

Review

# Lipid Peroxidation and Antioxidant Protection

Luca Valgimigli 

Department of Chemistry “G. Ciamician”, University of Bologna, Via Piero Gobetti 85, 40129 Bologna, Italy; luca.valgimigli@unibo.it; Tel.: +39-0512095683

**Abstract:** Lipid peroxidation (LP) is the most important type of oxidative-radical damage in biological systems, owing to its interplay with ferroptosis and to its role in secondary damage to other biomolecules, such as proteins. The chemistry of LP and its biological consequences are reviewed with focus on the kinetics of the various processes, which helps understand the mechanisms and efficacy of antioxidant strategies. The main types of antioxidants are discussed in terms of structure–activity rationalization, with focus on mechanism and kinetics, as well as on their potential role in modulating ferroptosis. Phenols, pyri(mi)dinols, antioxidants based on heavy chalcogens (Se and Te), diarylamines, ascorbate and others are addressed, along with the latest unconventional antioxidant strategies based on the double-sided role of the superoxide/hydroperoxyl radical system.

**Keywords:** autoxidation; peroxy radicals; hydroperoxyl radicals; kinetics; antioxidants; phenols; catechols; nitroxides; thiols; pyridinols

## 1. Introduction

*Lipid peroxidation* (LP) is a complex phenomenon, first investigated in the early 20th century, consisting in the uptake of molecular oxygen by lipids exposed to air, which was soon recognized as bearing remarkable similarity to hydrocarbon autoxidation [1,2], the formal insertion of one molecule of oxygen in the C-H bond of a hydrocarbon to afford a hydroperoxide:  $R-H + O_2 \rightarrow R-OO-H$ . Indeed, lipid peroxidation is one embodiment of hydrocarbon autoxidation.

While the direct reaction with ground state (triplet) oxygen is spin restricted and too slow to occur, the transformation of hydrocarbons (or lipids) by oxygen to hydroperoxides and further oxidized products occurs rapidly and efficiently via the intermediation of peroxy radicals ( $ROO\bullet$ ), in a chain reaction that can be triggered by a multitude of events in any chemical system, such as in food or in living organisms, and it can be blocked or prevented by *antioxidants* [1–3].

Over the last 70 years, extensive research efforts have shown the association of lipid peroxidation with an impressive number of pathological conditions, from arteriosclerosis and cardiovascular diseases to neurological disorders and cancer [1–7], stemming from the “Free Radical Theory of Aging”, which attributes the progressive decline of functionality associated with aging to the progressive accumulation of damage to biomolecules and essential biological structures, caused by radicals’ overproduction, also referred to as “oxidative stress” [6,7]. Yet the interest in lipid peroxidation has recently seen a boost after the formal recognition in 2012 of ferroptosis as a new form of programmed cell death [8], driven by LP and specifically linked to the pathophysiology of several degenerative diseases [9–11]. Even more interesting are pioneering observations that antioxidants can effectively inhibit ferroptosis [12,13] and that either antioxidants or pro-oxidant molecules can be used to modulate it, promising a handle on the connected pathologies [10,13].

Although several valuable reviews have been dedicated to LP and ferroptosis [14,15], the importance of lipid peroxidation is not limited to them. Damage to membrane lipids by radicals and reactive oxygen species (ROS) is perhaps the most important type of oxidative/radical damage in biological systems; not only because of the susceptibility of lipids to



**Citation:** Valgimigli, L. Lipid Peroxidation and Antioxidant Protection. *Biomolecules* **2023**, *13*, 1291. <https://doi.org/10.3390/biom13091291>

Academic Editor: Chryssostomos Chatgililoglu

Received: 13 July 2023

Revised: 17 August 2023

Accepted: 21 August 2023

Published: 24 August 2023



**Copyright:** © 2023 by the author. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

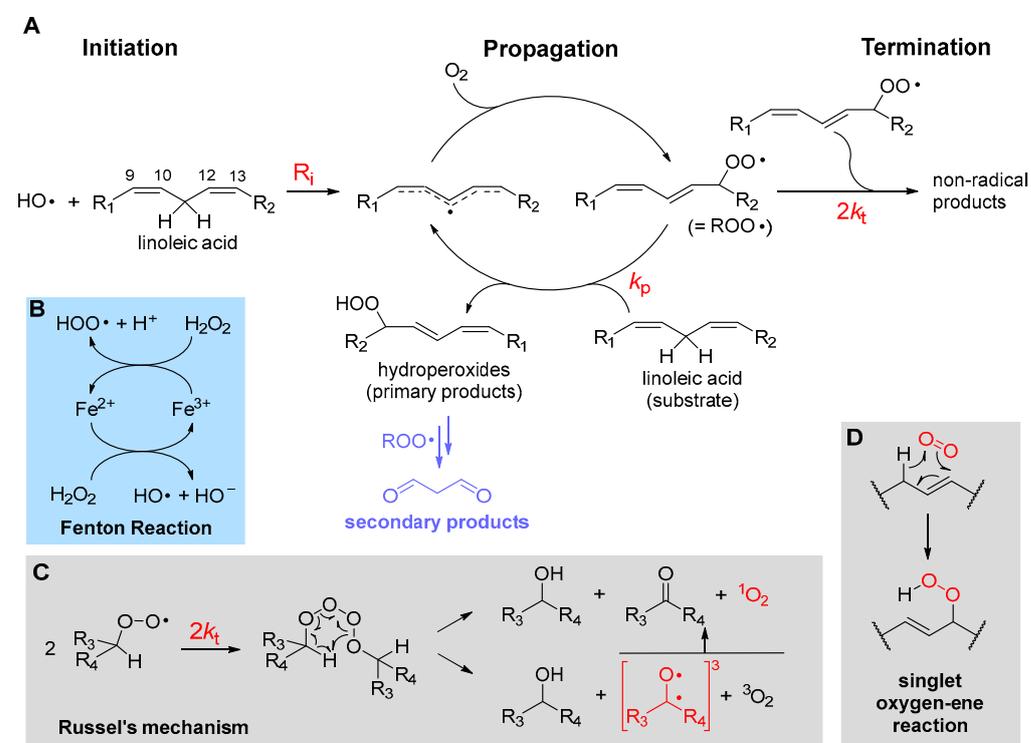
radical attack and because it produces the majority of biomarkers commonly used to monitor such damage but because such biomarkers—the LP secondary products—are also major effectors of the damage itself [16], owing to their toxicity and to their ability to alter other biomolecules such as proteins [17], thereby extending the radical/oxidative damage [18].

The aim of this review is to offer a brief up-to-date overview on lipid peroxidation, including the main processes leading to secondary products, with focus on the kinetic aspects regulating the different reactions, so to guide the understanding of the efficacy of the different antioxidant protection strategies. The main type of antioxidants will also be briefly reviewed addressing their efficacy, on kinetic bases, and their mechanism of action.

## 2. The Chemistry of Lipid Peroxidation

### 2.1. The Three Stages of Lipid Peroxidation

LP is a radical chain reaction composed of the canonical three stages of initiation, propagation and termination, summarized in Figure 1 using PUFA as the prototypical oxidizable substrate.



**Figure 1.** The main steps of lipid peroxidation (A), with indication of one main mechanism of initiation (B), details of Russel's mechanism for termination (C) and reaction of singlet oxygen with unsaturated lipids (D).

*Initiation*, i.e., the formation of the first lipid-derived radical, can consist in attack by a variety of radical species. In biological systems, these would most commonly be an alkoxy radical ( $\text{RO}\cdot$ ) or a hydroxyl radical ( $\text{HO}\cdot$ ), e.g., generated by Fenton-type decomposition of hydroperoxides or hydrogen peroxide, catalyzed by transition metals such as iron or copper. Other initiating species could be hydroperoxyl radicals ( $\text{HOO}\cdot$ ), the neutral form of metabolically produced superoxide radical anion ( $\text{O}_2^{\bullet-}$ ), which, however, is not prevailing at pH 7 or higher—the  $\text{pK}_a$  is 4.8 in water at 20 °C [19]. Excited triplet states produced by UV irradiation of carbonyl compounds, e.g., in the skin, or radicals produced from water by ionizing radiations would also serve the purpose. Since the attack to lipids occurs typically by formal hydrogen-atom transfer (HAT) from the  $>\text{CH}_2$  in allylic/bis-allylic positions, having a bond dissociation enthalpy (BDE) of about 77/87 kcal/mole, respectively, any

radical species  $X\bullet$  forming a product  $X-H$  with BDE higher than this value would be suitable for the task.

*Propagation* of the oxidative chain occurs by two alternating steps: the reaction of the lipid-derived C-centered radical ( $>C(\bullet)H$ ;  $=R\bullet$ ) with oxygen to form the alkylperoxyl radical ( $ROO\bullet$ ) is extremely fast (rate constant in the range  $2-5 \times 10^9 \text{ M}^{-1}\text{s}^{-1}$  [20]), hence the kinetics of propagation is governed by the second step, the reaction of  $ROO\bullet$  with a new lipid molecule to afford a new lipid-derived  $R\bullet$  radical. Its rate constant  $k_p$  is the most important parameter in evaluating antioxidant strategies (vide infra) and it depends dramatically on the structure of the lipid molecule. As summarized in Table 1,  $k_p$  is negligible for saturated fatty acids (or hydrocarbons) at close to physiologic temperature [21], but it grows by about two orders of magnitude in monounsaturated fatty acids (MUFAs, e.g., methyl oleate) and by about four orders of magnitude with two unsaturations, like in methyl linoleate, which reaches  $k_p = 62 \text{ M}^{-1}\text{s}^{-1}$  at  $30^\circ\text{C}$  [22]. This is due to resonance stabilization of the allyl and, particularly, of the bis-allyl radical, resulting by HAT from MUFA and PUFA, respectively. On increasing the degree of unsaturation in fatty acids, the number of  $>CH_2$  in the bis-allylic position also increases, which explains the almost linear increase in  $k_p$  with the number of double bonds [6], first reported by Porter's group [23] (Table 1), representing a significant challenge for antioxidant protection (vide infra).

**Table 1.** Rate constants for the peroxidation of lipids in bulk or in chlorobenzene at 303 K.

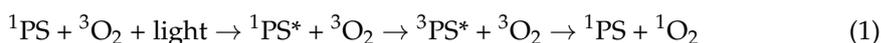
Lipid	$k_p/(2k_t)^{1/2}$ $10^{-5} \text{ M}^{-1/2}\text{s}^{-1/2}$	$k_p$ $\text{M}^{-1}\text{s}^{-1}$	$2k_t$ $10^5 \text{ M}^{-1}\text{s}^{-1}$	Ref.
Methyl stearate (18:0) <sup>1</sup>	~0.8	~0.01	15	[21]
Methyl oleate (18:1)	89.0	0.89	10	[22]
Methyl linoleate (18:2)	2100	62.0	88	[22]
Methyl linolenate (18:3)	3900	236.0	360	[22]
Linoleic acid (18:2)	-	62	-	[23]
Arachidonic acid (20:4)	-	197	-	[23]
Eicosapentaenoic ac. (20:5)	-	249	-	[23]
Docosahexaenoic ac. (22:6)	-	334	-	[23]
Cholesterol	-	11	-	[23]
7-Dehydrocholesterol	-	2260	-	[23]
Squalene	2500	68.0	74.0	[24]
Sunflower oil (60% of 18:2)	3600	66.9	34.5	[24]
PLPC <sup>2</sup>	-	16.6	1.27	[25]
DLPC <sup>3</sup>	-	13.6 <sup>4</sup>	1.02	[25]

<sup>1</sup> Data for hexadecane. <sup>2</sup> 1-Palmitoyl-2-linoleoyl-*sn*-glycero-3-phosphocholine; data at 310 K. <sup>3</sup> 1,2-Dilinoleoyl-*sn*-glycero-3-phosphocholine; data at 310 K. <sup>4</sup> For intermolecular propagation,  $k_p$  (intramolecular) =  $5.1 \text{ s}^{-1}$ .

The main products of the overall propagation stage are hydroperoxides ( $ROOH$ ), the *primary products* of LP [1], and new alkylperoxyl radicals, which are the chain-carrying species in LP. Depending on the lipid structure, formation of other side products like epoxides and endoperoxides might also gain importance (vide infra).

*Termination* of the chain reaction occurs when two radicals quench each other to afford non-radical (diamagnetic) species, which are unable to propagate the chain. Uninhibited LP is carried on by secondary peroxyl radicals, and the best-established termination process is Russel's mechanism [26], which consists in the formation of a tetroxide that decomposes to afford carbonyl compounds and molecular oxygen (Figure 1). It has lately been clarified that  $O_2$  released by decomposition of the tetroxide is predominantly in the excited singlet state  $^1O_2$  [27], meaning that oxygen molecules bear all electrons in pairs with antiparallel spin. While the singlet state is the lowest in energy for most molecules, oxygen is the exception and its ground state is a (paramagnetic) triplet biradical, i.e., it has two electrons with parallel spin in (degenerate) antibonding  $\pi^*$  orbitals. This, perhaps counterintuitively, makes it much less reactive toward "normal" diamagnetic molecules due to spin restriction, i.e.,  $^3O_2$  is much less oxidizing than  $^1O_2$ . Other processes have been shown to form singlet

oxygen in biological systems. Among them, the reaction of hydrogen peroxide ( $\text{H}_2\text{O}_2$ ) and lipid hydroperoxides (ROOH) by myeloperoxidase (MPO) in the presence of  $\text{Cl}^-$  ions, which is possibly a defensive mechanism occurring during phagocytosis. It was demonstrated that the process occurs by reaction of  $\text{H}_2\text{O}_2$  or ROOH with HOCl produced by MPO [27]. Earlier work indicated that  $^1\text{O}_2$  is quantitatively produced by HOCl reaction on  $\text{H}_2\text{O}_2$  but it is not observed with ROOH [28]. However, subsequent work, addressing the ability of different hydroperoxides to produce  $^1\text{O}_2$  upon HOCl reaction, showed that tertiary alkylhydroperoxides are ineffective, while typical lipid-derived secondary ROOH form  $^1\text{O}_2$  with an approximately 10% yield via Russel's mechanism [29]. The decomposition of lipid hydroperoxides in the presence of metal ions or peroxyxynitrite was also shown to produce  $^1\text{O}_2$  with an approximately 10% yield [27]. Decomposition of 7- $\alpha$ -OOH, 6- $\beta$ -OOH and 5- $\alpha$ -OOH cholesterol hydroperoxides, formed upon cholesterol autoxidation in biomembranes, is also a relevant source of  $^1\text{O}_2$  via a Russel's-type mechanism [27]. Cytochrome c causes the formation of  $^1\text{O}_2$  in mitochondria by promoting the autoxidation of cardiolipin [30]. Additionally, singlet oxygen can be produced by photoexcitation of ground state triplet oxygen in the presence of a photosensitizer [6], e.g., in the skin, as summarized in Equation (1). The photosensitizer (PS), such as a carbonyl compound in its ground singlet state, absorbs visible or UV light and is promoted to the excited singlet state, which undergoes intersystem crossing to convert into the excited triplet state. This finally transfers energy to ground state triplet oxygen via a triplet annihilation process, to go back to the ground state converting oxygen into the excited singlet state.



Singlet oxygen is unable to restart the autoxidation chain; however, it directly reacts with unsaturated lipids via an oxygen-ene reaction to afford the lipid hydroperoxides (Figure 1) [6].

## 2.2. The Rate of Lipid Peroxidation

Chain termination typically quenches two peroxy radicals in a fast radical–radical reaction with rate constant  $2k_t$ . Although  $2k_t$  is much higher than the rate constant  $k_p$  for propagation (see Table 1), the actual rate of termination also depends on the square of the steady-state concentration of the chain-carrying peroxy radical, which is very low, typically in the micromolar-to-nanomolar range. Therefore, it can be estimated that, following an initiation event, >100 propagation cycles occur before termination takes place in LP of unsaturated lipids, implying that one single initiating radical can damage hundreds of lipid molecules, which explains why antioxidants are necessary.

In the absence of antioxidants, the rate of peroxidation of a lipid substrate RH, at a constant rate of chain initiation  $R_i$ , follows Equation (2) [21,22,24,31], i.e., it is dictated by the ratio of the rate constant for propagation over the square root of the rate constant for termination. The kinetic quantity  $k_p/(2k_t)^{1/2}$  is named the “oxidizability” of the specific lipid substrate; it increases with unsaturation (Table 1) and it determines how important antioxidant protection is for the specific substrate.

$$-\frac{d[\text{O}_2]}{dt} = -\frac{d[\text{RH}]}{dt} = \frac{d[\text{ROOH}]}{dt} = \frac{k_p}{\sqrt{2k_t}} \times \sqrt{R_i} \times [\text{RH}] + R_i \quad (2)$$

## 2.3. Light as a Product of Lipid Peroxidation

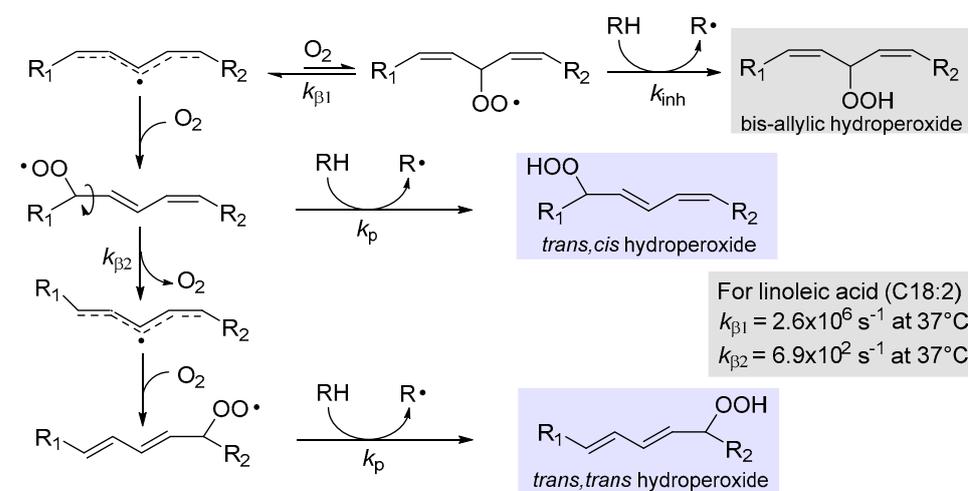
The termination stage of LP produces high-energy (transient) species that can deactivate to ground state or lose excess energy by emitting light, which can therefore be regarded as a product of LP. One important example is singlet oxygen,  $^1\text{O}_2$ , which can return to a ground state triplet by giving weak phosphorescence in the near infrared region at 1270 nm [27]. This chemoluminescence (CL) phenomenon has been extensively exploited to probe the formation of  $^1\text{O}_2$  during LP [27–29] and strategies have been developed to enhance CL in biological systems to improve detection sensitivity [32].

Emission of chemoluminescence in the visible region is also observed during LP and it is attributed to excited carbonyls in the triplet state, such as those formed in Russel's termination mechanism (Figure 1C), although singlet oxygen, e.g., formed in the same process, also contributes [33]. It has also been proposed that part of the excited triplet carbonyls affording CL are formed by an alternative termination mechanism consisting in cyclization of the alkylperoxyl radical to a dioxyethane followed by its thermolysis [34]. Emission can be enhanced by additives like 9,10-dibromoanthracene and it is proportional to the rate of LP [33]. Interestingly, the time-based monitoring of CL has been developed as a method to monitor the kinetics of LP both in animal and vegetable lipids [33,35], which has also been fruitfully applied to study the activity of antioxidants [36,37]

## 2.4. Further Insights into Chain-Propagation Reactions

### 2.4.1. $\beta$ -Fragmentation of the Peroxyl Radical

Addition of oxygen to lipid radical  $R\bullet$  to afford the peroxyl radical  $ROO\bullet$  is a reversible process and its back reaction, named  $\beta$ -fragmentation, has a rate constant  $k_{\beta}$ , which depends on the stability of the (reformed) carbon-centered radical (Figure 2). While it is generally too slow compared to the forward reaction to affect the rate of chain propagation, under some conditions it can compete with the subsequent propagation step, the reaction of  $ROO\bullet$  with the lipid molecule  $RH$  to afford the hydroperoxide primary product  $ROOH$  [38]. Therefore, it determines the distribution of isomeric hydroperoxide products, since *cis,trans* hydroperoxides are formed when propagation outcompetes  $\beta$ -fragmentation, hence they are the kinetic products of LP, while *trans,trans* hydroperoxides are the thermodynamic products, forming when  $\beta$ -fragmentation allows isomerization to take place [23,38].

