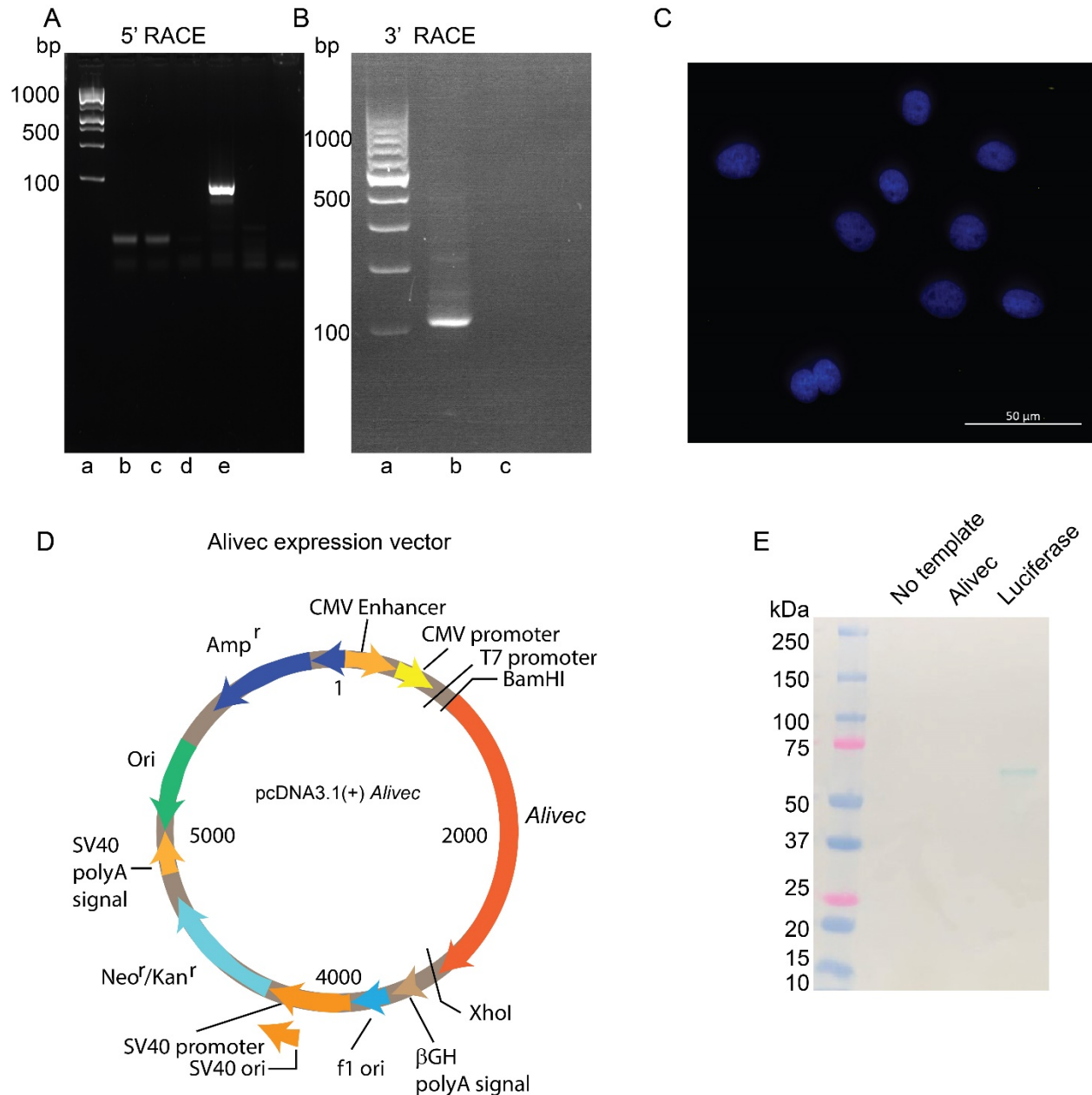


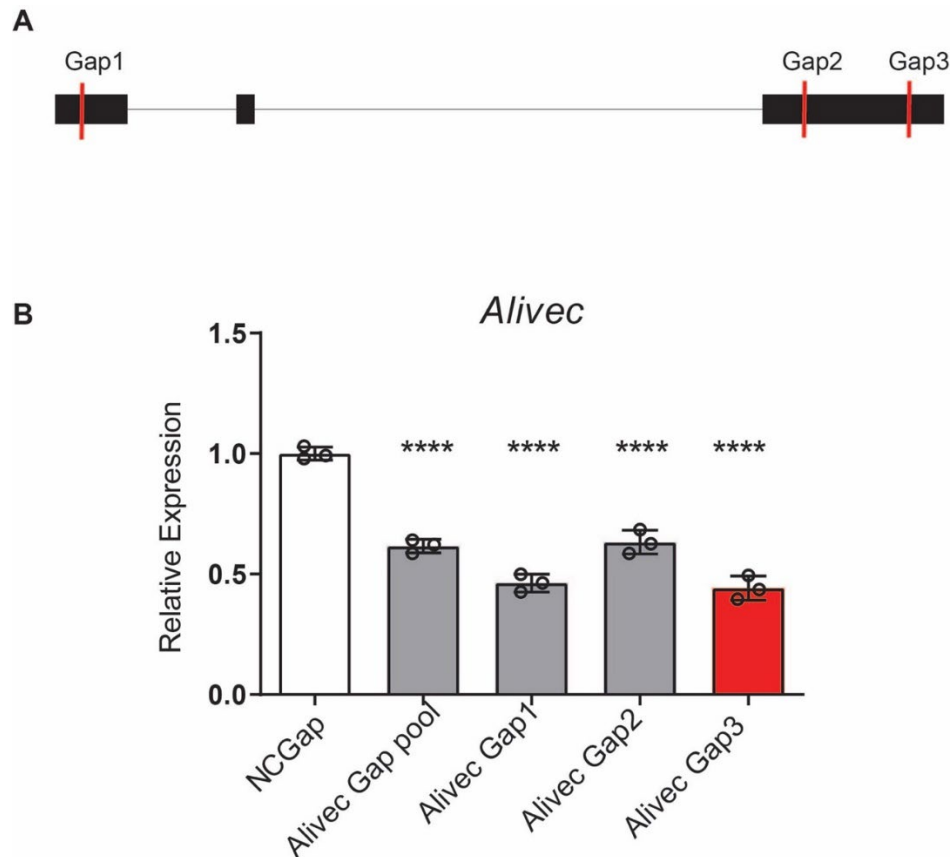
**Supplementary Materials (Figures and Tables) for Vishnu Amaram Samara, et al.**



