



Article **Circulating miRNAs Are Associated with Inflammation** Biomarkers in Children with Overweight and Obesity: Results of the I.Family Study

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Abstract: Increasing data suggest that overnutrition-induced obesity may trigger an inflammatory process in adipose tissue and upturn in the innate immune system. Numerous players have been involved in governing the inflammatory response, including epigenetics. Among epigenetic players, miRNAs are emerging as crucial regulators of immune cell development, immune responses, autoimmunity, and inflammation. In this study, we aimed at identifying the involvement of candidate miRNAs in relation to inflammation-associated biomarkers in a subsample of European children with overweight and obesity participating in the I.Family study. The study sample included individuals with increased adiposity since this condition contributes to the early occurrence of chronic low-grade inflammation. We focused on the acute-phase reagent C-reactive protein (CRP) as the primary outcome and selected cytokines as plausible biomarkers of inflammation. We found that chronic low-grade CRP elevation shows a highly significant association with miR-26b-3p and hsa-miR-576-5p in boys. Furthermore, the association of CRP with hsa-miR-10b-5p and hsa-miR-31-5p is highly significant in girls. We also observed major sex-related associations of candidate miRNAs with selected cytokines. Except for IL-6, a significant association of hsa-miR-26b-3p and hsa-miR-576-5p with TNF- α , IL1-Ra, IL-8, and IL-15 levels was found exclusively in boys. The findings of this exploratory study suggest sex differences in the association of circulating miRNAs with inflammatory response biomarkers, and indicate a possible role of miRNAs among the candidate epigenetic mechanisms related to the process of low-grade inflammation in childhood obesity.

Keywords: miRNAs; chronic low-grade inflammation; inflammation-associated biomarkers; overweight and obesity; children/adolescents; sex-related associations

1. Introduction

During the past decade, obesity has become a global epidemic with substantial health, social, and economic implications. According to the World Health Organization, there are



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nearly 2 billion overweight adults worldwide and approximately 600 million of them are obese [1,2]. Numerous studies have recognized a strong association between an increased body mass index (BMI) and quality and expectancy of life given its interconnected effects with type 2 diabetes, hyperlipidemia, non-alcoholic fatty liver disease, hypertension, heart disease, stroke, arthritis, cancer, depression, asthma, psychological problems, and other non-communicable diseases [3–5].

Despite extensive investigation, most of the genetic variability in obesity remains unresolved and the influence of candidate genes in this context appears limited. Numerous studies have suggested that genes that predispose to obesity are not causal but act in conjunction with a variety of individual, environmental, and lifestyle factors, including obesogenic environments, low levels of education, sedentary habits, and reduced sleep hours, among others [6]. A highly energy-dense diet, as well as low physical activity levels, are considered driving factors [3]. Evidence also indicates that offspring born to obese mothers are prone to a higher BMI and fat accumulation, thus supporting the conceivable transgenerational risk contribution based on epigenetic mechanisms [7].

The basic tissue for energy storage in humans is the white adipose tissue. Aside from its storing function, this tissue is metabolically active. It releases hundreds of different factors, such as hormones including adiponectin and leptin, growth factors including Insulin-Like Growth Factor-1 (IGF-1) and Platelet-Derived Growth Factor (PDGF), and inflammatory mediators including Tumor Necrosis Factor- α (TNF- α), Interleukin-6 (IL-6), and Interleukin-8 (IL-8), all of which contribute to insulin sensitivity and dietary control. Growing data suggest that overnutrition-induced obesity triggers an inflammatory process in adipose tissue [8], with even moderate weight gain linked to inflammatory activation [9] and an upturn in the innate immune system [10]. Moreover, chemokines and their receptors also contribute to obesity development by activating the resident immune surveillance system [11]. This state is firmly interconnected to the local recruitment of macrophages and enhanced immune cell infiltration/proliferation/activation [12]. All these processes collectively point toward adipocyte hypertrophy and impaired adipogenesis [13]. The resulting persistent chronic low-grade inflammation represents a hallmark of obesity that leads to and perpetuates the state of metabolic alterations and insulin resistance in target organs, including the adipose tissue, liver, muscles, and vascular system [14]. Among the inflammatory biomarkers, the acute-phase reactant C-reactive protein (CRP) is considered the major factor associated with overweight and obesity in both adults and children [15,16].

In recent years, numerous biochemical players have been recognized as being involved in the inflammatory response, including microRNAs (miRNAs), small non-coding RNAs expressed in a wide variety of organs and cells and capable of potentially influencing almost all cellular functions. miRNAs are constituents of the epigenetic mechanisms that finely regulate the expression of messenger RNAs [17]. Until now, 2599 different miRNAs have been documented in humans (miRTarBase, release 8.0) [18]. Growing evidence also underlines their relevance as stable, non-invasive, and reliable biomarkers for a variety of pathophysiological conditions [19–21], including the body's energy balance [22]. Many studies have established that altered miRNA profiles are interconnected with obesity [23] and other non-communicable diseases [24,25]. However, nowadays, epigenetics represents a critical but still poorly understood factor among the known molecular mechanisms related to the process of low-grade inflammation in obesity [26]. Among epigenetic players, miRNAs are progressively emerging as key regulators of immune cell development, immune responses, autoimmunity, and inflammation, capable of affecting both pro-and anti-inflammatory responses [26]. Indeed, by influencing definite signaling networks in the immune system, so-called immuno-miRs have been shown to impact both innate and adaptive immune responses in health and disease [27]. Of note, immuno-miRs may exert different functions in different cell types. Therefore, during inflammation, different cells undergo substantial transcriptional activation, thus presenting different sets of targets regulated by a given miRNA.