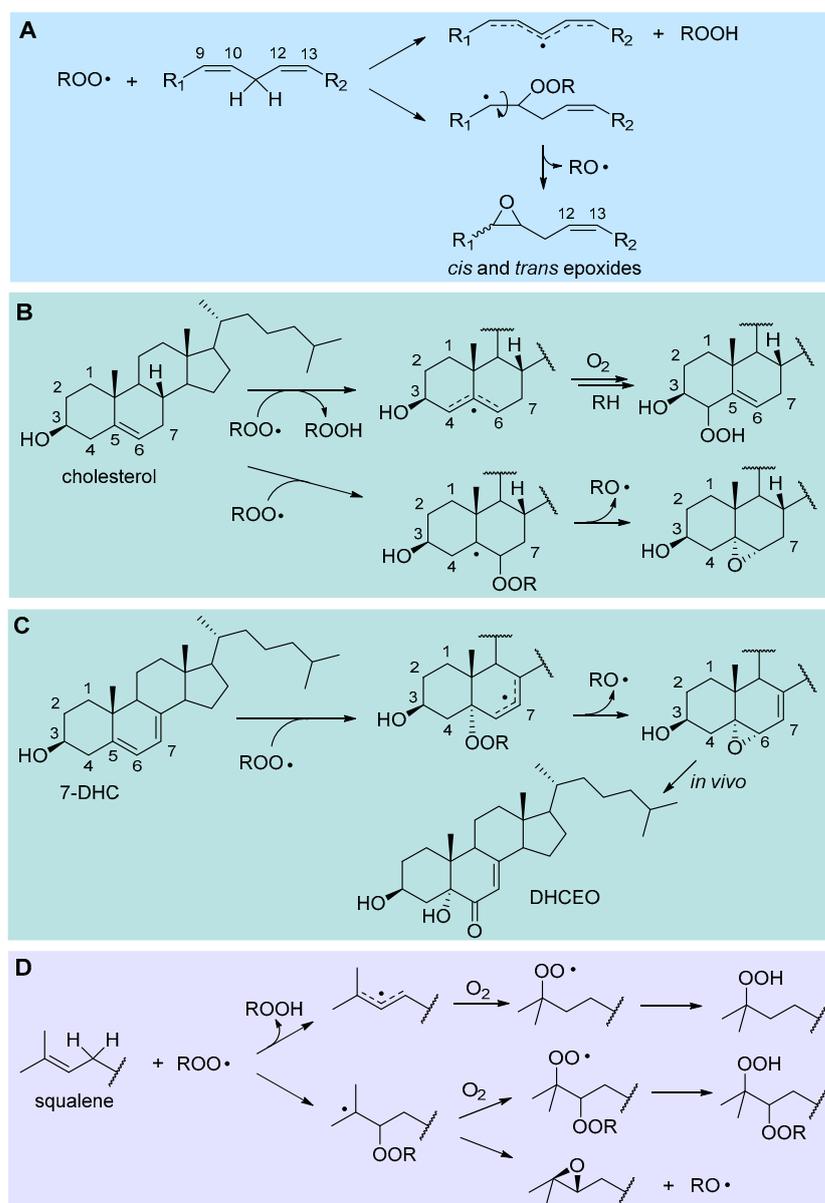


**Figure 2.** Schematic representation of  $\beta$ -fragmentation reactions in peroxyl radicals competing with H-atom abstraction from a donor  $RH$ , which is a lipid molecule ( $k_p$ ) or an antioxidant ( $k_{inh}$ ).

This reaction was thoroughly investigated in Porter's group who found that, upon calibration of the rate of  $\beta$ -fragmentations, these unimolecular reactions could be used as a "radical clock" to measure the kinetics of competing H-abstractions, by the analysis of the hydroperoxide product ratio from autoxidation [39]. For instance, they found that the rate constant for fragmentation of *cis,trans* hydroperoxides from linoleic acid (Figure 2) was  $k_{\beta 2} = 6.9 \times 10^2 \text{ s}^{-1}$  (at 37 °C) and would be useful to measure rate constants for propagation ( $k_p$ ). The fragmentation of bis-allyl hydroperoxide is instead much faster ( $k_{\beta 1} = 2.6 \times 10^6 \text{ s}^{-1}$  at 37 °C) and the corresponding hydroperoxide would never be formed in the absence of a much faster H-atom donor such as  $\alpha$ -tocopherol ( $\alpha$ -TOH) or other antioxidants; therefore, they used this radical clock to measure the rate constant of inhibition  $k_{inh}$  for antioxidants (vide infra) [39].

### 2.4.2. Hydrogen-Atom Abstraction vs. Radical Addition: Formation of Primary Epoxides

Although chain propagation in PUFAs such as linoleic acid is carried out mainly by H-atom abstraction from the bis-allylic position to afford the stabilized pentadienyl-type radical, addition of the ROO• radical to one of the C=C double bonds can also compete, to some extent, being favored by the contribution of polar effects in the transition state [40]. The resulting alkylperoxyalkyl radical would rapidly undergo a homolytic substitution ( $S_H^1$ ) reaction, forming an epoxide and releasing an alkoxy radical RO• (Figure 3), which would rapidly attack another lipid molecule, propagating the chain. Therefore, from a kinetic perspective, H-abstraction and addition are difficult to distinguish, besides the fact they afford different primary oxidation products, hydroperoxides or epoxides, respectively.



**Figure 3.** Formation of epoxides during the peroxidation of PUFA (A), of cholesterol (B), of 7-dehydrocholesterol (C) and of squalene (D).

The relative importance of addition might depend on the experimental conditions and would certainly depend on the lipid structure. Confirmation of its relevance comes from the identification of both 9,10- and 12,13-epoxides among the oxidation products of methyl

linoleate [41], while epoxyeicosatrienoic acids formed via peroxy radical addition were found in the peroxidation of arachidonic acid (C20:4) [42].

In the autoxidation of cholesterol, epoxides at C5,6 ( $\alpha+\beta$ ) were found to account for about 12% of the total oxidation products [43], demonstrating the importance of addition in chain propagation. A detailed study by Pratt's group showed that addition at C6 had similar activation energy ( $\Delta G^\ddagger = 17.6$  kcal/mol) to H-abstraction at C7-H (BDE = 83.2 kcal/mol,  $\Delta G^\ddagger = 17.5$  kcal/mol), being faster than H-abstraction at C4-H (BDE = 89.0 kcal/mol,  $\Delta G^\ddagger = 19.4$  kcal/mol) which would afford, respectively, the 7-OOH and 4-OOH hydroperoxides. Instead, the 6-OOR resulting from addition would undergo very rapid ( $\Delta G^\ddagger = 7.3$  kcal/mol) transformation to the 5,6-epoxide [43], as shown in Figure 3.

7-Dehydrocholesterol (7-HDC) is formed as the last step before cholesterol in the biosynthesis path that starts from squalene, and its metabolism is implicated in Smith–Lemli–Opitz syndrome (SLOS), a devastating autosomal disorder with a range of phenotypical expressions, including malformations and neurological deficits [38]. SLOS is characterized by much-elevated levels of 7-HDC (and low levels of cholesterol) due to defects in the gene encoding for 7-dehydrocholesterol reductase. 7-HDC is one the most oxidizable lipids in biological systems with a  $k_p$  200-fold higher than cholesterol and 10- to 40-fold higher than PUFA (Table 1), which reacts with peroxy radicals mainly H-abstraction, at C9-H or by addition at C5 (at variance with cholesterol) to afford a stabilized allylic radical [38]. This yields the 5 $\alpha$ ,6 $\alpha$ -epoxide (Figure 3) that, in vivo, is converted to 3 $\beta$ ,5 $\alpha$ -dihydroxycholest-7-en-6-one (DHCEO), which has been used as a biomarker in SLOS [38,43].

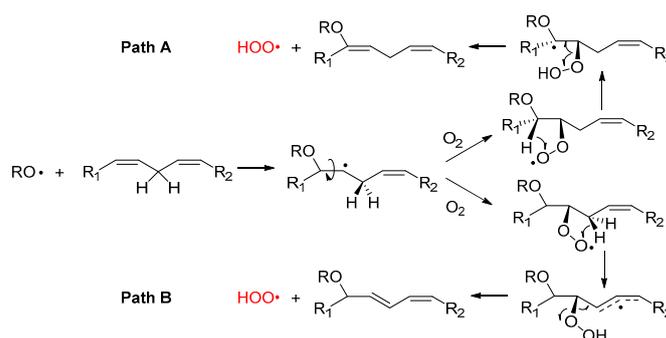
The autoxidation of squalene, the polyunsaturated triterpenic precursor of cholesterol and phytosterols, has recently been kinetically characterized in our group, finding a  $k_p$  of  $68 \text{ M}^{-1}\text{s}^{-1}$ , just slightly higher than that of methyl linoleate and 6-fold that of cholesterol (Table 1), which is due mainly to H-abstraction at repeating allylic positions, with significant contribution of ROO• addition, to afford a tertiary alkyl radical developing into an epoxide, as illustrated in Figure 3 [24].

A peroxy radical clock designed for measuring both H-atom abstraction and radical addition has recently been described and its application to a variety of unsaturated hydrocarbons indicates that addition grows in importance in conjugated dienes and polyenes; these include retinol (vitamin A) and carotenoids, conjugated linoleic acid (CLA 18:2) and conjugated linolenic acid (CLA 18:3) [44].

#### 2.4.3. Release of HOO• and Chain-Transfer Processes

Alkoxy radicals released via  $S_H^1$  reaction upon epoxide formation have very high reactivity toward unsaturated lipids and, besides H-atom abstraction, they undergo very fast addition to C=C double bonds [45,46], forming a  $\beta$ -alkoxyalkyl radical that rapidly reacts with oxygen (Figure 4). The resulting  $\beta$ -alkoxyalkylperoxy can undergo intramolecular H-atom abstraction (1,4-HAT) either in C $\alpha$  to the RO- group, to form a radical stabilized by resonance with the oxygen lone pair (Path A), or from the allylic  $>CH_2$  to form a radical stabilized by resonance with the double bond (Path B). In both cases, the resulting hydroperoxide can undergo fragmentation to release the hydroperoxy radical HOO• and reform the C=C double bond in the lipid structure. The release of HOO• has important consequences in the antioxidant protection of lipids (vide infra).

Although this process has never been demonstrated to occur in lipids like PUFA, similar reactions are known for simpler unsaturated hydrocarbons. The best established is 1,4-cyclohexadiene, which upon attack by alkylperoxy radicals releases HOO• and converts to benzene [47], and the monoterpene  $\gamma$ -terpinene, which undergoes identical chemistry affording HOO• and *p*-cymene [48]. In those hydrocarbons, the two steps, 1,4-HAT and elimination, are concerted and occur in a single transition state with a low barrier [49]. Pratt and coworkers were able to demonstrate that the reaction also occurs for simple monounsaturated hydrocarbons (like cyclooctene), albeit it does so stepwise, as depicted in Path A of Figure 4, and its kinetics is favored by quantum tunneling in the 1,4-HAT [49].



**Figure 4.** Mechanistic proposal for the release of HOO• during the autoxidation of methyl linoleate.

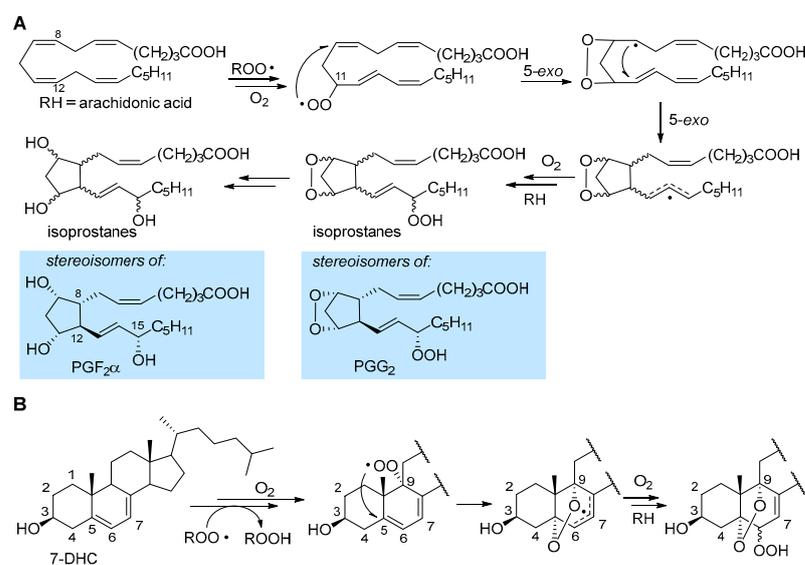
We have recently suggested the same mechanism to account for some non-classical behavior in the inhibited autoxidation of squalene [24], and ongoing studies in our group indicate a similar chemistry would take place during the autoxidation of methyl linoleate in micelles. In the case of PUFA, however, the possibility to form conjugated double bonds in the product suggests that Path B could also be a viable mechanism.

The release of HOO• has kinetic consequences, as these radicals can propagate the oxidative chain in bulk lipids—i.e., it represents a chain-transfer process—but, owing to their hydrosolubility, they could export the unpaired electron in the aqueous phase in heterogenous systems. As will be discussed in Section 5, it also has major importance for the antioxidant protection of lipids, under some conditions.

#### 2.4.4. Formation of Endoperoxides

Product distribution can become quite complex in highly unsaturated lipids as it can involve intramolecular radical addition to double bonds to yield cyclized products.

In the autoxidation of arachidonic acid in the absence of fast H-atom donors (e.g.,  $\alpha$ -TOH), the peroxy radical can undergo 5-*exo* cyclization occurring with a monomolecular rate constant of about  $800 \text{ s}^{-1}$  which outcompetes  $\beta$ -fragmentation with  $k_{\beta}$  of about  $140 \text{ s}^{-1}$  [1]. The result is the formation of an endoperoxide still bearing a C-centered radical (Figure 5), which undergoes another 5-*exo* cyclization, to form another alkyl radical, and further autoxidation steps to ultimately yield endoperoxide products or intermediates, which are stereoisomeric to some of the prostaglandins (e.g., PGG<sub>2</sub> and PGF<sub>2</sub> $\alpha$ ) produced by cyclooxygenase (COX) enzyme [31], therefore generally named isoprostanes.



**Figure 5.** Formation of endoperoxides during the propagation of lipid peroxidation of arachidonic acid (A) and 7-dehydrocholesterol (B).

In 7-HDC, autoxidation proceeds both by ROO• addition to yield the epoxide (Figure 3) and by H-atom abstraction preferably from C9-H [31,38]. The resulting peroxy radical was found to undergo 5-exo cyclization at C5 to yield a stabilized allyl radical that continues the autoxidation to afford a mixture of  $\alpha$ -5,9-endoperoxides- $\alpha$ & $\beta$ -6-hydroperoxide [38], as shown in Figure 5. In general, endoperoxide formation is observed when the unsaturated lipid geometry allows for 5-exo cyclization, possibly yielding an unstrained or resonance-stabilized radical.

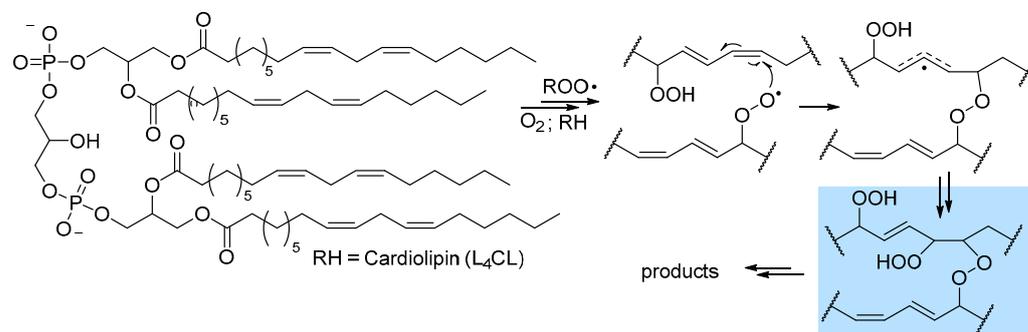
### 2.5. Peroxidation of Intact Triglycerides and Phospholipids

Most of the knowledge and understanding of the chemistry and related kinetics of lipid peroxidation have been developed using isolated PUFAs and MUFAs and their simple monoesters (e.g., methyl linoleate) as model compounds, along with biologically relevant yet specific compounds, like cholesterol and some of its congeners, or it has been mutated from knowledge on simpler hydrocarbons. However, on quantitative grounds, the majority of lipids in biological systems are phospholipids (e.g., in cell membranes) and triglycerides (e.g., in adipocytes and lipoproteins). Unfortunately, only a limited number of studies have specifically addressed their peroxidation chemistry, owing to their complexity, showing, however, some distinctive features.

A pioneering study by Antunes et al. investigated the kinetics of peroxidation of phosphatidylcholine multilamellar liposomes in buffered water. They studied the liposomes of both 1-palmitoyl-2-linoleoyl-sn-glycero-3-phosphocholine (PLPC) and 1,2-dilinoleoyl-sn-glycero-3-phosphocholine (DLPC), containing, respectively, one and two oxidizable linoleic acid residues per phospholipid molecule [25]. For PLPC, where only one chain is really oxidizable, they found  $k_p = 16.6 \text{ M}^{-1}\text{s}^{-1}$  and  $2k_t = 1.27 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$  at 37 °C, hence they were both significantly lower than those recorded for methyl linoleate in organic solution (Table 1). Interestingly, with DLPC, carrying two oxidizable chains per molecule, they were able to distinguish two rate constants  $k_p$  of  $13.6 \text{ M}^{-1}\text{s}^{-1}$  and of  $5.1 \text{ s}^{-1}$  for intermolecular and intramolecular chain propagation, respectively, while chain termination gave the rate constant  $2k_t = 1.02 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$  at 37 °C [25]. This clearly shows that the presence of two oxidizable chains in the same lipid molecules changes the kinetic behavior. Stemming from their results, Porter's group investigated, by the radical clock method, the kinetics of chain propagation for PC liposomes always containing palmitic acid esterified in position 1 of the glycerol and one PUFA in position 2: linoleic acid (C18:2; PLPC), used as reference, arachidonic acid (C20:4; PAPC), eicosapentaenoic acid (C20:5; PEPC) and docosahexanoic acid (C22:6; PDPC) [23]. The measured  $k_p$  values were proportional to the number of double bonds, as also seen for isolated PUFA in organic solution, but the absolute values were much lower (see Table 1), i.e., 35, 115, 145 and  $172 \text{ n}^{-1}\text{s}^{-1}$ , respectively, referring to the mole fraction of the oxidizable lipid in the liposomes, instead of the molar concentration in solution [23]. Interestingly, they found a different kinetic behavior of the peroxy radicals in C9 or in C13 in linoleoyl residue, indicating that oxidation in liposomes depends on the position of the formed peroxy radical. While the above results refer to lipids having only one oxidizable PUFA residue, subsequent studies on 1,2-dilinoleoyl-glycero-3-phosphocholine (DLPC) and on tetralinoleoyl cardiolipin (L<sub>4</sub>CL), a mitochondria-specific phospholipid carrying four linoleic acid residues, proved that arm-to-arm attack by peroxy radical is relevant and it can form interarm peroxide (-O-O-) bridges by addition to double bonds (Figure 6), with consequences on the formation of toxic electrophilic fragmentation products like 4-hydroxynonenal (vide infra) [50].