**Supplementary Figure S1. *Alivec* characterization and full-length cloning**

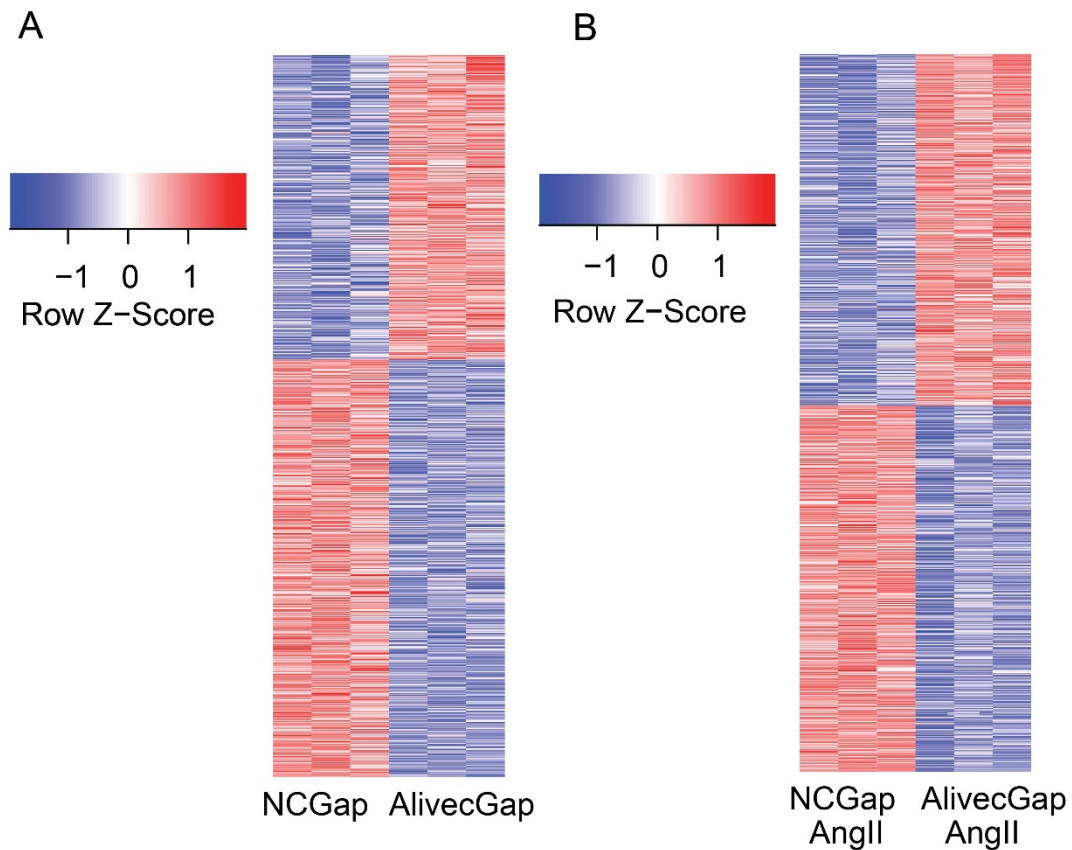
A) Representative agarose gel image of 5' RACE of *Alivec* (lane a - 100bp DNA ladder, b - no template PCR, c - 5' outer *Alivec* RACE-PCR, d - empty lane, e - 5' inner *Alivec* RACE-PCR that was chosen for Sanger sequencing. (the faint bands at lower mol wt. of ~20bp in lanes b and c are artifacts that could be from primer dimers or reaction mixture) B) Representative agarose gel image of 3' end RACE products of *Alivec* (a - 100 bp DNA ladder, b - 3' *Alivec* RACE-PCR chosen for

Sanger sequencing, c – no template PCR). C) Control image of RNA-FISH with no probes showing absence of spots in VSMC; nuclei are stained with DAPI (blue), Scale bars represent 50µm. D) Plasmid map of *Alivec* sequence cloned into pcDNA3.1+ overexpression vector to generate pcDNA*Alivec* plasmid. E) *In vitro* transcription/translation assay with pcDNA*Alivec* plasmid performed using T7 TNT Quick coupled transcription/translation system (Promega). T7 luciferase plasmid was used as positive control and no template pcDNA plasmid as negative control.



