

Supplementary Material

The MraY Inhibitor Muraymycin D2 and Its Derivatives Induce Enlarged Cells in Obligate Intracellular *Chlamydia* and *Wolbachia* and Break the Persistence Phenotype in *Chlamydia*

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Table S1. Cytotoxic concentration 50% (CC₅₀) of muraymycin (MRY) D2, its derivatives (MRHs), and carbacaprazamycin (cCPZ) on eukaryotic HEp-2 cells. Muraymycin D2, its derivatives, and cCPZ were incubated with HEp-2 cells for 28 h. The cytotoxicity was measured via alamarBlue normalized against dimethyl sulfoxide and the CC₅₀ was calculated by non-linear regression from the mean of two independent experiments. The CC₅₀ of muraymycin D2 and the derivatives MRH-22, -23, -38, and -82 was greater than the highest concentration tested (128 µg/mL). MRH-25, -76, -92, and cCPZ were more toxic with CC₅₀ values between 81 µg/mL and 124 µg/mL.

Compound	CC ₅₀ (µg/mL)
MRY D2	> 128
MRH-22	> 128
MRH-23	> 128
MRH-25	116
MRH-38	> 128
MRH-76	81
MRH-82	> 128
MRH-92	109
cCPZ	124

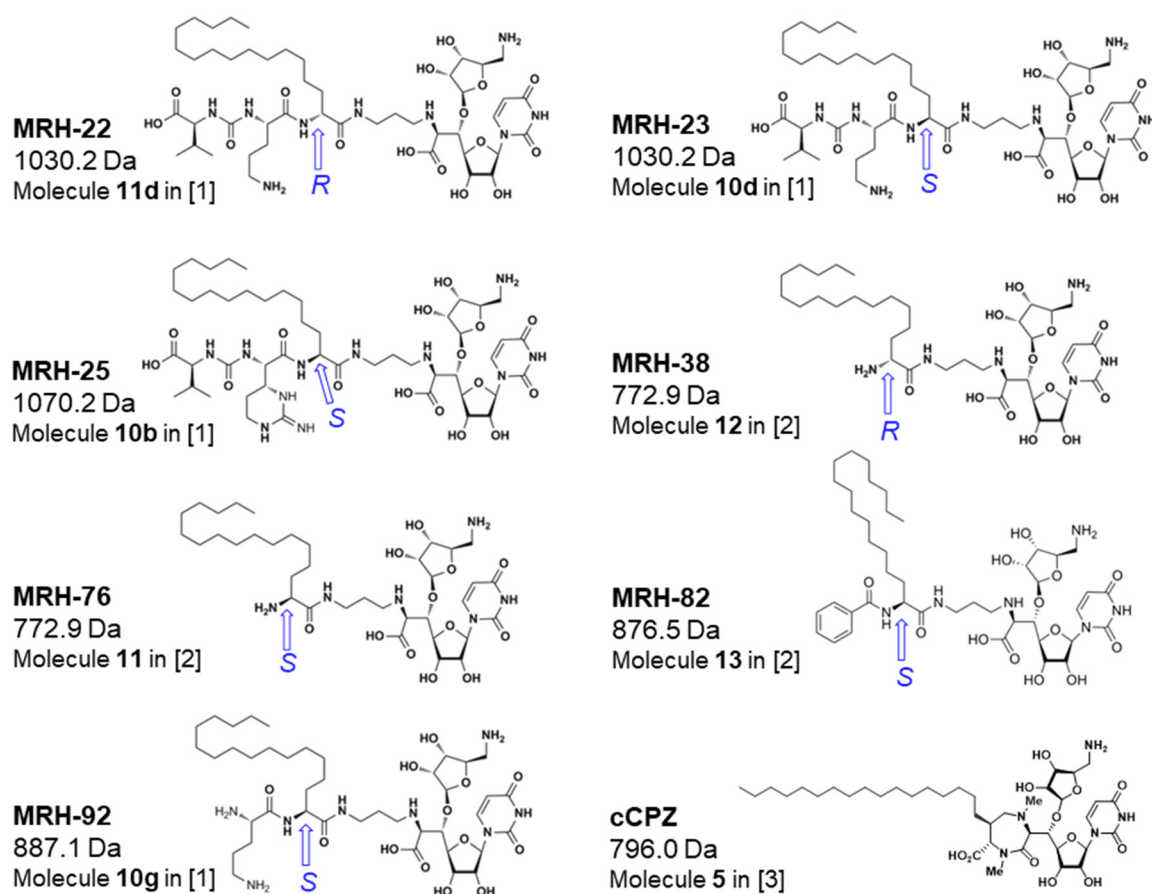


