

**Supplementary Information**

Supplementary Table 1. Original dataset containing 1002 patients with esophageal cancer that underwent surgical therapy. 110 patient characteristics are listed with amount and percentage if not otherwise depicted. Missing variables are not listed individually, but represent the remainder if the sum of the individual variables does not add up to 100 percent.

	No.	%
Total patients (<i>n</i>)	1002	
CLINICAL CHARACTERISTICS		
Age at surgical treatment, mean \pm SD (y)	62.2 ± 10.1	
Sex assigned at birth		
Female	157	15.7
Male	845	84.3
BMI, mean \pm SD (kg/m ²)	25.9 ± 4.6	
Height, mean \pm SD (cm)	176 ± 9.5	
Weight at primary staging (kg)	83.3 ± 18.3	
Weight loss		
yes/no	100/ 114	9.98/ 11.3
kg, mean \pm SD	4.9 ± 7.5	
months, mean \pm SD	2.7 ± 3.6	
Ogilvie Dysphagia Scale		
0	53	5.3
1	58	5.8
2	44	4.4
3	23	2.3
4	6	0.6
Duration dysphagia, mean \pm SD (y)	2.3 ± 2.4	
Preoperative WBC count ($\times 10^9/L$)		
< 4.4	33	3.3
> 4.4	668	66.7
Preoperative platelet count ($\times 10^9/L$)		
< 150	18	1.8
> 150	683	68.2
ASA-Score		
1	31	3.1
2	394	39.3
3	271	27.05
4	5	0.5
MEDICAL HISTORY		
ECOG-Score		
0	195	19.5

1	207	20.7
2	62	61.9
3	9	0.9
Arterial hypertension, yes/no	309/ 204	30.8/ 20.4
Antiplatelet agents		
<i>none</i>	453	45.2
<i>any</i>	15	1.5
<i>ASA</i>	93	9.3
AT-II receptor antagonist, yes/no	84/ 429	8.4/ 42.8
ACE-inhibitor, yes/no	113/ 399	11.3/ 39.8
Statins, yes/no	97/ 417	9.7/ 41.6
Diuretics, yes/no	116/ 398	11.6/ 39.7
Beta-blocker, yes/no	164/ 349	16.4/ 34.8
Calcium channel blocker, yes/no	66/ 448	6.6/ 44.7
Smoking history		
<i>Non-smoker</i>	279	27.8
<i>Smoker</i>	153	15.3
<i>Ex-smoker</i>	177	17.7
Alcohol consumption		
<i>No alcohol</i>	326	32.5
<i>1-3 times/week</i>	41	4.1
<i>4-5 times/week</i>	8	0.8
<i>daily</i>	35	3.5
MEDICAL TREATMENT		
Neoadjuvant therapy, yes/no	692/ 263	69.1/ 26.2
Type of neoadjuvant therapy		
<i>CROSS protocol [1]</i>	401	40.02
<i>FLOT protocol [2]</i>	109	10.9
<i>other</i>	204	20.4
One-/Two-stage procedure		
<i>One-stage</i>	667	66.6
<i>Two-stage</i>	34	3.4
Surgical procedure (Ivor-Lewis- Esophagectomy)		
<i>Open</i>	80	7.9
<i>Hybrid</i>	587	58.6
<i>Totally minimally invasive</i>	15	1.5
<i>Robot-Assisted</i>	18	1.8
<i>Minimally Invasive</i>		

COMPLICATIONS			
Clavien-Dindo-classification			
<i>none</i>	274	27.3	
1	30	2.9	
2	59	5.9	
3a	230	22.9	
3b	31	3.1	
4a	61	6.1	
4b	14	1.4	
5	1	0.1	
Postoperative pneumonia, yes/no	72/ 629	7.2/ 62.8	
Postoperative pylorospasm, yes/no	139/ 513	13.9/ 51.2	
Anastomosis leak, yes/no	50/ 651	4.9/ 64.9	
Secondary chest drainage, yes/no	57/ 545	5.7/ 54.4	
Re-intubation, yes/no	64/ 636	6.4/ 63.5	
Tracheostomy, yes/no	43/ 656	4.3/ 65.5	
Re-admission intensive-care unit, yes/no	50/ 650	4.9/ 64.9	
TUMOR ASSOCIATED FACTORS			
Histology			
<i>ACC</i>	842	84.03	
<i>SCC</i>	147	14.7	
<i>other</i>	12	1.2	
Adenocarcinoma of esophagogastric junction			
<i>Siewert I</i>	284	28.3	
<i>Siewert II</i>	174	17.4	
<i>Siewert III</i>	12	1.2	
Tumor distance to the incisors, mean ± SD (cm)			
<i>Proximal margin</i>	33.2 ± 5.2		
<i>Distal margin</i>	39.1 ± 4.4		
Tumor stenosis at primary staging (endoscopically), yes/no	135/ 53	13.5/ 5.3	
CLINICAL TNM-STAGING AT PRIMARY STAGING			
cT			
1	31	3.1	
2	49	4.9	
3	269	26.8	
4	3	2.9	
cN			
0	25	24.9	
1	168	16.8	

