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Review

Human Health Benefits through Daily Consumption of Jerusalem Artichoke (*Helianthus tuberosus* L.) Tubers

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Abstract: "Let food be thy medicine and medicine be thy food", is one of the most famous phrases attributed to Hippocrates, the father of medicine. Scientific research on superfoods has increased in the last six years. These foods have nutritional and pharmacological properties, such that they can help to fight against diseases and poor nutritional status. *Helianthus tuberosus* L., or Jerusalem artichoke, appears to be a superfood that provides benefits to human health at the level of the digestive, gastrointestinal, and dermatological systems, being fit for patients with diabetes mellitus due to its high content of inulin and use in an optimal hypocaloric diet due to its low carbohydrate content. In fact, 5 to 15 g per day is beneficial, with evidence of a prebiotic effect. Unfortunately, its consumption and cultivation are not well known worldwide. For this reason, the present review describes the benefits of *H. tuberosus* in human health to promote knowledge about its nutritional benefits.

Keywords: Helianthus tuberosus L.; human health; nutritional quality; gastronomic applications



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1. Introduction

Helianthus tuberosus L. or Jerusalem artichoke is a plant native to North America from the family Asteraceae. Genome skimming suggests the ancestor of Helianthus annuus gave rise to H. tuberosus through hybridization between two types of perennial sunflowers: Hairy and Sawtooth [1]. The morphological characteristics of this species, including size, rhizome morphology, cultivation practices, distribution, and adaptability, have been previously described [2,3]. At the physiological level, H. tuberosus is a perennial plant with tuberous roots, that can be propagated in different ways: rhizomes, grafts, tubers, cuttings, seeds, and tissue culture [4].

Tubers are the edible part of this plant. The morphology of the tubers of H. tuberosus might vary between these forms: pyriform, oval, clavate, rounded and elongated. Plants present a variable number of buds, scale-like rings, tuber colors, tuber surface types, tuber pith colors, possible daughter tubers, dents, and knobs [5]. In different countries, collections exist to study this species for its high nutritional and pharmacological value, and raw materials to produce bioethanol. In the INRA-Montpellier located in France, for example, a collection of 140 cultivated clones with peculiar typologies and genetics exists, including hexaploids (2n = 6x = 102), tetraploids (2n = 4x = 68) and hybrids with sunflower (2n = 34) [6].

Different factors could alter their development. Abiotic stress can be caused by environmental conditions such as soil type, wind, radiation, temperature, drought, salinity, and

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photoperiod [2,7]. Biotic stress may be imposed by saprophytic fungi and nematodes [2] (Table 1). Despite the above, *H. tuberosus* is an invasive plant that has been highly successful in invading new territories, with economic and cultural consequences, altering biodiversity [8].

Table 1. Different species that produce biotic stress in *H. tuberosus*. Summarizes information adapted from [2,7].

Fungi Annelids Ne		Nematode	Insects	
Sclerotinia sclerotiorum	Strauzia longipennis	Ditylenchus dipsaci	Macrosiphum euphorbiae	
Sclerotium rolfsii		Meloidogyne spp.	Trama penecaeca	
Botrytis cinerea		Heterodera schachtii	Trama troglodytes	
Alternaria helianthi		Heterodera marioni	Uroleucon compositae	
Rhizopus nigricans		Caconema radicicola	Uroleucon gobonis	
Erysiphe cichoracearum		Aphelenchoides ritzemabosi	Uroleucon helianthicola	
Puccinia helianthi			Cochylichroa hospe	
Bipolaris zeae			Homoeosoma electellum	
Fusarium spp.				
Penicillium spp.				

2. Main Uses of H. tuberosus

The chemical composition of *H. tuberosus* varies according to the timing of harvest (week) and between varieties. Additionally, storage conditions influence the quality, causing weight loss, changes in dry matter, soluble solids and total sugars, and sucrose contents [9]. Harvest time causes changes in volatile concentrations and sensory scores, as reported by Bach et al. [10], where tubers were collected at different times from planting in soil (30, 38, and 46 weeks).

The phytocomposition of *H. tuberosus* has been described in several publications [3,11–14]. In general, the authors have studied the compounds of *H. tuberosus* from three perspectives: bioethanol production, biological control, and human health.

2.1. Bioethanol

In comparison with corn, rice, sugarcane, and wheat, *H. tuberosus* has double the cellulose production, depending on the month of harvest, with differences of up to 88% in biomass productivity [15]. The principal carbohydrate of *H. tuberosus* is inulin (~50%), which is used as a substrate to generate bioethanol in two manners: after pretreatment, it can hydrolyze and ferment inulin or saccharify and ferment inulin [16]. Information about bioethanol production with *H. tuberosus* can be found in several publications [2,3,17–19].

2.2. Biological Control

The allelopathic potential of this crop has been demonstrated with shoot extracts of *H. tuberosus*, where diethyl ether and aqueous extract inhibited the growth of lettuce seedlings, and salicylic acid, p-hydroxybenzaldehyde, o-coumarin, and coumarin were identified in the extracts [20]. *Helianthus Tuberosus* produces chemicals with allelopathic effects on other plants or weeds, generating a long-term problem with crop rotation [21]. Additionally, plant pathogens and biological pests could be controlled with *H. tuberosus* secondary metabolites or compounds of different extracts [22].

2.3. Human Health

The effects of *H. tuberosus* on human health have been widely studied. The use of *H. tuberosus* as a source of chlorophyll [23], prebiotics [24], protein [25], and other benefits is described in the following sections.