Figure S1. Chemical structures of muraymycin derivatives and cCPZ. All compounds were previously described [1–3]. The core structure of muraymycins resembles uridine diphosphate *N*-acetyl-muramoyl-pentapeptide, the last cytoplasmic precursor molecule of lipid I. Muraymycin derivatives and cCPZ are more lipophilic due to an additional palmitoyl side chain. Important chemical configurations of muraymycin derivatives are indicated by a blue arrow.

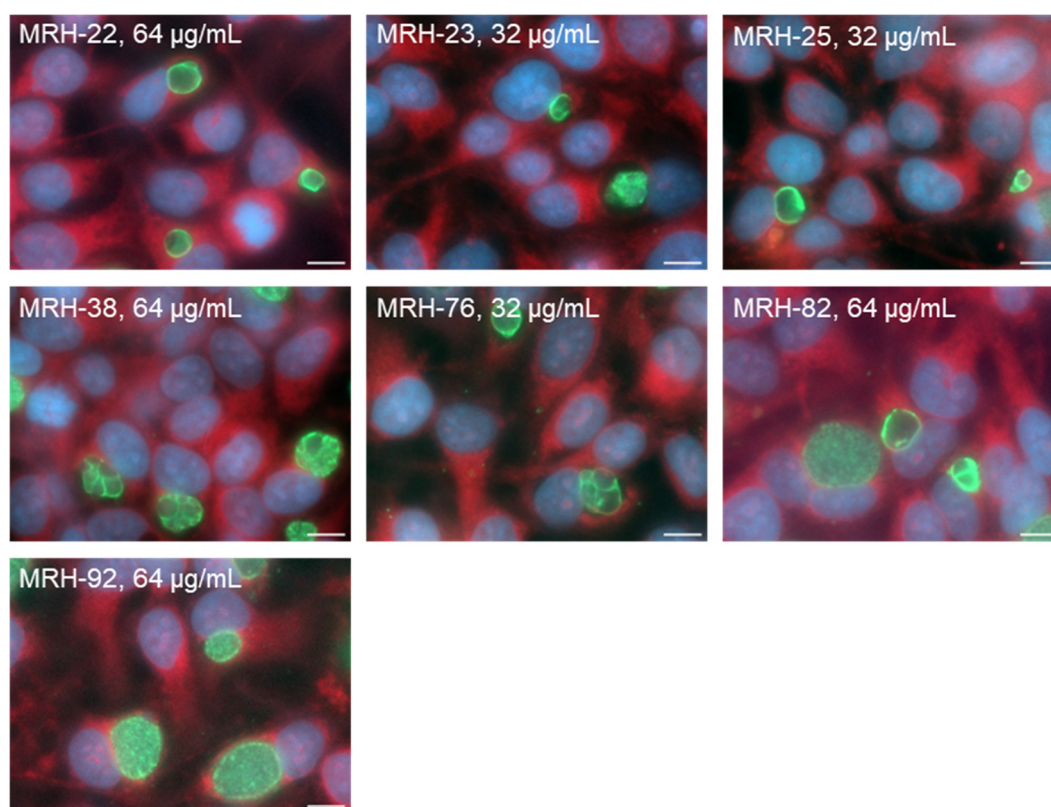


Figure S2. Most muraymycin derivatives induce the formation of enlarged chlamydial cells. HEp-2 cells infected with *C. trachomatis* D/UW-3/CX were treated with muraymycin derivatives at 2 h post infection (hpi). At 30 hpi, the anti-chlamydial activity was analyzed for concentrations which did not exert cytotoxicity towards the host cells in resazurin-based viability assays (Table S1) and in fluorescence microscopy analyses. Eukaryotic host cell cytoplasm from representative images was labelled with Evans Blue (red), DNA with DAPI (blue), and chlamydial lipopolysaccharide with fluorescein (green). Scale bar: 10 µm. All derivatives tested are shown at their minimal inhibitory concentration, which is reflected by the presence of aberrant bodies. MRH-92 did not induce a persistence-like phenotype and is shown at the highest concentration tested (64 µg/mL).

