

Article

Effects of Partial Replacement of Soybean with Local Alternative Sources on Growth, Blood Parameters, Welfare, and Economic Indicators of Local and Commercial Broilers

Muazzez Cömert Acar ¹, Berna Türkekul ², Özlem Karahan Uysal ², Sezen Özkan ¹ and Servet Yalcin ^{1,*}

¹ Department of Animal Science, Faculty of Agriculture, Ege University, 35100 İzmir, Türkiye; muazzez.comert@ege.edu.tr (M.C.A.); sezen.ozkan@ege.edu.tr (S.Ö.)

² Department of Agricultural Economics, Faculty of Agriculture, Ege University, 35100 İzmir, Türkiye; berna.turkekul@ege.edu.tr (B.T.); ozlem.uysal@ege.edu.tr (Ö.K.U.)

* Correspondence: servet.yalcin@ege.edu.tr

Simple Summary: While the demand for poultry meat is increasing, arable land for crop production is limited. Therefore, the use of locally produced alternative sources in chicken diets has become a necessity. The substitution of soybean with local by-products such as sunflower meal, brewers' dried grain, and wheat middlings or a combination of local by-products with black soldier larvae meal was evaluated in broilers. The substitution of soybean in broiler diets did not affect growth performance. However, the diet with black soldier fly larvae meal increased the production costs because of its high price. Therefore, lowering black soldier fly larvae meal price is the key issue hindering its inclusion in broiler diets.

Abstract: The effects of the partial replacement of soybean with alternative local agri-industry by-products and black soldier fly (BSF) larvae meal on broiler growth performance, blood biochemistry, welfare, and, subsequently, economic performance of these diets were evaluated. A total of 524 day-old chicks from a local and a commercial strain were fed one of the three diets from the day of hatch to the slaughter age. The diets were the following: a soybean-based control diet, a diet in which soybean was partially replaced (SPR) with agri-industrial by-products, or a diet with BSF larvae meal added to the SPR (SPR + BSF). There was no effect of the diets on the slaughter weight, total feed consumption, and feed conversion of the chickens. The SPR + BSF diet reduced the blood glucose, alanine aminotransferase, aspartate aminotransferase, gamma-glutamyl transferase, protein, triglycerides, and cholesterol levels in the local chickens and the gamma-glutamyl transferase, protein, and creatinine levels in the commercial broilers. The negative effect of the SPR diet on plumage cleanliness in the commercial broilers was alleviated by the SPR + BSF diet, whereas 100% of the local birds presented either slight or moderate soiling. The results showed that, due to the high cost of the BSF larvae meal, the SPR + BSF diet was not economically feasible. In a further study, the price trends of BSF larvae will be examined from the standpoint of economic profitability conditions.

Keywords: alternative feedstuffs; meat-type chicken strains; growth performance; feather cleanliness; economic profitability



Citation: Acar, M.C.; Türkekul, B.; Karahan Uysal, Ö.; Özkan, S.; Yalcin, S. Effects of Partial Replacement of Soybean with Local Alternative Sources on Growth, Blood Parameters, Welfare, and Economic Indicators of Local and Commercial Broilers. *Animals* **2024**, *14*, 314. <https://doi.org/10.3390/ani14020314>

Academic Editors: Vassilios Dotsa and George K. Symeon

Received: 11 December 2023

Revised: 12 January 2024

Accepted: 15 January 2024

Published: 19 January 2024



Copyright: © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

1. Introduction

Despite the negative impact of rising global feed prices on profitability, the market for chicken meat is projected to have steady growth due to its comparative advantages over beef and sheep meat in terms of affordability, health benefits, and environmental sustainability [1]. Soybean serves as the main source of plant protein utilized in conventional chicken diets. The increasing demand for soybean meal and its environmental impact has led to increased efforts to investigate alternative feedstuffs in chicken diet formulations [2]. Therefore, emphasis has been given to the use of locally available agro-industrial

by-products that are not intended for human consumption and can be used in chicken nutrition [3]. Several previous studies have demonstrated the potential of the inclusion of by-products such as sunflower meal, brewers' dried grain, and wheat middlings in broiler diets [4–6]. Although their use is limited due to their comparatively higher fiber and lower lysine and methionine levels than soybean meal, this limitation can be overcome by adding enzymes to the diets. The addition of enzymes has been shown to increase the nutritional value and digestibility of non-starch polysaccharides and proteins in broilers [7–9].

Sunflower meal is a byproduct of the sunflower seed oil extraction process. Moghadam et al. [10] reported that the inclusion of up to 140 g/kg of SFM in the diet improved body weight gain, feed intake, feed conversion, and the weight of the gastrointestinal tract and gizzard and reduced blood low-density lipoproteins in broilers. Salari et al. [11] found that adding up to 210 g/kg full-fat sunflower seed to the broiler diet improved body weight gain, feed intake, and feed conversion ratios, while neither low-density lipoprotein nor blood alkaline phosphatase, Ca, P, glucose, and protein concentrations were affected.

Brewers' dried grain is the most common by-product of the brewery industry. The chemical composition of this grain is subject to variation as a result of the brewing process, although it is known to contain proteins, fibers, and lipids [12]. The addition of 20% brewers' dried grain in local chickens' diets, partially replacing maize and soybean meal, contributed to gizzard development [7] and increased carcass yields and profit margins without affecting growth performance [13]. In contrast to these findings, the inclusion of brewers' dried grain at a 3 to 9% concentration was reported to be optimum, while an improvement in kidney and liver functions was reported by reducing the blood AST levels and creatinine levels [14]. It was also shown that wheat middlings, a by-product of the wheat milling process, could be incorporated into broiler diets up to a 30% concentration without any detrimental effects on body weight gain, slaughter weight, and feed conversion ratio [15].

Recently, there has been growing interest in using insects as an alternative protein source to partially replace soybeans in broiler diets [2]. The European Commission authorized the use of processed animal proteins obtained from insects in poultry diets (Commission Regulation 2021/1372). The nutritional content of black soldier fly (*Hermetia illucens* L.) (BSF) larvae makes them suitable for meeting the dietary requirements of animals [16]. BSF larvae can be considered an environmentally sustainable alternative feedstuff. This is primarily due to the relatively small land area required to produce one kg of protein, their use of organic waste, and their low levels of greenhouse gas emissions [17–20]. The inclusion of 100 g/kg of BSF larvae meal has been shown to provide a protein supply covering 33.01% of the total protein requirements and improve the carcass weight of broilers without affecting their blood parameters and health status [21,22]. De Souza Vilela et al. [23] showed that a concentration of up to 20% of full-fat BSF larvae in broiler diets increased body weight gain compared to the control diet. Murawska et al. [24] concluded that the replacement of soybean meal with full-fat BSF larvae meal should be lower than 50% as higher inclusion levels of full-fat BSF meal deteriorated growth performance. Gariglio et al. [25] reported that the inclusion of de-fatted BSF larvae meal up to a 9% concentration in Muscovy ducks' diets did not result in reduced growth and improved some of the blood traits. Moreover, it has been also suggested that the inclusion of whole larvae has been an effective mean of environmental enrichment for broiler chickens, positively affecting the welfare of birds by increasing foraging activity [26], improving footpad health, and reducing fear [27].

There is an increasing interest in using slower-growing and local chickens. Anadolu-T is a registered genotype for broiler production and is within the scope of the selection and breeding program in Türkiye. The pure lines of Anadolu-T and their crosses had lower body weights and impaired feed conversion ratios but a higher livability compared to commercial strains [28]. These chickens could be an alternative for niche markets and small local producers. On the other hand, feed is the main cost, with 65–75% of the total cost of broiler production. Therefore, the cost of a diet is also important, and the use of alternative protein sources depends not only on their impact on performance but also on their price.

Based on the above-mentioned background, we hypothesized that locally produced agri-industrial by-products with enzyme and BSF larvae supplementation could be used to partially replace soybean in broiler diets. Therefore, the effect of diets in which soybean was partly replaced with sunflower meal, brewers' dried grain, and wheat middlings as well as a combination of BSF larvae meal with these local agri-industrial by-products on broiler performance, blood biochemistry, animal welfare, and economic efficiency of production was investigated. A local and commercial line was used to evaluate the effects of the diets on the strains.

2. Materials and Methods

The present study is a part of the ongoing PRIMA project "Alternative animal feeds in Mediterranean poultry breeds to obtain sustainable products" (SUSTAvianFEED) (Grant number 2015). All procedures were approved by the Farm Animal Experiments Local Ethics Committee of the Faculty of Agriculture at Ege University (Approval no.: 2022/002, 7 April 2022).

2.1. Experimental Design and Diets

A total of 504 day-old chicks (252 chicks/strain) from local (Anadolu-T pure dam line) and commercial (Cobb 500) strains were used. The chicks from both sexes were individually weighed, wing-banded, and placed in 36-floor pens with 14 chicks in each pen in an environmentally controlled experimental house. Each 1.4 m × 1.2 m floor pen was furnished with wood-shaving litter, one round feeder, and nipple drinkers. The ambient temperature was adjusted to 32 ± 1 °C on the first day and gradually decreased to reach 24 ± 1 °C on day 21. The lighting program was 23L:1D for the first three days and then gradually reduced to 16L:8D (about 20 lx) by day seven, and kept until the end of the experiment.