The kinetics of peroxidation of triglycerides is more complex than that of the isolated fatty acids in solution, owing to arm-to-arm propagation processes, which are more relevant in dilute solution than at high concentration or in the bulk [51]. To date, the only complete kinetic characterization of an intact natural triglyceride is that of sunflower seed oil (SSO) recently reported by our group [24]. Each triglyceride molecule statistically contains 1.7 chains of linoleic acid and its oxidizability was found to be roughly 1.7-fold that of linoleic acid; however, to our surprise, the rate constant  $k_p$  was just slightly higher

than that of linoleic acid ( $67$  vs.  $62 \text{ M}^{-1}\text{s}^{-1}$ ), hence the higher oxidizability is due to slower termination (see Table 1), possibly owing to steric impairment in Russel's mechanism [24].



**Figure 6.** Formation of arm-to-arm peroxides in the peroxidation of mitochondrial cardiolipin.

Clearly, more data would be desirable for complex lipids like phospholipids and triglycerides, particularly in the light of their importance for the onset of antioxidant strategies in biological systems.

### 3. Secondary and Late Products of Lipid Peroxidation

Following the formation of primary oxidation products of LP, namely primary hydroperoxides and primary epoxides, formed directly in chain propagation, their further oxidation, followed or accompanied by other reactions, yields a multitude of secondary products. Several such products are electrophiles and can undergo other reactions, e.g., with proteins, and they have cell signaling functions. Hence, they are toxic or might have some active biological role and have been regarded as biomarkers of oxidative damage, prompting great attention and major efforts in the development of analytical techniques to detect them [16,52–61].

#### 3.1. Formation of Electrophilic Carbonyl Compounds

The most regarded secondary product of LP, which is also among the most relevant LP biomarkers in biological systems, is 4-hydroxynonenal (4-HNE), formed as a late product of oxidation of linoleic acid [52,55,60]. It is an  $\alpha$ - $\beta$  unsaturated aldehyde with very high reactivity towards biological nucleophiles (e.g., amino acid residues and DNA bases), which grants both its role as a signaling molecule and its toxicity. Despite its importance, 4-HNE is not unique but a member of a broad family of short-chain aldehydes and ketones, often bearing conjugated unsaturation in the hydrocarbon chain, all characterized by the marked electrophilic character and all deriving from fragmentation of the hydrocarbon backbone of oxidized lipids; some of these carbonyl compounds are collected in Figure 7. Among them, two arguably stand out: 4-oxononenal (4-ONE), the oxidized form of 4-HNE, being orders of magnitude more reactive as an electrophile [6,58], and malondialdehyde (MDA), a dicarbonyl compound that is arguably the best-known marker of LP, owing to its facile detection (by spectrophotometry or HPLC) upon derivatization with thiobarbituric acid (the so-called TBARS assay) [52]. The mechanism by which 4-HNE forms from oxidized lipids has been highly debated, and several hypotheses have been proposed, which are not mutually exclusive and, possibly, might all be relevant to some extent.

The most classical mechanism is perhaps the one based on Hock rearrangement and fragmentation of the primary hydroperoxide in the presence of an acid catalyst [6,31]. The formed carbonyl compounds can then undergo further autoxidation to afford 4-HNE, as depicted in Figure 7A. Another well-established mechanism is based on Fenton-type cleavage of one hydroperoxyl group in a multiply oxidized linoleyl residue to afford an alkoxy radical that undergoes fragmentation [6,55]. The reduction of the formed 4-hydroperoxynonenal affords 4-HNE (Figure 7B).



### 3.2. Formation of Isoprostanes

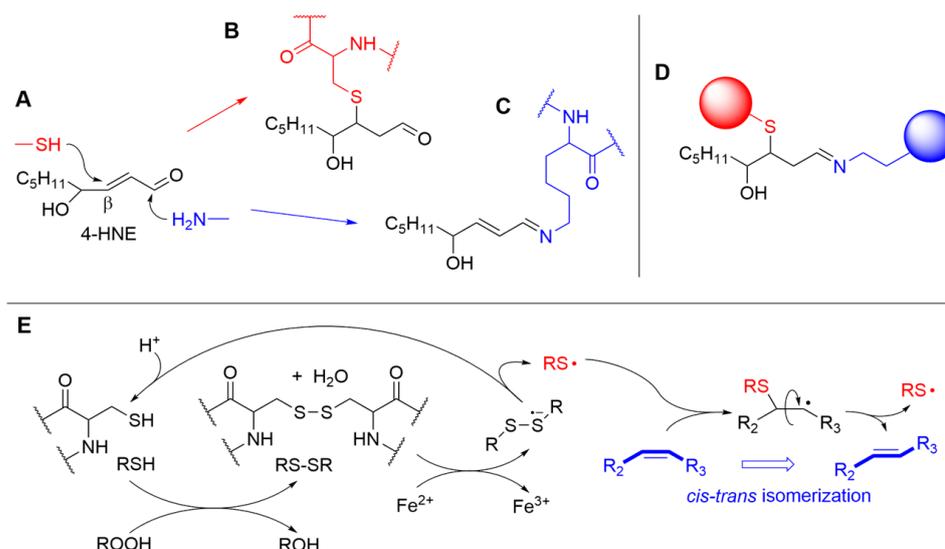
Formation of isoprostanes, from sequential steps following the formation of the peroxy radical in the peroxidation of arachidonic acid, is described in Section 2.3 and illustrated in Figure 5. Isoprostanes are stereoisomers of some of the otherwise identical prostaglandins, e.g.,  $\text{PGH}_2$ ,  $\text{PGG}_2$  and  $\text{PGF}_2\alpha$ , formed (instead) with full stereochemical control by cyclooxygenase (COX 1 and 2 in humans), still from arachidonic acid, released by a phospholipase during the inflammatory cascade [38]. Owing to their high specificity, isoprostanes have gained importance as the most reliable markers of LP and oxidative stress in biological systems, particularly in relation to some conditions which they directly affect [4,64–68]. Among them, 8-epi- $\text{PGF}_2\alpha$  (also called 8-isoprostane, 8-iso $\text{PGF}_2\alpha$ , iPF $_2\alpha$ -III and 15-F $_2$ t-IsoP), the epimer in C8 of  $\text{PGF}_2\alpha$ , is the best-established marker of oxidative stress in human studies, owing to its stability and selective urinary excretion, which makes its monitoring sensitive and convenient [4,67]. The autoxidation of arachidonic acid occurs with preferential formation of 8-, 9-, 11- or 12-peroxy radicals; each of them generates one class of F $_2$ -IsoP [67,68] and each of the four classes consists of 16 stereoisomers, since the autoxidation and 5-exo ring closures proceed with no stereochemical control (Figure 5), resulting in a total of 64 F $_2$ -IsoPs that can be formed during peroxidation of arachidonic acid [67].

### 3.3. Interaction of LP Products with Amino Acids and Proteins

Carbonyl secondary products show high reactivity with cell nucleophiles, namely DNA and proteins [6,58]. The reaction with proteins is particularly noteworthy as it interferes with their biological functions in both ways and can express either cytoprotective or damaging roles [6,18]. The most important nucleophiles in peptides are thiols (e.g., the -SH of cysteine residues) and amines (e.g., the side chain - $\text{NH}_2$  of lysine, the imidazolyl residue in histidine, the guanidine residue in arginine) [69]. Taking conjugated alkenals like 4-HNE as a model, amino groups preferably undergo Schiff-base formation with the carbonyl, while other nucleophiles preferably undertake 1,4-Michael-type addition to the C=C; the subsequent reaction of 4-HNE with both an amine residue and a thiol residue of different proteins can result in protein cross-linking [6,58], as exemplified in Figure 8. Second-order rate constants for the reaction of 4-HNE and 4-ONE with model amino acid derivatives shows that 4-ONE is more reactive by two orders of magnitude with thiols (e.g., reaction with GSH at 23 °C, pH 7.4:  $k_2$  is 1.33 and  $150 \text{ M}^{-1}\text{s}^{-1}$ , for 4-HNE and 4-ONE, respectively), while it is only about one order of magnitude more reactive with other nucleophiles (e.g., reaction with N-acetylhistamine at 23 °C, pH 7.4:  $k_2$  is  $2.1 \times 10^{-3}$  and  $2.2 \times 10^{-2} \text{ M}^{-1}\text{s}^{-1}$ , for 4-HNE and 4-ONE, respectively) [69]. Protein alkylation is the main process responsible for the biological activity of LP secondary products (vide infra).

Interestingly, in parallel to the influence of lipid oxidation products in the structure and functions of proteins, oxidative damage to peptides and proteins, particularly concerning cysteine and methionine residues, would cause radical damage to lipids, altering their structure and functions [70–73].

A main mechanism for peptide-to-lipid damage transfer was extensively investigated in Chatgililoglu-Ferreri's group and consists of cis–trans isomerization of C=C double bonds in MUFA and PUFA, caused by reversible addition of thiyl radicals ( $\text{RS}\bullet$ ) and sulfhydryl radicals ( $\text{HS}\bullet/\text{S}^{-\bullet}$ ) generated from cysteine, as exemplified in Figure 8E. Thiyl radicals undergo fast addition to cis C=C double bonds with a bimolecular rate constant of  $1.6 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$  (for methyl oleate), but the adduct undergoes rapid fragmentation, which yields the trans isomer approximately 10-fold faster than the cis isomer (viz.  $1.7 \times 10^7 \text{ s}^{-1}$  vs.  $1.6 \times 10^8 \text{ s}^{-1}$  for methyl oleate), thereby converting the less stable cis isomer of the lipids into the stable trans isomer [71]. Although formally this process does not belong to LP, as the lipids have not changed their oxidation state, isomerization, occurring, for instance, in cell membrane phospholipids, alters membrane function similarly to LP (vide infra) by affecting its fluidity; additionally, the two processes are interconnected, and both are involved in the development and control of ferroptosis [74].



**Figure 8.** Examples of lipid–peptide interaction in peroxidation: (A) favorite nucleophilic attack sites in 4-HNE; (B) resulting Cys-HNE adduct; (C) Lys-HNE adduct; (D) protein cross-linking by 4-HNE; (E) lipid *cis*–*trans* isomerization caused by the reversible addition of Cys-derived thiyl radical.

#### 4. Biological Consequences of Lipid Peroxidation

Lipid peroxidation, oxidative damage to other essential biomolecules, like proteins and DNA, and the generation in biological systems of oxidizing radicals not adequately balanced by antioxidant defenses have long been associated with different degenerative pathologies, particularly with cardiovascular diseases and cancer [1–3], and have been referred to as oxidative stress (OS), although the actual causal interplay has long escaped full rationalization. Understanding has enormously progressed in recent years and a number of valuable reviews have addressed it [4,6,10,14,15,18,57]. Therefore, we will limit this section to a brief overview, aimed at highlighting the potential of antioxidant strategies, presented in Section 5.

It is now recognized that LP is necessary to cell physiology, to maintain redox homeostasis, and it has essential signaling functions and protective roles. However, dysregulation causes alteration of cell metabolism and/or cell death and it is involved in a multitude of pathologic conditions.

##### 4.1. LP and Membrane Integrity and Functions

Modification of cell membranes is the most direct biological consequence of LP. Accumulation of hydroperoxides in the phospholipid bilayer alters their physical–chemical character by increasing the polarity of phospholipids and by altering their aggregation and hence membrane structure. Studies on giant unilamellar vesicles (GUVs) as model systems showed that accumulation of hydroperoxides causes an increase in the area which is proportional to the extent of peroxidation, and it is accompanied by a decrease in thickness of the membrane. Additionally, changes in the liquid–gel transition temperature and phase separations in the lipid domain were observed [75]. Oxidation of cholesterol extends those changes but, alone, is insufficient to cause them. On extending the oxidation and releasing small-chain secondary products, permeability increases, and the integrity is compromised with creation of pores [75]. The mechanical properties of the membrane are significantly altered, with the stretching module decreasing linearly with the conversion into peroxidized lipids, from  $200 \text{ mN m}^{-1}$  for native lipids to  $50 \text{ mN m}^{-1}$  for fully oxidized bilayers [76]. Mechanical changes and increases in the surface area were judged to be the consequence of changes in phospholipid chain conformation, caused by the introduction of polar -OOH groups, with increased probability to settle at the lipid–water interface [75]. When changes to membrane structure are sufficiently extended to cause pores or altered

permeability, the unavoidable consequence is cell death, which is one of the main effector strategies of ferroptosis (vide infra).

#### 4.2. LP and Cell Signaling

ROS and LP electrophiles (LPEs) are two main families of players in non-enzymatic cell signaling; among them, LPEs have a prominent role due to generally higher persistency and ability to diffuse from the immediate surroundings of the generation site [57]. Their main signaling mechanism is by forming protein adducts. Due to the different location of the reacting amino acid residue and to the protein to which the amino acid pertains, this would result in modifying cell metabolism in different ways, both by activating and deactivating specific pathways [18]. At low levels (0.1–1  $\mu\text{M}$ ), 4-HNE produces adducts that promote the biosynthesis of antioxidant and detoxifying enzymes, thereby having a protective role. The main mechanism is the reaction with Cys residues of cytoplasmic Keap1 protein, which is part of the nuclear factor erythroid 2-related factor 2 (Nrf2) signaling system. A dimer of Keap1 protein normally holds the transcription factor Nrf2 in the cytoplasm, inhibiting its function. Upon alkylation by 4-HNE, a change in Keap1 conformation causes the release of Nrf2, which translocates into the nucleus and binds to Maf small protein. The resulting heterodimer binds DNA at antioxidant-responsive element (ARE) sites to upregulate genes coding for antioxidant enzymes, such as thioredoxin reductase, and for proteins that cause increased glutathione (GSH) levels [6,18]. However, at higher doses (50  $\mu\text{M}$ ), 4-HNE also creates adducts with histones, modifying protein expression at the DNA level, e.g., it blocks histone H2A acetylation, thereby impairing gene expression and contributing to the vulnerability of DNA to apoptosis. 4-HNE also affects the activity of NF $\kappa$ B transcription factor in opposite directions depending on the dose, with resulting anti-inflammatory or proinflammatory effects. The formation of MDA–protein adducts is associated with a proinflammatory reaction throughout the entire organism, via the activation of Th17 lymphocytes, which triggers autoimmune reactions [77]. Additionally, MDA leads to collagen cross-linking causing a loss of elasticity and disturbance in tissue remodeling, with systemic consequences, particularly on the blood vessel system [78]. The above are just examples of the complex pattern of influence of the metabolic machinery elicited by LPEs' signaling role.

Lipid hydroperoxides themselves are signaling factors and are key players in inflammatory processes. Besides being the primary products of LP formed in the propagation stage, they are also formed by direct oxidation of lipids by singlet oxygen (see: Section 2.1) as a form of defense from such highly reactive species and by enzymatic processes, most notably by lipoxygenases (LOXs), a family of dioxygenases converting lipids to their hydroperoxides that can further transform into other hormone-like signaling molecules (e.g., thromboxanes, leucotrienes, lipoxines and resolvines), which modulate the activity of the innate immune system (e.g., the action of macrophages) in both ways, exerting proinflammatory or anti-inflammatory roles [79]. Also, untransformed LOX-generated ROOHs primarily exert a signaling role towards the immune systems (typically proinflammatory), they directly alter membrane structure and permeability (see: Section 4.1) and might indirectly modify cell gene expression by changing the overall cell redox state [79]. LOX-generated hydroperoxides have also been attributed a key role in regulation of ferroptosis (see: Section 4.5) [80].

#### 4.3. LP Association with Cancer and Apoptosis

Association between different markers of oxidative stress and cancer has been extensively reported in recent decades. Metabolic requirements of cancer cells are higher due to their rapid proliferation, which makes mitochondrial ROS generation higher than in non-transformed cells. Redox status and ROS are also of particular importance in tumor cell signaling associated with cancer progression. Therefore, compared to healthy non-malignant cells, cancer cells have altered redox homeostasis and higher levels of LP; in many types of tumor cells, these higher levels support their growth, proliferation and

survival [81]. Therefore, higher levels of LP products and biomarkers are expected in association with cancer.

For instance, in a large population-based cohort study with 14 years of follow-up, it was found that a positive association exists between 8-isoprostane urinary levels and occurrence of lung cancer but not of other cancers. Actually, in smokers 8-isoprostane levels were inversely correlated with occurrence of prostate cancer, indicating a protective effect, possibly because higher OS levels associated with smoking, combined with testosterone, activate apoptosis, thereby protecting from cancer development in testosterone-sensitive tissues [68]. A recent study on the relationship between 4-HNE and prostate cancer based on metabolic profiling using LC-ESI-QTOF-MS and GC-EI-Q-MS revealed increased 4-HNE-protein adduct levels in the plasma of cancer patients while there were no 4-HNE-protein adducts in prostate carcinoma tissue [60]. Interestingly, while higher levels of LPEs are typically associated with cancer, some of them also promote defensive mechanisms.

A main defense mechanism against cancer development is apoptosis, a physiological mechanism of programmed cell death that can be triggered both by *death receptor* activation and by metabolic changes in the cell [64]. 4-HNE can modify the structure of the MDM2 protein, breaking down the MDM2-p53 complex. As a consequence, the p53 factor is activated and translocates to the nucleus, where it promotes the transcription of proapoptotic proteins, such as Bax and caspase-3 [66]. 4-HNE may also interact with other proteins; for example, with AKT kinase, which has inhibiting roles, thereby leading to a significant increase in apoptosis. 4-HNE and 4-hydroxyhexenal further promote apoptosis by other mechanisms, which have recently been reviewed [66].

#### 4.4. LP and Neurological Disorders

The number of deaths associated with neurological disorders including neurodegenerative diseases like amyotrophic lateral sclerosis and Parkinson's, Alzheimer's and Huntington's diseases, as well as other diseases that involve neurodegenerative processes, like diabetes, have increased worldwide with time [82]. Additionally, these diseases have an important social impact as they compromise patients' quality of life. Although the causes of neurodegeneration are often unclear, OS and LP, along with inflammation, are recognized as prevailing molecular pathways leading to these diseases [82]. Some known inducers of neurodegeneration like arabin and methylmercury have been found to cause it through induction of LP and reduction of antioxidant defenses, or at least in parallel to inducing them [82,83], acrylamide induces neurodegeneration that can be reversed by antioxidant plant essential oils, while the link between retinal degeneration and OS is supported by the protective role of cytochrome b5 overexpression, which reduces lipid peroxidation [82]. Among the mechanisms linking LP to neurodegeneration, 4-HNE was identified as a trigger and its levels in an animal model increased following induction of neurodegenerative lesions [82].

In the animal model, both Parkinson's disease and Friedreich's ataxia were associated with dysregulated mitochondrial metabolism and increased lipid peroxidation, and they could be ameliorated with antioxidants. In Huntington's disease (HD), an increase in 4-HNE levels in different brain areas was also observed [82].