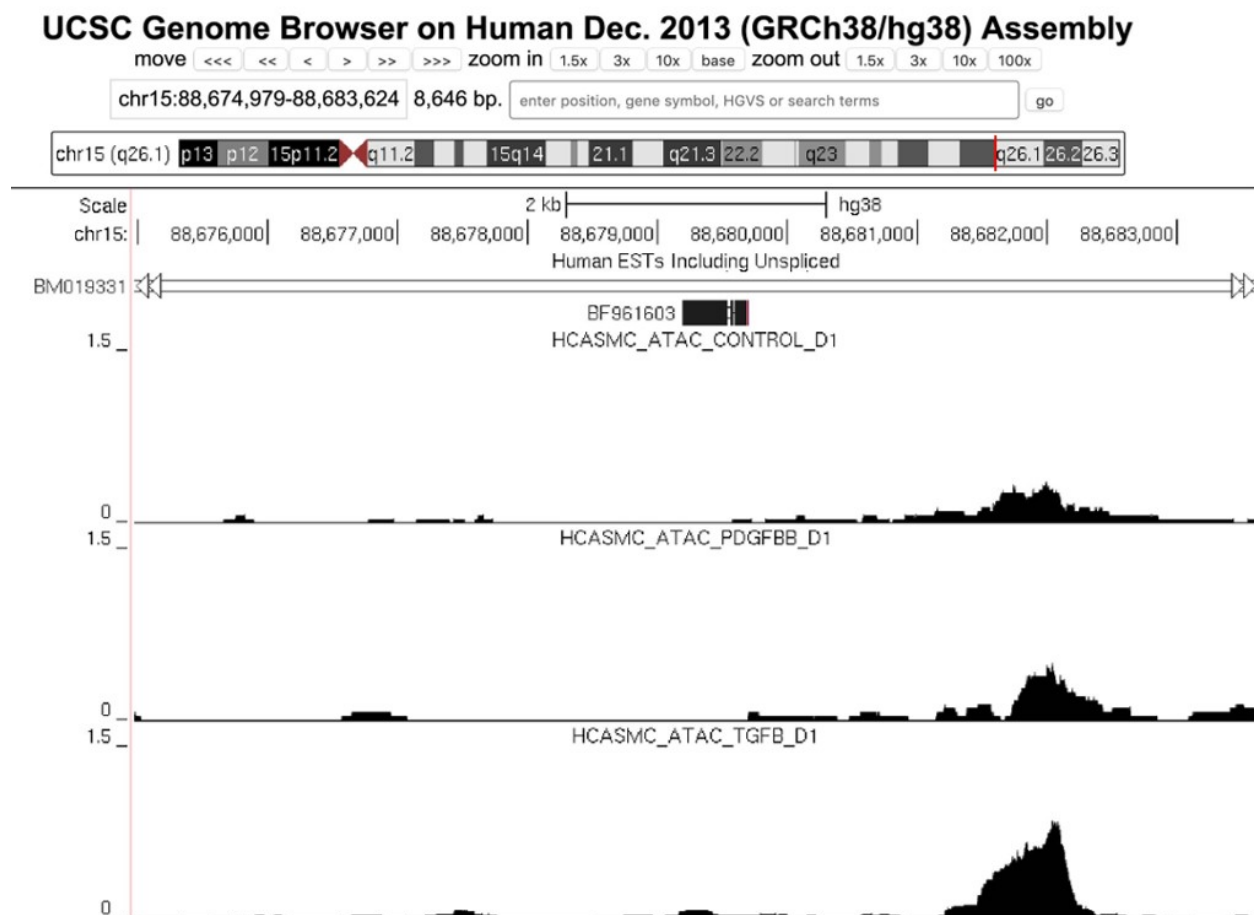
**Supplementary Figure S2. Design and efficacy of LNA-GapmeRs targeting *Alivec***

A) Three GapmeRs (Gap1, Gap2, Gap3) were designed using the Qiagen-Exiqon GapmeR designer tool to target three indicated regions of the *Alivec* transcript sequence. B) RT-qPCR analysis of *Alivec* in RVSMCs 48 hours post transfection with GapmeRs targeting *Alivec* or non-targeting GapmeR (NCGap) at 100nM. Individual GapmeRs and a pool of these GapmeRs were tested for knockdown efficiency of *Alivec* in the presence of Ang II (100nM, 3 hour). *Alivec*Gap3 was most effective and was used for subsequent knockdown experiments (*Alivec*Gap). Data presented as mean  $\pm$  SD, n=3 biological replicates, one-way ANOVA with Dunnett's multiple comparisons test, \*\*\*\*p<0.0001 versus NCGap.



**Supplementary Figure S3. Microarray profiling in RVSMCs treated with NCGap and AliveGap**

A, B) Heat map of differentially expressed genes from RVSMCs transfected with AliveGap or NCGap and treated without (A) or with (B) Ang II (100 nM). n=3 per group. Cut off value: fold change  $\geq 1.20$ , p value  $< 0.05$



**Supplementary Figure S4. Chromatin accessibility at putative human *ALIVEC* locus in Human Coronary Artery Smooth Muscle Cells (HCASMCs)**

Normalized ATAC-seq tracks generated from HCASMCs under control, and PDGF-BB or TGFB treated conditions. Tracks were adapted through UCSC genome browser from <https://pubmed.ncbi.nlm.nih.gov/27386823/>. ATACseq represents Assay for Transposase-Accessible Chromatin using sequencing, PDGFBB and TGFB indicates platelet derived growth factor and transforming growth factor-beta.