The chicks were fed one of three diets with six replications from the day of hatching to the slaughter age, which was 40 d for the commercial and 55 d for the local strains. The control diet was a typical soybean–corn diet. Soybean partial replacement (SPR) was formulated by partly replacing soybean with local agri-industrial by-products consisting of high-protein sunflower meal, brewers' dried grain, and wheat middlings. Dried BSF (5%) larvae meal was included in the SPR to obtain a SPR + BSF diet. An average of 24.9 and 42.2% of soybean meal was replaced with alternative agri-industrial by-products in the SPR and SPR + BSF diets, respectively. All the diets were formulated to be isocaloric and isonitrogenous. The proximate composition of the experimental diets and sunflower meal, brewers' dried grain, wheat middlings, and dried BSF larvae meal reared on vegetable and bakery by-products in Türkiye (Germina Tarım Teknolojileri Tic. Ltd., Şti., Ankara, Türkiye) are shown in Tables 1 and 2, respectively.

Table 1. Ingredients and nutrient composition of the experimental diets for the starter (0–10 d), grower (11–25 d), and finisher (26–slaughter age¹) periods.

| | Control | | | SPR | | | SPR + BSF | | |
|----------------------|---------|--------|----------|---------|--------|----------|-----------|--------|----------|
| | Starter | Grower | Finisher | Starter | Grower | Finisher | Starter | Grower | Finisher |
| Corn | 45.28 | 51.24 | 57.34 | 39.18 | 44.44 | 47.44 | 41.3 | 46.54 | 49.64 |
| Wheat | 11.86 | 14.86 | 15.00 | 12.5.0 | 14.50 | 15.50 | 12.18 | 14.50 | 15.50 |
| Soybean meal | 34.33 | 27.90 | 23.20 | 29.8 | 21.10 | 14.6 | 25.10 | 16.30 | 9.70 |
| Sunflower meal | - | - | - | 3.58 | 6.30 | 8.00 | 3.63 | 6.30 | 8.00 |
| Brewers' dried grain | - | - | - | 2.58 | 3.08 | 4.00 | 2.63 | 3.08 | 4.00 |
| Wheat middling | - | - | - | 2.58 | 3.08 | 4.00 | 2.63 | 3.08 | 4.00 |
| BSF larvae | - | - | - | - | - | - | 5.00 | 5.00 | 5.00 |
| Sunflower oil | 5.88 | 4.00 | 3.00 | 7.13 | 5.5.0 | 5.00 | 4.88 | 3.20 | 2.70 |
| Limestone | 0.50 | 0.30 | 0.20 | 0.50 | 0.30 | 0.20 | 0.50 | 0.30 | 0.20 |

Table 1. Cont.

| | Control | | | SPR | | | SPR + BSF | | |
|--|---------|--------|----------|---------|--------|----------|-----------|--------|----------|
| | Starter | Grower | Finisher | Starter | Grower | Finisher | Starter | Grower | Finisher |
| DCP | 1.00 | 0.80 | 0.60 | 1.00 | 0.80 | 0.60 | 1.00 | 0.80 | 0.60 |
| Vit+ min premix ² | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 | 0.25 |
| NaCl | 0.20 | 0.20 | 0.20 | 0.20 | 0.20 | 0.20 | 0.20 | 0.20 | 0.20 |
| Lysine (HCL—% 78) | 0.50 | 0.30 | 0.15 | 0.50 | 0.30 | 0.15 | 0.50 | 0.30 | 0.15 |
| Methionine dl (% 99) | 0.10 | 0.05 | 0.01 | 0.1 | 0.05 | 0.01 | 0.01 | 0.05 | 0.01 |
| Threonine | 0.05 | 0.05 | - | 0.05 | 0.05 | - | 0.05 | 0.05 | - |
| Enzyme ³ | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 | 0.05 |
| Analyzed Nutrient Composition ⁴ | | | | | | | | | |
| ME kcal/kg diet | 2984 | 2923 | 2904 | 2992 | 2921 | 2904 | 2991 | 2919 | 2903 |
| CP, % | 20.78 | 18.68 | 17.00 | 20.74 | 18.65 | 17.05 | 20.78 | 18.64 | 17.04 |
| EE, % | 8.49 | 6.63 | 5.79 | 9.41 | 8.11 | 7.52 | 9.38 | 7.72 | 7.52 |
| CF, % | 2.91 | 2.61 | 2.35 | 3.70 | 3.69 | 3.77 | 3.94 | 3.92 | 3.99 |
| Ca, % | 1.08 | 1.04 | 1.02 | 1.08 | 1.03 | 0.99 | 1.13 | 1.08 | 1.05 |
| Total P | 0.50 | 0.44 | 0.38 | 0.55 | 0.52 | 0.47 | 0.55 | 0.52 | 0.48 |

¹ Slaughter age was 55 and 40 d for the local and commercial strains, respectively. ² Vitamin + mineral premix provided per 2.5 kg feed of diet: Vitamin A, 15,000,000 IU; Vitamin D3, 3,000,000 IU; Vitamin E, 50,000 mg; Vitamin K3, 4000 mg; Vitamin B1, 3000 mg; Vitamin B2, 6000 mg; Niacinamid, 40,000 mg; Vitamin B6, 5000 mg; Vitamin B12, 30 mg; Calcium-D-Pantothenate, 15,000 mg; Biotin, 75 mg; Folic acid, 1000 mg; Choline Chloride, 400,000 mg; Manganese, 80,000 mg; Iron, 60,000 mg; Copper, 5000 mg; Zinc, 60,000 mg; Iodine, 2,000 mg; and Selenium, 150 mg. ³ Rovabio (50 gr) + Natuphos E (100 gr) BASF. ⁴ ME: metabolizable energy; CP: crude protein; EE: ether extract; CF: crude fiber; Ca: calcium; Total P: total phosphorus.

Table 2. Analyzed nutrient compositions of alternative feedstuffs (% on dry matter).

| | Sunflower Meal | Brewers' Dried Grain | Wheat Middlings | BSF Larvae |
|--------------------------------------|----------------|----------------------|-----------------|------------|
| Metabolizable energy, kcal/kg | 2108 | 1565 | 1837 | 5381 |
| Dry matter | 90.46 | 90.53 | 88.20 | 95.52 |
| Crude protein | 41.78 | 28.61 | 17.53 | 42.62 |
| Ether extract | 1.65 | 2.68 | 4.34 | 42.54 |
| Crude fiber | 14.37 | 20.20 | 7.94 | 11.04 |
| Crude ash | 6.92 | 4.11 | 5.98 | 6.29 |
| Starch | - | 2.71 | 17.87 | - |
| Total sugar | 8.18 | 2.07 | - | 1.79 |
| Neutral detergent fiber | 37.57 | 74.68 | 32.59 | 17.56 |
| Acid detergent fiber | 29.88 | 28.97 | 10.78 | 13.36 |
| Acid detergent insoluble nitrogen, % | - | - | - | 0.74 |
| Methionine | 0.70 | 0.24 | 0.26 | 0.65 |
| Lysine | 1.59 | 0.88 | 0.68 | 1.35 |
| Threonine | 1.91 | 1.14 | 0.56 | 2.73 |
| Calcium | 0.43 | 2.55 | 1.05 | 1.70 |
| Total phosphorus | 1.38 | 0.78 | 0.14 | 0.62 |
| Fatty acids | | | | |
| Σ SFA ¹ , g/100 g lipid | 29.45 | 27.43 | 19.21 | 71.89 |
| Σ MUFA ² , g/100 g lipid | 27.96 | 18.79 | 17.56 | 17.09 |
| Σ PUFA ³ , g/100 g lipid | 42.59 | 53.78 | 62.89 | 11.02 |

¹ SFA: saturated fatty acids; ² MUFA: monounsaturated fatty acids; ³ PUFA: polyunsaturated fatty acids.

All the chicks were vaccinated against Newcastle disease, Gumboro disease, and infectious bronchitis at the hatchery. On days 10 and 18, Newcastle and infectious bronchitis vaccine recalls were performed. Coccidiostats were not used during the experimental period.

2.2. Measurements

Individual body weights were measured on days 0, 10, and 25 and at the slaughter age. Feed consumption was measured on the same dates on a pen basis. Feed conversion was calculated considering the weight of the dead birds.