Alzheimer's disease (AD) is certainly the most investigated in connection with LP. In human studies, excessive ROS generation was reported under AD conditions, and it was found to be associated with  $\beta$ -amyloid aggregation, while, in turn,  $\beta$ -amyloid plaques produce ROS and LP, and increased OS biomarkers are found in association with AD [84]. Histological analysis revealed co-localization of LP secondary products and  $\beta$ -amyloid plaques in the brain, while there is a relationship with oxidized low-density lipoprotein (LDL) levels in AD [84]. Owing to this strong association, LP markers can be used both as diagnostic tools for AD and to monitor the efficacy of AD treatments [84]. Formation of Schiff bases of the cholesterol oxidation product 3- $\beta$ -hydroxy-5-oxo-5,6-secocholestan-6-al and  $\beta$ -amyloid is known to be amyloidogenic and it was found that this modification, occurring in specific sites, has a different influence on aggregation kinetics, particularly

adducts at Lys-16, obtained at physiological levels of  $\beta$ -amyloid, enabling both kinetic and thermodynamic aggregation that are sufficient to form neurotoxic lesions [65]. Based on the above, antioxidants are among the most promising therapeutic approaches for AD. Among them, metal chelators (acting as preventive antioxidants, *vide infra*), indirect antioxidants acting through the activation of the Nrf2 transcription factor, which induce *de novo* synthesis of antioxidant enzymes, and plant-based antioxidants like curcumin, resveratrol, capsaicin, epigallocatechin gallate (EGCG) from green tea and quercetin have been more extensively investigated, with favorable clinical outcomes [15]. Naringenin, a flavanone obtained mainly from grapefruit (*Citrus paradisi*) and other citrus fruits, possesses neuroprotective activity, along with anti-inflammatory effects, and it has beneficial effects on learning and memory in the AD model, through the mitigation of lipid peroxidation [15].

#### 4.5. LP and Ferroptosis

Ferroptosis, first identified by Stockwell in 2012 [8], is the last of several forms of programmed cell death, differing from apoptosis, which have been described in recent decades. It owes its name to the initial attribution as iron-dependent oxidative cell death, promoted by a void in the antioxidant defenses based on Cys, which could be artificially induced by erastin, an inhibitor of cystine uptake working by irreversible blocking of the cystine/glutamate antiporter system ( $x_c^-$ ), and it could be prevented by ferrostatin-1 [8], an iron-chelating aromatic amine that is instead unable to inhibit apoptosis. The discovery that, in mice, the knockout of selenoenzyme glutathione peroxidase 4 (GPX4)—a specific phospholipid hydroperoxide glutathione peroxidase that differs from other GPXs preferably reducing hydrogen peroxide—would cause cell death by inducing massive ferroptosis, which would be inhibited by liproxstatin-1, allowed recognizing the key role of GPX4 as a controller of ferroptosis [14,85]. The role of iron in ferroptosis appears linked to its ability to initiate LP via the Fenton reaction (Figure 1), by decomposing phospholipid hydroperoxides, which in turn could be the product of LP or formed enzymatically, e.g., by lipoxygenase (LOX) [11]. Although it was initially identified as a key player in the definition of ferroptosis, the actual necessity or dominance of iron in control of ferroptosis remains unclear [86]. Instead, the subsequent recognition that ferrostatin-1 and lipoxstatin-1 would act essentially as chain-breaking antioxidants (*vide infra*) in the inhibition of ferroptosis [12], and that other potent antioxidants like phenoxazines, diarylamines and nitroxides would also inhibit ferroptosis, points toward the prominent role of LP in ferroptosis, besides it being initiated by iron or by other processes [11,12,86]. Other antioxidants such as vitamin E [86] and vitamin K [87] were also found to inhibit ferroptosis. In other words, it is LP that drives the cell death [11] and the many degenerative diseases that are linked to ferroptosis. Regulation of LOX biosynthesis and of the LOX-mediated accumulation of hydroperoxides was identified as a key point in the control of ferroptosis [80]; however, its actual significance has lately been challenged by showing that, in different LOX-overexpressing cells, only some well-known LOX inhibitors were able to counteract ferroptosis and, incidentally, they were all effective chain-breaking antioxidants, likely acting through inhibition of LP [88]. Ferroptosis is related to cancer, which can be modulated by ferroptosis inducers or inhibitors [89]. Ferroptosis is also associated with neurodegenerative conditions such as AD, Parkinson's disease, Friedreich's ataxia and Huntington's disease [90], cardiovascular diseases and diseases of the urinary system [91]. Therefore, modulating ferroptosis in both directions, either by triggering it or by inhibiting it with antioxidants, might offer a potent tool in the therapy of many such diseases [90,91].

### 5. Antioxidants

Antioxidants are a very heterogeneous class of compounds, small molecules and enzymes that share the task of protecting oxidizable molecules or materials from oxidative transformation. In the biological context, the reference oxidative process is lipid peroxidation (LP); therefore, antioxidants are typically defined and discussed on the basis of their ability to prevent, slow down or block LP [3,21]. Based on where and how they

interfere with the LP radical chain, antioxidants are classified as *preventive*, if they impair the initiation process, and *chain-breaking* if they block or slow down the propagation, while a new category, the *termination-enhancing* antioxidants, was recently introduced by our group, to include those molecules, such as some terpenes and terpenoids from essential oils, which act by favoring the radical-chain termination without actually impairing propagation [21,92,93].

### 5.1. Preventive Antioxidants

Antioxidants in this class are extremely heterogenous, ranging from small molecules to complex enzyme systems. They prevent the onset of LP by preventing the formation of the initial alkyl radical, which would give rise to the propagation cycle. Since triggering events can be different, their prevention occurs by a multitude of mechanisms, each addressing one specific mode of radical initiation. For instance, UV filters (sunscreens) might protect the skin by photoinitiated LP (i.e., preventing the action of photosensitizers) [21], but they would not affect radical initiation due to metal-catalyzed decomposition of peroxides and hydroperoxides (e.g., via the Fenton reaction, see Section 2.1). The latter is a prevailing mechanism of initiation, which explains the evolutionary development of enzyme systems like peroxidases aimed at clearing the biological medium of peroxides and hydroperoxides. Among them, catalase (CAT), which catalyzes the dismutation of  $H_2O_2$  into water and molecular oxygen, and thiol peroxidases like GPX, which reduce hydroperoxides and hydrogen peroxide to alcohols or water, respectively, using a cysteine-derived thiol (e.g., the tripeptide glutathione) as the sacrificial reducing agent, can be classified as preventive antioxidants, since they “fuel down” the initiation process [21]. Along this line, superoxide dismutases (SODs), clearing superoxide radicals, and glutathione reductase (GR), which reduces oxidized glutathione using NADPH as the reducing agent, can also be assigned to the same class [21].

Metal chelators are also important members of this community. They block transition metal ions like iron and copper in a less redox-active form, allowing their indispensable presence in biological systems, yet neutralizing their ability to undergo Fenton-like chemistry. They can do so in case the two states participating in the redox cycle (e.g.,  $Fe^{2+}$  and  $Fe^{3+}$ ) form with the chelator complexes with sufficiently different stability. For instance, most chelators have higher affinity for  $Fe^{3+}$  than for  $Fe^{2+}$ , therefore chelated  $Fe^{3+}$  would have a reduction potential diminished by the difference in formation free energy ( $\Delta\Delta G_{form} = \Delta G_{form}^{3+} - \Delta G_{form}^{2+}$ ) between  $Fe^{3+}$  and  $Fe^{2+}$  complexes, which would make the redox cycle thermodynamically not viable, blocking iron in the oxidized form. Examples of chelating agents of biological relevance are transferrin and ceruloplasmin [21].

### 5.2. Chain-Breaking Antioxidants

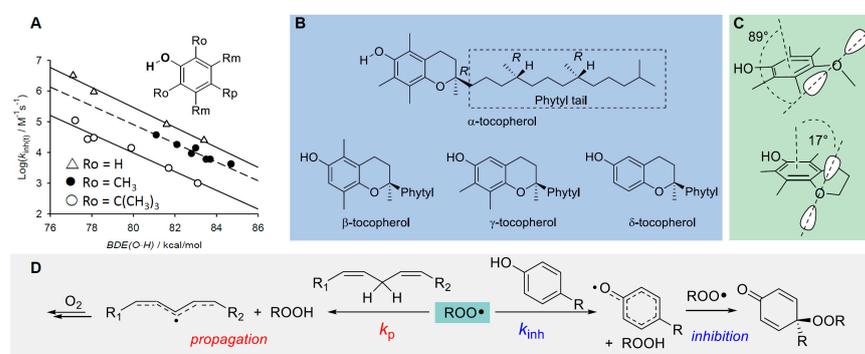
Chain-breaking antioxidants, also named radical trapping antioxidants (RTAs), are certainly the biggest class of small-molecule antioxidants in all fields of application, and *phenols* are the prototypical members [94,95]. In cells,  $\alpha$ -tocopherol ( $\alpha$ -TOH) is certainly the most important representative and the most potent lipophilic antioxidant, often used as the benchmark in antioxidant research [3]. The identification of the mechanism of its antioxidant action and of the relative reactivity of different tocopherols ( $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$ ) highlighting the parallelism between vitamin E activity *in vivo* and the relative antioxidant activity [3], allowed understanding of the structure–activity relationship governing its properties and linked vitamin E to the knowledge developed with simpler synthetic phenolic antioxidants [3,21,94–97].

RTAs act by trapping alkylperoxyl radicals, thereby competing with chain propagation. In order to be effective, the reaction of RTAs with  $ROO\bullet$  must be (much) faster than the rate of chain propagation, i.e., the rate at which peroxyl radicals attack the lipids (Equation (3)). Therefore, chain-breaking antioxidant activity works on purely kinetic bases, being a competition between two reactions. The rate constant of inhibition  $k_{inh}$  for trapping  $ROO\bullet$  by the antioxidant (Equation (3)) is the key parameter deciding the effectiveness of

an antioxidant and, since the antioxidant has normally much lower concentration than the oxidizable substrate it is called to protect (e.g., membrane lipids),  $k_{inh} \gg k_p$  is necessary. For instance, to protect linoleic acid residues ( $k_p = 62 \text{ M}^{-1}\text{s}^{-1}$ , see Table 1) with 1% antioxidant relative to lipids,  $k_{inh} \geq 10^4 \text{ M}^{-1}\text{s}^{-1}$  is required [21].

$$V_{trapping} > V_{propagation} \Rightarrow k_{inh}[Antioxidant] > k_p[Substrate] \quad (3)$$

It should be noted that only the trapping of  $\text{ROO}\bullet$  is relevant to afford antioxidant protection, as they are the sole chain-propagating species (with the exception of  $\text{HOO}\bullet$ , see Section 2.2). In the case of phenols like  $\alpha$ -TOH, the reaction with  $\text{ROO}\bullet$  occurs by formal HAT from the phenolic  $-\text{OH}$  group, to afford a stabilized phenoxyl radical  $\alpha$ - $\text{TO}\bullet$ , which is normally unable to propagate the radical chain. Instead, it is sufficiently long-lived to “wait” in solution to trap a second  $\text{ROO}\bullet$ , normally by addition to the aromatic ring, as illustrated in Figure 9. Therefore, one molecule of (mono)phenolic antioxidant can break two radical chains, or it has a *stoichiometric factor*  $n = 2$ . This is the second most important parameter in quantifying antioxidant performance.



**Figure 9.** Linear free energy correlations between rate constant for  $\text{ROO}\bullet$  trapping and  $\text{BDE}_{\text{OH}}$  for phenolic antioxidants bearing different ring substituents (A); structure of the main tocopherol congeners forming vitamin E (B); role of stereoelectronics in maximizing the ED contribution of  $-\text{O}-$  substituent included in the chroman ring or free to rotate (C); and representation of the kinetic competition involving chain-breaking antioxidants (D).

The rate of peroxidation during full inhibition by an antioxidant  $\text{AH}$  is given by Equation (4), where  $R_i$  is the rate of radical initiation.

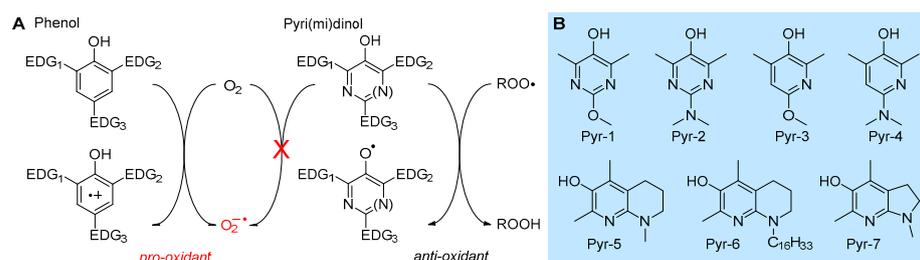
$$-\frac{d[\text{O}_2]}{dt} = \frac{d[\text{ROOH}]}{dt} = \frac{k_p[\text{RH}]R_i}{nk_{inh}[\text{AH}]} + R_i \quad (4)$$

The rate constant  $k_{inh}$  depends on the BDE of the phenolic O-H being broken and of the  $\text{ROO-H}$  bond of newly formed hydroperoxide, which is worth about 88 kcal/mol [21,94]. Hence, phenols with lower BDE would give a more exothermic and faster reaction (BDE is 77.1 kcal/mol for  $\alpha$ -TOH [94]). There exist linear free energy correlations between the BDE and  $k_{inh}$  for phenolic antioxidants, which also account for steric hindrance in *ortho* to the reactive  $-\text{OH}$ , as depicted in Figure 9 [21,94,95]. Substituents in the phenolic ring determine the reactivity with peroxy radicals, according to their electronic properties: electron-donating groups (EDGs) decrease the bond dissociation enthalpy (BDE) of the phenolic O-H, making the HAT to peroxy radicals faster, while electron-withdrawing groups (EWGs) have the opposite effects [21,94]. The effect is more marked when the substituent is in the *ortho* or *para* position, conjugated with the  $-\text{OH}$  group, while it is less important in the *meta* position and, in a first approximation, the effect of each substituent in decreasing the BDE is additive [94]. Stereoelectronic effects are also important; indeed,  $\alpha$ -TOH has lower BDE and much higher  $k_{inh}$  than equivalently substituted phenols, due to the almost parallel alignment of one lone pair of the  $\text{RO}-$  substituent with respect to the axis of the aromatic  $\pi$ -system, forced by the geometric constraints in the chroman structure

(Figure 9). The performance of synthetic or natural monophenolic antioxidants, including currently popular compounds like bakuchiol [95] and curcumin derivatives [37], can be rationalized with the above concepts. The values of BDE,  $k_{inh}$  and  $n$  for some representative antioxidants are listed in Table 2.

### 5.2.1. Insertion of N(s) in the Phenolic Ring: 3-Pyridinols and 5-Pyrimidinols

Research aimed at the rational design of phenolic antioxidants that would be more effective than  $\alpha$ -TOH, using the strategies outlined above, e.g., by inserting stronger ED substituents like the amino group and forcing its conjugation by geometrical constraints, eventually showed its limits. In parallel to decreasing the BDE and hence increasing the reactivity with ROO•, ED substituents also decrease the ionization potential (IP) of the molecule, to the point it directly reacts with molecular oxygen by electron transfer (ET) to yield the superoxide radical ( $O_2^{\bullet-}$ ) (Figure 10). Therefore, these compounds are pro-oxidant and toxic, besides being terribly unstable under normal handling conditions [98]. In a joint effort with Pratt's group, we found that replacing one or two >CH moieties in the aromatic ring with nitrogens in non-conjugated positions (i.e., 3 and 5), to afford the corresponding 3-pyridinols and 5-pyrimidinols, would increase the BDE<sub>OH</sub> by about 1.1 kcal/mole per nitrogen ( $\Delta$ BDE of +1.1 kcal/mol and +2.5 kcal/mol for 3-pyridinols and 5-pyrimidinols, respectively), while the IP would increase 10-fold more ( $\Delta$ IP of +11 kcal/mol and +24 kcal/mol for 5-pyrimidinol) [98,99]. In other words, insertion of heteroaromatic nitrogen(s) expands the thermodynamic gap between the two competing reaction pathways, HAT to ROO• and ET to  $O_2$ , while maintaining the same substituent effects known for phenols [99]. Two features favored this strategy in antioxidant design: (1) the slightly higher BDE<sub>OH</sub> that would disfavor the reactivity can be compensated with stronger ED substituents, like the amino group (inaccessible in the phenolic series), without compromising the stability toward oxygen [99]; (2) 3-pyridinols and 5-pyrimidinols are more reactive toward ROO• than phenols having an identical BDE, owing to the intervention of polar effects stabilizing the transition state [100]. This led to the discovery of a wealth of potent RTAs (some examples in Table 2), many of which easily outperformed  $\alpha$ -TOH despite the much simpler structure and easier synthetic accessibility [98–106], which also included the most potent chain-breaking antioxidant ever reported, able to quench peroxy radicals at a diffusion-controlled rate (Pyr-7 in Figure 10) [101].

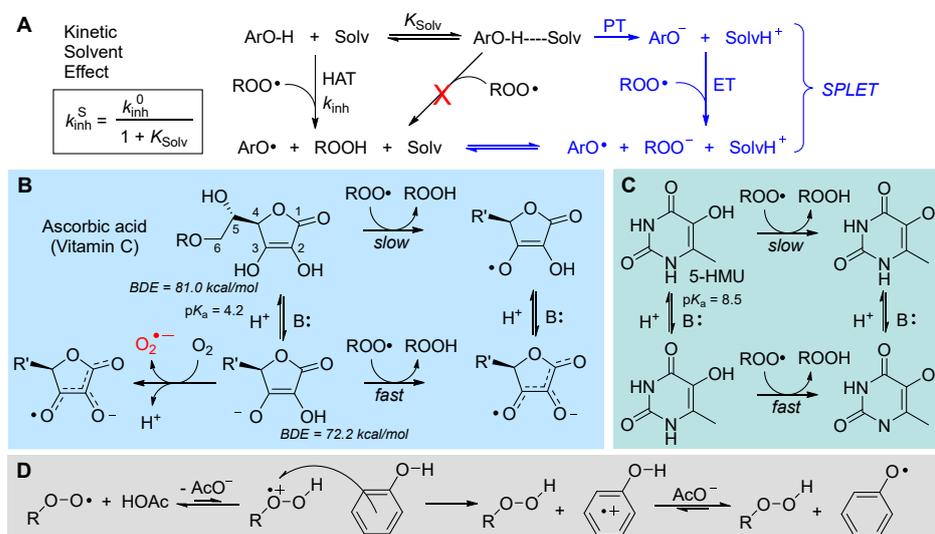


**Figure 10.** Different reactivity with oxygen and ROO• of electron-rich phenols and pyri(mi)dinols (A) and examples of 3-pyridinol and 5-pyrimidinol antioxidants (B).

### 5.2.2. Solvent and Medium Effects in Chain-Breaking Antioxidant Activity

The rate of radical trapping by phenols and most other RTAs is subject to solvent effects [107], which must be carefully considered in testing antioxidant activity [108] or in planning antioxidant strategies. In hydrogen-bond-accepting (HBA) solvents,  $k_{inh}$  is apparently decreased by as much as two orders of magnitude, depending on the actual HBA ability of the solvent (not its polarity) [107]. The effect does not depend on the radical reacting with the antioxidant (e.g., R•, RO•, ROO•) but on the ability of the antioxidant to act as a hydrogen-bond donor (HBD) to the solvent [109], since the effect is due to formation of an antioxidant–solvent complex in which the “active” -OH group (or, in general, the -XH group transferring the H-atom to radicals) is “blocked” by H-bonding to the solvent,

and only the fraction of non-H-bonded antioxidant at equilibrium is available to react with radicals (Figure 11) [106,107]. Different antioxidants will have different sensitivity to solvent effects.



**Figure 11.** Solvent effects on the reactivity of antioxidants, showing the possible mechanistic change in protic solvents (A), pH-dependent antioxidant chemistry of ascorbic acid (B) and of 5-hydroxymethyluracil (C) and acid catalysis in the antioxidant activity of phenols in polar solvents (D).