In this study, we aimed to identify the potential involvement of candidate miRNAs in relation to inflammation-associated biomarkers in a subsample of European children with overweight and obesity (OW/Ob) participating in the I.Family study (www.ifamilystudy.eu, last accessed on 21 February 2022) [28,29]. The primary outcome of this study was the association of miRNA expression with CRP. The association of miRNAs with the selected cytokines constituted a secondary outcome of the study. The study sample included individuals with OW/Ob since this condition is a known risk factor contributing to the early occurrence of chronic low-grade inflammation and interconnected metabolic changes.

2. Materials and Methods

2.1. Participants

The study was conducted on a subgroup of European children/adolescents with OW/Ob belonging to the I.Family study, an EC-funded project aiming at investigating determinants of food choice, lifestyle, and related health outcomes in children and adolescents of eight European countries (Belgium, Cyprus, Estonia, Germany, Hungary, Italy, Spain, and Sweden). A full description of the study designs, selection criteria, and participants' characteristics, from which the subsample data are drawn, has been previously published [30]. A complete explanation of the I.Family study (registration number IS-RCTN62310987) has been earlier reported [29]. The study was conducted according to the criteria set by the Declaration of Helsinki. Approval by the national ethics committees was obtained by each of the participating centers. Anthropometric characteristics, pubertal status, and dietary intake data were collected using standardized procedures; a full description of these methods has been previously published [31,32]. In each country, we first selected 20 children (n = 160) who retained overweight or obesity, i.e., who had a BMI z-score of more than + 1 at baseline and after 2 years at follow-up, respectively, and did not change more than ± 0.1 in BMI z-score per year (defined as overweight/obese) [33]. Out of 160 subjects, the current analysis was performed on a subsample of 79 overweight and obese children/adolescents (Belgium n = 7, Cyprus n = 6, Estonia n = 13, Germany n = 16, Hungary n = 8, Italy n = 12, Spain n = 5, and Sweden n = 12), with a complete dataset for the variables of interest, after the exclusion of 19 participants with hemolyzed samples.

2.2. Biochemical Analysis

The fasting venous blood draws were collected in BD Vacutainer[®] blood tubes according to standardized operative procedures. A complete description of the sample collection and investigative procedures has been earlier published [34]. Clinical chemistry tests were determined as part of routine laboratory testing, in a central laboratory (University of Bremen, Centre for Biomolecular Interactions Bremen—CBIB). Serum samples stored at -80 °C were used to detect levels of CRP, Interleukin-1 Receptor Antagonist (IL1-Ra), IL-6, IL-8, Interleukin-15 (IL-15), and TNF- α using an electrochemiluminescent multiplex assay (using either single or MULTI-SPOT[®] Assay Systems, Meso Scale Discovery).

2.3. miRNA Profiling

Taking advantage of the qPCR array technology, we previously reported that an altered circulating miRNA profile is associated with OW/Ob in children and adolescents [30]. In that study, we also identified four circulating miRNAs, hsa-miR-10b-5p (MIMAT0000254), hsa-miR-26b-3p (MIMAT0004500), hsa-miR-31-5p (MIMAT000089), hsa-miR-576-5p (MI-MAT0003241), potentially linked to increased CRP levels in subjects with OW/Ob (unpublished data). Of note, among the four miRNAs characterized, only hsa-miR-10b-5p was confirmed to be associated with OW/Ob. In the current investigation, we aimed to confirm the association of the candidate miRNAs with levels of CRP and the selected inflammatory biomarkers through validation by SYBR green-based real-time quantitative RT-PCR (RT-qPCR) in the new sample of children and adolescents with OW/Ob. Protocols for miRNA extraction and screening from plasma samples have been earlier published [30,32]. Individual plasma samples were first tested for hemoglobin levels and hemolyzed samples

were excluded from the analysis [30]. Different assays were performed in triplicate employing the miScript Primer Assays according to the manufacturer's instructions (Qiagen, Germany). miRNA relative levels were determined using the Cel-miR-39 spike-in as the endogenous normalizer [30]. Levels were calculated using the Data Assist v3.1 software package (Life Technologies, Thermo Fisher Scientific, Milan, Italy).