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3. Nutritional Value of H. tuberosus

The nutritional value of *H. tuberosus* has been addressed in recent publications and reviews [26–29]. Table 2 summarizes the principal components found in *H. tuberosus*. However, as described before, harvest time (weeks after planting) and seasonal changes contribute to changing nutritional components such as soluble carbohydrates [30]. This effect is very important because properties such as the high content of inulin are crucial to human health.

H. tuberosus Variety	Composition	Total Protein (%)	Amino Acids	Vitamins	Minerals	Reference
Rote zonenkugel (red variety)		6.36%	H, I, L, K, M + C, F + Y, T, W, V	-	-	[26]
Red and white varieties	Water, ash, protein, fat, crude fiber, inulin-type fructans	-	-	Thiamin, niacin, pantothenic acid, pyridoxine, vitamin C	Fe, Ca, Mg, P, K	[27]
Albik and Rubik varieties	Soluble dry mass, inulin, crude fiber, crude fat, crude protein, true protein, total amino acids	-	-	Ascorbic acid	N, P, K, Mg, Ca, Na	[28]

Table 2. Principal components with nutritional value found in *H. tuberosus* varieties.

Inulin is the principal nutritional component in *H. tuberosus*, and fresh tubers have 80% water, 15% sugar, and 2% protein. The difference between *H. tuberosus* and another tubers as potatoes is the inulin content, because in *H. tuberosus* starch and sucrose are replaces by inulin [31]. It has been demonstrated that inulin is an antimicrobial and anti-inflammatory agent and can heal wounds [32]. Unfortunately, this bioactive ingredient has been underestimated in comparison to those of other crops, such as wheat or rice. Rubel et al. [33] reported a thorough analysis of the biosynthesis and biotechnological applications of inulin from *H. tuberosus* and its effects on human health.

Inulin extraction is approached with different experimental methodologies. Jantaharn et al. [34] suggested an inulin extraction method through frequency and vibration, producing fragments with different molecular weights. On the other hand, Bedzo et al. [35] reported that in the protein purification of *H. tuberosus*, the fraction with inulin was 82.3%, but *H. tuberosus* slices were more efficient in inulooligosaccharide production. Srinameb et al. [36] proposed a fast laboratory methodology to obtain an inulin production yield of 92.5%.

Essential amino acids can be found in *H. tuberosus*: histidine, isoleucine, leucine, lysine, methionine + cystine, phenylalanine + tyrosine, threonine, tryptophan, and valine [26]. The content and concentration of different amino acids depend on the cultivar and even the storage time, where the principal essential amino acids are arginine, asparagine, glutamine and alanine, and non-essential amino acids are conserved [37]. Regarding protein content, proteome analysis of *H. tuberosus* was recently reported, revealing the existence of proteins related to health and controlling human metabolism, with applications in the management of Parkinson's, Huntington's, and Alzheimer's diseases [25]. In the same research, they found two Kunitz-type and serine hydroxy-methyltransferase proteins, which were antifungal and anticancer as well as antimicrobial and antineoplastic agents, respectively. The high protein content of *H. tuberosus* (5.82–13.36% crude protein) makes it a candidate for feeding ruminants [38].

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4. Applications of *H. tuberosus* in Human Health

4.1. Antioxidant Capacity

A wide range of compounds and secondary metabolites from $H.\ tuberosus$ can be used in human health and can be extracted from its tubers, leaves, and flowers. A 2018 study by Wang et al. [39] found 23 compounds in its flowers. As a result, the compound feradiol demonstrated antiproliferative activity against colon cancer cell lines and ent-kaur-16-en-19-oic acid and β -sitostenone showed antimicrobial activity against $Enterococcus\ faecium$. Similar research found 21 compounds in flowers, where 3,4,2'-tri-hydroxychalcone4'-O- β -D-glucoside was the principal compound in the water-soluble fraction. Tyrosol has powerful activity in scavenging free radicals, and quercetin-3-O- β -D-glucopyranoside inhibited the α -glucosidase enzyme [40]. Using the nematode $Caenorhabditis\ elegans$, the antioxidant activity of the flowers of this species has been proven, whereby the ethyl acetate fraction increases catalase and superoxide dismutase enzyme activity [40].

The tubers also present antioxidant activity, as was demonstrated by Mariadoss et al. [41]. In the ethyl acetate fraction, phenolic compounds with benefits to human health, such as chlorogenic acid, were detected, proving antidiabetic and wound-healing properties. The quantities of antioxidant compounds are influenced by the environmental conditions because overwintering tubers have a minor antioxidant capacity, which could be related to mass concentration [42].

The leaf extract inhibited lipid peroxidation, protecting Chang cells from oxidative stress [43]. Using DPPH and ABTS assays revealed 5,8-diOH-6,7diMeO-2-(3,4-diMeOPh)-4-benzopyrone and 5,8-diOH-6,7,4'-triOMe had high antioxidant activity, which was better than that of the positive control [44].

Showkat et al. [45] analyzed and compared phenolic contents from different organs of *H. tuberosus*: tubers, flowers, stems, and leaves. These organs demonstrated major quantities of phenolic acids with a high content of chlorogenic acid and its isomers. A comparison of *H. tuberosus* with *Helianthus salicifolius* was made using bulked and dried aerial parts demonstrated that the two extracts have potential application as natural antioxidants with biocidal activity [46].