2	13	1.3
3	3	0.3
χ	111	11.1
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cM		
0	241	24.1
1	27	2.7
χ	24	2.4
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PATHOLOGICAL TNM CLASSIFICATION		
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pT		
0	135	13.5
1a/1b	248	24.8
2	157	15.7
3	440	43.9
4a/b	17	1.7
Tis	1	0.1
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pN		
0	513	511,9
1	250	24.9
2	135	13.5
3	100	9.9
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pL		
0	600	59.9
1	226	22.6
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pV		
0	762	76.04
1	59	5.9
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R		
0	885	88.3
1	40	3.9
2	3	0.3
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Positive lymph nodes, mean \pm SD (n)	2.3 ± 4.7	
Total resected lymph nodes, mean \pm SD (n)	30.8 ± 11.3	
Positive lymph node quotient, mean \pm SD (n)	0.07 ± 0.14	
Pathological tumor margin distance, mean \pm SD (mm)		
<i>Proximal margin</i>	36.7 ± 18.1	
<i>Distal margin</i>	45.2 ± 23.4	
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Tumor regression grade		
1: > 50% rest vital tumor cells (RTV)	127	12.7
2: 10 - 50% RTV	146	14.6
3: < 10 % RTV	130	12.9
4: no RTV	118	11.8
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BIOMARKER		
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Insulin-like growth factor II mRNA-binding protein 3 (IMP-3) [3]			
<i>0: no expression</i>	59	58.9	
<i>1: weak expression</i>	61	6.1	
<i>2: moderate expression</i>	125	12.5	
E-Cadherin			
<i>0: no expression</i>	8	0.8	
<i>1: weak expression</i>	60	5.9	
<i>2: moderate expression</i>	175	17.5	
Carbonic anhydrase IX (CAIX)			
<i>0: no expression</i>	128	12.8	
<i>1: weak expression</i>	62	6.2	
<i>2: moderate expression</i>	43	4.3	
Class III β-tubulin (TUBB3) [4]			
<i>0: no expression</i>	55	5.5	
<i>1: weak expression</i>	66	6.6	
<i>2: moderate expression</i>	55	5.5	
<i>3: strong expression</i>	52	5.2	
TP-53 mutation [5]			
<i>0: mutant = no expression</i>	97	9.7	
<i>1: wild-type protein</i>	111	11.07	
<i>2: mutant = overexpression</i>	282	28.1	
Ki-67 (expression, %)			
<i>1: 1% – 9%</i>	97	9.7	
<i>2: 10% – 19%</i>	42	4.2	
<i>3: 20% – 39%</i>	84	8.4	
<i>4: ≥40%</i>	66	6.6	
Murine double minute-2 (MDM-2), IHC			
<i>0: negative</i>	267	26.6	
<i>1: weak/moderate, few cells</i>	16	15.9	
<i>2: weak/moderate, many cells</i>	16	15.9	
<i>3: strong expression</i>	6	0.6	
MDM2, FISH [5]			
<i>0: negative amplification</i>	288	28.7	
<i>1: positive amplification</i>	16	1.6	
Lymphocyte activation gene-3 (LAG-3)			
<i>0: negative</i>	200	19.9	
<i>1: 1-2%</i>	59	5.9	
<i>2: 3-5%</i>	15	1.5	