2.4. Statistical Analysis

Statistical analyses were achieved by using IBM SPSS Statistics software (v24.0. Armonk, NY, USA: IBM Corp.). Data collected were calculated as means and 95% confidence intervals (CIs). Associations of miRNA expression with CRP and the different interleukins were assessed using linear regression analyses, adjusting for covariates including country of residence, age, BMI *z*-score, pubertal status. Since potential sex disparities in CRP levels have been previously reported [35], we considered boys and girls separately in the statistical analysis. To control for the false discovery rate (FDR), the Benjamini–Hochberg (BH) method was adopted. The level of statistical significance was set at $\alpha < 0.05$.

3. Results

3.1. Anthropometric Characteristics and Biochemical Markers of the Study Sample

The anthropometric and metabolic characteristics and levels of inflammatory markers of the 79 participants are reported in Table 1. There were no obvious differences regarding the anthropometric characteristics and tested biochemical markers between males and females.

| Ow/Ob | Boys (25/11) | Girls (31/12) | | |
|-----------------------|---------------------|---------------------|--|--|
| Age (years) | 12.1 (11.5–12.8) | 12.4 (11.9–12.9) | | |
| Puberty (% yes) | 47.8 | 52.2 | | |
| BMI z-score | 1.8 (1.6–2.0) | 1.7 (1.5–1.9) | | |
| CRP (mg/dL) | 0.36 (0.08–0.63) | 0.45 (0.14-0.77) | | |
| TNF- α (pg/mL) | 4.3 (2.7-6.0) | 3.9 (2.4–5.4) | | |
| IL-1Ra (pg/mL) | 422.9 (339.7–506.2) | 527.8 (398.0-657.6) | | |
| IL-6 (pg/mL) | 1.2 (-0.1-2.5) | 0.8 (0.5–1.0) | | |
| IL-8 (pg/mL) | 38.5 (-23.1-100.0) | 17.6 (1.7–33.6) | | |
| IL-15 (pg/mL) | 2.6 (2.2–3.0) | 3.0 (2.5–3.4) | | |
| hsa-miR-10b-5p | 2.9 (2.1–3.6) | 3.3 (2.7–3.9) | | |
| hsa-miR-26b-3p | 2.4 (1.6–3.3) | 2.7 (0.7-4.6) | | |
| hsa-miR-31-5p | 1.1 (0.3–1.8) | 1.5 (0.8–2.3) | | |
| hsa-miR-576-5p | 7.3 (5.4–9.1) | 6.4 (5.0–7.8) | | |

Table 1. Anthropometric and chemical characteristics of the study sample.

Data are expressed as mean (CIs) or as frequency (%).

3.2. RT-qPCR Validation in Individual Plasma Samples

After plasma extraction, the single candidate miRNAs were determined in individual assays by RT-qPCR. Differences in miRNA signatures with respect to anthropometric and biochemical variables were investigated. Associations of miRNA expression levels with CRP, the primary outcome of this study, and the selected interleukins were assessed using linear regression analysis stratified by sex. Results reported in Table 2 are adjusted for covariates including country of residence, age, BMI z-score, and pubertal status. CRP values show a significant association with miR-26b-3p and hsa-miR-576-5p exclusively in boys. Moreover, their associations with hsa-miR-10b-5p and hsa-miR-31-5p are highly significant only in girls. In Figure S1 is reported the distribution of selected miRNAs in relation to CRP levels in the girls' and boys' subgroups.

| | Boys (36) | <i>q</i> -Value | Girls (43) | q-Value |
|----------------|----------------|-----------------|----------------|---------|
| hsa-miR-10b-5p | 2.7 (1.7–3.7) | 0.399 | 3.6 (3.0-4.2) | 0.008 |
| hsa-miR-26b-3p | 3.2 (3.0-3.4) | 0.004 | 2.1 (-0.9-5.0) | 0.553 |
| hsa-miR-31-5p | 1.1 (-0.3-2.1) | 0.914 | 1.3 (0.9–1.8) | 0.02 |
| hsa-miR-576-5p | 8.3 (6.8–9.8) | 0.006 | 6.8 (5.7-8.0) | 0.187 |

Table 2. Association of miRNA expression levels with CRP.

Data are expressed as mean (CIs). Covariates: Country of residence, age, BMI z-score, pubertal status. *q*-values are BH-adjusted *p*-values. Values in bold indicate statistically significant results.

Table 3 reports the results of the associations of candidate miRNAs with the cytokines selected as additional inflammation biomarkers. A significant association of both hsa-miR10b-5p and hsa-miR-26b-3p with TNF- α , IL1-Ra, IL-8, and IL-15 levels was found exclusively in boys. No association of candidate miRNAs with IL-6 has been established. Moreover, none of the candidate miRNAs was associated with cytokine levels in girls.