4.2. Dermatological Treatments

Helianthus tuberosus can also be applied externally and internally. Ethanol extracts applied in mice with atopic dermatitis caused by the mite *Dermatophagoides farina* and HaCaT keratinocytes demonstrated relief and attenuation of symptoms and a decrease in epidermal thickness, alongside enhanced expression of filaggrin [47]. Inulin isolated from *H. tuberosus* has been used to develop cosmetic products, such as a body wash gel, decreasing the skin irritation [48].

4.3. Digestive System

The principal beneficial of *H. tuberosus* is inulin, a fructan type compound stored in plants as a reservoir; in *H. tuberosus*, carbohydrate content range between 65–82%, and inulin corresponds to 22.4% [49]. Inulin is useful as a prebiotic and probiotic in the human body because it influences probiotic adhesion to intestinal epithelial cells, allowing the survival of probiotics under gastrointestinal conditions [50,51]. For this reason, inulin incorporation in foods could help to improve health. For example, studies in the development of yogurt with added inulin from *H. tuberosus* as a prebiotic at a 5% concentration show that this treatment reduced blood glucose, cholesterol, and total lipids [52]. Another study with sixty-six volunteers proved that inulin increases the levels of *Bifidobacteria*, *Lactobacillus*, and *Enterococcus* genera [53]. Additionally, supplying older adults with 5 g/day of *H. tuberosus* powder in the morning was beneficial to intestinal microbiota [54].

In animals, the efficacy of *H. tuberosus* has also been demonstrated. In rats, supplementation of daily food with *H. tuberosus* powder at different concentrations increased *Lactobacillus* spp. and *Bifidobacterium* spp. and aided the absorption of calcium and phos-

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phorous [55]. The addition of *H. tuberosus* to the boar diet decreased the levels of pathogenic bacteria and consequently skatole, a toxic compound [56].

4.4. Improvement of Biochemical Parameters

Helianthus tuberosus consumption, at a 10% administration rate, induces gene expression related to enzymes involved in fatty acid synthesis, fibrosis, and inflammation, improving the prevention of type 2 diabetes and nonalcoholic fatty liver disease in rats [57]. Similar results in sugar blood and hepatic lipids have been described [58,59], where a biochemical analysis revealed a decrease in blood glucose level, liver triglyceride, and total cholesterol in rats fed with 10 w/w% *H. tuberosus*. In older adults, *H. tuberosus* consumption reduces glucose levels [40]. Despite these encouraging results, an excessive intake of *H. tuberosus* is not recommended because it could aggravate diabetes [43].

4.5. Superfood

Due to its nutritional qualities, investigations are focusing on the applications of *H. tuberosus* in gastronomic preparations such as flour, chips, noodles, and pastries, among others. Figure 1 shows the potential applications for *H. tuberosus* in human health.

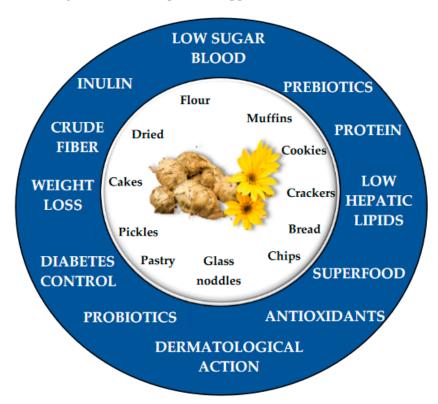


Figure 1. Summarize gastronomic preparations and human health benefits of *H. tuberosus*.

5. Helianthus tuberosus: Increasing Quality of Foods for Cooking

Helianthus tuberosus varieties may differ in their contents of nutritional components. For example, Wang et al. [44] analyzed the components of seven varieties from Lithuania, China, and Thailand harvested at different weeks. Crop relocation changes the chemical composition of tubers so that edaphic and genetic factors are key in the nutritional components of *H. tuberosus* [28]. Pinar et al. [38] recommended genetic analysis to choose the best variety according to the given use; for example, *H. tuberosus* destined to a source of roughage for ruminants or animals with offspring must have high quality, with appropriate and necessary nutrients for the developmental stage of the animal [38]. The above was verified in Abd El-Mola and Aboulfotoh [60], where Ossimi rams were fed with *H. tuberosus*

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in different proportions. As a result, nutrient digestibility, feeding values, and some blood parameters were better than those of the control.

The applications of *H. tuberosus* in the food industry are wide (Table 3). Slapkauskaite et al. [61] fermented tubers with lactic acid bacteria to improve the nutritional quality of dairy products. Helianthus tuberosus was well accepted because its organoleptic qualities were better than those of the other plants tested. In addition, the powder of H. tuberosus was used to optimize glass noodles, a very common food in the Asian diet. Powder incorporation in glass noodle production provided acceptable organoleptic parameters including good hardness, cohesiveness, springiness, and gumminess, and nutritionally, the glass noodles had increased fiber and sugar quantities [62]. In China, H. tuberosus is prepared as a pickle and produces a smooth, sweet, smelly, and crisp product [63]. Bread enrichment with inulin from *H. tuberosus* at a low concentration (2.5 g/100 g flour) increases crumb hardness and chewiness [64]. Another report suggests that adding 5% H. tuberosus powder results in bread with good organoleptic qualities, long shelf life, and high nutrient contents [65]. Helianthus tuberosus powder has been investigated for its ability to make pastry products healthier. Similar to the addition of the powder to the bread, changes were observed in appearance, organoleptic qualities, and nutritional value. A large quantity of H. tuberosus powder decreased the content of kcal in cakes, butter biscuits, and honey biscuits [66]. Sensorial studies of cookies, muffins, cracker bread, and mash with H. tuberosus powder generally indicated a good reception, especially by women [67].