At day 40 for the commercial and day 55 for the local chickens, eighteen broilers (three birds/replicate, nine from each sex) from each treatment close to the average weight were randomly selected. After 8 h of feed withdrawal, blood samples were collected in a tube during the slaughtering process. The breast (including the bones) and the legs (including the hip and the bones) were separated from the carcass of each bird and weighed. The weights of the liver, the whole intestine, the spleen, and the bursa of Fabricius were measured. The relative weights were calculated by dividing the individual organ weights by the live body weight of the birds. The serum was separated after centrifugation at 4 °C and 2750 rpm for 10 min and stored at −80 °C until the analysis of blood metabolites. Blood protein, triglycerides, uric acid, alanine aminotransferase (ALT), aspartate aminotransferase (AST), gamma-glutamyl transferase (GGT), creatinine, cholesterol, corticosterone, and the blood minerals of Ca, P, and Mg concentrations were measured using commercial kits (Mindray, China) and an autoanalyzer (Mindray BS-240 Vet, Nanshan, Shenzhen 518057, China). The ELISA kits (Chicken corticosterone ELISA kit; Bioassay Technology Lab., Shanghai, China) were used to assess and quantify the corticosterone concentration of the blood samples following the manufacturer's instructions. The absorbance was measured at 450 nm, and the concentration of corticosterone was determined relative to the absorbance of the calibration curve.

Tonic immobility testing was conducted at 28 d of age and at the slaughter age. Two birds (one male and one female) from each replicate pen were randomly chosen (twelve in total), placed on their backs on a table, and held for 15 s. Then, hands were slowly removed from the birds, and the timer was turned on to measure the latency to the right, with a maximum of 120 s [29]. If the birds righted themselves in less than 15 s, the same procedure was repeated for up to three times. The birds were given a score of zero if all three attempts for TI induction failed. Longer tonic immobility durations were associated with a higher fearfulness [29,30]. On days 40 and 55, for the commercial and local chickens, respectively, plumage cleanliness and footpad dermatitis (FPD) were scored on all the birds. A four-point scoring scale for plumage cleanliness (0: intact; 1: slight soiling; 2: moderate soiling; and 3: severe soiling) [31] and a three-point scale for footpad dermatitis (0: intact footpad; 1: moderate FPD; and 3: severe FPD) were used [32].

2.3. Chemical Analysis

The diets were analyzed for dry matter, crude protein, ether extract, crude fiber, total starch, total sugar, and total phosphorus and calcium [33]. The metabolizable energy content (ME) of the diet was calculated from the chemical composition using the equation $\text{ME kcal/kg} = 38 \times [(\text{Crude Protein}) + (2.25 \times \text{Ether Extract}) + (1.1 \times \text{Starch}) + (1.05 \times \text{Total Sugar})] + 53$ [32]. A Fiber-Tec system was used to determine the neutral detergent fiber (NDF) value with alpha amylase treatment and the acid detergent fiber (ADF) value [34]. Moreover, BSF larvae meal was analyzed for acid detergent insoluble nitrogen (ADIN) by Kheldahl [35] and for ash-free ADF in the residues of acid detergent fibers. The acid detergent fiber-linked protein (ADIP) was calculated by multiplying ADIN with 6.25. The chitin level in the BAF larvae meal, which was 8.32%, was calculated according to the following equation: $\text{Chitin (\%)} = \text{ash free ADF (\%)} - \text{ADIP (\%)} [36]$.

The fatty acid composition was determined using gas-chromatography after the lipid was extracted according to Folch et al. [37]. The analyses were performed on an Agilent

6890 Network Gas Chromatograph (USA). The column was an Agilent HP-88 capillary (100 m × 0.25 mm ID × 0.2 µm). An amino acid analysis was carried out via high-performance liquid chromatography using HPLC (Agilent 1260 Infinity II, Santa Clara, CA 95051, USA).

2.4. Cost–Benefit Analysis

The economic implications of substituting soybean meal with local ingredients and BSF larvae were evaluated using key criteria such as gross profit margin (GPM), cost–benefit ratio (CBR), and return on investment (RoI) [38]. The costs of feed consumed were determined using the market prices of the ingredients for 2023 and considering the quantities of each item used in the diets. The GPM was calculated using the following formula: $GPM = \text{total selling price of carcass (SP)} - \text{cost of feed consumed (CF)}$ (in euros). The total selling price of a carcass is estimated by multiplying the market price of chicken meat for 2023 by the average carcass weight. The utilization of CBR within the framework of a cost–benefit analysis was employed to succinctly assess the economic worth of substituting soybean meal with local ingredients. The CBR was determined by dividing the SP by the CF. A CBR value above 1 indicated that the benefits derived from production surpassed the associated production costs, whereas a value below 1 suggested the opposite. The RoI is a metric used to evaluate the profitability of an investment by comparing the gain or loss achieved to the amount of money initially invested. The RoI was calculated using the following formula: $RoI = (GPM/CF) \times 100$. The greater the value of the RoI, the more favorable the financial gains of the project being evaluated [38].

2.5. Statistical Analyses

Several models were used to analyze the data. A two-way factorial model including diet, strain, and their interaction was used to analyze performance and blood parameters. The experimental unit for the body and organ weights, blood parameters, tonic immobility, footpad, and feather cleanliness was individual birds. A pen was the experimental unit for feed consumption and feed conversion. The responses of the strains to the diets were similar for the performance parameters but differed for the blood parameters. Our purpose was to estimate the diet effect for each strain; therefore, all the parameters were also analyzed separately. The Shapiro–Wilk test was used to assess the normality of distribution before the analysis. When there are significant interactions, the means were separated using Student’s *t*-test. *p* values < 0.05 indicate statistical significance.

A nonparametric Kruskal–Wallis test was used for the statistical analysis of tonic immobility duration and induction numbers. The footpad dermatitis and feather cleanliness scores at the slaughter age for each strain were analyzed with a chi-square test of independence. For the footpad dermatitis scoring, there were very few birds with a score of 1 (moderate) among the Cobb strain; therefore, scores 1 and 2 were pooled, resulting in a binomial scaling for intact and severely (score 1 + 2) affected birds. For the plumage cleanliness of the Cobb broilers, there were no birds with a score of 3, and there were only a few birds that scored 2. Therefore, the data were pooled as a one–zero scaling, as 0 indicated clean plumage and 1 indicated slight soiling. In the local line, there were no birds who scored 0 (clean) or 3 (severe soiling). Therefore, all the birds from the local strain showed slight (1) or moderate (2) discoloration. When the chi-square test indicated a statistically significant effect, the cell chi-squares and the adjusted standardized residuals for each cell in the contingency table were calculated for a post hoc evaluation. The cells associated with adjusted standardized residuals greater than ±2 indicated a significant contribution of that specific cell to the omnibus chi-square [39].

3. Results

3.1. Performance of the Birds

The analyses of variance (significance of p -values) for body weight, feed consumption, and feed conversion ratio by diet and strain and the related means are presented in Tables 3 and 4, respectively.

Table 3. Analyses of variance (significance of p -values) for body weight, feed consumption, and feed conversion ratio by strain and diet.

| | | Strain | Diet | Strain \times Diet |
|------------------|--------------------|--------|--------|----------------------|
| Body weight | 0 d | <0.001 | 0.298 | 0.279 |
| | 10 d | <0.001 | <0.001 | 0.348 |
| | 25 d | <0.001 | 0.806 | 0.565 |
| | Slaughter age | 0.317 | 0.134 | 0.672 |
| Feed consumption | 0–10 d | 0.167 | <0.001 | 0.816 |
| | 11–25 d | <0.001 | 0.530 | 0.567 |
| | 26 d—Slaughter age | <0.001 | 0.530 | 0.567 |
| | Total | <0.001 | 0.493 | 0.586 |
| Feed conversion | 0–10 d | <0.001 | 0.128 | 0.667 |
| | 11–25 d | <0.001 | 0.276 | 0.388 |
| | 26 d—Slaughter age | <0.001 | 0.864 | 0.674 |
| | Total | <0.001 | 0.908 | 0.688 |

Table 4. Effects of the diets ¹ on body weight, feed consumption, and feed conversion ratio in local and commercial strains.