Table 3. Association of miRNA expression levels with selected cytokines (secondary outcome).

| Cytokine | Sex | miR-10b-5p | q-Value | miR-26b-3p | q-Value | miR-31-5p | q-Value | miR-576-5p | q-Value |
|----------|-------|---------------|---------|----------------|---------|-----------------|---------|----------------|---------|
| TNF-α | Boys | 2.5 (2.0-3.0) | 1.000 | 3.0 (2.6–3.4) | 0.006 | 1.3 (0.3–2.2) | 0.963 | 7.7 (6.4–9.0) | 0.005 |
| | Girls | 3.2 (2.4-4.0) | 0.635 | 2.4 (-0.4-5.2) | 1.000 | 1.5 (1.0-2.1) | 0.485 | 7.2 (6.1-8.2) | 0.730 |
| IL1-Ra | Boys | 2.6 (2.1-3.1) | 0.810 | 3.0 (2.6-3.4) | 0.005 | 1.1 (-0.03-2.2) | 0.393 | 7.9 (6.4–9.4) | 0.023 |
| | Girls | 3.3 (2.5-4.1) | 0.558 | 2.3 (-0.8-5.5) | 1.000 | 1.6 (0.9-2.2) | 1.000 | 6.6 (5.3–7.8) | 0.735 |
| IL-6 | Boys | 2.6 (2.0-3.2) | 0.743 | 3.0 (2.1-4.0) | 0.461 | 0.8 (-0.5-2.0) | 0.963 | 7.1 (4.2–10.0) | 0.932 |
| | Girls | 3.3 (2.5-4.1) | 0.949 | 2.2 (-0.9-5.3) | 0.732 | 1.6 (0.9-2.2) | 0.692 | 6.8 (5.7-8.0) | 0.625 |
| IL-8 | Boys | 2.4 (1.8-3.0) | 0.927 | 2.9 (2.5-3.3) | 0.005 | 1.2 (0.2–2.2) | 0.410 | 7.2 (5.6-8.9) | 0.021 |
| | Girls | 3.3 (2.4-4.1) | 0.649 | 2.5 (-0.5-5.4) | 1.000 | 1.6 (1.1-2.1) | 1.000 | 7.3 (6.2–8.4) | 0.856 |
| IL-15 | Boys | 2.6 (2.1-3.1) | 0.588 | 2.9 (2.5-3.3) | 0.005 | 1.2 (0.1–2.3) | 0.890 | 7.5 (5.9–9.1) | 0.015 |
| | Girls | 3.3 (2.6–4.0) | 0.512 | 2.3 (0.9–2.3) | 0.912 | 1.6 (0.9–2.2) | 0.856 | 6.8 (5.7–8.0) | 0.908 |

Data are expressed as mean (CIs). Covariates: Country of residence, age, BMI z-score, pubertal status. *q*-values are BH-adjusted *p*-values. Values in bold indicate statistically significant results.

4. Discussion

Inflammation is a physio-pathological process, commonly triggered by injuries and infections, and characterized by a complex flow of dynamically and finely regulated responses. The degree, the dynamics of pro-and anti-inflammatory networks, and the course of an inflammatory reaction may decisively impact the onset, progression, and development of health disorders. Inflammation and its supporting mechanisms are considered closely related to numerous diseases' progression. Various studies have established that specific miRNAs participate in the development of innate and adaptive immunity, acting as crucial players in the fine-tuning of the inflammatory network. In this context, several miRNAs attenuate the response, while others, by depressing specific inhibitors, are capable of intensifying the immune reaction [36], with certain miRNAs essential for mounting the inflammatory response [37]. Notably, chronic inflammation is one of the main factors involved in the process of obesity progression [9,38], and several studies have reported that even limited weight gain is interconnected with the sustained inflammatory process [38].

Previously, we identified specific circulating miRNAs associated with childhood obesity in a subsample of the I.Family study [28,30]. Moreover, we also recognized in that study candidate miRNAs possibly related to CRP levels, among which only hsa-miR-10b-5p was associated with OW/Ob. In the present analysis, we report sex-related associations of these miRNAs with inflammatory biomarkers in children and adolescents with OW/Ob. Of note, the subjects enrolled for the current investigation belong to a different subgroup of the common I.Family population.

Given the different fat distribution and the influence of sex hormones, it is conceivable that the relationship between inflammatory markers and obesity may differ by sex. Accordingly, we stratified the study population by sex. Association analyses were corrected for confounding factors including the country of residence, age, BMI z-score, and pubertal status. We found a clear indication pointing towards a cross-talk between candidate cmiRNAs and inflammatory biomarkers in the context of raised adiposity. Among the numerous inflammatory biomarkers, we focused on the acute-phase reactant CRP as the primary outcome of the study. CRP has been considered the strongest factor associated with OW/Ob in epidemiological studies [15]. Several reports consider CRP to be a consequence of an obesity condition rather than the cause [15]; conversely, increasing evidence establishes a causal role of CRP elevation in the onset and development of obesity by causing extensive tuning in the innate immune system and energy expenditure system [39].

We also found sex-related associations of the candidate miRNAs with the selected cytokines as plausible inflammation biomarkers. Except for IL-6, a significant association of both hsa-miR-26b-3p and hsa-miR-576-5p with TNF- α , IL1-Ra, IL-8, and IL-15 levels was found exclusively in boys. Moreover, no association of candidate miRNAs with cytokine levels was established in the girls' subgroup.