Table 3. Summary of cooking products with *H. tuberosus*.

Product	Organoleptic Qualities	Reference	
	Cookies with antioxidant capacity.		
Cookies	Hard cookies with more H. tuberosus flour.	[68]	
Cookies	Darker cookies with <i>H. tuberosus</i> flour, in comparison with potato flour.	[69]	
	Elastic dough with more H. tuberosus flour.		
Cake	30% of H. tuberosus powder improves organoleptic qualities such as aroma,		
	texture, softness, porosity, appearance, and color, among others.	[70]	
	5-10% of <i>H. tuberosus</i> powder change cake color, decreases softness, but	[71]	
	evaluators qualified well as a source of inulin.		
Muffins	The addition of 10.99% of <i>H. tuberosus</i> powder and 71.40% of oligosaccharide is	[72]	
	the optimum formulation to prepare muffins with good organoleptic qualities.	[/2]	
Pickles	Inactivation of inulinase enzyme and marination to produce a better taste.	[63]	
	A percentage of <i>H. tuberosus</i> flour in the bread preparation (5%), adds essential		
Bread	amino acids, micro, and macronutrients, extends its useful life, and improves	[65]	
	its organoleptic qualities.		
Chips	Chips with low or without calories and sugar. Probing the best preparation of	[73]	
	H. tuberosus chips in a deep fat fryer or microwave oven.		
Glass noodles	Define optimum concentrations for glass noodles with more fiber and sugar,	[62]	
	with other good organoleptic qualities such as cohesiveness and gumminess.	[02]	
Other cooking	Inulin extracted from <i>H. tuberosus</i> is used in preparations such as ice porridge,	[74]	
preparations	instant cereal drinks, ready mixed soya power chocolate malt mixed beverage	[/ =]	

6. Challenges and Perspectives

This review demonstrates that *H. tuberosus* is a tuber with a great potential in the pharmaceutical and gastronomic areas. As Hippocrates said, consumption of quality food, such as *H. tuberosus*, can improve health. Cultural barriers and lack of knowledge are the principal problems with *H. tuberosus* use in the feeding of the population because being an unknown food generates mistrust. Government campaigns and measures or the Ministry of Health could help people incorporate this plant into their regular diet.

Interestingly, in Chile and other South American countries, *H. tuberosus* is not a well-known superfood. However, cooperatives located in southern Chile and people who live on farms have cultivated this crop for personal consumption or recently for commercialization in small local markets. Additionally, in recent years, *H. tuberosus* has received particular attention from research centers with efforts focused on increasing the

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quality of the extraction and determining the nutritional compounds present in these tubers; producers have received support from the government to increase the quality of the production process. For this reason, this review attempts to promote the important quality traits to maintain the health benefits of incorporating this tuber into foods, which might even be better if added to high-consumption foods such as bread. In addition, because it is a very low-cost food, this crop could serve as an important nutritional source in poor regions of the world, while in more privileged regions, it is now consumed as gourmet food. This would help to significantly improve the health of the population, especially elderly individuals, according to the points that we have discussed and presented above.

