

Abstract

Experimental Model for High-Throughput Screening of Microalgae Strains Useful for CO₂ Fixation [†]

Eliza-Gabriela Mihaila ^{1,2}, Daria Gabriela Popa ^{1,3}, Maria Daria Dima ⁴, Ioana Marcela Stoian ⁵, Cristian Florian Dinca ², Diana Constantinescu-Aruxandei ¹  and Florin Oancea ^{1,3,*} 

¹ INCDCP-ICECHIM Bucharest, 202 Spl. Independentei, 6th District, 060021 Bucharest, Romania; eliza-gabriela.mihaila@icechim.ro (E.-G.M.); daria.popa@icechim.ro (D.G.P.); diana.constantinescu@icechim.ro (D.C.-A.)

² Power Engineering Faculty, University Politehnica Bucharest, 060042 Bucharest, Romania; cristian.dinca@upb.ro

³ Faculty of Biotechnologies, University of Agronomic Sciences and Veterinary Medicine of Bucharest, 011464 Bucharest, Romania

⁴ International Computer High School of Bucharest, 032622 Bucharest, Romania; maria.daria0033@gmail.com

⁵ Faculty of Applied Chemistry and Materials Science, University Politehnica Bucharest, 060042 Bucharest, Romania; ioana_stoian18@yahoo.com

* Correspondence: florin.oancea@icechim.ro

[†] Presented at the 17th International Symposium “Priorities of Chemistry for a Sustainable Development” PRIOCHEM, Bucharest, Romania, 27–29 October 2021.

Keywords: CO₂; microalgae; experimental model; high throughput screening



Citation: Mihaila, E.-G.; Popa, D.G.; Dima, M.D.; Stoian, I.M.; Dinca, C.F.; Constantinescu-Aruxandei, D.; Oancea, F. Experimental Model for High-Throughput Screening of Microalgae Strains Useful for CO₂ Fixation. *Chem. Proc.* **2022**, *7*, 25. <https://doi.org/10.3390/chemproc2022007025>

Academic Editors: Mihaela Doni, Zina Vuluga and Radu Claudiu Fierăscu

Published: 7 March 2022

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In this study, we developed an experimental model for microalgae cultivation and CO₂ fixation. We used three different species of microalgae and several cultivation media. The industrial gas emissions contain a significant proportion of CO₂ (3–30%) [1]. Addition of extra CO₂ to microalgae culture initially boosts its development, but further acidification processes limit microalgae development [2]. In this study, three strains of microalgae were cultivated: *Chlorella sorokiniana* NIVA-CHL 176, *Desmodesmus communis* NIVA-CHL 7, and *Raphidocelis subcapitata* ATCC22662, using three different cultivation media, BG11 [3], BBM, and Z8, respectively. The experimental model used for the cultivation of the microalgae was developed using a GLS80 glass reactor with an LED stripe for illumination. Nitrogen containing 7% CO₂ was discontinuously added (90 min/day), at 25 °C, 200 RPM and approximately 10 μE, in order to avoid acidification of the cultivation medium [4]. Microalgae growth was monitored via optical density, turbidity, chlorophyll content, biomass, pH, and cell number. The best cultivation protocol was selected after the initial experiments. *C. sorokiniana* NIVA-CHL 176 was raised in BG11 medium and *D. communis* NIVA CHL-7 in Z8 medium. *R. subcapitata* ATCC22662 had around half the growth rate of the other two microalgae, and experiments were discontinued. Subsequent to CO₂ bubbling, the pH dropped with one unit after the first 7 days (from pH 6 to 5), from day 7 to day 10 the values were constant (pH = 4), and, for 4 more days, the pH increased (to 6 and 6.5, depending on the microalgae). The control culture had a constant pH of 8. Based on optical density, the growth rates of the studied microorganisms were monitored. *D. communis* had the best results, and showed eight times greater growth rate than the control, and *C. sorokiniana* was two times greater compared to the control, not supplemented with CO₂. A higher rate of CO₂ significantly increases growth rates, but pH monitoring is needed. Our experimental model is efficient for high-throughput screening of microalgae strains useful for CO₂ fixation.

Author Contributions: Conceptualization, F.O. and D.C.-A.; methodology, E.-G.M. and D.G.P.; software, C.F.D.; validation, E.-G.M., D.C.-A. and F.O.; formal analysis, M.D.D.; investigation, I.M.S.; resources, F.O.; data curation, D.C.-A.; writing—original draft preparation, E.-G.M.; writing—review and editing, D.C.-A. and F.O.; visualization, D.G.P.; supervision, F.O.; project administration, C.F.D.; funding acquisition, C.F.D. and F.O. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the Government of Romania, Ministry of Research and Innovation, UEFISCDI Contract296PED/2020 ASOCIAT.

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Data Availability Statement: Not applicable.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Bholá, V.; Swalaha, F.; Ranjith Kumar, R.; Singh, M.; Bux, F. Overview of the potential of microalgae for CO₂ sequestration. *Int J. Environ. Sci. Technol.* **2014**, *11*, 2103–2118. [[CrossRef](#)]
2. Ho, S.H.; Chena, C.Y.; Lee, D.J.; Chang, J.S. Perspectives on microalgal CO₂-emission mitigation systems—A review. *Biotechnol. Adv.* **2011**, *29*, 189–198. [[CrossRef](#)] [[PubMed](#)]
3. Rosa, G.M.; Morais, M.G.; Costa, J.V.A. Fed-batch cultivation with CO₂ and monoethanolamine: Influence on *Chlorella fusca* LEB 111 cultivation, carbon biofixation and biomolecules production. *Bioresour. Technol.* **2019**, *273*, 627–633. [[CrossRef](#)] [[PubMed](#)]
4. Qiu, R.; Gao, S.; Lopez, P.A.; Ogden, K.O. Effects of pH on cell growth, lipid production and CO₂ addition of microalgae *Chlorella sorokiniana*. *Algal Res.* **2017**, *28*, 192–199. [[CrossRef](#)]