Among the characterized miRNAs, miR-10b-5p is one of the first identified as abnormal in human cancer and, since its first description, it has been widely studied in this context. Recent papers also suggest that it participates in inflammation control and inflammation-associated diseases by regulating T cells [40,41]. Similarly, miR-26b has been described as a key regulator in carcinogenesis and cancer progression, acting as a tumor suppressor gene in several types of cancer. miR-26b has also been shown to play a role in inflammation as in cytokine secretion [42]. miR-26b also targets the inflammatory factor prostaglandin-endoperoxide synthase 2, which plays relevant roles in inflammatory diseases by inducing the production of prostaglandin E2 [43]. However, abnormal expression of miR-31-5p has been described in various cancers, where this miRNA plays a significant role in tumorigenesis, acting as either an oncogene or tumor suppressor, in a context-dependent manner, although the underlying mechanism remains unclear [44]. Moreover, miR-31 is involved in several inflammation-associated disorders. Interestingly, the molecular role of miR-31-5p activation in early inflammation has been recently defined [45]. Several studies have demonstrated that miR-576-5p acts as a tumor-promoting miRNA in several types of human tumors, highlighting its potential role as a predictor of cancer prognosis [46]. However, in silico studies have shown that miR-576-5p is involved in the regulation of inflammatory, growth, and proliferation signaling pathways.

Our apparently controversial results are in line with recent studies reporting sex influences on the severity and evolution of various inflammatory conditions [47,48]. Numerous studies have confirmed the role of sex hormones in the immune response and recent clinical data have shown significant sex differences in inflammatory markers also in prepubertal children, supporting a genetic contribution [47]. Sex differences occur in both innate and adaptive immune responses and are evolutionarily conserved across species [49]. Overall, there is accumulating evidence that sex is a critical variable that influences innate and adaptive immune responses, resulting in sex-specific outcomes, but the main molecular mechanisms remain elusive [48].

The findings of this exploratory study suggest major differences in the association of circulating miRNAs and inflammatory response biomarkers across sexes, pointing to a conceivable role of miRNAs among the candidate epigenetic mechanisms related to the lowgrade inflammation process in childhood obesity, calling for more attention in this largely underexplored area. However, evidence concerning how these molecules may act remains questioned since the experimental design of our cross-sectional analysis, explorative in essence, cannot answer this question.

5. Conclusions

Many regulatory steps are relevant in the transformation and delivery of genetic information to cellular effectors. This network orchestration is finely regulated by both transcriptional and post-transcriptional mechanisms. Sex differences can provide substantial support in defining the course of these supervisory mechanisms [50]. The fascinating emergence of circulating miRNAs as stable and affordable molecules has opened up a promising

opportunity for the identification of new crucial players in the onset and progression of inflammation in childhood obesity, as well as their potential application as non-invasive biomarkers. However, it is still uncertain whether the identified miRNAs are drivers of sex-related disparities in obesity-related inflammation or represent epiphenomena. Further molecular-oriented studies are needed to explore the functional relevance of the miRNA species identified.

Supplementary Materials: The following supporting information can be downloaded at: https://www.mdpi.com/article/10.3390/genes13040632/s1, Figure S1. The different panels show the distribution of reported miRNAs in relation to CRP levels in girls and boys.

Author Contributions: G.I., F.L. and A.S. conceived, designed, and oversaw the analyses and drafted the manuscript. A.V. and P.M. conducted the molecular analyses. P.R. contributed to the interpretation of data and provided critical input during the drafting and revision of the manuscript. W.A., S.D.H., G.E., R.F., A.H., Y.K., D.M., L.A.M. and T.V. contributed to the critical revision of the manuscript. All authors have read and agreed to the published version of the manuscript.

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Institutional Review Board Statement: This study was conducted according to the standards of the Declaration of Helsinki. Approval by the appropriate ethics committees was obtained by each of the eight participating centers carrying out the fieldwork (Ethics Committee of the Ghent University Hospital—Belgium, National Bioethics Committee—Cyprus, Tallinn Medical Research Ethics Committee —Estonia, Ethics Committee of the University of Bremen—Germany, Scientific and Research Ethics Committee of the Medical Research Council of Pécs (TUKEB) and Baranja County Public Health and Medical Officer Service (ANTSZ)—Hungary, Ethics Committee of the Local Health Institute in Avellino (ASL)—Italy, Ethics Committee of Clinical Research of Aragon (CEICA)—Spain, Regional Ethics Committee of the University of Gothenburg—Sweden).

Informed Consent Statement: Participants were not subjected to any study procedure before both the children and their parents gave their oral (children) and written (parents) informed consent for examinations, collection of samples, subsequent analysis, and storage of personal data and collected samples.

Data Availability Statement: All data produced or analyzed during this study are included in this article.

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Conflicts of Interest: The authors declare no conflict of interest.