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References

- 1. Bock, D.G.; Kane, N.C.; Ebert, D.P.; Rieseberg, L.H. Genome skimming reveals the origin of the Jerusalem Artichoke tuber crop species: Neither from Jerusalem nor an artichoke. *New Phytol.* **2014**, 201, 1021–1030. [CrossRef]
- 2. Rossini, F.; Provenzano, M.E.; Kuzmanovic, L.; Ruggeri, R. Jerusalem artichoke (*Helianthus tuberosus* L.): A versatile and sustainable crop for renewable energy production in Europe. *Agronomy* **2019**, *9*, 528. [CrossRef]
- 3. Liava, V.; Karkanis, A.; Danalatos, N.; Tsiropoulos, N. Cultivation practices, adaptability and phytochemical composition of Jerusalem Artichoke (*Helianthus tuberosus* L.): A weed with economical value. *Agronomy* **2021**, *11*, 914. [CrossRef]
- 4. Alla, N.A.; Domokos-Szabolcsy, E.; El-Ramady, H.; Hodossi, S.; Fári, M.; Ragab, M.; Taha, H. Jerusalem artichoke (*Helianthus tuberosus* L.): A review of *in vivo* and *in vitro* propagation. *Int. J. Hortic. Sci.* **2014**, 20, 131–136. [CrossRef]
- 5. Smekalova, T.N.; Lebedeva, N.V.; Novikova, L.Y. Morphological analysis of Jerusalem Artichoke (*Helianthus tuberosus* L.) accessions of different origin from VIR collection. *Proc. Latv. Acad. Sci.* **2011**, 73, 502–512.
- 6. Serieys, H.; Souyris, I.; Gil, A.; Poinso, B.; Bervillé, A. Diversity of Jerusalem artichoke clones (*Helianthus tuberosus* L.) from the INRA-Montpellier collection. *Genet. Resour. Crop Evol.* **2010**, *57*, 1207–1215. [CrossRef]
- 7. Kays, S.J.; Nottingham, S.F. Pollinators, pests, and diseases. In *Biology and Chemistry of Jerusalem Artichoke: Helianthus tuberosus L.*; CRC Press: Boca Raton, FL, USA; Taylor & Francis Group: Tokyo, Japan, 2008.
- 8. Pacanoski, Z.; Mehmeti, A. The first report of the invasive alien weed Jerusalem Artichoke (*Helianthus tuberosus* L.) in the Republic of North Macedonia. *Agric. For.* **2020**, *66*, 115–127.
- 9. Danilčenko, H.; Jariené, E.; Aleknavičiene, P.; Gajewski, M. Quality of Jerusalem Artichoke (*Helianthus tuberosus* L.) Tubers in relation to storage conditions. *Not. Bot. Horti Agrobot. Cluj-Napoca* **2008**, *36*, 23–27.
- 10. Bach, V.; Kidmose, U.; Bjørn, G.; Edelenbos, M. Effects of harvest time and variety on sensory quality and chemical composition of Jerusalem artichoke (*Helianthus tuberosus*) tubers. *Food Chem.* **2012**, *133*, 82–89.
- 11. Yuan, X.; Gao, M.; Xiao, H.; Tan, C.; Du, Y. Free radical scavenging activities and bioactive substances of Jerusalem artichoke (*Helianthus tuberosus* L.) leaves. *Food Chem.* **2012**, *133*, 10–14.
- 12. Kapusta, L.; Krok, E.S.; Jamro, D.B.; Cebulak, T.; Kaszuba, J.; Salach, R.T. Identification and quantification of phenolic compounds from Jerusalem artichoke (*Helianthus tuberosus* L.) tubers. *J. Food Agric. Environ.* **2013**, *11*, 601–606.
- 13. Barkhatova, T.V.; Nazarenko, M.N.; Kozhukhova, M.A.; Khripko, I.A. Obtaining and identification of inulin from Jerusalem artichoke (*Helianthus tuberosus*) tubers. *Foods Raw Mater.* **2015**, *3*, 13–22. [CrossRef]
- 14. Ozgoren, E.; Isik, F.; Yapar, A. Effect of Jerusalem artichoke (*Helianthus tuberosus* L.) supplementation on chemical and nutritional properties of crackers. *J. Food Meas. Charact.* **2019**, *13*, 2812–2821. [CrossRef]
- 15. Gunnarsson, I.B.; Svensson, S.E.; Johansson, E.; Karakashev, D.; Angelidaki, I. Potential of Jerusalem artichoke (*Helianthus tuberosus* L.) as biorefinery crop. *Ind. Crops Prod.* **2014**, *56*, 231–240. [CrossRef]
- 16. Yang, L.; He, Q.S.; Corscadden, K.; Udenigwe, C.C. The prospects of Jerusalem artichoke in functional food ingredients and bioenergy production. *Biotechnol. Rep.* **2015**, *5*, 77–88. [CrossRef]
- 17. Kim, S.; Kim, C.H. Evaluation of whole Jerusalem artichoke (*Helianthus tuberosus* L.) for consolidated bioprocessing ethanol production. *Renew. Energy* **2014**, *65*, 83–91. [CrossRef]

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18. Song, Y.; Wi, S.G.; Ki, H.M.; Bae, H. Cellulosic bioethanol production from Jerusalem artichoke (*Helianthus tuberosus* L.) using hydrogen peroxide-acetic acid (HPAC) pretreatment. *Bioresour. Technol.* **2016**, 214, 30–36. [CrossRef]