Table S2. Sequence identities and confidence of *C. trachomatis*, *C. pneumoniae*, and *Wolbachia* endosymbionts of *A. albopictus* or *B. malayi* MraY protein models against *A. aeolicus* apoMraY or MraY_{Aae} bound to muraymycin D2. Bacterial strains and primary protein accession numbers: *A. aeolicus* VF5 (Aae, O66465), *C. trachomatis* D/UW-3/CX (Ctr, O84762), *C. pneumoniae* GiD (Cpn, A0A0F7WR30), and *Wolbachia* endosymbiont of *A. albopictus* (wAlbB, A0A4S2QUK2) and *B. malayi* (wBm, Q5GRZ3). Predictions of MraY_{Cpn/Ctr/wAlbB/wBm} protein models were made with one-to-one threading against crystal structures of apoMraY_{Aae} chain A [4] or MraY_{Aae} bound to MRY D2 [5] using phyre2 software (Imperial College London, London, UK; [6]).

Query	MraY _{Aae} template	Sequence identity (%)	Aligned residues	Confidence (%)
MraY _{Ctr}	apoMraY _{Aae} chain A	33	306/336 (= 91 %)	100
	MraY _{Aae} bound to MRY D2	33	311/336 (= 93 %)	100
MraY _{Cpn}	apoMraY _{Aae} chain A	37	308/349 (= 88 %)	100
	MraY _{Aae} bound to MRY D2	37	316/349 (= 91 %)	100
MraY _{wAlbB}	apoMraY _{Aae} chain A	41	301/326 (= 92 %)	100
	MraY _{Aae} bound to MRY D2	42	305/326 (= 94 %)	100
MraY _{wBm}	apoMraY _{Aae} chain A	40	302/326 (= 93 %)	100
	MraY _{Aae} bound to MRY D2	40	303/326 (= 93 %)	100

References

1. Tanino, T.; Al-Dabbagh, B.; Mengin-Lecreulx, D.; Bouhss, A.; Oyama, H.; Ichikawa, S.; Matsuda, A. Mechanistic Analysis of Muraymycin Analogues: A Guide to the Design of MraY Inhibitors. *J Med Chem* **2011**, *54*, 8421–8439, doi:10.1021/jm200906r.
2. Takeoka, Y.; Tanino, T.; Sekiguchi, M.; Yonezawa, S.; Sakagami, M.; Takahashi, F.; Togame, H.; Tanaka, Y.; Takemoto, H.; Ichikawa, S.; et al. Expansion of Antibacterial Spectrum of Muraymycins toward *Pseudomonas Aeruginosa*. *ACS Med Chem Lett* **2014**, *5*, 556–560, doi:10.1021/ml5000096.
3. Ichikawa, S.; Yamaguchi, M.; Hsuan, L.S.; Kato, Y.; Matsuda, A. Carbaprazamycins: Chemically Stable Analogues of the Caprazamycin Nucleoside Antibiotics. *ACS Infect Dis* **2015**, *1*, 151–156, doi:10.1021/id5000376.
4. Chung, B.C.; Zhao, J.; Gillespie, R.A.; Kwon, D.-Y.; Guan, Z.; Hong, J.; Zhou, P.; Lee, S.-Y. Crystal Structure of MraY, an Essential Membrane Enzyme for Bacterial Cell Wall Synthesis. *Science* **2013**, *341*, 1012–1016, doi:10.1126/science.1236501.
5. Chung, B.C.; Mashalidis, E.H.; Tanino, T.; Kim, M.; Matsuda, A.; Hong, J.; Ichikawa, S.; Lee, S.-Y. Structural Insights into Inhibition of Lipid I Production in Bacterial Cell Wall Synthesis. *Nature* **2016**, *533*, 557–560, doi:10.1038/nature17636.
6. Kelley, L.A.; Mezulis, S.; Yates, C.M.; Wass, M.N.; Sternberg, M.J.E. The Phyre2 Web Portal for Protein Modeling, Prediction and Analysis. *Nat Protoc* **2015**, *10*, 845–858, doi:10.1038/nprot.2015.053.