| Performance | | Local Strain | | | | | Commercial Strain | | | | |
|------------------|----------------------------|------------------|------------------|------------------|------------------|------------|-------------------|-------------------|-------------------|-------|------------|
| | | Control | SPR | SPR + BSF | SEM ² | p -Value | Control | SPR | SPR + BSF | SEM | p -Value |
| Body weight | 0 d | 35.32 | 35.37 | 35.34 | 0.27 | 0.993 | 36.96 | 36.63 | 36.38 | 0.29 | 0.368 |
| | 10 d | 218 ^b | 213 ^b | 230 ^a | 3.1 | <0.001 | 236 ^b | 242 ^{ab} | 251 ^a | 3.4 | 0.012 |
| | 25 d | 792 | 777 | 805 | 12.6 | 0.280 | 998 | 1007 | 989 | 15.2 | 0.692 |
| | Slaughter age ³ | 2223 | 2283 | 2236 | 30.7 | 0.353 | 2161 | 2247 | 2181 | 31.3 | 0.112 |
| Feed consumption | 0–10 d | 366 ^b | 390 ^b | 444 ^a | 15.6 | 0.008 | 356 | 372 | 414 | 18.6 | 0.106 |
| | 11–25 d | 975 | 968 | 985 | 19.8 | 0.844 | 1120 | 1150 | 1161 | 19.4 | 0.344 |
| | 25 d—slaughter age | 3820 | 3855 | 3703 | 96.4 | 0.519 | 2292 | 2438 | 2372 | 109.0 | 0.645 |
| | Total | 5162 | 5214 | 5133 | 101.4 | 0.849 | 3769 | 3960 | 3948 | 118.1 | 0.458 |
| Feed conversion | 0–10 d | 2.03 | 2.21 | 2.28 | 0.09 | 0.191 | 1.79 | 1.81 | 1.91 | 0.092 | 0.629 |
| | 11–25 d | 1.71 | 1.68 | 1.70 | 0.042 | 0.918 | 1.48 ^b | 1.49 ^b | 1.57 ^a | 0.020 | 0.021 |
| | 25 d—slaughter age | 2.65 | 2.58 | 2.60 | 0.064 | 0.736 | 1.93 | 1.96 | 1.92 | 0.086 | 0.932 |
| | Total | 2.35 | 2.32 | 2.33 | 0.0336 | 0.844 | 1.76 | 1.78 | 1.79 | 0.052 | 0.869 |

^{a,b} The means in the same row within a strain with different superscripts differ significantly ($p < 0.05$). ¹ Diets: control: corn-soybean-based diet; SPR: the soybean in the control diet was partially replaced with sunflower meal, brewers' dried grain, and wheat middlings; SPR + BSF: black soldier fly dried larvae were added to the SPR diet. ² SEM: standard error of means. ³ Slaughter age was 55 and 40 d for the local and commercial strains, respectively.

On day 10, the birds from both experiments who had been fed the SPR + BSF diet had the heaviest body weight, whereas the body weights of the birds who had been fed the SPR diet were similar to the control. The effect of the diets on the body weights of the birds on day 25 and at the slaughter age was not significant. The feed consumption of the local and Cobb birds fed the SPR + BSF diet increased from day 0 to day 10 compared to the feed consumption of those fed the SPR and control diets. The feed consumption of the chickens from both experiments was similar from days 11 to 25 and from day 26 to the slaughter age (Table 4). The total feed consumption was 5162, 5214, and 5133 g for the local chickens and 3769, 3960, and 3948 g for the Cobb chickens who had been fed the control, SPR, and SPR + BSF diets. From day 11 to day 25, the feed conversion ratio of the Cobb chickens was impaired with the SPR + BSF diet. There was no significant effect of the diets on the feed

conversion ratio from day 0 to day 10, from day 25 to the slaughter age, or from day 0 to the slaughter age (Table 4).

The commercial broilers had a higher breast yield compared to the local chickens. The SPR and SPR + BSF diets significantly increased the breast yield of the local line. The diets did not affect the breast and leg yield of the Cobb broilers (Tables 5 and 6).

Table 5. Analyses of variance (significance of *p*-values) for the relative weights of breast, leg, liver, intestine, spleen, and bursa of Fabricius by strain and diet.

| | Strain | Diet | Strain × Diet |
|--------------------|--------|-------|---------------|
| Breast | <0.001 | 0.017 | 0.027 |
| Leg | 0.215 | 0.651 | 0.974 |
| Liver | <0.001 | 0.147 | 0.391 |
| Intestine | 0.583 | 0.016 | 0.064 |
| Spleen | 0.374 | 0.342 | 0.953 |
| Bursa of Fabricius | <0.001 | 0.199 | 0.261 |

Table 6. Effects of the diets ¹ on the relative weights of breast, leg, digestive, and immune system organs in local and commercial strains at the slaughter age ².

| | Local Strain | | | | | Commercial Strain | | | |
|--------------------|--------------------|--------------------|--------------------|------------------|------------------------------|-------------------|-----------|---------|-----------------|
| | Control | SPR | SPR + BSF | SEM ³ | <i>p</i> -Value ⁴ | SPR | SPR + BSF | SEM | <i>p</i> -Value |
| Breast | 19.54 ^b | 20.70 ^a | 21.30 ^a | 0.399 | 0.016 | 23.08 | 23.69 | 0.370 | 0.518 |
| Leg | 18.90 | 18.69 | 18.97 | 0.268 | 0.744 | 19.01 | 19.19 | 0.238 | 0.867 |
| Liver | 1.62 ^a | 1.49 ^b | 1.52 ^b | 0.043 | 0.007 | 1.48 | 1.46 | 0.033 | 0.853 |
| Intestine | 4.25 | 4.62 | 4.09 | 0.17 | 0.122 | 3.64 | 3.71 | 0.126 | 0.891 |
| Spleen | 0.112 | 0.116 | 0.112 | 0.0064 | 0.922 | 0.099 | 0.083 | 0.0055 | 0.104 |
| Bursa of Fabricius | 0.0516 | 0.0467 | 0.0506 | 0.04177 | 0.701 | 0.0484 | 0.0543 | 0.00442 | 0.476 |

^{a,b} The means in the same row within a strain with different superscripts differ significantly ($p < 0.05$). ¹ Diets: control: corn–soybean-based diet; SPR: the soybean in the control diet was partially replaced with sunflower meal, brewer's dried grain, and wheat middlings; SPR + BSF: black soldier fly dried larvae were added to the SPR diet.

² Slaughter age was 55 and 40 d for the local and commercial strains, respectively. ³ SEM: standard error of means.

⁴ *p*-Value: Significance

3.2. Blood Parameters and Organ Weights

The relative weights of the liver and bursa of Fabricius were higher in the commercial strain than in the local chickens. The SPR and SPR + BSF diets reduced the relative liver weights in both strains. The relative weights of the intestine, spleen, and bursa of Fabricius were not influenced by the diets (Tables 5 and 6).

Tables 7 and 8 present analyses of variance (significance of *p*-values) and means for the blood parameters.

At the slaughter age, all the blood parameters measured, except for corticosterone, were higher in the commercial strain than in the local chickens. The local birds who had been fed the SPR + BSF diets had the lowest glucose levels, followed by those who had been fed the SPR and control diets (Table 8). The SPR diet increased the blood ALT levels of the local chickens. The blood AST, GGT, protein, triglycerides, and cholesterol levels of the local chickens were reduced by the SPR + BSF diet, whereas the birds who had been fed the SPR and control diets had similar levels. The diets had no significant effect on the blood glucose, AST, triglycerides, total cholesterol, uric acid, and Ca levels of the Cobb broilers (Table 8). The SPR diet tended to reduce the blood ALT levels and significantly increase the blood GGT levels of the Cobb broilers. While the blood total protein levels of the Cobb broilers decreased in the birds who had been fed the SPR + BSF diet, similar blood protein levels were obtained in the birds who had been fed the control and SPR diets. The highest creatinine levels were observed for the Cobb birds who had been fed the control diet. The diets had no significant effect on the blood corticosterone and blood uric acid levels of the chickens from either strain (Table 8).

Table 7. Analyses of variance (significance of *p*-values) for blood parameters by diet and strain.

| Blood Parameters ¹ | Strain | Diet | Strain × Diet |
|-------------------------------|--------|--------|---------------|
| Corticosterone | 0.926 | 0.926 | 0.971 |
| Glucose | <0.001 | 0.007 | <0.001 |
| ALT | <0.001 | 0.518 | <0.001 |
| AST | <0.001 | 0.030 | 0.048 |
| GGT | <0.001 | 0.030 | 0.021 |
| Protein | <0.001 | <0.001 | 0.004 |
| Triglycerides | <0.001 | 0.358 | 0.034 |
| Cholesterol | <0.001 | <0.001 | 0.073 |
| Creatinine | <0.001 | 0.041 | 0.094 |
| Uric acid | 0.003 | 0.038 | 0.281 |
| Mg | <0.001 | <0.001 | <0.001 |
| Ca | <0.001 | <0.001 | <0.001 |
| P | <0.001 | <0.001 | 0.012 |

¹ ALT: alanine aminotransferase; AST: aspartate aminotransferase; GGT: gamma-glutamyl transferase.