High mobility group box 1 (HMGB-1)			
0: no expression	84	8.4	
1: weak expression	59	5.9	
2: moderate expression	65	6.5	
3: strong expression	137	13.7	
MutL homolog 1 (MLH-1)			
0: Microsatellite instability	15	1.5	
1: Microsatellite stable	493	49.3	
CD-3			
1: low expression	198	19.8	
2: high expression	72	7.2	
CD3, expression tumor center [6]			
0: peritumoral	248	24.8	
2: intratumoral	22	2.2	
X-linked inhibitor of apoptosis protein (XIAP) [7]			
0: no expression	25	2.5	
1: weak expression	118	11.8	
2: moderate expression	84	8.4	
3: strong expression	38	3.8	
Mesothelin (Novocastra antibody) [8]			
0: no expression	208	20.8	
1: low/moderate	74	7.4	
2: high	42	4.2	
Mesothelin (Ventana antibody) [8]			
0: no expression	155	15.4	
1: low/moderate	92	9.2	
2: high	73	7.3	
Claudin (expression, %)			
0: no expression	267	26.6	
1: ≥5% - 49%	38	3.8	
2: ≥50%	26	2.6	
AT-rich interactive domain 1A (ARID1A) [9]			
0: mutated/deleted gene	43	4.3	
1: intact gene	464	46.3	
SMARCA4 (BRG-1) [9]			
0: mutated/deleted gene	11	1.1	
1: intact gene	432	43.1	
SMARCA2 (BRM) [9]			
0: mutated/deleted gene	39	3.9	
1: intact gene	414	41.3	

Indoleamine 2,3-dioxygenase (IDO) expressed on tumor infiltrating lymphocytes [10]			
0: negative expression	168	16.8	
1: low inflammation	123	12.3	
2: high inflammation	52	5.2	
Indoleamine 2,3-dioxygenase (IDO) expressed on tumor cells			
0: no expression	300	29.9	
1: weak expression	30	2.9	
2: high expression	14	1.4	
T-cell immunoglobulin and mucin-domain containing-3 (TIM-3)			
0: negative expression	158	15.8	
1: low inflammation	109	10.9	
2: high inflammation	67	6.7	
Aldo-keto reductase family 1 (AKR1)			
0: negative	35	3.5	
1: low level	120	11.9	
2: intermediate level	86	8.6	
3: high level	52	5.2	
Gremlin1 (GREM1)			
0: negative	22	2.2	
1: low level	142	14.2	
2: high level	64	6.4	
c-MYC			
0: no amplification	294	29.3	
1: low amplification	22	2.2	
2: high amplification	41	4.1	
KRAS			
0: negative amplification	296	29.5	
1: positive amplification	70	6.9	
GATA-binding factor (GATA6) [11]			
0: negative amplification	311	31.03	
1: positive amplification	33	3.3	
Her2/neu, IHC [12]			
0: negative staining	438	43.7	
1: very weak	1	0.1	
2: weak/moderate	15	1.5	
3: strong	29	2.9	
Her2/neu, FISH [12]			
0: negative amplification	326	32.5	
1: positive amplification	31	3.1	

Loss of Y-chromosome (LoY), long arm [13]			
0: negative	170	16.9	
1: positive	125	12.5	
LoY, short arm [13]			
0: negative	164	16.4	
1: positive	131	13.1	
Phosphatase and tensin homolog deleted on chromosome 10 (PTEN)			
0: negative	32	3.2	
1: weak signal	74	7.4	
2: normal signal	296	29.5	
Fructose-1,6-bisphosphatase 1 (FBP1) [14]			
0: no expression	65	6.4	
1: weak	94	9.4	
2: moderate	133	13.3	
3: strong	78	7.8	
Ubiquilin-4 (UBQLN4, dots/tumorcells)			
0: 0 or < 1	71	7.1	
1: 1-3	125	12.5	
2: 4-9	105	10.5	
3: ≥ 10	18	1.8	
Trimethylation of lysine 27 on histone H3 (H3K27me3)			
0: negative	42	4.2	
1: partly positive	67	6.7	
2: completely positive	185	18.5	
p16, IHC			
0: negative	276	27.5	
1: partly positive	44	4.4	
2: completely positive	63	6.3	
p16, FISH			
0: no deletion	202	20.2	
1: hemizygous deletion	35	3.5	
2: homozygous deletion	78	7.8	
Trophoblast Cell Surface Antigen 2 (TROP2) [15], (H-Score expression level)			
0: no expression	43	4.3	
1: weak expression	85	8.5	
2: moderate expression	183	18.3	
3: strong expression	76	7.6	
AKR			
0: no expression	28	2.8	
1: weak expression	88	8.8	