The HBD ability of the antioxidant and the HBA ability of the solvent can be quantified by Abraham's solvatochromic parameters  $\alpha_2^H$  and  $\beta_2^H$ , respectively [110,111], and solvent effects can be accounted for quantitatively by the Ingold–Snelgrove equation (Equation (5)), which allows predicting  $k_{inh}$  in any solvent ( $k_{inh}^S$ ) once it is known in a non-H-bonding solvent ( $k_{inh}^0$ ) [112].

$$\text{Log} \left( k_{inh}^S / M^{-1} s^{-1} \right) = \text{Log} \left( k_{inh}^0 / M^{-1} s^{-1} \right) - 8.3 \times \alpha_2^H \beta_2^H \quad (5)$$

Triglycerides [24] and phospholipids [107] have strong HBA groups (i.e., the C=O and P=O), which visibly decrease the antioxidant protection by RTAs. In a heterogenous system of lipid particles in water, however, an additional factor affects the kinetics of antioxidant protection: the compartmentalization, which may render the rate of exchange of radicals and antioxidants among particles rate limiting. For instance, the value of  $k_{inh}$  for  $\alpha$ -TOH measured in micelles of methyl linoleate and in phospholipid liposomes is, respectively, two orders of magnitude and three orders of magnitude lower than in homogenous non-H-bonding solution [21,107].

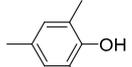
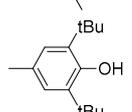
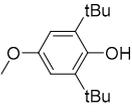
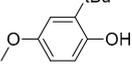
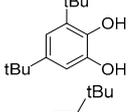
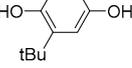
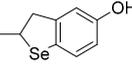
The decrease in the rate of formal HAT reaction to radicals in HBA solvents could favor other reaction mechanisms [113,114]. It has been shown that in protic solvents like alcohols, able to solvate anions, the reaction of acidic phenols (or other antioxidants) with radicals could take place via a mechanism named sequential proton loss electron transfer (SPLET), depicted in Figure 11, consisting in proton transfer (PT) to the solvent followed by ET from the electron-rich anion of the antioxidant to oxidizing radicals [113]. In water, this mechanism is favored in alkaline pH and can result in an increase in the rate of peroxy radical trapping [114].

This change in the reaction mechanism does not occur only with phenols but controls the antioxidant activity of other biological antioxidants like 5-hydroxymethyluracil (5-HMU) [115] and, most notably, of *ascorbic acid* and its derivatives [116,117], boosting their performance in the presence of a base (Figure 11). In the case of ascorbic acid, deprotonation also opens the way to its direct reaction with  $O_2$  to yield superoxide radicals, accounting for its instability in aqueous solution. This is also a pro-oxidant process that partly counteracts

the antioxidant behavior; as a consequence, the overall antioxidant performance of ascorbic acid largely depends on the experimental settings, and it can be significantly improved by inclusion in inert nanocarriers [117].

Opposite to the base effect outlined above, phenolic (and pyridinolic/pyrimidinolic) antioxidants have been found to undergo acid catalysis in their antioxidant activity. The behavior is only observed in polar solvents and can largely boost the reactivity, depending on the phenol. It is due to a change in reaction mechanism involving the partial protonation of the ROO• radical to form a highly oxidizing species that takes one electron from the phenol and forms the antioxidant's radical cation, which undergoes rapid acid–base exchange to afford the usual reaction products [118], as illustrated in Figure 11.

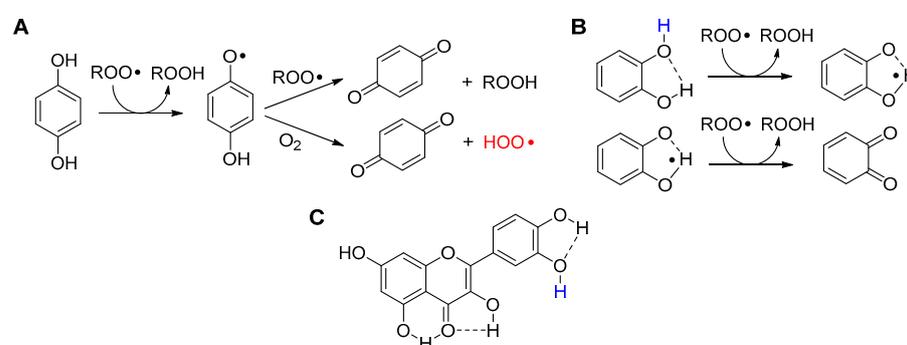
**Table 2.** Bond dissociation enthalpy (BDE), rate constants for ROO• radical trapping  $k_{inh}$  and stoichiometric factor  $n$  for representative antioxidants at 303 K in chlorobenzene.

Entry	Compound	BDE kcal/mol	$k_{inh}$ M <sup>-1</sup> s <sup>-1</sup>	$n$	Ref.
1	$\alpha$ -Tocopherol	77.1	$3.2 \times 10^6$	2.0	[3,21]
2	$\beta$ -Tocopherol	-	$1.3 \times 10^6$	2.0	[3,21]
3	$\gamma$ -Tocopherol	-	$1.4 \times 10^6$	2.0	[3,21]
4	$\delta$ -Tocopherol	-	$4.4 \times 10^5$	2.0	[3,21]
5		81.7	$2.7 \times 10^5$	2.0	[21,94]
6		81.6	$8.5 \times 10^4$	2.0	[21,94]
7		79.9	$1.4 \times 10^4$	2.0	[21,94]
8		77.2	$1.1 \times 10^5$	2.0	[21,94]
9		80.3	$6.4 \times 10^5$	1.8	[21,94]
10		78.2	$1.1 \times 10^6$	2.0	[94,119]
11		79.2	$1.6 \times 10^{6.1}$	0.3	[120]
12	Quercetin	-	$5.5 \times 10^5$	2.1	[119]
13	Pyr-1 <sup>2</sup>	81.4	$2.1 \times 10^5$	2.0	[100]
14	Pyr-2 <sup>2</sup>	77.1	$8.6 \times 10^6$	2.0	[100]
15	Pyr-3	78.9	$4.4 \times 10^5$	2.1	[106]
16	Pyr-4	75.9	$1.6 \times 10^7$	2.0	[101]
17	Pyr-5	75.2	$8.8 \times 10^7$	1.3–2.0	[101]
18	Pyr-6	75.2	$8.8 \times 10^7$	~2	[103]
19	Pyr-7	74.3	$2.8 \times 10^8$	~2	[101]
20	$\alpha$ -Selenotocopherol	78.1	$1.2 \times 10^6$	1.9	[121]
21		81.6	$3.8 \times 10^5$	2.0	[122]
22	Te-1	-	$9.2 \times 10^6$	0.4 <sup>6</sup>	[123]
23	Te-2	78.9	$1.0 \times 10^7$	0.4 <sup>6</sup>	[124]
24	Te-3	-	$1.6 \times 10^6$	0.3 <sup>6</sup>	[125]
25	Te-4	-	$1.0 \times 10^7$	0.4 <sup>6</sup>	[125]
26	Phenoxazine <sup>2,3</sup>	76.1	$2.9 \times 10^7$	5	[94]
27	Phenothiazine <sup>2,4</sup>	78.2	$8.8 \times 10^6$	1.8	[94]
28	Dia-1	78.8	$3.4 \times 10^7$	>2	[126]
29	Dia-2	79.0	$3.7 \times 10^7$	>2	[126]
30	Ferrostatin-1 <sup>5</sup>	-	$3.5 \times 10^5$	2.0	[11]
31	Liproxstatin-1 <sup>5</sup>	-	$2.4 \times 10^5$	1.9	[11]

<sup>1</sup>  $k_{inh}$  is doubled by a statistical factor. <sup>2</sup> Data at 50 °C. <sup>3</sup> Values at 37 °C are:  $k_{inh}$   $4.1 \times 10^7$  M<sup>-1</sup>s<sup>-1</sup>,  $n = 2.3$  from ref. [11]. <sup>4</sup> Values at 37 °C are:  $k_{inh}$   $8.0 \times 10^6$  M<sup>-1</sup>s<sup>-1</sup>,  $n = 2.1$  from ref. [11]. <sup>5</sup> Data at 37 °C. <sup>6</sup> Without thiols.

### 5.2.3. Polyphenols and Flavonoids

Hydroquinone (1,4-dihydroxybenzene) and catechol (1,2-dihydroxybenzene) are the two key structural motifs in polyphenols and flavonoids. Even the simple unsubstituted molecules have high reactivity with  $\text{ROO}\bullet$  radicals due to the strong ED character of each -OH group toward the other in the conjugated position [21,120]. Both hydroquinones and catechols are nominally able to trap two  $\text{ROO}\bullet$  radicals by stepwise transfer of the two O-H. However, their relative position (*ortho* or *para*) deeply affects the antioxidant behavior [120], as shown in Figure 12. In hydroquinones (e.g., ubiquinol or coenzyme  $\text{QH}_2$ ), the semiquinone radical formed upon transfer of the first H-atom has a very low BDE for the second O-H ( $\sim 50$  kcal/mol) and it is rather acidic ( $\text{pK}_a \sim 4$ ) [120]; therefore, via different mechanisms which have been discussed in detail [120] the semiquinone radical can react directly with  $\text{O}_2$  to form a hydroperoxyl radical ( $\text{HOO}\bullet$ ) or superoxide radical anion ( $\text{O}_2^{\bullet-}$ ) in lipidic medium or in water, respectively. This process, which parallels the generation of superoxide in the mitochondrial respiratory chain, has pro-oxidant action starting new oxidative chains; therefore, it partly counteracts (by shortening  $n$ ) the otherwise very good antioxidant behavior of hydroquinones [120].



**Figure 12.** Radical reactivity of hydroquinone (A), of catechol (B) and structure of quercetin (C).

In catechols, the intramolecular H-bond makes one -OH unreactive, but it decreases the BDE of the unbound O-H, increasing its reactivity. The resulting semiquinone radical is stabilized by the intermolecular H-bonding, which prevents its reaction with  $\text{O}_2$ . Overall, even the unsubstituted catechol traps two peroxy radicals with  $k_{\text{inh}}$  similar to  $\delta$ -TOH, being an excellent antioxidant [119,127]. Not surprisingly, the catechol moiety is highly conserved in natural flavonoids (Figure 12) and, in a lipidic environment, their antioxidant activity is largely dictated by the catechol ring. Indeed, despite the large number of phenolic -OH groups in their structure, the reactivity of most polyphenols is modulated by the occurrence of intramolecular H-bonds [128]. As a consequence, flavonoids can trap more than two  $\text{ROO}\bullet$  radicals, but with different rate constants  $k_{\text{inh}}$ , hence the trapping of the first two contributes most to the overall antioxidant performance [129]. In an aqueous environment, their reactivity is instead dictated by pH, and multiple mechanisms (PT-ET vs. HAT) can take place, as previously discussed in detail for quercetin [127].

### 5.2.4. Synergy among Antioxidants and Tocopherol-Mediated Peroxidation (TMP)

In nature, as in the protection of human-made materials, antioxidants are never used alone and cooperative effects among antioxidants are often the key to successful protection. The most relevant example in cells is certainly the synergistic interplay between vitamin E and vitamin C.  $\alpha$ -TOH is confined in the lipid bilayers and protects phospholipids from  $\text{ROO}\bullet$  attack, being oxidized to an  $\alpha$ -TO• radical, while water-soluble ascorbate reacts with  $\alpha$ -TO• at the lipid–water interface, reducing it back to the starting  $\alpha$ -TOH and extending its protective duration [130]. On doing so, ascorbate is oxidized to the corresponding radical; therefore, besides the recycling of  $\alpha$ -TOH, the importance of this process resides in the “exporting of the unpaired electron” outside the lipid membrane, out of the reach of highly oxidizable lipid components. Indeed, it has been demonstrated by Bowry and

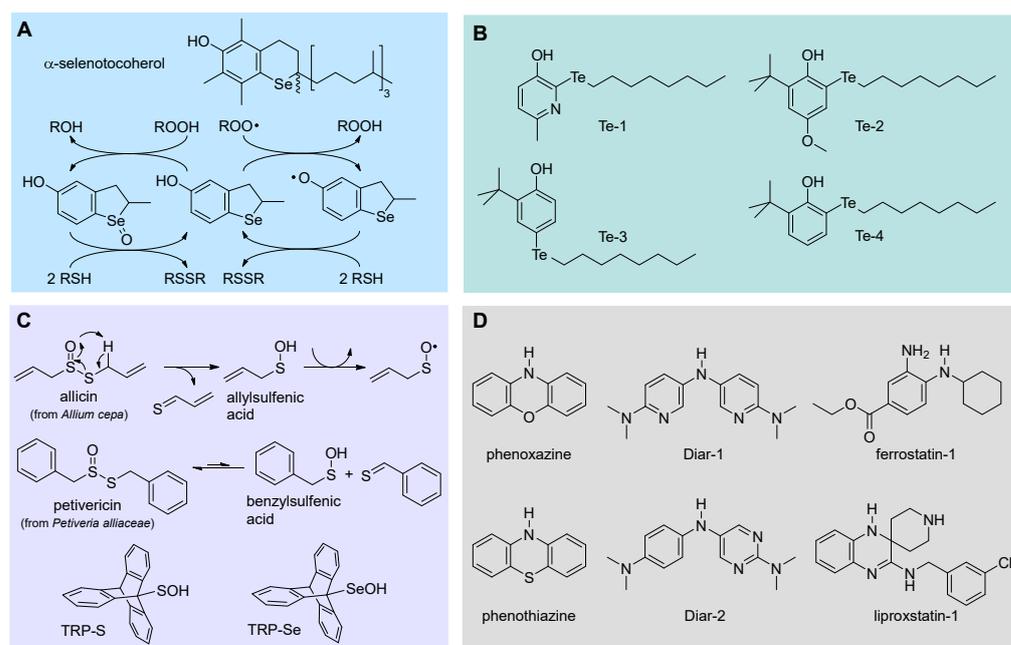
Ingold [131] that, in the autoxidation of human low-density lipoprotein (LDL), in the absence of ascorbate in the aqueous medium, the presence of  $\alpha$ -TOH in the lipid core accelerates the peroxidation instead of blocking it—an apparently paradoxical behavior that they named tocopherol-mediated peroxidation (TMP)—due to the “excessive” resident time of  $\alpha$ -TO• inside the lipid particle before it encounters a second ROO•, which gives time for the occurrence of even a slow and unfavored reaction of  $\alpha$ -TO• with lipids, to (re)start the oxidative chain [131].

Interestingly, when pyridinol Pyr-6 was probed in place of  $\alpha$ -TOH in the protection of LDL in the absence of vit C or co-antioxidants, it showed no sign of TMP, along with much higher antioxidant protection [103]. Cooperative effects similar to the vit E/vit C example also occur among phenolic and/or other antioxidants in homogenous solution, and their mechanism and efficiency have been thoroughly investigated [106,132].

### 5.2.5. Phenols Bearing Organochalcogen Substituents

The insertion of chalcogens heavier than oxygen (S, Se and Te) as substituents in the *ortho* or *para* position in phenolic (and pyridinolic) structures is an important strategy to afford novel antioxidants with a distinctive reactivity toward oxidizing radicals [121,133]. It has highlighted how intramolecular H-bonding can be used to afford antioxidants whose reactivity can predictably be modulated by conformational constraints and by the solvent [119]. Notable examples are lipoic acid adducts of natural catechols like hydroxytyrosol or ditocopheryl sulfides and disulfides [121].

Phenolic antioxidants cannot be recycled by thiols (unlike by ascorbate), which somewhat “wastes” the most abundant reducing source in biological systems. Looking for antioxidants that would boost this ability, we carried out a long cooperative project with Engman’s group focused on Se- and Te-substituted phenols. All-*rac*- $\alpha$ -selenotocopherol, in which the chromanol -O- was replaced by -Se-, disappointingly showed slightly higher BDE<sub>OH</sub> and slightly lower  $k_{inh}$  than the natural counterpart, with no additional property toward thiols [121]. However, some of its simplified congeners like Se-1 (Figure 13), besides being potent chain-breaking antioxidants, could be recycled by *N*-acetylcysteine (NAC) at the water–lipid interface in a biphasic lipid peroxidation model system [122].



**Figure 13.** Examples of organochalcogen-containing phenolic antioxidants (A,B), sulfenic and selenenic acids (C) and diarylamine antioxidants (D).

Insertion of alkyltelluro (R<sub>Te</sub>-) substituents in the *ortho* or *para* position in the phenol or in the 3-pyridinol structure affords even more interesting antioxidants (Figure 13). These compounds can be recycled by thiols like cysteine derivatives both in homogenous lipid solution and heterogenous (aqueous biphasic) systems [123–125]. Additionally, even simple structures have a  $k_{inh}$  for trapping ROO• radicals much higher than reference  $\alpha$ -TOH and their reactivity does not respond to the correlation with the BDE<sub>OH</sub> reported in Figure 9 or to the solvent effect described in Figure 11. This proves a different reaction mechanism involving an oxygen-atom transfer from ROO• to R<sub>Te</sub>-, for which we refer to the original literature [125]. Interestingly, the efficiency of regeneration by thiols is much lower than 1, meaning that the same compounds could also be used as pro-oxidant agents in biological systems, acting through the depletion of Cys-derived thiols like glutathione [125]. This is particularly intriguing in the light of the role that this mechanism has in triggering ferroptosis [8].

#### 5.2.6. Sulfenic and Selenenic Acids

*Allium* plants have long been regarded as a rich source of antioxidants, particularly owing to the distinctive content of sulfurated volatile components. Among them, thio-sulfonates like allicin and petivericin (Figure 13) have been demonstrated to be potent antioxidants [134,135]. This is due to the release of unstable sulfenic acids (RSOH), which would rapidly trap ROO• radicals to afford stabilized sulfenyl radicals (RSO•). Owing to their instability that makes their isolation prohibitive, the reactivity of sulfenic acids has remained largely unknown, despite their major relevance in biological systems, since they are involved in cellular redox homeostasis, being formed upon oxidation (e.g., by H<sub>2</sub>O<sub>2</sub>) and fragmentation of cystine. Only recently did the synthesis of stable tripticonesulfenic acid (TRP-S, Figure 13) shed light on their antioxidant chemistry [134]. With a BDE<sub>OH</sub> as low as 71.9 kcal/mol, they trap ROO• radicals with  $k_{inh}$  in the range of  $3 \times 10^6 \text{ M}^{-1}\text{s}^{-1}$  from sterically hindered TRP-S to  $3 \times 10^7 \text{ M}^{-1}\text{s}^{-1}$  for benzylsulfenic acid from petivericin and up to  $\sim 1 \times 10^8 \text{ M}^{-1}\text{s}^{-1}$  for allylsulfenic acid from allicin, i.e., they are among the most potent antioxidants in nature [135].

Their heavier homologues, the selenenic acids (RSeOH), are involved in the redox cycle of the GPX enzyme; however, they are even less stable than sulfenic acids. Synthesis of hindered tripticoneselenenic acid (TRP-Se, Figure 13) allowed clarifying their properties and reactivity, showing a higher BDE<sub>OH</sub> (80.9 kcal/mol) compared to sulfenic acids, yet having unexpectedly high reactivity toward ROO•, i.e.,  $k_{inh} = 1.7 \times 10^5 \text{ M}^{-1}\text{s}^{-1}$  for sterically hindered TRP-Se (in PhCl at 30 °C) [136]. This is 18-fold lower than that of TRP-S despite the 9 kcal/mol higher BDE [136]. Unfortunately, the instability of these compounds hampers their use as antioxidants, unless they can be generated in situ from suitable precursors.