- 19. Bhagia, S.; Akinosho, H.; Ferreira, J.F.S.; Ragauskas, A.J. Biofuel production from Jerusalem artichoke tuber inulins: A review. *Biofuel Res. J.* **2017**, *14*, 587–599. [CrossRef]
- 20. Tesio, F.; Weston, L.A.; Ferrero, A. Allelochemicals identified from Jerusalem artichoke (*Helianthus tuberosus* L.) residues and their potential inhibitory activity in the field and laboratory. *Sci. Hortic.* **2011**, *129*, 361–368. [CrossRef]
- 21. Vidotto, F.; Tesio, F.; Ferrero, A. Allelopathic effects of *Helianthus tuberosus* L. on germination and seedling growth of several crops and weeds. *Biol. Agric. Hortic.* **2007**, *26*, 55–68. [CrossRef]
- 22. Chen, F.; Long, X.; Liu, Z.; Shao, H.; Liu, L. Analysis of phenolics acids of Jerusalem artichoke (*Helianthus tuberosus* L.) responding to salt-stress by liquid chromatography/tandem mass spectrometry. *Sci. World J.* **2014**, 568043. [CrossRef]
- 23. Kaszás, L.; Kovács, Z.; Nagy, E.; Elhawat, N.; Abdalla, N.; Domokos-Szabolcsy, E. Jerusalem artichoke (*Helianthus tuberosus* L.) as a potential chlorophyll source for humans and animals nutrition. *Environ. Biodivers. Soil Secur.* **2018**, 2, 1–9. [CrossRef]
- 24. Gupta, D.; Chaturvedi, N. Prebiotic potential of underutilized Jerusalem artichoke in human health: A comprehensive review. *Int. J. Environ. Agric. Biotechnol.* **2020**, *5*, 97–103. [CrossRef]
- 25. Bakku, R.K.; Gupta, R.; Min, C.; Kim, S.; Takahashi, G.; Shibato, J.; Shioda, S.; Takenoya, F.; Agrawal, G.K.; Rakwal, R. Unravelling the *Helianthus tuberosus* L. (Jerusalem artichoke, Kiku-Imo) tuber proteome by label-free quantitative proteomics. *Molecules* 2022, 27, 1111. [CrossRef] [PubMed]
- 26. Cieslik, E.; Gębusia, A.; Florkiewicz, A.; Mickowska, B. The content of protein and of amino acids in Jerusalem artichoke tubers (*Helianthus tuberosus* L.) of red variety Rote Zonenkugel. *Acta Sci. Pol. Technol. Aliment.* **2011**, 10, 433–441.
- 27. Catana, L.; Catana, M.; Iorga, E.; Lazar, A.; Lazar, M.; Teodorescu, R.I.; Asanica, A.C.; Belc, N.; Iancu, A. Valorification of Jerusalem artichoke tubers (*Helianthus tuberosus*) for achieving of functional ingredient with high nutritional value. *Conf. Proc. Agric. Life Life Agric.* 2018, 1, 276–283. [CrossRef]
- 28. Sawicka, B.; Danilčencko, H.; Jariene, E.; Skiba, D.; Rachon, L.; Barbas, P.; Pszczólkowski, P. Nutritional value of Jerusalem artichoke tubers (*Helianthus tuberosus* L.) grown in organic system under Lithuanian and Polish conditions. *Agriculture* **2021**, *11*, 440. [CrossRef]
- 29. Wang, Y.; Zhao, Y.; Xue, F.; Nan, X.; Wang, H.; Hua, D.; Liu, J.; Yang, L.; Jiang, L.; Xiong, B. Nutritional value, bioactivity, and application potential of Jerusalem artichoke (*Helianthus tuberosus* L.) as a neotype feed resource. *Anim. Nutr.* **2020**, *6*, 429–437. [CrossRef]
- 30. Kocsis, L.; Liebhard, P.; Praznik, W. Effect of seasonal changes on content and profile of soluble carbohydrates in tubers of different varieties of Jerusalem artichoke (*Helianthus tuberosus* L.). *J. Agric. Food Chem.* **2007**, *55*, 9401–9408. [CrossRef]
- 31. Brkljača, J.; Bodroza-Solarov, M.; Krulj, J.; Terzić, S.; Mikić, A.; Marjanović Jeromela, A. Quantification of inulin content in selected accessions of Jerusalem artichoke (*Helianthus tuberosus* L.). *Helia* **2014**, 37, 105–112. [CrossRef]
- 32. Rolnik, A.; Olas, B. Plants of the Asteraceae family as agents in the protection of human health. *Int. J. Mol. Sci.* **2021**, 22, 3009. [CrossRef] [PubMed]
- 33. Rubel, I.A.; Iraporda, C.; Manrique, G.D.; Genovese, D.B.; Abraham, A.G. Inulin from Jerusalem artichoke (*H. tuberosus* L.): From its biosynthesis to its application as bioactive ingredient. *Bioact. Carbohydr. Diet. Fibre* **2021**, *26*, 100281. [CrossRef]
- 34. Jantaharn, P.; Mongkoltharnaruk, W.; Senawong, T.; Jogloy, S.; McCloskey, S. Bioactive compounds from organic extracts of *Helianthus tuberosus* L. flowers. *Ind. Crops. Prod.* **2018**, *119*, 57–63. [CrossRef]
- 35. Bedzo, O.K.K.; van Rensburg, E.; Görgens, J.F. Investigating the effect of different inulin-rich substrate preparations from Jerusalem artichoke (*Helianthus tuberosus* L.) tubers on efficient inulooligosaccharides production. *Prep. Biochem. Biotechnol.* **2020**, 51, 440–449. [CrossRef]
- 36. Srinameb, B.; Nuchadomrong, S.; Jogloy, S.; Patanothai, A.; Srijaranai, S. Preparation of inulin powder from Jerusalem artichoke (*Helianthus tuberosus* L.) tuber. *Plant Foods Hum. Nutr.* **2015**, 70, 221–226. [CrossRef]
- 37. Danilcenko, H.; Jariene, E.; Gajewski, M.; Sawicka, B.; Kulaitiene, J.; Cerniauskiene, J. Changes in amino acids content in tubers of Jerusalem artichoke (*Helianthus tuberosus* L.) cultivars during storage. *Acta Sci. Pol.-Hortorum Cultus* **2013**, *12*, 97–105.
- 38. Pinar, H.; Kara, K.; Hanci, F.; Kaplan, M. Nutritional composition of herbage of different Jerusalem artichoke genotypes. *J. Anim. Feed Sci.* **2021**, 30, 141–148. [CrossRef]
- 39. Wang, Y.; Zhao, J.; Yang, J.; Idong, P.T.; Mei, L.; Tao, Y.; Shi, Y. Antioxidant and α-glucosidase inhibitory ingredients identified from Jerusalem artichoke flowers. *Nat. Prod. Res.* **2019**, *33*, 584–588. [CrossRef]
- 40. Kim, J.H.; Lee, Y.B.; Han, S.Y.; Kim, S.J.; Hwang, I.H.; Kim, D.K. Antioxidant activity of *Helianthus tuberosus* L. flower in *Caenorhabditis elegans*. *Kor. J. Pharmacogn.* **2019**, *50*, 96–101.
- 41. Mariadoss, A.V.A.; Park, S.; Saravanakumar, K.; Sathiyaseelan, A.; Wang, M. Ethyl acetate fraction of *Helianthus tuberosus* L. induces anti-diabetic, and wound-healing activities in insulin-resistant human liver cancer and mouse fibroblast cells. *Antioxidants* **2021**, *10*, 99. [CrossRef]
- 42. Zhou, H.; Li, B.; Wu, M.; Liu, Y. Evaluation of antioxidant capacity of polysaccharide in Jerusalem artichoke (*Helianthus tuberosus* L.) during overwintering. *E3S Web Conf.* **2019**, *78*, 02008. [CrossRef]
- 43. Kim, Y.; Lee, S.; Hwang, J.; Kim, E.; Park, P.; Jeon, B. Antioxidant activity and protective effects of extracts from *Helianthus tuberosus* L. leaves on t-BHP induced oxidative stress in Chang cells. *J. Korean Soc. Food Sci. Nutr.* **2011**, 40, 1525–1531. [CrossRef]

