTGAGGAGTACCAGGACAG |

Supplementary Table S3: NHEJ GFP construct sequence comprised between primers used for amplification.

tcctgaggctacctccagttc taaggctgcactccattctcacagccaggtctgctcagggcacagggcacagggtagggcccttact
ttcccggcatttaaacaatagccctggaaagagaagctgacagttgcaaagctgcttgctgccttggttcagtgcccttctgacggtag
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gttgaggacaaaactcttcgcggtcttccagtagtcttgatcggaaccctcggcctccgaacggtaagagcctagcatgtagaactgg
ttcgacctgcagattaccctgttatXccctaagcttgtcacagaccctcctgctctggttctaagcatcccagaaatgtaactgaaa
gtgttctgggagtgacctggcctgtttccctctctggaactgtttgtaaactctgaccactgctgtcctgggtactcctcactatg

aaa: Primers

aaa: Intron sequence

aaa: I-Sce1 recognition sequence

aaa: Adenoviral exon sequence

X: Expected I-Sce1 cut sites