References

- 1. WHO. Library Cataloguing in Publication Data Report of the Commission on Ending Childhood Obesity; World Health Organization: Geneva, Switzerland, 2016.
- Gómez, L.A.; Abden, Z.A.; Hamid, Z.A.; Rmeileh, N.M.A.; Cazares, B.A.; Acuin, C.; Adams, R.J.; Aekplakorn, W.; Afsana, K.; Salinas, C.A.A.; et al. Worldwide trends in body-mass index, underweight, overweight, and obesity from 1975 to 2016: A pooled analysis of 2416 population-based measurement studies in 128.9 million children, adolescents, and adults. *Lancet* 2017, 390, 2627–2642. [CrossRef]
- 3. Hruby, A.; Hu, F.B. The epidemiology of obesity: A big picture. *Pharmacoeconomics* **2015**, *33*, 673–689. [CrossRef] [PubMed]
- Iacomino, G.; Lauria, F.; Venezia, A.; Iannaccone, N.; Russo, P.; Siani, A. microRNAs in Obesity and metabolic diseases. In *Obesity* and *Diabetes: Scientific Advances and Best Practice*; Faintuch, J., Faintuch, S., Eds.; Springer Nature: Cham, Switzerland, 2020; Chapter 6, pp. 71–95. [CrossRef]
- 5. Farag, Y.M.; Gaballa, M.R. Diabesity: An overview of a rising epidemic. *Nephrol. Dial. Transplant. Off. Publ. Eur. Dial. Transpl. Assoc.-Eur. Ren. Assoc.* **2011**, *26*, 28–35. [CrossRef] [PubMed]