Table 8. Effects of the diets ¹ on the blood corticosterone and blood metabolites in local and commercial strains at slaughter age ².

| Blood Metabolites ⁴ | Local Strain | | | | | Commercial Strain | | | | |
|--------------------------------|--------------------|--------------------|--------------------|------------------|-----------------|--------------------|--------------------|--------------------|--------|-----------------|
| | Control | SPR | SPR + BSF | SEM ³ | <i>p</i> -Value | Control | SPR | SPR + BSF | SEM | <i>p</i> -Value |
| Corticosterone, ng/m | 8.1 | 7.44 | 8.18 | 0.592 | 0.222 | 8.77 | 7.59 | 8.31 | 0.740 | 0.531 |
| Glucose, mg/dL | 183 ^a | 159 ^b | 110 ^c | 9.5 | <0.001 | 212 | 226 | 221 | 7.0 | 0.354 |
| ALT, U/L | 1.57 ^b | 2.47 ^a | 1.40 ^b | 0.252 | 0.015 | 3.17 | 2.54 | 3.18 | 0.288 | 0.062 |
| AST, U/L | 239 ^a | 231 ^a | 146 ^b | 13.3 | <0.001 | 411 | 396 | 386 | 28.1 | 0.837 |
| GGT, U/L | 15.82 ^a | 15.05 ^a | 9.21 ^b | 1.107 | <0.001 | 18.39 ^b | 20.75 ^a | 17.44 ^b | 0.803 | 0.017 |
| Protein, g/L | 22.75 ^a | 24.11 ^a | 14.12 ^b | 1.463 | <0.001 | 28.32 ^a | 29.49 ^a | 26.28 ^b | 0.555 | <0.001 |
| Triglyceride, mg/dL | 14.51 ^a | 14.06 ^a | 8.55 ^b | 1.402 | 0.008 | 22.39 | 21.93 | 23.93 | 1.825 | 0.728 |
| Cholesterol, mg/dL | 88.39 ^a | 79.82 ^a | 55.43 ^b | 5.507 | <0.001 | 133 | 134 | 126 | 4.1 | 0.304 |
| Creatinine, mg/dL | 0.183 | 0.197 | 0.153 | 0.0163 | 0.175 | 0.259 ^a | 0.224 ^b | 0.221 ^b | 0.0111 | 0.044 |
| Uric acid, mg/dL | 1.71 | 1.64 | 1.17 | 0.187 | 0.094 | 2.24 | 1.96 | 1.94 | 0.158 | 0.365 |
| Mg, mg/dL | 2.32 ^a | 2.06 ^a | 1.32 ^b | 0.114 | <0.001 | 2.52 ^{ab} | 2.74 ^a | 2.43 ^b | 0.089 | 0.051 |
| Ca, mg/dL | 7.80 ^a | 7.55 ^a | 4.92 ^b | 0.415 | <0.001 | 8.33 | 8.66 | 8.22 | 0.169 | 0.187 |
| P, mg/dL | 6.53 ^a | 5.85 ^a | 4.06 ^b | 0.323 | <0.001 | 7.49 ^a | 7.50 ^a | 6.61 ^b | 0.171 | <0.001 |

^{a,b} The means in the same row within a strain with different superscripts differ significantly ($p < 0.05$). ¹ Diets: control: corn–soybean-based diet; SPR: the soybean in the control diet was partially replaced with sunflower meal, brewer’s dried grain, and wheat middlings; SPR + BSF: black soldier fly dried larvae were added to the SPR diet.

² Slaughter age was 55 and 40 d for the local and commercial strains, respectively. ³ SEM: standard error of means.

⁴ Alanine aminotransferase (ALT), aspartate aminotransferase (AST), and gamma-glutamyl transferase (GGT).

The blood Mg and P levels of the birds from both strains who had been fed the SPR + BSF diet were reduced in the birds who had been fed the SPR + BSF diet. The lowest P level was observed for the birds who had been fed the SPR + BSF diet (Table 8). The blood Ca levels of the local chickens were affected by the diets, and the SPR + BSF diet reduced the Ca levels, but this effect was not significant in the Cobb chickens.

3.3. Welfare Traits of the Birds

The effect of the diet on tonic immobility duration and induction numbers for each strain is presented in Table 9. The diets did not affect tonic immobility duration and the number of inductions in either strain.

Table 9. The effect of the diets ¹ on tonic immobility (TI) test responses of local and commercial strains on day 28 and at the slaughter age ².

| | | Local Strain | | | | | Commercial Strain | | | | |
|---------------|------------------|--------------|-------|-----------|------------------|-----------------|-------------------|-------|-----------|-------|-----------------|
| | | Control | SPR | SPR + BSF | SEM ³ | <i>p</i> -Value | Control | SPR | SPR + BSF | SEM | <i>p</i> -Value |
| D28 | TI duration (s) | 68.50 | 52.83 | 63.08 | 12.7 | 0.594 | 81.67 | 89.67 | 98.83 | 9.20 | 0.375 |
| | Induction number | 2.17 | 1.83 | 1.58 | 0.26 | 0.249 | 1.08 | 1.33 | 1.42 | 0.15 | 0.346 |
| Slaughter age | TI duration (s) | 79.25 | 81.08 | 79.08 | 12.51 | 0.977 | 104.08 | 97.00 | 87.67 | 10.57 | 0.441 |
| | Induction number | 2.08 | 1.83 | 1.67 | 0.24 | 0.489 | 1.42 | 1.17 | 1.17 | 0.15 | 0.738 |

¹ Diets: control: corn–soybean-based diet; SPR: the soybean in the control diet was partially replaced with sunflower meal, brewers' dried grain, and wheat middlings; SPR + BSF: black soldier fly dried larvae were added to the SPR diet. ² Slaughter age was 55 and 40 d for the local and commercial strains, respectively. ³ SEM: standard error of means.

The severity of footpad dermatitis was independent of the diets for both strains ($p > 0.05$). The distribution of the footpad scores of the chickens at the slaughter age is presented in Table 10. The percentage of birds with a score of "0" was 68.95% in the local strain and 78.48% in the Cobb one. The percentage of mildly affected birds was 19.35% in the local strain and ranged between 29.17 and 39.58% within the dietary groups. The percentage of birds with a score of "2" was 11.69%. For the commercial strain, footpad dermatitis incidence was 21.52% and ranged between 27.08 and 43.75% within the diets.

Table 10. Distribution (%) of the footpad dermatitis scores of chickens from local and commercial strains by the diets ¹ at slaughter age ² (observed numbers are given in parenthesis).

| Diets | Footpad Dermatitis (%) | | | | |
|-----------|------------------------|-----------------------|-----------------------|-----------------------|-----------------------|
| | Local Strain | | | Commercial Strain | |
| Control | 32.75 ($n = 56$) | 29.17 ($n = 14$) | 37.93 ($n = 11$) | 32.57 ($n = 57$) | 43.75 ($n = 21$) |
| | Cell $\chi^2 = 0.000$ | Cell $\chi^2 = 0.179$ | Cell $\chi^2 = 0.247$ | Cell $\chi^2 = 0.289$ | Cell $\chi^2 = 1.056$ |
| SPR | 29.82 ($n = 51$) | 39.58 ($n = 19$) | 48.28 ($n = 14$) | 34.86 ($n = 61$) | 27.08 ($n = 13$) |
| | Cell $\chi^2 = 0.826$ | Cell $\chi^2 = 0.462$ | Cell $\chi^2 = 1.176$ | Cell $\chi^2 = 0.148$ | Cell $\chi^2 = 0.53$ |
| SPR + BSF | 37.43 ($n = 64$) | 31.25 ($n = 15$) | 13.75 ($n = 4$) | 32.57 ($n = 57$) | 29.17 ($n = 14$) |
| | Cell $\chi^2 = 0.800$ | Cell $\chi^2 = 0.070$ | Cell $\chi^2 = 3.354$ | Cell $\chi^2 = 0.029$ | Cell $\chi^2 = 0.107$ |

¹ Diets: control: corn–soybean-based diet; SPR: the soybean in the control diet was partially replaced with sunflower meal, brewers' dried grain, and wheat middlings; SPR + BSF: black soldier fly dried larvae were added to the SPR diet. ² Slaughter age was 55 and 40 d for the local and commercial strains, respectively.

The distribution of the feather cleanliness scores among the diets for each strain is presented in Table 11.

Table 11. Distribution (%) of the feather cleanliness scores of chickens from local and commercial strains by the diets ¹ at slaughter age ² (observed numbers are given in parenthesis).

| Diets | Local Strain | | Commercial | |
|--|-----------------------|-----------------------|--|-----------------------|
| | 1 (Slight Soiling) | 2 (Moderate Soiling) | 0 (Clean Plumage) | 1 (Slight Soiling) |
| Control | 35.48 ($n = 66$) | 23.33 ($n = 14$) | 32.61 ($n = 15$) | 35.06 ($n = 61$) |
| | Cell $\chi^2 = 0.502$ | Cell $\chi^2 = 1.557$ | Cell $\chi^2 = 0.049$ | Cell $\chi^2 = 0.013$ |
| SPR | 32.80 ($n = 61$) | 38.33 ($n = 23$) | 13.04 ($n = 6$) | 38.51 ($n = 67$) |
| | Cell $\chi^2 = 0.099$ | Cell $\chi^2 = 0.308$ | Cell $\chi^2 = 5.622$ | Cell $\chi^2 = 1.486$ |
| SPR + BSF | 31.72 ($n = 59$) | 38.33 ($n = 23$) | 54.35 ($n = 25$) | 26.44 ($n = 46$) |
| | Cell $\chi^2 = 0.145$ | Cell $\chi^2 = 0.450$ | Cell $\chi^2 = 6.946$ | Cell $\chi^2 = 1.836$ |
| Total (%) | 75.61 ($n = 186$) | 24.39 ($n = 60$) | 20.9 ($n = 46$) | 79.09 ($n = 174$) |
| Pearson $\chi^2 = 3.062$, $p = 0.216$ | | | Pearson $\chi^2 = 15.954$, $p = 0.0003$ | |

¹ Diets: control: corn–soybean-based diet; SPR: the soybean in the control diet was partially replaced with sunflower meal, brewer's dried grain, and wheat middlings; SPR + BSF: black soldier fly dried larvae were added to the SPR diet. ² Slaughter age was 55 and 40 d for the local and commercial strains, respectively.