	<i>2: moderate expression</i>	120	11.9
	<i>3: strong expression</i>	104	10.4
F-box and WD repeat domain containing 7 (FBXW7)			
	<i>0: negative</i>	28	2.8
	<i>1: weak signal</i>	59	5.9
	<i>2: normal signal</i>	294	29.3
Cyclin D1, IHC			
	<i>0: no expression</i>	161	16.1
	<i>1: weak expression</i>	166	16.6
	<i>2: moderate expression</i>	59	5.9
	<i>3: strong expression</i>	4	0.4
Cyclin D1, FISH			
	<i>0: no amplification</i>	274	27.3
	<i>1: polyribosome</i>	12	1.2
	<i>2: amplification</i>	13	1.3
Cyclin Dependent Kinase 6 (CDK6), FISH			
	<i>0: no amplification</i>	169	16.9
	<i>2: amplification</i>	13	1.3
Periostin			
	<i>0: negative</i>	93	9.3
	<i>1: >5% - 30%</i>	178	17.8
	<i>2: >30%</i>	226	22.6
Matrix metalloproteinase-2 (MMP-2)			
	<i>0: no expression</i>	99	9.9
	<i>1: weak expression</i>	226	22.6
	<i>2: moderate expression</i>	159	15.9
Homeobox protein NANOG			
	<i>0: no expression</i>	130	12.9
	<i>1: weak expression</i>	127	12.7
	<i>2: moderate expression</i>	235	23.5
Cytokeratin 6 (CK6)			
	<i>0: negative</i>	369	36.8
	<i>1: weak positive</i>	74	7.4
	<i>2: strong positive</i>	46	4.6
N-myc down-regulated gene 1 (NDRG1)			
	<i>0: no expression</i>	46	4.6
	<i>1: weak expression</i>	109	10.9
	<i>2: moderate expression</i>	92	9.2
	<i>3: strong expression</i>	134	13.4
Thymidine phosphorylase (TYMP)			
	<i>0: no expression</i>	152	15.2
	<i>1: weak expression</i>	160	15.9

	<i>2: moderate expression</i>	120	11.9
	<i>3: strong expression</i>	57	5.7
Growth differentiation factor 15 (GDF-15), tumor cells			
	<i>0: negative</i>	80	7.9
	<i>1: low</i>	96	9.6
	<i>2: medium</i>	76	7.6
	<i>3: strong</i>	6	0.6
GDF15, stroma			
	<i>0: negative</i>	54	5.4
	<i>1: low</i>	111	11.1
	<i>2: medium</i>	87	8.7
	<i>3: strong</i>	6	0.6
Mast cells			
	<i>0: negative</i>	171	17.1
	<i>1: weak positive</i>	278	27.8
	<i>2: strong positive</i>	102	10.2
Natural killer cells (NK cells)			
	<i>0: negative</i>	329	32.8
	<i>1: weak positive</i>	217	21.7
	<i>2: strong positive</i>	8	0.8
Tumor-infiltrating plasma cells (TIP)			
	<i>0: negative</i>	285	28.4
	<i>1: weak positive</i>	196	19.6
	<i>2: strong positive</i>	72	7.2

Supplementary Table 2. Hyperparameters for each model after optimization. Randomized and Grid Search Cross Validation was used for Random Forest (RF), XG-Boost (XG) and Logistic Regression (LR). Best hyperparameters for Artificial Neural Network (ANN) and TabNet (TN) were selected using *Optuna*, an open-source optimization framework.