**Supplementary Table S1. Primers sequences used in the study**

<b>Rat gene qPCR primers</b>	Forward	Reverse
<i>Alivec</i>	CTGCCCTTCTTTTCACCATGC	GCGGTACACCTGAAGCTAGT
<i>Acan</i>	ACTCAGGACATTAGCTGCATG	CACGTGTTCCCATTCACTTTG
<i>Runx1</i>	CGGGCAATGACGAAACTAC	GGGTTTGTAAGACGGTGATG
<i>Spp1</i>	TGGCTTACGGACTGAGGTCA	GACCTCAGAAGATGAACTCT
<i>Tnfaip6</i>	GTAGGAAGATACTGCGGTGAT GAA	GACGGACGCATCACTCAGAA
<i>Sox9</i>	AGGAAGCTGGCAGACCAGTA	ACGAAGGGTCTCTTCTCGCT
<i>Olr1</i>	ACGAGAAATCCAAAGAGCAGG	TCCCATTCACTTCCAGTTG
<i>Ppia</i>	TATCTGCACTGCCAAGACTGA GG	CTTCTTGCTGGTCTTGCCATCC
<i>Neat1</i>	GCAAGACCATGTGCCCTAGT	TCTGGAATTGGCCGGAAGTC
<i>H19</i>	TTGTCTGGCTTCATCCTGTG	TGGACTTGGTGCCTGTATG
<b>Rat ChIP qPCR primers</b>		
<i>Alivec</i> promoter	TTGTACCTTGTCCCCAGTTG	ATGCCCAACCCCATAGTTAG
<b>Human gene qPCR primers</b>		
<i>ALIVEC</i>	TGCCAAGCACCACTCTCAAT	AAGGTCAATGCCAGACAGGG
<i>ACAN</i>	TGCGGGTCAACAGTGCCTATC	CACGATGCCTTTCACCACGAC
<i>GAPDH</i>	CTTTTGCGTCGCCAGCCGAG	CCAGGCGCCCAATACGACCA

**Supplementary Table S2. Complete sequence of *Alivec***

ATAAAAGCCTACGGTCCTTTTGTAGACATAGTTTCATTGCCAGACTGGCCTTGAATTC  
GCAATCATGAGAAGACACAGGTACCTTCAGTCCAACGACTGTTGCATTAATTAGCCA  
GGGTGAAACTTGGGGATGTCAAGGATAGAGAGTTAAAGTGATTTTCATGAAGAAAAG  
GTATTGTAGTGACTATCCCTGTTGTCAACTTGACTATCCAGAATTGGAAGGTTACCT  
GTGATCGAGGTCTTGGGGGCTGGGAGATACAAGTTTCTGACCTTGATCTTGTTCATGA  
AGATCTTCTATATCTGGAGTGAAGTAGAGGGCACAGTGGCTATGAATCCCAGGCAAG  
AAGATCTCTGAGTTCAAGGTCATCTGGGACAAAGCAAGTCCCAGATCCAGAAGTCT  
GGATTACACATCTTTAATCTGGGCTACACCTTCTGCTGGAGACCTACATAAGGACAT  
TGGAAGAAGGGAGATGCTCTCTGCCTGCTTGCTTGCCTTGTAGGACTGAGCCACTGC  
TAGATCTTTGGACTTCCAATCATAGCTGCTGCTGACCATTGTTGGGGAGTTGGACTA  
CAGACTGTAAGCCGTCAACAATTTCCCTTACATATAAAGACTACTCATAAGTTCTGT  
GACTCTAGAGAACCCTGACTAATATAGTGTTGCCTTCTGTTCTCGGCATCTGTCTTCA  
GATTCCTATTGCTGTGATAAAACACCATGGCCAAAAGCAACTTGGAAGAAAAGGGT  
TTGTGTCCCATCATCCAGGGACTTCGGCCAACAGCCAGCAGTGCAGACAAGGGCGG  
CATGGACCTGCAGCCTGTGGCTGGCCCCCGAGGGCGAGCAGCCAGATGCCGGCAGC  
CGCAGGAGAGAAGGGCTGCTCTTCTCTTCACTCAAAGGTGGATTGAGGAAGGTTGG  
CTCTGAGATTTTTTTTTTCAAGAAGCTCGTCGGGAGTGGGAGAATATGAACATCTGTG  
CCCTGGGTTTCAGTTCTGGACTCGGCTCAGGCCACAGTCTGGAGCTGGCGTCAGCTGG  
AGACAAGCCTGCATCCTGCTGGCGGGTGGGTGGGGCTGCTTCTGGCCTGCTGGTTGA  
GGAACGCAGACACTAGCCTCCACCCTCTGGAGCCCAGGAGAGCGGCGGGCATGGGA  
GGGCACACAGAGCTCTCAGCGCTCAGGCACCAGGGTGCCTGCATGTACTGTGTTTAC  
AGAAGGGCTGGGGGCAGGTCCTGGAGTCTCGCAGGTACACTCCACAGGGAAAGGCA  
CCCCAGGTCTTTCCCTCCCCCTCTGCCTTTGGTGACAGGCATTTGTGGCCAATACTG  
ATGTTCTCTCTACACTTTATTCCCAGTCCCAGTTGTTTCATCAAGGCTGCAAGACAAT  
CCAGAAATTACCTAGTCAACTGGTTTTTCCCTGACCTTTGCTTTTTTTTAACTTAGCC  
TATTTTTTTTAACTTAAATTTTAGTGTGAAAAGGTAGTATATTTTGCTGGGGATATAGC  
TCAGTTGGCAGAGGGCTTGCTTAGCATGTACCACTAGGCCCTGGATTTGAGCCCCAA  
CACCACACAACTAGGCATGTTGCTGTAAGTCAATAAACCCAATCAATACTCTGGAA  
GTTCAAGATCTTCCTCAACTATCTATCCAGTTCAAAAACCAGTTTAAAAGAAAACAGT