In the local strain, there were no birds who scored “0” (clean) or “3” (severe dirtiness). All the birds showed slight (score 1) or moderate (score 2) discoloration and, thus, impaired cleanliness at the slaughter age. Of the birds examined, 75.61% had a score of “1”, which indicated slight discoloration, while the percentage of birds with moderate soiling (score “2”) was 24.39%. A chi-square analysis revealed that the feather cleanliness scores of the birds did not depend on the diets in the local strain. However, in the commercial strain, the feather cleanliness scores were dependent on the diet ($\chi^2 = 15.954$; $p = 0.0003$; df: 2). The percentage of clean birds at the slaughter age for the commercial strain was 20.9%. The lowest (%13.4) and the highest (54.35%) number of birds with a score of “0” were found in the birds who had been fed the SPR and SPR + BSF diets, respectively. The higher cell χ^2 values of the SPR and SPR + BSF diets by score “0” combinations revealed that the SPR diet reduced the number of clean birds, which was indicated by a greater adjusted standardized residual value (> -2). However, higher than expected numbers of clean birds were found in those who had been fed the SPR + BSF diet, with the highest contribution to the total χ^2 and a higher adjusted standardized residual (> 2).

3.4. Economic Performance

Tables 12 and 13 display the statistical analysis and mean values of the GPM, CBR, and RoI estimations by diet and strain.

Table 12. Analyses of variance (significance of p -values) for economic indicators by diet and strain.

| | Strain | Diet | Strain \times Diet |
|-----------------------|--------|--------|----------------------|
| Cost of feed consumed | <0.001 | <0.001 | 0.953 |
| Total selling price | <0.001 | 0.408 | 0.219 |
| Gross profit margin | <0.001 | <0.001 | 0.185 |
| Cost–benefit ratio | 0.001 | <0.001 | 0.675 |
| Return on investment | 0.001 | <0.001 | 0.675 |

Table 13. Effects of the diets ¹ on the economic indicators ² of production by strains.

| Economic Indicators | Local Strain | | | | | Commercial Strain | | | | |
|---------------------------------------|--------------------|--------------------|-------------------|-------|--------|--------------------|--------------------|-------------------|-------|--------|
| | Control | SPR | SPR + BSF | SEM | p | Control | SPR | SPR + BSF | SEM | p |
| Cost of feed (EUR /kg) | 0.4460 | 0.4474 | 0.5610 | | | 0.4519 | 0.4532 | 0.5666 | | |
| CF | 2.30 ^b | 2.33 ^b | 2.87 ^a | 0.043 | <0.001 | 1.70 ^b | 1.74 ^b | 2.25 ^a | 0.043 | <0.001 |
| Selling price (EUR per kg of carcass) | 2.83 | 2.83 | 2.83 | | | 2.26 | 2.26 | 2.26 | | |
| SP | 4.65 | 4.71 | 4.62 | 0.046 | 0.345 | 3.69 | 3.75 | 3.79 | 0.036 | 0.207 |
| GPM | 2.35 ^a | 2.38 ^a | 1.74 ^b | 0.032 | <0.001 | 1.99 ^a | 2.01 ^a | 1.54 | 0.060 | <0.001 |
| CBR | 2.03 ^a | 2.02 ^a | 1.61 ^b | 0.026 | <0.001 | 2.17 ^a | 2.16 ^a | 1.68 ^b | 0.054 | <0.001 |
| RoI | 102.7 ^a | 102.2 ^a | 60.8 ^b | 2.62 | <0.001 | 117.5 ^a | 115.7 ^a | 68.4 ^b | 5.43 | <0.001 |

^{a,b} The means in the same row within a strain with different superscripts differ significantly ($p < 0.05$). ¹ Diets: control: corn–soybean-based diet; SPR: the soybean in the control diet was partially replaced with sunflower meal, brewer’s dried grain, and wheat middlings; SPR + BSF: black soldier fly dried larvae were added to the SPR diet. ² CF: cost of feed consumed (euro/bird); SP: Total selling price of carcass (euro/bird); GPM: gross profit margin; CBR: cost–benefit ratio; and RoI: return on investment.

While the cost of the diet was about the same for the control and SPR diets, the SPR + BSF diet was 25.3% more expensive than the control diet (Table 13).

The cost of the feed consumed ranged from 2.30 to 2.87 euro/bird and from 1.70 to 2.25 euro/bird for the local and commercial strains, respectively. The total cost of the feed

consumed for the control, SPR, and SPR + BSF diets was 26.0, 25.3, and 21.3% lower in the commercial strain compared to the local strain. The GPM, CBR, and RoI values achieved for the birds who had been fed the control and SPR diets were similar, higher than those of the birds who had been fed the SPR + BSF larvae diet (Table 13). Despite the current price of BSF larvae being too high to be an economically competitive ingredient, the observed CBR for the SPR + BSF diet appears to be higher than one. On the other hand, all the economic indicators (GPM, CPR, and RoI) are deteriorated significantly by the addition of 5% BSF larvae in the SPR diet.

4. Discussion

Soybeans are the main source of protein in broiler diets. However, soybean cultivation depends on extensive land use, leading to deforestation in Brazil and Argentina [40,41]. This has stimulated interest in nonconventional feedstuffs, such as agro-industrial by-products and BSF larvae, that offer an alternative source of protein in broiler nutrition and an opportunity for environmental sustainability. High-protein sunflower meal, brewers' dried grain, and wheat middlings are valuable protein sources for broiler diets with an average protein content of 35, 20, and 18%, respectively [12,42,43]. BSF larvae, rich in protein (37–63%), are a promising feedstuff for replacing soybeans in broiler diets and do not compete with human nutrition [16]. Although it has been shown that agro-industrial by-products and BSF larvae can be used in broiler diets without affecting broiler performance, to our knowledge, there are no studies using a combination of them. Therefore, this study aimed to investigate the effect of diets in which soybean meals had been partially replaced with sunflower meal, brewers' dried grain, wheat middlings, and BSF larvae meal on the performance, blood parameters, and welfare traits of commercial and local broiler chickens and evaluate the economic impacts of using alternative feedstuffs.

4.1. Performance of the Birds

In the present study, the body weights of the day-old chicks were similar among the groups, indicating that the chicks were distributed randomly. The average slaughter weight and feed conversion ratio were 2247 and 2.33 for the local and 2196 g 1.77 for the commercial chickens at 55 and 40 d of age, respectively. The results obtained for the Cobb chickens were lower than the body weight and the feed values specified for Cobb chickens at 40 days [43]. This was because they were fed diets containing less protein than the dietary protein values recommended for Cobb500 (21–22, 19–20, 18–19, and 17–18% of protein for starter, grower 1, grower 2, and finisher diets) [44].

Considering the growth and feed conversion ratio, brewers' dried grain can be included in a diet in concentrations by up to 20% [7,14,45]. Ramo Rao et al. [4] found that the replacement of soybean with sunflower meal up to 67% in the starter and 100% in the finisher diets increased feed consumption and discouraged feed efficiency. On the other hand, the replacement of soybean meal with high-protein sunflower meal by up to 50% using multi-enzyme mixtures did not affect the growth performance of broilers [9]. In our study, the SPR diet did not affect the body weight, feed consumption, and feed conversion rate of broilers. This result may be related to the use of enzymes and the lower rates of soybean replacement in our study.

The effect of BSF larvae meal inclusion in the broiler diets on body weight, feed consumption, and feed conversion may vary due to the level of inclusion [24,46,47], larvae form (live, dried, full-fat, or de-fatted) [46,48,49], and BSF larvae composition, which can be modified by the diet of the larvae [17,50]. The soybean-based diets replaced with BSF larvae by up to 50 and 100% reduced the body weight of Ross broilers [24]; thus, low levels would be more suitable [21]. The higher body weight obtained on day 10 in the chicks who had been fed the SPR + BSF diet showed that a 5% substitution of soybean with BSF larvae meal the addition of 5% BSF larval meal to the SPR diet improved the early growth performance of chicks from both strains. This result was similar to the findings reported by Dabbou et al. [21]. It is possible to attribute this result to two reasons, as follows: (1) The occurrence

of initial colonization of the intestinal microbiota has an impact on growth performance and daily gain [51]. BSF larvae meal may have affected the weight gain in this period by contributing to the formation of intestinal microbiota in the chicks. (2) The increase in body weight was accompanied by the increased feed consumption and appetite of the chicks during the same period. Naturally, chickens like to consume larvae [52]. Heuel et al. [53] reported that diets containing BSF larvae meal were slightly superior in palatability to a diet including soybean meal, which may increase feed consumption. However, this effect disappeared at later ages; birds from different dietary groups measured similar body weights at day 25 and at the slaughter age. An impaired feed conversion was obtained for the commercial birds consuming the SPR + BSF diet from day 11 to day 25; however, the diets did not affect the final feed conversion of the local and commercial birds. These results showed that SPR could be used as an alternative diet to soybean-based diets without affecting broiler performance and that adding 5% of BSF to the SPR diet would not affect growth performance and the feed conversion ratio.