#### 5.2.7. Aromatic Amines and Diarylamines as RTAs

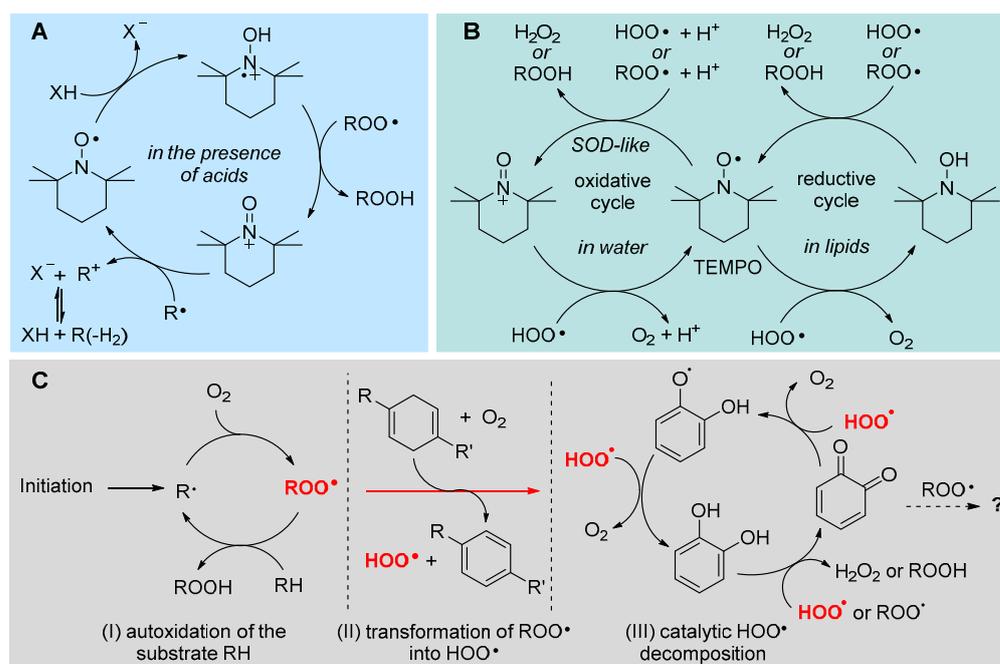
Aromatic amines such as diarylamines are, after phenols, the main class of chain-breaking antioxidants. Owing to the higher thermal stability and lower reactivity compared to phenols, the most common substituted diphenylamines have mainly been used as antioxidants in high-temperature processes. However, their tricyclic analogues, such as phenoxazine and phenothiazine, have instead very high antioxidant activity at close to ambient temperature, owing to better conjugative stabilization of the aminyl radical formed upon quenching ROO• [21]. Insertion of heterocyclic nitrogens in the structure of diphenylamine, i.e., extending the same design strategy previously described for phenols (see Section 5.2.1), has created a large family of outstanding antioxidants, containing up to four heterocyclic nitrogens per molecule and a wealth of ED substituents, with large structural variability [98,126,137,138]. Many such molecules largely outperform reference  $\alpha$ -TOH in terms of  $k_{inh}$ , in some cases having very low to negligible activation energy for ROO• trapping [126], implying negligible temperature dependence of the antioxidant activity [98]. Perhaps most notably, they outperform  $\alpha$ -TOH in terms of the number of trapped ROO• radicals, having large stoichiometric factors that depend

on temperature, due to a regeneration cycle involving the transient formation of the corresponding nitroxyl radicals [49].

Interestingly, recent work from Pratt's group has demonstrated that several antioxidants belonging to this class, namely diarylamines, phenoxazines and phenothiazines, including those displayed in Figure 13, are potent inhibitors of ferroptosis, able to match or surpass the performance of reference inhibitors liproxtatin-1 and ferrostatin-1 (Figure 13), which incidentally also belong to the class of aromatic amines [11].

#### 5.2.8. Unconventional Antioxidant Mechanisms and HOO• as Co-Antioxidant

Sterically hindered dialkyl nitroxides like the 2,2,6,6-tetramethylpiperidin-1-oxyl radical (TEMPO) are sufficiently persistent to be manipulated like a "normal" molecule, which has allowed an enormous body of investigation. Among the many applications, a wealth of studies have shown a beneficial activity in a multitude of pathological conditions commonly associated with oxidative stress, which has suggested an antioxidant activity that has long escaped rationalization [139,140]. In water, TEMPO was demonstrated to behave as a SOD mimic, being able to decompose superoxide and peroxy radicals via a catalytic cycle with the intermediation of the oxoammonium ion [141,142] (Figure 14). However, early knowledge indicated that nitroxides are unable to trap peroxy radicals in a lipidic environment and have negligible antioxidant activity toward lipids.



**Figure 14.** Non-conventional antioxidant mechanisms involving persistent nitroxides in acidic medium (A), or in the presence of HOO• (B), or involving *ortho*-quinones in the presence of HOO• (C).

We were surprised to find that they can indeed be excellent antioxidants in lipidic or organic media, but only in the presence of acids [143]. Of interest, depending on the strength of the acid, they could behave as stoichiometric antioxidants (trapping one radical per nitroxide) or they could be regenerated and work in a catalytic fashion if the acid is weak and forms a nucleophilic conjugated base, like acetic acid [144]. This chemistry, summarized in Figure 14, clearly helps explain their redox-related bioactivity, as carboxylic acids are abundant in biological systems, e.g., in proteins [144]. Even more surprising was the recent discovery of another catalytic antioxidant cycle of nitroxides, which is only expressed in lipidic or apolar environments, although it apparently resembles the SOD mimic behavior expressed only in water. The overall catalytic cycle occurs only in the presence of HOO• radicals (the neutral form of superoxide) and it does not involve the intermediacy of the oxoammonium ion from the nitroxide [145]. It was clarified, independently by our

group and Pratt's group, that the cycle occurs by *reduction* of the nitroxide ( $>NO\bullet$ ) to the parent hydroxylamine ( $>NOH$ ), which is then reoxidized to the nitroxide by  $HOO\bullet$  or by  $ROO\bullet$  [49,145] (Figure 14). Therefore,  $HOO\bullet$ , normally regarded as an oxidizing chain-carrying radical, here works as sacrificial *reducing* agent or as a *co-antioxidant*. It can be formed spontaneously during the peroxidation of lipids (see Section 2.4.3), or a dedicated source like cyclohexadiene or  $\gamma$ -terpinene can be added along with the nitroxide, to form a co-antioxidant system [145]. Interestingly, this co-antioxidant system is among the most potent ever reported, largely outperforming  $\alpha$ -TOH [145]. This chemistry is likely to have major relevance in the control of ferroptosis and some preliminary success in this direction has recently been reported [12].

Unfortunately, no nitroxide is currently approved for medicinal use; however, we were intrigued to find that *ortho* quinones, the oxidized "waste" of catechol-type polyphenolic antioxidants, can undergo a similar chemistry (Figure 14). In the presence of spontaneously generated  $HOO\bullet$ , or of an added source, *ortho*-quinones (*o*-Q) (more efficiently than *para*) are stepwise reduced to the semiquinone radical (*o*-QH $\bullet$ ) then to the catechol (*o*-QH<sub>2</sub>), which can then perform its antioxidant action trapping both  $HOO\bullet$  and  $ROO\bullet$ . Albeit not as efficient as that involving nitroxides, the synergic combination of *o*-Q/ $\gamma$ -terpinene or *o*-QH<sub>2</sub>/ $\gamma$ -terpinene has shown superior protection of highly oxidizable polyunsaturated lipids [146]. The kinetic aspects regulating this chemistry have been clarified and found to be key to the purported antioxidant behavior of melanin biopolymers [47]. Possibly, this chemistry would show its potential in the control of ferroptosis, an issue currently under investigation in our group.

### 5.3. Termination-Enhancing Antioxidants

Some hydrocarbons, aldehydes and other simple highly oxidizable molecules, such as many terpenic components of plant essential oils (e.g.,  $\gamma$ -terpinene, limonene, linalool and citral), when subjected to some radical source, undergo autoxidation that is characterized by very fast chain termination [147]. As a consequence, if they are mixed with other highly oxidizable substrates, they will promptly co-oxidize but increase the overall rate of chain termination, thereby reducing the overall rate of autoxidation and slowing down the degradation of the oxidizable substrate [147]. These compounds are unable to block the chain propagation, nonetheless they have antioxidant behavior, which we classified as termination-enhancing [147]. This mechanism, which is typical of many non-phenolic essential oil components is, however, less effective than the chain-breaking activity possessed, for instance, by phenolic essential oil components [148] and it largely depends on the experimental conditions. It has non-monotonic dependence on the concentration of the antioxidant and it can become pro-oxidant at a high concentration [147].

### 5.4. Indirect Antioxidants

This class is often used to group those mechanisms that cannot take place within a material or in a test tube but require a living cell or organism. Indeed, many molecules which might show antioxidant behavior in cells, e.g., by reducing the markers of lipid peroxidation or the related biological damage, do not trap peroxy radicals at a significant rate and would be unable to directly protect lipids from peroxidation; however, they are able to enhance the antioxidant defenses in a living organism, e.g., by inducing the biosynthesis of antioxidant enzymes. Typical examples are isothiocyanates from brassica vegetables [149,150], but the list can be extended to many plant-derived compounds, such as essential oil components [150,151]. Indeed, even flavonoids like quercetin, having good direct (chain-breaking) antioxidant activity, can also boost the antioxidant enzymes in cells [152], therefore direct and indirect activity can co-exist and have different weight depending on the experimental settings. One main mechanism by which indirect activity is accomplished is via the Nrf2 signaling system [150], which has been illustrated in Section 4.2.

## 6. Conclusive Remarks and Future Perspective

Research in the broad area of lipid peroxidation and antioxidant protection has taken place during the past 70 years, with variable emphasis. In the biomedical area, the disappointing outcomes of early clinical studies aimed at using antioxidants as drugs in the treatment of those degenerative conditions lacking other curative pharmacological solutions had somewhat, gradually, curbed the enthusiasm. Lack of success might have been due to a combination of factors, such as incomplete understanding of the role of oxidative stress and lipid peroxidation in physiology and disease, incomplete understanding of the double-sided antioxidant–pro-oxidant behavior of some molecules in biological systems, a misjudgment in the strategy of using an antioxidant when a pro-oxidant would have been the more effective choice and, last but not least, the choosing of antioxidants that were not fit for purpose. The recognition of ferroptosis as a new LP-based form of programmed cell death has revived the area and enormously boosted the search for molecules able to address OS-related pathologies via the modulation of ferroptosis. Now, understanding of the interplay between LP and pathophysiology is more mature and a wealth of extremely potent new antioxidants have become available. Pyridinols, diarylamines and heavy organochalcogen-substituted phenols have underexplored potential in this regard. Additionally, non-conventional antioxidant systems based on the exploitation of the endogenous  $\text{HOO}\bullet/\text{O}_2^-\bullet$  reducing system appear very promising in the modulation of ferroptosis, a research area we are currently set to address.

**Funding:** This research was funded by MUR (Rome, Italy), project FEROX, code number 20227XZKBY, call PRIN2022, and by the University of Bologna, grant RFO2022.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

1. Porter, N.A. Mechanism for the Autoxidation of Polyunsaturated Lipids. *Acc. Chem. Res.* **1986**, *19*, 262–268. [[CrossRef](#)]
2. Ingold, K.U. Peroxyl radicals. *Acc. Chem. Res.* **1969**, *2*, 1–9. [[CrossRef](#)]
3. Burton, G.W.; Ingold, K.U. Vitamin E: Application of the Principles of Physical Organic Chemistry to the Exploration of Its Structure and Function. *Acc. Chem. Res.* **1986**, *19*, 194–201. [[CrossRef](#)]
4. Ito, F.; Sono, Y.; Ito, T. Measurement and Clinical Significance of Lipid Peroxidation as a Biomarker of Oxidative Stress: Oxidative Stress in Diabetes, Atherosclerosis, and Chronic Inflammation. *Antioxidants* **2019**, *8*, 72. [[CrossRef](#)]
5. Tarozzi, A.; Bartolini, M.; Piazzini, L.; Valgimigli, L.; Amorati, R.; Bolondi, C.; Djemil, A.; Mancini, F.; Andrisano, V.; Angela Rampa, A. From the dual function lead AP2238 to AP2469, a multi-target-directed ligand for the treatment of Alzheimer’s disease. *Pharma. Res. Per.* **2014**, *2*, e00023. [[CrossRef](#)] [[PubMed](#)]
6. Foret, M.K.; Lincoln, R.; Do Cormo, S.; Cuello, A.C.; Cosa, G. Connecting the “Dots”: From Free Radical Lipid Autoxidation to Cell Pathology and Disease. *Chem. Rev.* **2020**, *120*, 12757–12787. [[CrossRef](#)]
7. Canistro, D.; Boccia, C.; Falconi, R.; Bonamassa, B.; Valgimigli, L.; Vivarelli, F.; Soleti, A.; Genova, M.L.; Lenaz, G.; Sapone, A.; et al. Redox-Based Flagging of the Global Network of Oxidative Stress Greatly Promotes Longevity. *J. Gerontol. A Biol. Sci. Med. Sci.* **2015**, *70*, 936–943. [[CrossRef](#)]
8. Dixon, S.J.; Lemberg, K.M.; Lamprecht, M.R.; Skouta, R.; Zaitsev, E.M.; Gleason, C.E.; Patel, D.N.; Bauer, A.J.; Cantley, A.M.; Yang, W.S.; et al. Ferroptosis: An Iron Dependent Form of Nonapoptotic Cell Death. *Cell* **2012**, *149*, 1060–1072. [[CrossRef](#)]
9. von Krusenstiern, A.N.; Robson, R.N.; Qian, N.; Qiu, B.; Hu, F.; Reznik, E.; Smith, N.; Zandkarimi, F.; Estes, V.E.; Dupont, M.; et al. Identification of essential sites of lipid peroxidation in ferroptosis. *Nat. Chem. Biol.* **2023**, *19*, 719–730. [[CrossRef](#)]
10. Cheng, X.; Zhang, J.; Xiao, Y.; Wang, Z.; He, J.; Ke, M.; Liu, S.; Wang, Q.; Zhang, L. Mitochondrial Regulation of Ferroptosis in Cancer Therapy. *Int. J. Mol. Sci.* **2023**, *24*, 10037. [[CrossRef](#)]
11. Shah, R.; Margison, K.; Pratt, D.A. The Potency of Diarylamine Radical-Trapping Antioxidants as Inhibitors of Ferroptosis Underscores the Role of Autoxidation in the Mechanism of Cell Death. *ACS Chem. Biol.* **2017**, *12*, 2538–2545. [[CrossRef](#)]
12. Poon, J.-F.; Zilka, O.; Pratt, D.A. Potent Ferroptosis Inhibitors Can Catalyze the Cross-Dismutation of Phospholipid-Derived Peroxyl Radicals and Hydroperoxyl Radicals. *J. Am. Chem. Soc.* **2020**, *142*, 14331–14342. [[CrossRef](#)]
13. Conrad, M.; Lorenz, S.M.; Proneth, B. Targeting Ferroptosis: New Hope for As-Yet-Incurable Diseases. *Trends Mol. Med.* **2020**, *27*, 113–122. [[CrossRef](#)] [[PubMed](#)]
14. Ursini, F.; Maiorino, M. Lipid peroxidation and ferroptosis: The role of GSH and GPx4. *Free Radic. Biol. Med.* **2020**, *152*, 175–185. [[CrossRef](#)] [[PubMed](#)]
15. Angelova, P.R.; Esteras, N.; Abramov, A.Y. Mitochondria and lipid peroxidation in the mechanism of neurodegeneration: Finding ways for prevention. *Med. Res. Rev.* **2021**, *41*, 770–784. [[CrossRef](#)]

16. Gabbanini, S.; Matera, R.; Valvassori, A.; Valgimigli, L. Rapid liquid chromatography–tandem mass spectrometry analysis of 4-hydroxynonenal for the assessment of oxidative degradation and safety of vegetable oils. *Anal. Chim. Acta* **2015**, *869*, 50–58. [[CrossRef](#)]
17. Yang, S.; Takeuchi, M.; Friedrich, H.; van Duynhoven, J.P.M.; Hohlbein, J. Unravelling mechanisms of protein and lipid oxidation in mayonnaise at multiple length scales. *Food Chem.* **2023**, *402*, 134417. [[CrossRef](#)] [[PubMed](#)]
18. Gegotek, A.; Skrzydlewska, A. Biological effect of protein modifications by lipid peroxidation products. *Chem. Phys. Lipids* **2019**, *221*, 46–52. [[CrossRef](#)] [[PubMed](#)]
19. Christensen, H.; Sehested, K. HO<sub>2</sub> and O<sub>2</sub> Radicals at Elevated Temperatures. *J. Phys. Chem.* **1988**, *92*, 3007–3011. [[CrossRef](#)]
20. Maillard, B.; Ingold, K.U.; Scaiano, J.C. Rate constants for the reactions of free radicals with oxygen in solution. *J. Am. Chem. Soc.* **1983**, *105*, 5095–5099. [[CrossRef](#)]
21. Valgimigli, L.; Pratt, D.A. Antioxidants in Chemistry and Biology. In *Encyclopedia of Radicals in Chemistry, Biology and Materials*; Chatgililoglu, C., Studer, A., Eds.; John Wiley & Sons, Ltd.: Chirchester, UK, 2012; Volume 3, pp. 1623–1678. [[CrossRef](#)]
22. Howard, J.A.; Ingold, K.U. Absolute rate constants for hydrocarbon autoxidation. VI. Alkyl aromatic and olefinic hydrocarbons. *Can. J. Chem.* **1967**, *45*, 793–802. [[CrossRef](#)]
23. Xu, L.; Davis, T.A.; Porter, N.A. Rate Constants for Peroxidation of Polyunsaturated Fatty Acids and Sterols in Solution and in Liposomes. *J. Am. Chem. Soc.* **2009**, *131*, 13037–13044. [[CrossRef](#)]
24. Baschieri, A.; Pizzol, R.; Guo, Y.; Amorati, R.; Valgimigli, L. Calibration of Squalene, p-Cymene, and Sunflower Oil as Standard Oxidizable Substrates for Quantitative Antioxidant Testing. *J. Agric. Food Chem.* **2019**, *67*, 6902–6910. [[CrossRef](#)] [[PubMed](#)]
25. Antunes, F.; Barclay, L.R.C.; Vinqvist, M.R.; Pinto, R.E. Determination of propagation and termination rate constants by using an extension to the rotating-sector method: Application to PLPC and DLPC bilayers. *Int. J. Chem. Kinet.* **1998**, *30*, 753–767. [[CrossRef](#)]
26. Russell, G.A. Deuterium-Isotope Effects in the Autoxidation of Aralkyl Hydrocarbons. Mechanism of the Interaction of Peroxy Radicals. *J. Am. Chem. Soc.* **1957**, *79*, 3871–3877. [[CrossRef](#)]
27. Miyamoto, S.; Martinez, G.R.; Medeiros, M.H.; Di Mascio, P. Singlet Molecular Oxygen Generated by Biological Hydroperoxides. *J. Photochem. Photobiol. B* **2014**, *139*, 24–33. [[CrossRef](#)]
28. Noguchi, N.; Nakada, A.; Itoh, Y.; Watanabe, A.; Niki, E. Formation of Active Oxygen Species and Lipid Peroxidation Induced by Hypochlorite. *Arch. Biochem. Biophys.* **2002**, *397*, 440–447. [[CrossRef](#)] [[PubMed](#)]
29. Miyamoto, S.; Martinez, G.R.; Rettori, D.; Augusto, O.; Medeiros, M.H.G.; Di Mascio, P. Linoleic acid hydroperoxide reacts with hypochlorous acid, generating peroxy radical intermediates and singlet molecular oxygen. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 293–298. [[CrossRef](#)]
30. Miyamoto, S.; Nantes, I.L.; Faria, P.A.; Cunha, D.; Ronsein, G.E.; Medeiros, M.H.G.; Di Mascio, P. Cytochrome c-promoted cardiolipin oxidation generates singlet molecular oxygen. *Photochem. Photobiol. Sci.* **2012**, *11*, 1536–1546. [[CrossRef](#)]
31. Zielinski, Z.A.M.; Pratt, D.A. Lipid Peroxidation: Kinetics, Mechanisms, and Products. *J. Org. Chem.* **2017**, *82*, 2817–2825. [[CrossRef](#)]
32. Miranda-Apodaca, J.; Hananya, N.; Velázquez-Campoy, A.; Shabat, D.; Arellano, J.B. Emissive Enhancement of the Singlet Oxygen Chemiluminescence Probe after Binding to Bovine Serum Albumin. *Molecules* **2019**, *24*, 2422. [[CrossRef](#)]
33. Lissi, E.A.; Caceres, T.; Videla, L.A. Visible chemiluminescence from rat brain homogenates undergoing autoxidation. II Kinetics of the luminescence decay. *Free Radic. Biol. Med.* **1988**, *4*, 93–97. [[CrossRef](#)] [[PubMed](#)]
34. Timmins, G.S.; dos Santos, R.E.; Whitwood, A.C.; Catalani, L.H.; Di Mascio, P.; Gilbert, B.C.; Bechara, E.J.H. Lipid peroxidation-dependent chemiluminescence from the cyclization of alkylperoxy radicals to dioxetane radical intermediates. *Chem. Res. Toxicol.* **1997**, *10*, 1090–1096. [[CrossRef](#)]
35. Fedorova, G.F.; Menshov, V.A.; Trofimov, A.V.; Vasil'ev, R.F. Facile chemiluminescence assay for antioxidative properties of vegetable lipids: Fundamentals and illustrative examples. *Analyst* **2009**, *134*, 2128–2134. [[CrossRef](#)]
36. Vasil'ev, R.F.; Veprintsev, T.L.; Dolmatova, L.S.; Naumov, V.V.; Trofimov, A.V.; Tsaplev, Y.B. Kinetics of ethylbenzene oxy-chemiluminescence in the presence of antioxidants from tissues of the marine invertebrate *Eupentacta fraudatrix*: Estimating the concentration and reactivity of the natural antioxidants. *Kinet. Catal.* **2014**, *55*, 148–153. [[CrossRef](#)]
37. Kancheva, V.D.; Dettori, M.A.; Fabbri, D.; Alov, P.; Angelova, S.E.; Slavova-Kazakova, A.K.; Carta, P.; Menshov, V.A.; Yablonskaya, O.I.; Trofimov, A.V.; et al. Natural Chain-Breaking Antioxidants and Their Synthetic Analogs as Modulators of Oxidative Stress. *Antioxidants* **2021**, *10*, 624. [[CrossRef](#)]
38. Porter, N.A. A Perspective on Free Radical Autoxidation: The Physical Organic Chemistry of Polyunsaturated Fatty Acid and Sterol Peroxidation. *J. Org. Chem.* **2013**, *78*, 3511–3524. [[CrossRef](#)]
39. Pratt, D.A.; Tallman, K.A.; Porter, N.A. Free Radical Oxidation of Polyunsaturated Lipids: New Mechanistic Insights and the Development of Peroxy Radical Clocks. *Acc. Chem. Res.* **2011**, *44*, 458–467. [[CrossRef](#)] [[PubMed](#)]
40. Howard, J.A.; Ingold, K.U. Absolute rate constants for hydrocarbon oxidation. XII. Rate constants for secondary peroxy radicals. *Can. J. Chem.* **1968**, *46*, 2661–2666. [[CrossRef](#)]
41. Maerker, G.; Haebler, E.T.; Ault, W.C. Epoxidation of methyl linoleate. I. The question of positional selectivity in monoepoxidation. *J. Am. Oil Chem. Soc.* **1966**, *43*, 100–104. [[CrossRef](#)]
42. Aliwarga, T.; Raccor, B.S.; Lemaitre, R.N.; Sotoodehnia, N.; Gharib, S.A.; Xu, L.; Totah, R.A. Enzymatic and free radical formation of cis- and trans-epoxyeicosatrienoic acids in vitro and in vivo. *Free Radic. Biol. Med.* **2017**, *112*, 131–140. [[CrossRef](#)] [[PubMed](#)]