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44. Wang, M.; Ma, Z.; He, C.; Yuan, X. The antioxidant activities of flavonoids in Jerusalem artichoke (*Helianthus tuberosus* L.) leaves and their quantitative analysis. *Nat. Prod. Res.* **2020**, *36*, 1009–1013. [CrossRef]

- 45. Showkat, M.M.; Falck-Ytter, A.B.; Straetkvern, O. Phenolic acids in Jerusalem Artichoke (*Helianthus tuberosus* L.): Plant organ dependent antioxidant activity and optimized extraction from leaves. *Molecules* **2019**, 24, 3296. [CrossRef]
- 46. Malm, A.; Grzegorczyk, A.; Biernasiuk, A.; Baj, T.; Rój, E.; Tyskiewicz, K.; Debczak, A.; Stolarski, M.J.; Krzyzaniak, M.; Olba-Ziety, E. Could supercritical extracts from the aerial parts of *Helianthus salicifolius* A. Dietr and *Helianthus tuberosus* L. be regarded as potential raw materials for biocidal purposes? *Agriculture* 2020, 11, 10. [CrossRef]
- 47. Kang, Y.; Lee, K.; An, H. Inhibitory effects of *Helianthus tuberosus* ethanol extract on *Dermatophagoides farina* body-induced atopic dermatitis mouse model and human keratinocytes. *Nutrients* **2018**, *10*, 1657. [CrossRef]
- 48. Niziol-Lukaszewska, Z.; Bujak, T.; Wasilewski, T.; Szmuc, E. Inulin as an effectiveness and safe ingredient in cosmetics. *Pol. J. Chem. Technol.* **2019**, 21, 44–49. [CrossRef]
- 49. Gupta, K.; Talwar, G.; Jain, V.; Dhawan, K.; Jain, S. Salad crops: Root, bulb and tuber crops. In *Encyclopedia of Food Sciences and Nutrition*, 2nd ed.; Academic Press: Cambridge, MA, USA, 2003; pp. 5060–5073.
- 50. Elaheh, M.; Mohamadi, S.A.; Milani, E.; Ladan, N. Prebiotic effect of Jerusalem artichoke (*Helianthus tuberosus*) fructans on the growth performance of *Bifiobacterium bifium* and *Escherichia coli*. *Asian Pac. J. Trop. Dis.* **2016**, *6*, 385–389. [CrossRef]
- 51. Iraporda, C.; Rubel, I.A.; Manrique, G.D.; Abraham, A.G. Influence of inulin rich carbohydrates from Jerusalem artichoke (*Helianthus tuberosus* L.) tubers on probiotic properties of Lactobacillus strains. *LWT* **2019**, *101*, 738–746. [CrossRef]
- 52. El-Kholy, W.M.; Mahrous, H. Biological studies on Bio-Yoghurt fortified with prebiotic obtained from Jerusalem artichoke. *Food Nutr. Sci.* **2015**, *6*, 1552–1564. [CrossRef]
- 53. Ramnani, P.; Gaudier, E.; Bingham, M.; van Bruggen, P.; Tuohy, K.M.; Gibson, G.R. Prebiotic effect of fruit and vegetable shots containing Jerusalem artichoke inulin: A human intervention study. *Br. J. Nutr.* **2010**, *104*, 233–240. [CrossRef] [PubMed]
- 54. Kim, H.; Chijiki, H.; Nanba, T.; Ozaki, M.; Sasaki, H.; Takahashi, M.; Shibata, S. Ingestion of *Helianthus tuberosus* at breakfast rather than at dinner is more effective for suppressing glucose levels and improving the intestinal microbiota in older adults. *Nutrients* **2020**, *12*, 3035. [CrossRef] [PubMed]
- 55. Samal, L.; Chaturvedi, V.B.; Pattanaik, A.K. Effects of dietary supplementation with Jerusalem artichoke (*Helianthus tuberosus* L.) tubers on growth performance, nutrient digestibility as well as activity and composition of large intestinal microbiota in rats. *J. Anim. Feed Sci.* **2017**, 26, 50–58. [CrossRef]
- 56. Okrouhlá, M.; Cítek, J.; Svejstil, R.; Zadinová, K.; Pokorná, K.; Urbanová, D.; Stupka, R. Populations of pig faecal bacteria and the prevalence of skatole. *Animals* **2020**, *10*, 693. [CrossRef]
- 57. Chang, W.; Jia, H.; Aw, W.; Saito, K.; Hasegawa, S.; Kato, H. Beneficial effects of soluble dietary Jerusalem artichoke (*Helianthus tuberosus*) in the prevention of the onset of type 2 diabetes and non-alcoholic fatty liver disease in high-fructose diet-fed rats. *Br. J. Nutr.* **2014**, *112*, 709–717. [CrossRef]
- 58. Okada, N.; Kobayashi, S.; Moriyama, K.; Miyataka, K.; Abe, S.; Sato, C.; Kawazoe, K. *Helianthus tuberosus* (Jerusalem artichoke) tubers improve glucose tolerance ande hepatic lipid profile in rats fed a high-fat diet. *Asian Pac. J. Trop. Med.* **2017**, *10*, 439–443. [CrossRef]
- 59. Kim, S.H.; Kim, B.K.; Park, B.Y.; Kim, J.M.; Lee, Y.J.; Lee, M.K.; Yee, S.; Kang, M.Y. Effects of Jerusalem artichoke extract and inulin on blood glucose levels and insulin secretion in streptozotocin induced diabetic mice. *Korean Diabetes J.* 2021, 22, 60–70. [CrossRef]
- 60. Abd El-Mola, A.E.; Aboulfotoh, G. Effect of feeding *Helianthus tuberosus* (*Jerusalem artichoke*) on rations nutritive value and some blood parameters of Ossimi rams. *Egypt. J. Nutr. Feed.* **2018**, 21, 3365–3371. [CrossRef]
- 61. Slapkauskaite, J.; Sekmokiene, D.; Kabasinskiene, A.; Bartkiene, E.; Juodeikiene, G.; Sarkinas, A. Influence of lactic acid bacteria-fermented *Helianthus tuberosus* L. and *Lupinus luteus* on quality of milk products. *CyTA J. Food* **2015**, *14*, 482–488.
- 62. Singthong, J.; Thongkaew, C. Effect of Jerusalem artichoke (*Helianthus tuberosus*) powder on quality of glass noodles. *Food Res.* **2020**, *4*, 17–26. [CrossRef]
- 63. Zhang, L.; Liu, W.; Ji, J.; Deng, L.; Feng, Q.; Shi, W.; Gao, J. Inactivation of inulinase and marination of High-Quality Jerusalem Artichoke (*Helianthus tuberosus* L.) pickles with screened dominant strains. *Front. Bioeng. Biotechnol.* **2021**, *8*, 626861. [CrossRef] [PubMed]
- 64. Rubel, I.A.; Pérez, E.E.; Manrique, G.D.; Genovese, D.B. Fiber enrichment of wheat bread with Jerusalem Artichoke inulin: Effect on dough rheology and bread quality. *Food Struct.* **2015**, *3*, 21–29. [CrossRef]
- 65. Chirsanova, A.; Capcanari, T.; Gincu, E. Jerusalem artichoke (*Helianthus tuberosus*) flour impact on bread quality. *J. Food Eng.* **2021**, 28, 131–143. [CrossRef]
- 66. Gedrovica, I.; Karklina, D. Influence of Jerusalem Artichoke powder on the nutritional value of pastry products. *Int. J. Food Sci. Nutr.* **2012**, *6*, 524–527.
- 67. Ibarguren, L.; Calderon, M.; Tessaro, S.; Bertona, A.; Rebora, C. Evaluación sensorial del topinambur (*Helianthus tuberosus* L.) como alimento. *RIA Rev. Investig. Agropecu.* **2019**, 45, 204–210.
- 68. Lee, Y.; Kim, D.; Lee, O.; Yoon, W.B. Characterizing texture, color and sensory attributes of cookies made with Jerusalem Artichoke (*Helianthus tuberosus* L.) flour using a mixture design and browning reaction kinetics. *Int. J. Food Eng.* **2016**, *12*, 107–126. [CrossRef]
- 69. Lee, Y.J.; Lee, O.; Yoon, W.B. Effect of inulin in Jerusalem Artichoke (*Helianthus tuberosus* L.) flour on the viscoelastic behavior of cookie dough and quality of cookies. *FoodOPS* **2017**, *7*, 35–44.