GTATTTACATGGTTCACATTTTTTAAACGCTTAAATTGGTATCAATCTATATTTATATC  
TATCTATATATCTATATATCATCTATATCCATCTTTATCTCCATCTCTCTCTCCTCTCT  
GTCATCTATCTATATATCTCACACTTATCTGGATTTCATCTGCCCTTCTTTTCACCATG  
CCTTCCTACCTCATAGCCACAGTAAGATATGTACTAGCTTCAGGTGTACCGCTTCTT  
GGCTGGGTTTGTTTAGTAGCACACACCTTTAATCTCAGTACTGGGAGGCAGAAGAAG  
ACAGATCTTATGAATGTGAGGCCAGCCTGGTCTACATAGAAAGTTCTAGGCCAGCCA  
AGACTACATAATGAGATTCTGTCTCTAACAAACAAATCCTTATACAGTTTTGAAATG  
AAAATTGAGAATTAAAAATTTTCATATCAAAAAGTAGCTTTTACTTCATTACTTTATTT  
TATTATGTAATAAATTGTGTACAATATATTTTCTATATTTTGTTTTGTGGATAAAC  
ATTATTAGCACTAAGATGCAATCCACAGACCACAGGAAGCTCAAGAAGAAGGATGA  
CCAAAATGCGGATGCTCTCACTCCTTCTTAAAAGGGGCAAAAAAAAAA



**Supplementary Table S3. Sequences of GapmeRs and siRNAs used in the study**

GapmeR or siRNA	Sequence	Vendor or source
Alivec Gap1	GCAACAGTCGTTGGAC	QIAGEN
Alivec Gap2	TTGAACTTCCAGAGTA	QIAGEN
Alivec Gap3 ( <i>AlivecGap</i> )	GATTGCATCTTAGTGC	QIAGEN
Negative control Gap (NCGap)	AACACGTCTATACGC	QIAGEN
Sox9 siRNA pool	GCGUCAAACGGCUCCAGCAA; GCUCGGAACUGUCUGGAAA; GCCAGGUGCUGAAGGGCUA; GUAAGUGAAGGUAACGAUU	Horizon

**Supplementary Table S4. Antibodies used in the study and their source**

Target protein	Vendor or source	Antibody & Cat No.
Tnfaip6	Proteintech	Anti-TSG6, 13321-1-AP
Runx1	Proteintech	Anti-RUNX1,19555-1-AP
Tpm3	Genetex	Anti-Tpm3, GTX113568
hnRNPA2B1	Origene	Anti-hnRNPA2B1, TA3140
Alpha-SMA	Abcam	Anti-SMA, ab5694
Tagln (SM22)	Proteintech	Anti-TAGLN, 10493-1-AP
Acan	Proteintech	Anti-aggrecan, 13880-1-AP
Sox9	Millipore	Anti-Sox9, AB5535
Beta-actin	Sigma-Aldrich	Anti-b-Actin, A5441