43. Zielinski, Z.A.M.; Pratt, D.A. H-Atom Abstraction vs Addition: Accounting for the Diverse Product Distribution in the Autoxidation of Cholesterol and Its Esters. *J. Am. Chem. Soc.* **2019**, *141*, 3037–3051. [[CrossRef](#)]
44. Do, Q.; Lee, D.D.; Dinh, A.N.; Seguin, R.P.; Zhang, R.; Xu, L. Development and Application of a Peroxyl Radical Clock Approach for Measuring Both Hydrogen-Atom Transfer and Peroxyl Radical Addition Rate Constants. *J. Org. Chem.* **2021**, *86*, 153–168. [[CrossRef](#)] [[PubMed](#)]
45. Denisov, E.T. The role of triplet repulsion in alkyl radical addition to a  $7\pi$ -C-O bond and alkoxy radical addition to a  $\pi$ -C-C bond. *Kinet. Catal.* **2000**, *41*, 293–297. [[CrossRef](#)]
46. Degirmenci, I.; Coote, M.L. Comparison of Thiyl, Alkoxy, and Alkyl Radical Addition to Double Bonds: The Unusual Contrasting Behavior of Sulfur and Oxygen Radical Chemistry. *J. Phys. Chem. A* **2016**, *120*, 1750–1755. [[CrossRef](#)]
47. Guo, Y.; Baschieri, A.; Mollica, F.; Valgimigli, L.; Cedrowski, J.; Litwinienko, G.; Amorati, R. Hydrogen Atom Transfer from HOO• to ortho-Quinones Explains the Antioxidant Activity of Polydopamine. *Angew. Chem. Int. Ed.* **2021**, *60*, 15220–15224. [[CrossRef](#)]
48. Foti, M.C.; Ingold, K.U. Mechanism of Inhibition of lipid peroxidation by  $\gamma$ -terpinene, an unusual and potentially useful hydrocarbon antioxidant. *J. Agric. Food Chem.* **2003**, *51*, 2758–2765. [[CrossRef](#)]
49. Harrison, K.A.; Haidasz, E.A.; Griesser, M.; Pratt, D.A. Inhibition of hydrocarbon autoxidation by nitroxide-catalyzed cross-dismutation of hydroperoxyl and alkylperoxyl radicals. *Chem. Sci.* **2018**, *9*, 6068–6079. [[CrossRef](#)]
50. Liu, W.; Porter, N.A.; Schneider, C.; Brash, A.R.; Yin, H. Formation of 4-Hydroxynonenal from Cardiolipin Oxidation: Intramolecular Peroxyl Radical Addition and Decomposition. *Free Radic. Biol. Med.* **2011**, *50*, 166–178. [[CrossRef](#)]
51. Bowry, V.W. Arm-to-Arm Autoxidation in a Triglyceride: Remote Group Reaction Kinetics. *J. Org. Chem.* **1994**, *59*, 2250–2252. [[CrossRef](#)]
52. Esterbauer, H.; Schaur, R.J.; Zollner, H. Chemistry and biochemistry of 4-hydroxynonenal, malonaldehyde and related aldehydes. *Free Radic. Biol. Med.* **1991**, *11*, 81–128. [[CrossRef](#)]
53. Ullery, J.C.; Marnett, L.J. Protein modification by oxidized phospholipids and hydrolytically released lipid electrophiles: Investigating cellular responses. *Biochim. Biophys. Acta Biomembr.* **2012**, *1818*, 2424–2435. [[CrossRef](#)]
54. Codreanu, S.G.; Ullery, J.C.; Zhu, J.; Tallman, K.A.; Beavers, W.N.; Porter, N.A.; Marnett, L.J.; Zhang, B.; Liebler, D.C. Alkylation Damage by Lipid Electrophiles Targets Functional Protein Systems. *Mol. Cell. Proteom.* **2014**, *13*, 849–859. [[CrossRef](#)] [[PubMed](#)]
55. Spickett, C.M. The lipidperoxidation product 4-hydroxy-2-nonenal: Advances in chemistry and analysis. *Redox Biol.* **2013**, *2013*, 145–152. [[CrossRef](#)] [[PubMed](#)]
56. Liang, X.; Qian, R.; Ou, Y.; Wang, D.; Lin, X.; Sun, C. Lipid peroxide-derived short-chain aldehydes promote programmed cell death in wheat roots under aluminum stress. *J. Hazard. Mater.* **2023**, *443*, 130142. [[CrossRef](#)]
57. Parvez, S.; Long, M.J.C.; Poganik, J.R.; Aye, Y. Redox Signaling by Reactive Electrophiles and Oxidants. *Chem. Rev.* **2018**, *118*, 8798–8888. [[CrossRef](#)] [[PubMed](#)]
58. Schopfer, F.J.; Cipollina, C.; Freeman, B.A. Formation and Signaling Actions of Electrophilic Lipids. *Chem. Rev.* **2011**, *111*, 5997–6021. [[CrossRef](#)]
59. Codreanu, S.G.; Liebler, D.C. Novel Approaches to identify protein adducts produced by lipid peroxidation. *Free Radic Res.* **2015**, *49*, 881–887. [[CrossRef](#)]
60. Perkovic, M.N.; Jaganjac, M.; Milkovic, L.; Horvat, T.; Rojo, D.; Zarkovic, K.; Coric, M.; Hudolin, T.; Waeg, G.; Orehovec, B.; et al. Relationship between 4-Hydroxynonenal (4-HNE) as Systemic Biomarker of Lipid Peroxidation and Metabolomic Profiling of Patients with Prostate Cancer. *Biomolecules* **2023**, *13*, 145. [[CrossRef](#)]
61. Riahi, Y.; Cohen, G.; Shamni, O.; Sasson, S. Signaling and cytotoxic functions of 4-hydroxyalkenals. *Am. J. Physiol. Endocrinol. Metab.* **2010**, *299*, E879–E886. [[CrossRef](#)]
62. Onyango, A.N.; Baba, N. New hypotheses on the pathways of formation of malondialdehyde and isofurans. *Free Radic. Biol. Med.* **2010**, *49*, 1594–1600. [[CrossRef](#)] [[PubMed](#)]
63. Takeuchi, C.; Galve, R.; Nieva, J.; Witter, D.P.; Wentworth, A.D.; Troseth, R.P.; Lerner, R.A.; Wentworth, P. Proatherogenic Effects of the Cholesterol Ozonolysis Products, Atheronal-A and Atheronal-B. *Biochemistry* **2006**, *45*, 7162–7170. [[CrossRef](#)] [[PubMed](#)]
64. Nieva, J.; Song, B.-D.; Rogel, J.K.; Kujawara, D.; Altobel, L., III; Izharudin, A.; Boldt, G.E.; Grover, R.K.; Wentworth, A.D.; Wentworth, P., Jr. Cholesterol Secosterol Aldehydes Induce Amyloidogenesis and Dysfunction of Wild-Type Tumor Protein p53. *Chem. Biol.* **2011**, *18*, 920–927. [[CrossRef](#)]
65. Usui, K.; Hulleman, J.D.; Paulsson, J.F.; Siegel, S.J.; Powers, E.T.; Kelly, J.W. Site-specific modification of Alzheimer’s peptides by cholesterol oxidation products enhances aggregation energetics and neurotoxicity. *Proc. Natl. Acad. Sci. USA* **2009**, *106*, 18563–18568. [[CrossRef](#)] [[PubMed](#)]
66. Wojcik, P.; ˙Zarkovic, N.; Gegotek, A.; Skrzydlewska, E. Involvement of Metabolic Lipid Mediators in the Regulation of Apoptosis. *Biomolecules* **2020**, *10*, 402. [[CrossRef](#)]
67. Graille, M.; Wild, P.; Sauvain, J.-J.; Hemmendinger, M.; Guseva Canu, I.; Hopf, N.B. Urinary 8-isoprostane as a biomarker for oxidative stress. A systematic review and meta-analysis. *Toxicol. Lett.* **2020**, *328*, 19–27. [[CrossRef](#)]
68. Gao, X.; Brenner, H.; Holleccek, B.; Cuk, K.; Zhang, Y.; Anusruti, A.; Xuan, Y.; Xu, Y.; Schottker, B. Urinary 8-isoprostane levels and occurrence of lung, colorectal, prostate, breast and overall cancer: Results from a large, population-based cohort study with 14 years of follow-up. *Free Radic. Biol. Med.* **2018**, *123*, 20–26. [[CrossRef](#)]
69. Yang, J.; Tallman, K.A.; Porter, N.A.; Liebler, D.C. Quantitative Chemoproteomics for Site-Specific Analysis of Protein Alkylation by 4-Hydroxy-2-Nonenal in Cells. *Anal. Chem.* **2015**, *87*, 2535–2541. [[CrossRef](#)]

70. Ferreri, C.; Ferocino, A.; Batani, G.; Chatgililoglu, C.; Randi, V.; Riontino, M.V.; Vetica, F.; Sansone, A. Plasmalogens: Free Radical Reactivity and Identification of Trans Isomers Relevant to Biological Membranes. *Biomolecules* **2023**, *13*, 730. [[CrossRef](#)]
71. Chatgililoglu, C.; Ferreri, C.; Melchiorre, M.; Sansone, A.; Torreggiani, A. Lipid geometrical isomerism: From chemistry to biology and diagnostics. *Chem. Rev.* **2014**, *114*, 255–284. [[CrossRef](#)]
72. Torreggiani, A.; Tinti, A.; Jurasekova, Z.; Capdevila, M.; Saracino, M.; Foggia, M.D. Structural Lesions of Proteins Connected to Lipid Membrane Damages Caused by Radical Stress: Assessment by Biomimetic Systems and Raman Spectroscopy. *Biomolecules* **2019**, *9*, 794. [[CrossRef](#)] [[PubMed](#)]
73. Chatgililoglu, C.; Ferreri, C. Reductive Stress of Sulfur-Containing Amino Acids within Proteins and Implication of Tandem Protein–Lipid Damage. *Int. J. Mol. Sci.* **2021**, *22*, 12863. [[CrossRef](#)]
74. Hirata, Y.; Ferreri, C.; Yamada, Y.; Inoue, A.; Sansone, A.; Vetica, F.; Suzuki, W.; Takano, S.; Noguchi, T.; Matsuzawa, A.; et al. Geometrical isomerization of arachidonic acid during lipid peroxidation interferes with ferroptosis. *Free Radic. Biol. Med.* **2023**, *204*, 374–384. [[CrossRef](#)]
75. Itri, R.; Junqueira, H.C.; Mertins, O.; Baptista, M.S. Membrane Changes under Oxidative Stress: The Impact of Oxidized Lipids. *Biophys. Rev.* **2014**, *6*, 47–61. [[CrossRef](#)]
76. Weber, G.; Charitat, T.; Baptista, M.S.; Uchoa, A.F.; Pavani, C.; Junqueira, H.C.; Guo, Y.; Baulin, V.A.; Itri, R.; Marques, C.M.; et al. Lipid Oxidation Induces Structural Changes in Biomimetic Membranes. *Soft Matter* **2014**, *10*, 4241–4247. [[CrossRef](#)] [[PubMed](#)]
77. Wang, G.; Wang, J.; Fan, X.; Ansari, G.A.S.; Khan, M.F. Protein adducts of malondialdehyde and 4-hydroxynonenal contribute to trichloroethene-mediated autoimmunity via activating Th17 cells: Dose- and time-response studies in female MRL +/+ mice. *Toxicology* **2012**, *292*, 113–122. [[CrossRef](#)] [[PubMed](#)]
78. Yamada, T.; Place, N.; Kosterina, N.; Östberg, T.; Zhang, S.J.; Grundtman, C.; Erlandsson-Harris, H.; Lundberg, I.E.; Glenmark, B.; Bruton, J.D.; et al. Impaired myofibrillar function in the soleus muscle of mice with collagen-induced arthritis. *Arthritis Rheumat.* **2009**, *60*, 3280–3289. [[CrossRef](#)] [[PubMed](#)]
79. Kuhn, H.; Banthiya, S.; van Leyen, K. Mammalian lipoxygenases and their biological relevance. *Biochim. Biophys. Acta* **2015**, *1851*, 308–330. [[CrossRef](#)]
80. Yang, W.S.; Kim, K.J.; Gaschler, M.M.; Patel, M.; Shchepinov, M.S.; Stockwell, B.R. Peroxidation of Polyunsaturated Fatty Acids by Lipoxygenases Drives Ferroptosis. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, E4966–E4975. [[CrossRef](#)]
81. Kennedy, L.; Sandhu, J.K.; Harper, M.-E.; Cuperlovic-Culf, M. Role of Glutathione in Cancer: From Mechanisms to Therapies. *Biomolecules* **2020**, *10*, 1429. [[CrossRef](#)]
82. Peña-Bautista, C.; Vento, M.; Baquero, M.; Cháfer-Pericás, C. Lipid peroxidation in neurodegeneration. *Clin. Chim. Acta* **2019**, *497*, 178–188. [[CrossRef](#)] [[PubMed](#)]
83. da Santana, L.N.; Bittencourt, L.O.; Nascimento, P.C.; Fernandes, R.M.; Teixeira, F.B.; Fernandes, L.M.P.; Freitas Silva, M.C.; Nogueira, L.S.; Amado, L.L.; Crespo-Lopez, M.E.; et al. Low doses of methylmercury exposure during adulthood in rats display oxidative stress, neurodegeneration in the motor cortex and lead to impairment of motor skills. *J. Trace Elem. Med. Biol.* **2019**, *51*, 19–27. [[CrossRef](#)] [[PubMed](#)]
84. Peña-Bautista, C.; Baquero, M.; Vento, M.; Cháfer-Pericás, C. Free radicals in Alzheimer’s disease: Lipid peroxidation biomarkers. *Clin. Chim. Acta* **2019**, *491*, 85–90. [[CrossRef](#)] [[PubMed](#)]
85. Friedmann Angeli, J.P.; Schneider, M.; Proneth, B.; Tyurina, Y.Y.; Tyurin, V.A.; Hammond, V.J.; Herbach, N.; Aichler, M.; Walch, A.; Eggenhofer, E.; et al. Inactivation of the ferroptosis regulator Gpx4 triggers acute renal failure in mice. *Nat. Cell Biol.* **2014**, *16*, 1180–1191. [[CrossRef](#)] [[PubMed](#)]
86. Conrad, M.; Pratt, D.A. The chemical basis of ferroptosis. *Nat. Chem. Biol.* **2019**, *15*, 1137–1147. [[CrossRef](#)]
87. Mishima, E.; Ito, J.; Wu, Z.; Nakamura, T.; Wahida, A.; Doll, S.; Tonnus, W.; Nepachalovich, P.; Eggenhofer, E.; Aldrovandi, M.; et al. A non-canonical vitamin K cycle is a potent ferroptosis suppressor. *Nature* **2022**, *608*, 778–783. [[CrossRef](#)]
88. Shah, R.; Shchepinov, M.S.; Pratt, D.A. Resolving the Role of Lipoxygenases in the Initiation and Execution of Ferroptosis. *ACS Cent. Sci.* **2018**, *4*, 387–396. [[CrossRef](#)]
89. Soula, M.; Weber, R.A.; Zilka, O.Z.; Alwaseem, H.; La, K.; Yen, F.; Molina, H.; Garcia-Bermudez, J.; Pratt, D.A.; Birsoy, K. Metabolic determinants of cancer cell sensitivity to canonical ferroptosis inducers. *Nat. Chem. Biol.* **2020**, *16*, 1351–1360. [[CrossRef](#)]
90. Friedmann Angeli, J.P.; Shah, R.; Derek, A.; Pratt, D.A.; Conrad, M. Ferroptosis Inhibition: Mechanisms and Opportunities. *Trends Pharmacol. Sci.* **2017**, *38*, 489–498. [[CrossRef](#)] [[PubMed](#)]
91. Zhang, L.; Jia, R.; Li, H.; Yu, H.; Ren, K.; Jia, S.; Li, Y.; Wang, Q. Insight into the Double-Edged Role of Ferroptosis in Disease. *Biomolecules* **2021**, *11*, 1790. [[CrossRef](#)]
92. Amorati, R.; Valgimigli, L. Methods to measure the antioxidant activity of phytochemicals and plant extracts. *J. Agric. Food Chem.* **2018**, *66*, 3324–3329. [[CrossRef](#)]
93. Amorati, R.; Foti, M.C.; Valgimigli, L. Antioxidant activity of essential oils. *J. Agric. Food Chem.* **2013**, *61*, 10835–10847. [[CrossRef](#)] [[PubMed](#)]
94. Lucarini, M.; Pedulli, G.F. Free radical intermediates in the inhibition of the autoxidation reaction. *Chem. Soc. Rev.* **2010**, *39*, 2106–2119. [[CrossRef](#)]
95. Cariola, A.; El Chami, M.; Granatieri, J.; Valgimigli, L. Anti-tyrosinase and antioxidant activity of meroterpenoid bakuchiol from *Psoralea corylifolia* (L.). *Food Chem.* **2023**, *405*, 134953. [[CrossRef](#)] [[PubMed](#)]
96. Helberg, J.; Pratt, D.A. Autoxidation vs. antioxidants—The fight for Forever. *Chem. Soc. Rev.* **2021**, *50*, 7343–7358. [[CrossRef](#)]