Horticulturae **2022**, *8*, 620

70. Gedrovica, I.; Karklina, D.; Straumite, E. Sensory and qualitative indices (hardness and color) evaluation of cakes with Jerusalem artichoke (*Helianthus tuberosus* L.) powder. *Res. Rural Dev.* **2010**, *1*, 138–144.

- 71. Celik, I.; Isik, F.; Gursoy, O.; Yilmaz, Y. Use of Jerusalem Artichoke (*Helianthus tuberosus*) tubers as a natural source of inulin in cakes. *J. Food Process. Preserv.* **2012**, 37, 483–488. [CrossRef]
- 72. Park, G. Optimization of muffin preparation upon addition of Jerusalem artichoke powder and oligosaccharide by response surface methodology. *J. Korean Soc. Food Cult.* **2014**, 29, 101–110. [CrossRef]
- 73. Baltacioglu, C.; Esin, A. Chips production from Jerusalem artichoke (*Helianthus tuberosus* L.). Food Nutr. Sci. **2012**, 3, 1321–1328. [CrossRef]
- 74. Khuenpet, K.; Jittanit, W.; Sirisansaneeyakul, S.; Srichamnong, W. Inulin powder production from Jerusalem artichoke (*Helianthus tuberosus* L.) tuber powder and its application to commercial food products. *J. Food Process. Preserv.* **2016**, *41*, e13097. [CrossRef]