	Baseline Dataset	Primary Staging Dataset (PS)	PS Dataset including tumor biomarker
RF	n_estimators= 62, max_depth = 19, min_samples_split = 2, min_samples_leaf = 3, bootstrap=True, max_features='auto'	n_estimators= 65, max_depth = 15, min_samples_split = 2, min_samples_leaf = 3, bootstrap=False, max_features='auto'	n_estimators= 45, max_depth = 13, min_samples_split = 3, min_samples_leaf = 2, bootstrap=False, max_features='auto'
<i>AI-driven Reduced Datasets</i>	n_estimators= 45, max_depth = 17, min_samples_split = 4, min_samples_leaf = 3, bootstrap=True, max_features='auto'		n_estimators= 36, max_depth = 22, min_samples_split = 2, min_samples_leaf = 3, bootstrap=True, max_features='auto'
XG	n_estimators = 79, max_depth = 12, subsample = 0.7, colsample_bytree = 0.6, learning_rate = 0.05, reg_lambda = 0.7, gamma = 0.1	n_estimators = 68, max_depth = 8, subsample = 0.8, colsample_bytree = 0.7, learning_rate = 0.05, reg_lambda = 1, gamma = 0.1	n_estimators = 75, max_depth = 5, subsample = 0.8, colsample_bytree = 0.5, learning_rate = 0.05, gamma = 0.4, reg_lambda = 0.5
<i>AI-driven Reduced Datasets</i>	n_estimators = 29, max_depth = 9, subsample = 0.5, colsample_bytree = 0.6, learning_rate = 0.05, reg_lambda = 0.5, gamma = 0.1,		n_estimators = 63, max_depth = 3, subsample = 0.6, colsample_bytree = 0.5, learning_rate = 0.1, reg_lambda = 0.1, gamma = 0.0
ANN	embed_p=0.35, ps=0.5, layers=[120], n_epochs = 8, lr_max=0.00898	embed_p=0.2, ps=0.3, layers=[25, 140], n_epochs = 15, lr_max=0.003804471	embed_p=0.6, ps=0.45, layers=[100, 75], n_epochs = 8, lr_max=0.00500528
<i>AI-driven Reduced Datasets</i>	embed_p=0.25, ps=0.55, layers=[140, 140], n_epochs = 16, lr_max=0.006370		embed_p=0.8, ps=0.7, layers=[75], n_epochs = 17, lr_max=0.0044824595
TN	mask_type = entmax, cat_emb_dim = 5, optimizer_fn=torch.optim.Adam, n_a = 16, n_d = 60, n_shared = 4,	mask_type = entmax, cat_emb_dim = 4, optimizer_fn=torch.optim.Adam, n_a = 60, n_d = 16, n_shared = 1,	mask_type = entmax, cat_emb_dim = 2, optimizer_fn=torch.optim.Adam, n_a = 40, n_d = 36, n_shared = 5,

	n_steps = 3, n_independent = 1, optimizer_params=dic t(lr=2e-2), scheduler_params= step_size:12, gamma:1.0, scheduler_fn=torch.op tim.lr_scheduler.StepL R,	n_steps = 1, n_independent = 1, optimizer_params=dic t(lr=2e-2), scheduler_params=ste p_size:12, gamma":1.4, scheduler_fn=torch.op tim.lr_scheduler.StepL R	n_steps = 5, n_independent = 1, optimizer_params=dic t(lr=2e-2), scheduler_params=ste p_size:20, gamma:1.4, scheduler_fn=torch.op tim.lr_scheduler.StepL R
<i>AI- driven Reduced Datasets</i>	mask_type = 'entmax', cat_emb_dim = 4, optimizer_fn=torch.op tim.Adam, n_a = 12, n_d = 48, n_shared = 1, n_steps = 5, n_independent = 3, optimizer_params=dic t(lr=2e-2), scheduler_params=ste p_size:10, gamma:1.4, scheduler_fn=torch.op tim.lr_scheduler.StepL R,	mask_type = 'entmax', cat_emb_dim = 1, optimizer_fn=torch.op tim.Adam, n_a = 52, n_d = 28, n_shared = 3, n_steps = 9, n_independent = 2, optimizer_params=dic t(lr=2e-2), scheduler_params=ste p_size:6, gamma:1.2, scheduler_fn=torch.op tim.lr_scheduler.StepL R	mask_type = 'entmax', cat_emb_dim = 1, optimizer_fn=torch.op tim.Adam, n_a = 52, n_d = 28, n_shared = 3, n_steps = 9, n_independent = 2, optimizer_params=dic t(lr=2e-2), scheduler_params=ste p_size:6, gamma:1.2, scheduler_fn=torch.op tim.lr_scheduler.StepL R
LR	penalty = none, max_iter = 2000, solver = saga	penalty = none, max_iter = 3000, solver = sag	penalty = none, max_iter = 5000, solver = saga

Supplementary References

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