A concentration of Brewers' dried grain up to 20% increased the eviscerated yield but reduced the drumstick yield [13]. Araujo et al. [54] reported that the inclusion of sunflower meal with enzymes in concentrations up to 24% in broiler diets from day 21 to day 42 reduced carcass parameters. Murawska et al. [24] noted that replacing soybeans with full-fat BSF larvae by up to 100% reduced breast yield. Since soybeans were substituted with three feedstuffs in our study, it is difficult to directly compare them with these studies. In the present study, although the breast yield of the local chickens increased with the SPR and SPR + BSF diets, this effect was not significant for the commercial broilers. This difference between the response of breast yield to the diets in the commercial and the local strain may be related to the chickens' growth rate and carcass composition.

4.2. Blood Parameters and Organ Weights

Bongiorno et al. [48] reported that live BSF larvae increased the weights of the spleen and bursa of Fabricius and reduced GGT activity in the blood indicating positive effects on liver health and the immune system. Schiavone et al. [55] also showed that BSF larvae fat would not affect the liver health of commercial broilers, confirming its nutritional adequacy. The liver is a multi-purpose supportive organ of the digestive system and plays a role in digestion, with an involvement in lipid, carbohydrate, and protein metabolism, and glucose homeostasis to meet energy demands [56]. Glycolysis is the main pathway for glucose catabolism, which depends on the activities of enzymes [57]. The lower liver and blood glucose levels obtained in the local broilers who had been fed the SPR + BSF diet indicated that glycolysis was inhibited. Similarly, Chen et al. [58] reported that the glucose metabolism changed in shrimps who had been fed BSF larvae meal. However, this effect was not observed in the commercial broilers in our study. The present study showed that the blood parameters of the two strains differed in their response to the diets: i.e., the diets did not affect the blood glucose, ALT, AST, triglyceride, cholesterol, and uric acid levels of the commercial chickens. This result might be due to the feeding period, which was longer for the local chickens compared to the commercial chickens, and/or their different growth rates and genetic backgrounds.

In the literature, the results on the effects of by-products and BSF larvae on the blood profile of chickens are inconsistent and depend on the supplementation level. Parpinelli et al. [59] indicated that the inclusion of up to 8% brewers' dried grain did not affect the triglyceride, uric acid, and creatinine levels; however, it increased the ALT and AST levels of broilers. An increase in the blood ALT and AST levels due to the inclusion of BSF larvae meal was also reported in [47,60]. In contrast to these findings, Dabbou et al. [49] and Attia et al. [61] reported that a soybean-based diet partially substituted with BSF larvae did not affect the serum ALT, AST, triglycerides, or uric acid levels. ALT and AST are specific indicators of liver health. The increased blood ALT levels of the local chickens who had been fed the SPR diet indicated liver cell damage. However, this situation was thought to be tolerable because the mortality rates were very low for the dietary groups. On the

other hand, the decreased blood ALT levels with the inclusion of BSF in the SPR diets of local chickens may indicate a normal liver metabolism, which was accompanied by lower GGT levels. These results indicated that adding BSF to the SPR diet protected the liver of the local chickens. The blood cholesterol and triglyceride levels also decreased in the local chickens, agreeing with Bongiorno et al. [48]. This result may be related to the level of chitin in the BSF diet, which reduces lipid absorption in the intestine by binding lipids or fatty acids [61]. In our study, the chitin level was found to be 8.32% and was consistent with Marono et al. [36]. On the other hand, the creatinine levels of local chickens were not influenced by the diets, showing that the diets did not significantly affect renal functions, while the creatinine levels in the commercial chickens decreased in the diets implementing alternative feedstuffs.

There are conflicting results related to the effect of insect larvae on blood minerals. Loponte et al. [62] found no effect of BSF larvae on blood minerals when 25 and 50% of soybean meal were substituted with larvae protein. Gariglio et al. [25] reported that the inclusion of 0 to 9% of partially de-fatted BSF larvae meal in Muscovy ducks' diets did not affect the levels of blood Ca and P, while the blood Mg levels decreased linearly. In our study, the blood Mg and P levels decreased with BSF larvae meal inclusion in both strains, which agreed with Bovera et al.'s study [63]. The response of the strains to the BSF larvae meal-supplemented diet differed in the blood Ca levels; the BSF larvae diet reduced the blood Ca content of the local chickens. In contrast to this finding, Marono et al. [64] reported higher blood Ca levels in laying hens who had been fed BSL larvae meal.

4.3. Welfare Traits of the Birds

In this study, tonic immobility test responses, footpad dermatitis, and plumage cleanliness were measured to evaluate the effect of SPR and SPR + BSF diets on the welfare of broilers from local and commercial strains. There was not any significant effect on the tonic immobility responses. Ipema et al. [65] reported a reduced tonic immobility duration and improved activity, but no further improvement in health-related welfare traits, such as lameness and feather cleanliness, were observed when live BSF larvae were administered with a replacement level of either 5% or 10% of the dietary intake.

The plumage cleanliness score was the only welfare measure that was significantly affected by the diet in the commercial broilers. Our results revealed that the SPR diet had a negative impact on the plumage score in the commercial strain. However, the inclusion of BSF larvae meal into the SPR diet seemed to alleviate this negative effect. Footpad health and other welfare indicators including plumage cleanliness, breast irritation, hock burn, and gait were negatively related to litter quality [66]. The positive effect of BSF inclusion into the diet might have been related to the health of the digestive system and microbiota composition, which directly affect litter quality. Indeed, BSF larvae meal utilization has been shown to positively influence cecal microbiota and the gut's mucin dynamics, as indicated by an increased level of beneficial bacterial population, particularly lactic acid, and an increase in villi mucins [67,68]. Therefore, we may speculate that the inclusion of BSF larvae at a concentration of 5% into the SPR diet accounted for better plumage conditions. Another possible explanation for the improved cleanliness score in the broilers who had been fed the SPR + BSF diet could be the reduced serum Mg and P levels of the birds from both strains. This may modify the moisture content of the excreta and, thus, plumage cleanliness, as increasing dietary Mg levels have been reported to increase excreta moisture in broilers [69]. The cleanliness of the local broilers was not influenced by the SPR and SPR + BSF treatments. However, 100% of the birds presented either slight or moderate soiling, and this might be associated to the overall higher moisture content of the excreta from the local strain. Indeed, excreta dry matter was affected by strain, being higher in the Cobb chickens ($30.04 \pm 1.35\%$) than in the local ($25.99 \pm 0.89\%$) strain in this study (authors' unpublished data).

In our study, the footpad dermatitis scores were not affected by diet. Contrary to our findings, positive effects on the footpad health and hock burn conditions of broilers have

been reported after the inclusion of BSF larvae meal into the diet or the administration of live larvae separately from the diet [65]. The differences between the two studies might be related to the inclusion level of BSF larvae, which was 5% of the diet in our case and 8% of the diet's dry matter in their study. The differences in the inclusion levels of BSF larvae might have modified the magnitude of their effect on the footpad dermatitis scores of the broilers from both strains.

4.4. Economic Performance

The economic performance was assessed utilizing key indicators such as GPM, CBR, and RoI. Beyond chicken performance, the economy of production forms the basis for the inclusion of alternative feedstuffs as a protein source into chicken diets to partly substitute soybean meal in the sector. The price of feedstuffs has a great impact on diet cost and on the profitability of production. The findings of our economic analysis indicate that the SPR diet had similar economic results to the control diet. However, adding BSF larvae meal into the SPR diet resulted in a negative economic effect on chicken meat production. Nevertheless, the CBR of this particular diet demonstrated a relatively lower value compared to both the control and SPR diets. Contrary to our findings, Onsongo et al. [38] reported that the inclusion of BSF larvae meal as a substitute for soybean meal in chicken diets resulted in 16 and 25% higher CBR and RoI, respectively, compared to the control diet, the unit price of which was 19% more expensive. This effect was observed when the BSF larvae meal constituted 42.0% and 55.5% of the crude protein in the starter and finisher diets, respectively. The controversial state of the findings in our research can be attributed to the high prices for BSF in Türkiye, due to the low availability of this resource and, therefore, the small scale of production. The cost of the larvae also surged in our study due to the usage of dried larvae. Using live larvae, produced within one's own company, will likely reduce the costs. Another issue is that the cost of production with a local strain is high, which is an expected result. Increasing profitability can be achieved by modifying the selling prices of local products.