- Huls, A.; Wright, M.N.; Bogl, L.H.; Kaprio, J.; Lissner, L.; Molnar, D.; Moreno, L.A.; De Henauw, S.; Siani, A.; Veidebaum, T.; et al. Polygenic risk for obesity and its interaction with lifestyle and sociodemographic factors in European children and adolescents. *Int. J. Obes.* 2021, 45, 1321–1330. [CrossRef]
- Li, Y.; Pollock, C.A.; Saad, S. Aberrant DNA methylation mediates the transgenerational risk of metabolic and chronic disease due to maternal obesity and overnutrition. *Genes* 2021, 12, 1653. [CrossRef] [PubMed]
- Kammoun, H.L.; Kraakman, M.J.; Febbraio, M.A. Adipose tissue inflammation in glucose metabolism. *Rev. Endocr. Metab. Disord.* 2014, 15, 31–44. [CrossRef]
- Christ, A.; Günther, P.; Lauterbach, M.A.R.; Duewell, P.; Biswas, D.; Pelka, K.; Scholz, C.J.; Oosting, M.; Haendler, K.; Baßler, K.; et al. Western diet triggers NLRP3-dependent innate immune reprogramming. *Cell* 2018, 172, 162–175.e14. [CrossRef]
- 10. Thorburn, A.N.; Macia, L.; Mackay, C.R. Diet, Metabolites, and "Western-Lifestyle" Inflammatory Diseases. *Immunity* **2014**, *40*, 833–842. [CrossRef] [PubMed]
- 11. Yao, L.; Heuser-Baker, J.; Barlic-Dicen, J. Chemokine receptors on the defensive—The surprising role of CXCR4 in brown adipose tissue. *Recept. Clin. Investig.* 2015, 2, e397. [CrossRef]
- 12. Kraakman, M.J.; Murphy, A.J.; Jandeleit-Dahm, K.; Kammoun, H.L. Macrophage polarization in obesity and type 2 diabetes: Weighing down our understanding of macrophage function? *Front. Immunol.* **2014**, *5*, 470. [CrossRef] [PubMed]
- 13. Schaffler, A.; Muller-Ladner, U.; Scholmerich, J.; Buchler, C. Role of adipose tissue as an inflammatory organ in human diseases. *Endocr. Rev.* **2006**, *27*, 449–467. [CrossRef] [PubMed]
- 14. Saltiel, A.R.; Olefsky, J.M. Inflammatory mechanisms linking obesity and metabolic disease. J. Clin. Investig. 2017, 121, 2111–2117. [CrossRef]
- Timpson, N.J.; Nordestgaard, B.G.; Harbord, R.M.; Zacho, J.; Frayling, T.M.; Tybjærg-Hansen, A.; Davey Smith, G. C-reactive protein levels and body mass index: Elucidating direction of causation through reciprocal Mendelian randomization. *Int. J. Obes.* 2011, 35, 300–308. [CrossRef] [PubMed]
- Nappo, A.; Iacoviello, L.; Fraterman, A.; Gonzalez-Gil, E.M.; Hadjigeorgiou, C.; Marild, S.; Molnar, D.; Moreno, L.A.; Peplies, J.; Sioen, I.; et al. High-sensitivity C-reactive protein is a predictive factor of adiposity in children: Results of the identification and prevention of dietary- and lifestyle-induced health effects in children and infants (IDEFICS) study. *J. Am. Heart Assoc.* 2013, 2, e000101. [CrossRef] [PubMed]
- 17. Cavalieri, V. The expanding constellation of histone post-translational modifications in the epigenetic landscape. *Genes* **2021**, 12, 1596. [CrossRef]
- Huang, H.Y.; Lin, Y.C.; Li, J.; Huang, K.Y.; Shrestha, S.; Hong, H.C.; Tang, Y.; Chen, Y.G.; Jin, C.N.; Yu, Y.; et al. miRTarBase 2020: Updates to the experimentally validated microRNA-target interaction database. *Nucleic Acids Res.* 2020, 48, D148–D154. [CrossRef]
- 19. Kim, Y.K. Extracellular microRNAs as biomarkers in human disease. *Chonnam Med. J.* 2015, 51, 51–57. [CrossRef]
- 20. Condrat, C.E.; Thompson, D.C.; Barbu, M.G.; Bugnar, O.L.; Boboc, A.; Cretoiu, D.; Suciu, N.; Cretoiu, S.M.; Voinea, S.C. miRNAs as biomarkers in disease: Latest findings regarding their role in diagnosis and prognosis. *Cells* **2020**, *9*, 276. [CrossRef]
- 21. Lauria, F.; Iacomino, G. The landscape of circulating miRNAs in the post-genomic era. Genes 2022, 13, 94. [CrossRef]
- 22. Dumortier, O.; Hinault, C.; Van Obberghen, E. MicroRNAs and metabolism crosstalk in energy homeostasis. *Cell Metab.* **2013**, *18*, 312–324. [CrossRef]
- 23. Iacomino, G.; Siani, A. Role of microRNAs in obesity and obesity-related diseases. *Genes Nutr.* 2017, 12, 23. [CrossRef]
- 24. Kunej, T.; Jevsinek Skok, D.; Zorc, M.; Ogrinc, A.; Michal, J.J.; Kovac, M.; Jiang, Z. Obesity gene atlas in mammals. *J. Genom.* 2013, 1, 45–55. [CrossRef] [PubMed]
- Hartig, S.M.; Hamilton, M.P.; Bader, D.A.; McGuire, S.E. The miRNA interactome in metabolic homeostasis. *Endocrinol. Metab.* 2015, 26, 733–745. [CrossRef] [PubMed]
- Raghuraman, S.; Donkin, I.; Versteyhe, S.; Barres, R.; Simar, D. The emerging role of epigenetics in inflammation and immunometabolism. *Trends Endocrinol. Metab.* 2016, 27, 782–795. [CrossRef] [PubMed]
- 27. Kroesen, B.J.; Teteloshvili, N.; Smigielska-Czepiel, K.; Brouwer, E.; Boots, A.M.; van den Berg, A.; Kluiver, J. Immuno-miRs: Critical regulators of T-cell development, function and ageing. *Immunology* **2015**, *144*, 1–10. [CrossRef]
- Iacomino, G.; Russo, P.; Stillitano, I.; Lauria, F.; Marena, P.; Ahrens, W.; De Luca, P.; Siani, A. Circulating microRNAs are deregulated in overweight/obese children: Preliminary results of the I.Family study. *Genes Nutr.* 2016, 11, 7. [CrossRef] [PubMed]
- Ahrens, W.; Siani, A.; Adan, R.; De Henauw, S.; Eiben, G.; Gwozdz, W.; Hebestreit, A.; Hunsberger, M.; Kaprio, J.; Krogh, V.; et al. Cohort profile: The transition from childhood to adolescence in European children-how I.Family extends the IDEFICS cohort. *Int. J. Epidemiol.* 2017, 46, 1394j–1395j. [CrossRef] [PubMed]
- Iacomino, G.; Russo, P.; Marena, P.; Lauria, F.; Venezia, A.; Ahrens, W.; De Henauw, S.; De Luca, P.; Foraita, R.; Gunther, K.; et al. Circulating microRNAs are associated with early childhood obesity: Results of the I.Family study. *Genes Nutr.* 2019, 14, 2. [CrossRef] [PubMed]
- Bammann, K.; Lissner, L.; Pigeot, I.; Ahrens, W. (Eds.) Instruments for Health Surveys in Children and Adolescents; Springer International Publishing: Berlin/Heidelberg, Germany, 2019.
- Iacomino, G.; Lauria, F.; Russo, P.; Marena, P.; Venezia, A.; Iannaccone, N.; De Henauw, S.; Foraita, R.; Heidinger-Felso, R.; Hunsberger, M.; et al. Circulating miRNAs are associated with sleep duration in children/adolescents: Results of the I.Family study. *Exp. Physiol.* 2020, 105, 347–356. [CrossRef]