97. Ingold, K.U.; Pratt, D.A. Advances in Radical-Trapping Antioxidant Chemistry in the 21st Century: A Kinetics and Mechanisms Perspective. *Chem. Rev.* **2014**, *114*, 9022–9046. [[CrossRef](#)] [[PubMed](#)]
98. Valgimigli, L.; Pratt, D.A. Maximizing the Reactivity of Phenolic and Aminic Radical-Trapping Antioxidants: Just Add Nitrogen! *Acc. Chem. Res.* **2015**, *48*, 966–975. [[CrossRef](#)]
99. Pratt, D.A.; DiLabio, G.A.; Brigati, G.; Pedulli, G.F.; Valgimigli, L. 5-Pyrimidinols: Novel Chain-Breaking Antioxidants More Effective Than Phenols. *J. Am. Chem. Soc.* **2001**, *123*, 4625–4626. [[CrossRef](#)]
100. Valgimigli, L.; Brigati, G.; Pedulli, G.F.; DiLabio, G.A.; Mastragostino, M.; Arbizzani, C.; Pratt, D.A. The Effect of Ring Nitrogen Atoms on the Homolytic Reactivity of Phenolic Compounds: Understanding the Radical-Scavenging Ability of 5-Pyrimidinols. *Chem. Eur. J.* **2003**, *9*, 4997–5010. [[CrossRef](#)]
101. Wijtmans, M.; Pratt, D.A.; Valgimigli, L.; DiLabio, G.A.; Pedulli, G.F.; Porter, N.A. 6-Amino-3-Pyridinols: Towards Diffusion-Controlled Chain-Breaking Antioxidants. *Angew. Chem. Int. Ed.* **2003**, *42*, 4370–4373. [[CrossRef](#)]
102. Wijtmans, M.; Pratt, D.A.; Brinkhorst, J.; Serwa, R.; Valgimigli, L.; Pedulli, G.F.; Porter, N.A. Synthesis and Reactivity of Some 6-Substituted-2,4-Dimethyl-3-Pyridinols, a Novel Class of Chain-Breaking Antioxidants. *J. Org. Chem.* **2004**, *69*, 9215–9223. [[CrossRef](#)]
103. Kim, H.-Y.; Pratt, D.A.; Seal, J.R.; Wijtmans, M.; Porter, N.A. Lipid-Soluble 3-Pyridinol Antioxidants Spare  $\alpha$ -Tocopherol and Do Not Efficiently Mediate Peroxidation of Cholesterol Esters in Human Low-Density Lipoprotein. *J. Med. Chem.* **2005**, *48*, 6787–6789. [[CrossRef](#)]
104. Nam, T.-G.; Nara, S.J.; Zagol-Ikapitte, I.; Cooper, T.; Valgimigli, L.; Oates, J.A.; Porter, N.A.; Boutaud, O.; Pratt, D.A. Pyridine and Pyrimidine Analogs of Acetaminophen as Inhibitors of Lipid Peroxidation and Cyclooxygenase and Lipoxygenase Catalysis. *Org. Biomol. Chem.* **2009**, *7*, 5103–5112. [[CrossRef](#)]
105. Serwa, R.; Nam, T.-g.; Valgimigli, L.; Culbertson, S.; Rector, C.L.; Jeong, B.-S.; Pratt, D.A.; Porter, N.A. Preparation and Investigation of Vitamin B 6-Derived Aminopyridinol Antioxidants. *Chem. Eur. J.* **2010**, *16*, 14106–14114. [[CrossRef](#)]
106. Valgimigli, L.; Bartolomei, D.; Amorati, R.; Haidasz, E.; Hanthorn, J.J.; Nara, S.J.; Brinkhorst, J.; Pratt, D.A. 3-Pyridinols and 5-Pyrimidinols: Tailor-Made for Use in Synergistic Radical-Trapping Co-antioxidant Systems. *Beilstein J. Org. Chem.* **2013**, *9*, 2781–2792. [[CrossRef](#)]
107. Valgimigli, L.; Ingold, K.U.; Luszyk, J. Antioxidant activities of vitamin E analogues in water and a Kamlet-Taft  $\beta$ -value for water. *J. Am. Chem. Soc.* **1996**, *118*, 3545–3549. [[CrossRef](#)]
108. Valgimigli, L.; Ingold, K.U.; Luszyk, J. Solvent effects on the reactivity and free spin distribution of 2,2-diphenyl-1-picrylhydrazyl radicals. *J. Org. Chem.* **1996**, *61*, 7947–7950. [[CrossRef](#)]
109. Lucarini, M.; Pedulli, G.F.; Valgimigli, L. Do Peroxyl Radicals Obey the Principle That Kinetic Solvent Effects on H-Atom Abstraction Are Independent of the Nature of the Abstracting Radical? *J. Org. Chem.* **1988**, *63*, 4497–4499. [[CrossRef](#)]
110. Abraham, M.H.; Grellier, P.L.; Prior, D.V.; Taft, R.W.; Morris, J.J.; Taylor, P.J.; Laurence, C.; Berthelot, M.; Doherty, R.M.; Kamlet, M.J.; et al. A general treatment of hydrogen bond complexation constants in tetrachloromethane. *J. Am. Chem. Soc.* **1988**, *110*, 8534–8536. [[CrossRef](#)]
111. Abraham, M.H.; Grellier, P.L.; Prior, D.V.; Morris, J.J.; Taylor, P.J. Hydrogen bonding. Part 10. A scale of solute hydrogen-bond basicity using log K values for complexation in tetrachloromethane. *J. Chem. Soc. Perkin Trans.* **1990**, *2*, 521–529. [[CrossRef](#)]
112. Snelgrove, D.W.; Luszyk, J.; Banks, J.T.; Mulder, P.; Ingold, K.U. Kinetic Solvent Effects on Hydrogen-Atom Abstractions: Reliable, Quantitative Predictions via a Single Empirical Equation. *J. Am. Chem. Soc.* **2001**, *123*, 469–477. [[CrossRef](#)]
113. Litwinienko, G.; Ingold, K.U. Solvent Effects on the Rates and Mechanisms of Reaction of Phenols with Free Radicals. *Acc. Chem. Res.* **2007**, *40*, 222–230. [[CrossRef](#)] [[PubMed](#)]
114. Amorati, R.; Baschieri, A.; Morroni, G.; Gambino, R.; Valgimigli, L. Peroxyl Radical Reactions in Water Solution: A Gym for Proton-Coupled Electron-Transfer Theories. *Chem. Eur. J.* **2016**, *22*, 7924–7934. [[CrossRef](#)]
115. Amorati, R.; Valgimigli, L.; Pedulli, G.F.; Grabovskiy, S.A.; Kabalnova, N.N.; Chatgililoglu, C. Base-promoted reaction of 5-hydroxyuracil derivatives with peroxyl radicals. *Org. Lett.* **2010**, *12*, 4130–4133. [[CrossRef](#)]
116. Amorati, R.; Pedulli, G.F.; Valgimigli, L. Kinetic and thermodynamic aspects of the chain-breaking antioxidant activity of ascorbic acid derivatives in non-aqueous media. *Org. Biomol. Chem.* **2011**, *9*, 3792–3800. [[CrossRef](#)] [[PubMed](#)]
117. Baschieri, A.; Amorati, R.; Benelli, T.; Mazzocchetti, L.; D'Angelo, E.; Valgimigli, L. Enhanced antioxidant activity under biomimetic settings of ascorbic acid included in halloysite nanotubes. *Antioxidants* **2019**, *8*, 30. [[CrossRef](#)] [[PubMed](#)]
118. Valgimigli, L.; Amorati, R.; Petrucci, S.; Pedulli, G.F.; Hu, D.; Hanthorn, J.J.; Pratt, D.A. Unexpected acid catalysis in reactions of peroxyl radicals with phenols. *Angew. Chem. Int. Ed.* **2009**, *48*, 8348–8351. [[CrossRef](#)]
119. Amorati, R.; Valgimigli, L.; Panzella, L.; Napolitano, A.; D'Ischia, M. 5-S-lipoylhydroxytyrosol, a multidefense antioxidant featuring a solvent-tunable peroxyl radical-scavenging 3-thio-1,2-dihydroxybenzene motif. *J. Org. Chem.* **2013**, *78*, 9857–9864. [[CrossRef](#)] [[PubMed](#)]
120. Valgimigli, L.; Amorati, R.; Fumo, M.G.; DiLabio, G.A.; Pedulli, G.F.; Ingold, K.U.; Pratt, D.A. The unusual reaction of semiquinone radicals with molecular oxygen. *J. Org. Chem.* **2008**, *73*, 1830–1841. [[CrossRef](#)] [[PubMed](#)]
121. Alfieri, M.L.; Panzella, L.; Amorati, R.; Cariola, A.; Valgimigli, L.; Napolitano, A. Role of Sulphur and Heavier Chalcogens on the Antioxidant Power and Bioactivity of Natural Phenolic Compounds. *Biomolecules* **2022**, *12*, 90. [[CrossRef](#)]
122. Kumar, S.; Johansson, H.; Engman, L.; Valgimigli, L.; Amorati, R.; Fumo, M.G.; Pedulli, G.F. Regenerable chain-breaking 2,3-dihydrobenzo[b]selenophene-5-ol antioxidants. *J. Org. Chem.* **2007**, *72*, 2583–2595. [[CrossRef](#)]

123. Kumar, S.; Johansson, H.; Kanda, T.; Engman, L.; Muller, T.; Jonsson, M.; Pedulli, G.F.; Petrucci, S.; Valgimigli, L. Catalytic chain-breaking pyridinol antioxidants. *Org. Lett.* **2008**, *10*, 4895–4898. [[CrossRef](#)]
124. Johansson, H.; Shanks, D.; Engman, L.; Amorati, R.; Pedulli, G.F.; Valgimigli, L. Long-lasting antioxidant protection: A regenerable BHA analogue. *J. Org. Chem.* **2010**, *75*, 7535–7541. [[CrossRef](#)] [[PubMed](#)]
125. Amorati, R.; Valgimigli, L.; Dinér, P.; Bakhtiari, K.; Saeedi, M.; Engman, L. Multi-faceted reactivity of alkyltellurophenols towards peroxy radicals: Catalytic antioxidant versus thiol-depletion effect. *Chem. Eur. J.* **2013**, *19*, 7510–7522. [[CrossRef](#)]
126. Hanthorn, J.J.; Amorati, R.; Valgimigli, L.; Pratt, D.A. The reactivity of air-stable pyridine- and pyrimidine-containing diarylamine antioxidants. *J. Org. Chem.* **2012**, *77*, 6895–6907. [[CrossRef](#)]
127. Amorati, R.; Baschieri, A.; Cowden, A.; Valgimigli, L. The Antioxidant Activity of Quercetin in Water Solution. *Biomimetics* **2017**, *2*, 9. [[CrossRef](#)] [[PubMed](#)]
128. Lucarini, M.; Pedulli, G.F.; Valgimigli, L.; Amorati, R.; Minisci, F. Thermochemical and kinetic studies of a bisphenol antioxidant. *J. Org. Chem.* **2001**, *66*, 5456–5462. [[CrossRef](#)]
129. Matera, R.; Gabbanini, S.; Berretti, S.; Amorati, R.; De Nicola, G.R.; Iori, R.; Valgimigli, L. Acylated anthocyanins from sprouts of *Raphanus sativus* cv. Sango: Isolation, structure elucidation and antioxidant activity. *Food Chem.* **2015**, *166*, 397–406. [[CrossRef](#)] [[PubMed](#)]
130. Niki, E.; Saito, T.; Kawakami, A.; Kamiya, Y. Inhibition of oxidation of methyl linoleate in solution by vitamin E and vitamin C. *J. Biol. Chem.* **1984**, *259*, 4177–4182. [[CrossRef](#)]
131. Bowry, V.W.; Ingold, K.U. The Unexpected Role of Vitamin E ( $\alpha$ -Tocopherol) in the Peroxidation of Human Low-Density Lipoprotein. *Acc. Chem. Res.* **1999**, *32*, 27–34. [[CrossRef](#)]
132. Valgimigli, L.; Lucarini, M.; Pedulli, G.F.; Ingold, K.U. Does  $\beta$ -carotene really protect vitamin E from oxidation? *J. Am. Chem. Soc.* **1997**, *119*, 8095–8096. [[CrossRef](#)]
133. Amorati, R.; Pedulli, G.F.; Valgimigli, L.; Johansson, H.; Engman, L. Organochalcogen substituents in phenolic antioxidants. *Org. Lett.* **2010**, *12*, 2326–2329. [[CrossRef](#)]
134. McGrath, A.J.; Garrett, G.E.; Valgimigli, L.; Pratt, D.A. The redox chemistry of sulfenic acids. *J. Am. Chem. Soc.* **2010**, *132*, 16759–16761. [[CrossRef](#)] [[PubMed](#)]
135. Amorati, R.; Lynett, P.T.; Valgimigli, L.; Pratt, D.A. The reaction of sulfenic acids with peroxy radicals: Insights into the radical-trapping antioxidant activity of plant-derived thiosulfinates. *Chem. Eur. J.* **2012**, *18*, 6370–6379. [[CrossRef](#)]
136. Zielinski, Z.; Pesseau, N.; Amorati, R.; Valgimigli, L.; Pratt, D.A. Redox chemistry of selenenic acids and the insight it brings on transition state geometry in the reactions of peroxy radicals. *J. Am. Chem. Soc.* **2014**, *136*, 1570–1578. [[CrossRef](#)]
137. Hanthorn, J.J.; Valgimigli, L.; Pratt, D.A. Preparation of highly reactive pyridine- and pyrimidine-containing diarylamine antioxidants. *J. Org. Chem.* **2012**, *77*, 6908–6916. [[CrossRef](#)]
138. Shah, R.; Haidasz, E.A.; Valgimigli, L.; Pratt, D.A. Unprecedented inhibition of hydrocarbon autoxidation by diarylamine radical-trapping antioxidants. *J. Am. Chem. Soc.* **2015**, *137*, 2440–2443. [[CrossRef](#)] [[PubMed](#)]
139. Vaz, S.M.; Augusto, O. Reactive Oxygen Species Special Feature: Inhibition of myeloperoxidase-mediated protein nitration by tempol: Kinetics, mechanism, and implications. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 8191–8196. [[CrossRef](#)] [[PubMed](#)]
140. Bi, W.; Bi, Y.; Gao, X.; Li, P.; Hou, S.; Zhang, Y.; Bammert, C.; Jockusch, S.; Legalley, T.D.; Gibson, K.M.; et al. Indole-TEMPO conjugates alleviate ischemia-reperfusion injury via attenuation of oxidative stress and preservation of mitochondrial function. *Bioorg. Med. Chem.* **2017**, *5*, 2545–2568. [[CrossRef](#)]
141. Goldstein, S.; Merenyi, G.; Russo, A.; Samuni, A. The Role of Oxoammonium Cation in the SOD-Mimic Activity of Cyclic Nitroxides. *J. Am. Chem. Soc.* **2003**, *125*, 789–795. [[CrossRef](#)]
142. Goldstein, S.; Samuni, A. Kinetics and Mechanism of Peroxy Radical Reactions with Nitroxides. *J. Phys. Chem. A* **2007**, *111*, 1066–1072. [[CrossRef](#)]
143. Amorati, R.; Pedulli, G.F.; Pratt, D.A.; Valgimigli, L. TEMPO reacts with oxygen-centered radicals under acidic conditions. *Chem. Commun.* **2010**, *46*, 5139–5141. [[CrossRef](#)] [[PubMed](#)]
144. Haidasz, E.A.; Meng, D.; Amorati, R.; Baschieri, A.; Ingold, K.U.; Valgimigli, L.; Pratt, D.A. Acid Is Key to the Radical-Trapping Antioxidant Activity of Nitroxides. *J. Am. Chem. Soc.* **2016**, *138*, 5290–5298. [[CrossRef](#)] [[PubMed](#)]
145. Baschieri, A.; Valgimigli, L.; Gabbanini, S.; DiLabio, G.A.; Romero-Montalvo, E.; Amorati, R. Extremely Fast Hydrogen Atom Transfer between Nitroxides and HOO $\cdot$  Radicals and Implication for Catalytic Coantioxidant Systems. *J. Am. Chem. Soc.* **2018**, *140*, 10354–10362. [[CrossRef](#)] [[PubMed](#)]
146. Guo, Y.; Baschieri, A.; Amorati, R.; Valgimigli, L. Synergic antioxidant activity of  $\gamma$ -terpinene with phenols and polyphenols enabled by hydroperoxy radicals. *Food Chem.* **2021**, *345*, 128468. [[CrossRef](#)]
147. Baschieri, A.; Daci Ajvazi, M.; Folifack Tonfack, J.L.; Valgimigli, L.; Amorati, R. Explaining the antioxidant activity of some common non-phenolic components of essential oils. *Food Chem.* **2017**, *232*, 656–663. [[CrossRef](#)]
148. Guo, Y.; Pizzol, R.; Gabbanini, S.; Baschieri, A.; Amorati, R.; Valgimigli, L. Absolute Antioxidant Activity of Five Phenol-Rich Essential Oils. *Molecules* **2021**, *26*, 5237. [[CrossRef](#)]
149. Khoobchandani, M.; Ganesh, N.; Gabbanini, S.; Valgimigli, L.; Srivastava, M.M. Phytochemical potential of *Eruca sativa* for inhibition of melanoma tumor growth. *Fitoterapia* **2011**, *82*, 647–653. [[CrossRef](#)]
150. Cores, Á.; Piquero, M.; Villacampa, M.; León, R.; Menéndez, J.C. NRF2 Regulation Processes as a Source of Potential Drug Targets against Neurodegenerative Diseases. *Biomolecules* **2020**, *10*, 904. [[CrossRef](#)]

151. Matera, R.; Lucchi, E.; Valgimigli, L. Plant Essential Oils as Healthy Functional Ingredients of Nutraceuticals and Diet Supplements: A Review. *Molecules* **2023**, *28*, 901. [[CrossRef](#)]
152. Zamanian, M.Y.; Soltani, A.; Khodarahmi, Z.; Alameri, A.A.; Alwan, A.M.R.; Ramírez-Coronel, A.A.; Obaid, R.F.; Abosooda, M.; Heidari, M.; Golmohammadi, M.; et al. Targeting Nrf2 signaling pathway by quercetin in the prevention and treatment of neurological disorders: An overview and update on new developments. *Fundam. Clin. Pharmacol.* **2023**, 1–15. [[CrossRef](#)] [[PubMed](#)]

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.