- Cole, T.J.; Lobstein, T. Extended international (IOTF) body mass index cut-offs for thinness, overweight and obesity. *Pediatr. Obes.* 2012, 7, 284–294. [CrossRef]
- Peplies, J.; Fraterman, A.; Scott, R.; Russo, P.; Bammann, K. Quality management for the collection of biological samples in multicentre studies. *Eur. J. Epidemiol.* 2010, 25, 607–617. [CrossRef]
- Shanahan, L.; Copeland, W.E.; Worthman, C.M.; Erkanli, A.; Angold, A.; Costello, E.J. Sex-differentiated changes in C-reactive protein from ages 9 to 21: The contributions of BMI and physical/sexual maturation. *Psychoneuroendocrinology* 2013, *38*, 2209–2217. [CrossRef] [PubMed]
- 36. Lindsay, M.A. microRNAs and the immune response. Trends. Immunol. 2008, 29, 343–351. [CrossRef]
- 37. Nejad, C.; Stunden, H.J.; Gantier, M.P. A guide to miRNAs in inflammation and innate immune responses. *FEBS J.* **2018**, *285*, 3695–3716. [CrossRef]
- Piening, B.D.; Zhou, W.; Contrepois, K.; Rost, H.; Gu Urban, G.J.; Mishra, T.; Hanson, B.M.; Bautista, E.J.; Leopold, S.; Yeh, C.Y.; et al. Integrative personal omics profiles during periods of weight gain and loss. *Cell Syst.* 2018, 6, 157–170.e8. [CrossRef]
- 39. Li, Q.; Wang, Q.; Xu, W.; Ma, Y.; Wang, Q.; Eatman, D.; You, S.; Zou, J.; Champion, J.; Zhao, L.; et al. C-reactive protein causes adult-onset obesity through chronic inflammatory mechanism. *Front. Cell Dev. Biol.* **2020**, *8*, 18. [CrossRef]
- Chen, L.; Al-Mossawi, M.H.; Ridley, A.; Sekine, T.; Hammitzsch, A.; de Wit, J.; Simone, D.; Shi, H.; Penkava, F.; Kurowska-Stolarska, M.; et al. miR-10b-5p is a novel Th17 regulator present in Th17 cells from ankylosing spondylitis. *Ann. Rheum. Dis.* 2017, 76, 620–625. [CrossRef]
- Tu, J.; Han, D.; Fang, Y.; Jiang, H.; Tan, X.; Xu, Z.; Wang, X.; Hong, W.; Li, T.; Wei, W. MicroRNA-10b promotes arthritis development by disrupting CD4⁺ T cell subtypes. *Mol. Ther.-Nucleic Acids* 2022, 27, 733–750. [CrossRef]
- 42. Sun, J.; Yan, P.; Chen, Y.; Chen, Y.; Yang, J.; Xu, G.; Mao, H.; Qiu, Y. MicroRNA-26b inhibits cell proliferation and cytokine secretion in human RASF cells via the Wnt/GSK-3β/β-catenin pathway. *Diagn. Pathol.* **2015**, *10*, 72. [CrossRef]
- Ji, Y.; He, Y.; Liu, L.; Zhong, X. MiRNA-26b regulates the expression of cyclooxygenase-2 in desferrioxamine-treated CNE cells. FEBS Lett. 2010, 584, 961–967. [CrossRef]
- 44. Yu, T.; Ma, P.; Wu, D.; Shu, Y.; Gao, W. Functions and mechanisms of microRNA-31 in human cancers. *Biomed. Pharmacother.* **2018**, 108, 1162–1169. [CrossRef] [PubMed]
- 45. Tian, Y.; Xu, J.; Li, Y.; Zhao, R.; Du, S.; Lv, C.; Wu, W.; Liu, R.; Sheng, X.; Song, Y.; et al. MicroRNA-31 reduces inflammatory signaling and promotes regeneration in colon epithelium, and delivery of mimics in microspheres reduces colitis in mice. *Gastroenterology* 2019, 156, 2281–2296.e6. [CrossRef] [PubMed]
- 46. Zhang, L.; Chen, J.; Wang, L.; Chen, L.; Du, Z.; Zhu, L.; Cui, M.; Zhang, M.; Song, L. Linc-PINT acted as a tumor suppressor by sponging miR-543 and miR-576-5p in esophageal cancer. *J. Cell Biochem.* **2019**, *120*, 19345–19357. [CrossRef] [PubMed]
- 47. Lefèvre, N.; Corazza, F.; Duchateau, J.; Desir, J.; Casimir, G. Sex differences in inflammatory cytokines and CD99 expression following in vitro lipopolysaccharide stimulation. *Shock* **2012**, *38*, 37–42. [CrossRef] [PubMed]
- Wegner, A.; Benson, S.; Rebernik, L.; Spreitzer, I.; Jäger, M.; Schedlowski, M.; Elsenbruch, S.; Engler, H. Sex differences in the pro-inflammatory cytokine response to endotoxin unfold in vivo but not ex vivo in healthy humans. *Innate Immun.* 2017, 23, 432–439. [CrossRef]
- 49. Klein, S.L.; Flanagan, K.L. Sex differences in immune responses. Nat. Rev. Immunol. 2016, 16, 626–638. [CrossRef]
- 50. Sharma, S.; Eghbali, M. Influence of sex differences on microRNA gene regulation in disease. *Biol. Sex Differ.* 2014, *5*, 3. [CrossRef]