5. Conclusions

The availability of alternative protein sources that can partly replace soybean at acceptable prices is important for sustainable poultry meat production. The findings of this study indicated that the SPR diet, in which soybean meal was partially substituted with agro-industrial by-products, met the nutritional requirements of chickens without negatively affecting the slaughter weight and feed conversion of both local and commercial chickens. The incorporation of BSF larvae meal into the SPR diet improved the growth of the chicken during the first 10 d post hatching. Furthermore, the SPR + BSF diet seemed to alleviate the negative impact of the SPR diet on the plumage score of the commercial broilers at the slaughter age. The local strain, which had a longer production period of 55 d, exhibited compromised cleanliness scores and plumage conditions across all individuals, irrespective of their diet. This response may be related to differences in the diet response and gut health between strains, as gut health is associated with plumage cleanliness. Future research should explore how the microbiota is affected in chickens who are fed SPR and SPR + BSF diets. On the other hand, the economic analysis in this study showed that the SPR + BSF diet had a negative impact on the GPM, CBS, and RoI. The profitability of broiler production largely depends on the price of BSF larvae meal. In future studies, economic sustainability will be better assessed by making cost estimates under various price scenarios.

Author Contributions: M.C.A.: experimental design, carrying out the experiment, and writing the draft; B.T.: economic analysis, review, and editing; Ö.K.U.: economic analysis, review, and editing; S.Ö.: experimental design, carrying out the experiment, data analysis, review, and editing; S.Y.: experimental design, data analysis, review and editing, project administration, and funding acquisition. All authors have read and agreed to the published version of the manuscript.

Funding: This study was funded by the PRIMA Foundation (SUSTAvianFEED, Grant number 2015).

Institutional Review Board Statement: All the procedures were approved by the Farm Animal Experiments Local Ethics Committee of the Faculty of Agriculture at Ege University (Approval no:2022/002, 7 April 2022).

Informed Consent Statement: Not applicable.

Data Availability Statement: The datasets of the current study and the models used are available from the corresponding author upon reasonable request.

Acknowledgments: We are grateful to Nagehan Nur Altan and Enes Hoş for helping with data collection.

Conflicts of Interest: The authors declare no conflicts of interest.

References

1. Font-i-Furnols, M. Meat consumption, sustainability, and alternatives: An overview of motives and barriers. *Foods* **2023**, *12*, 2144. [\[CrossRef\]](#) [\[PubMed\]](#)
2. Van Huis, A.; Oonincx, D.G.A.B. The environmental sustainability of insects as food and feed. a review. *Agron. Sustain. Dev.* **2017**, *37*, 43. [\[CrossRef\]](#)
3. Georganas, A.; Giamouri, E.; Pappas, A.C.; Zoidis, E.; Goliomytis, M.; Simitzis, P. Utilization of agro-industrial by-products for sustainable poultry production. *Sustainability* **2023**, *15*, 3679. [\[CrossRef\]](#)
4. Rama Rao, S.V.; Raju, M.V.L.N.; Panda, A.K.; Reddy, M.R. Sunflower seed meal as a substitute for soybean meal in commercial broiler chicken diets. *Br. Poult. Sci.* **2006**, *47*, 592–598. [\[CrossRef\]](#)
5. Adedokun, S.A.; Jaynes, P.; Payne, R.L.; Applegate, T.J. Standardized ileal amino acid digestibility of corn, corn distillers' dried grains with solubles, wheat middlings, and bakery by-products in broilers and laying hens. *Poult. Sci.* **2015**, *94*, 2480–2487. [\[CrossRef\]](#)
6. Adams, C.B.; Souza, O.; Agilar, J.C.; Muller, G.; Rodrigues, B.; Stefanello, C. Energy values of brewer's grains and olive pomace waste for broiler chickens determined using the regression method. *Agriculture* **2022**, *12*, 444. [\[CrossRef\]](#)
7. Denstadli, V.; Balance, S.; Knutsen, S.H.; Westereng, B.; Svihus, B. Influence of graded levels of brewers dried grains on pellet quality and performance in broiler chickens. *Poult. Sci.* **2010**, *89*, 2640–2645. [\[CrossRef\]](#)
8. Kocher, A.; Choct, M.; Porter, M.D.; Broz, J. The effects of enzyme addition to broiler diets containing high concentrations of canola and sunflower meal. *Poult. Sci.* **2000**, *79*, 1767–1774. [\[CrossRef\]](#)
9. Waititu, S.M.; Sanjayan, N.; Hossain, M.M.; Leterme, P.; Nyachoti, C.M. Improvement of the nutritional value of high-protein sunflower meal for broiler chickens using multi-enzyme mixtures. *Poult. Sci.* **2018**, *97*, 1245–1252. [\[CrossRef\]](#)
10. Moghaddam, H.N.; Salari, S.; Arshami, J.; Golian, A.; Maleki, M. Evaluation of the nutritional value of sunflower meal and its effect on performance, digestive enzyme activity, organ weight, and histological alterations of the intestinal villi of broiler chickens. *J. Appl. Poult. Res.* **2012**, *21*, 293–304. [\[CrossRef\]](#)
11. Salari, S.; Moghaddam, H.N.; Arshami, J.; Golian, A. Nutritional evaluation of full-fat sunflower seed for broiler chickens. *Asian-Aust. J. Anim. Sci.* **2009**, *22*, 557–564. [\[CrossRef\]](#)
12. Mussatto, S.I.; Dragone, G.; Roberto, I. Brewers' spent grain: Generation, characteristics and potential applications. *J. Cereal Sci.* **2006**, *43*, 1–14. [\[CrossRef\]](#)
13. Swain, B.J.; Naik, P.K.; Chakurkar, E.; Singh, N.P. Effect of feeding brewers' dried grain on the performance and carcass characteristics of Vanaraja chicks. *J. Appl. Anim. Res.* **2012**, *40*, 163–166. [\[CrossRef\]](#)
14. Ashour, E.A.; Abd El-Hack, M.E.; El-Hindawy, M.M.; Attia, A.I.; Osman, A.O.; Swelum, A.A.; Alowaime, A.N.; Saadeldin, I.M.; Laudadio, V. Impacts of dietary inclusion of dried brewers' grains on growth, carcass traits, meat quality, nutrient digestibility and blood biochemical indices of broilers. *S. Afr. J. Anim. Sci.* **2019**, *49*, 573–574. [\[CrossRef\]](#)
15. Ahmadi, H.; Karimov, T. A study on wheat middlings usage on broiler performance. *Aust. J. Basic Appl. Sci.* **2010**, *4*, 5642–5648.
16. Barragan Fonseca, K.B.; Dicke, M.; van Loon, J.J.A. Nutritional value of the black soldier fly (*Hermetia illucens* L.) and its suitability as animal feed. *J. Insects Food Feed.* **2017**, *3*, 105–120. [\[CrossRef\]](#)
17. Meneguz, M.; Schiavone, A.; Gai, F.; Dama, A.; Lussiana, C.; Renna, M.; Gasco, L. Effect of rearing substrate on growth performance, waste reduction efficiency and chemical composition of black soldier fly (*Hermetia illucens*) larvae. *J. Sci. Food Agric.* **2018**, *98*, 5776–5784. [\[CrossRef\]](#)
18. Sogari, G.; Amato, M.; Biasato, I.; Chiesa, S.; Gasco, L. The potential role of insects as feed: A multi-perspective review. *Animals* **2019**, *9*, 119. [\[CrossRef\]](#)
19. Dörper, A.; Veldkamp, T.; Dicke, M. Use of black soldier fly and house fly in feed to promote sustainable poultry production. *J. Insects Food Feed.* **2020**, *7*, 761–780. [\[CrossRef\]](#)

20. Ferronato, N.; Paoli, R.; Romagnoli, F.; Tettamanti, G.; Bruno, D.; Torretta, V. Environmental impact scenarios of organic fraction municipal solid waste treatment with black soldier fly larvae based on a life cycle assessment. *Environ. Sci. Pollut. Res.* **2023**. [\[CrossRef\]](#)
21. Dabbou, S.; Gai, F.; Biasato, I.; Capucchio, M.T.; Biasibetti, E.; Dezzutto, D.; Meneguz, M.; Plachà, I.; Gasco, L.; Schiavone, A. Black soldier fly defatted meal as a dietary protein source for broiler chickens: Effects on growth performance, blood traits, gut morphology and histological features. *J. Anim. Sci. Biotechnol.* **2018**, *9*, 1–10. [\[CrossRef\]](#) [\[PubMed\]](#)
22. Schiavone, A.; Dabbou, S.; Petracci, M.; Zampiga, M.; Sirri, F.; Biasato, I.; Gai, F.; Gasco, L. Black soldier fly defatted meal as a dietary protein source for broiler chickens: Effects on carcass traits, breast meat quality and safety. *Animal* **2019**, *13*, 2397–2405. [\[CrossRef\]](#) [\[PubMed\]](#)
23. De Souza Vilela, J.; Andronicos, N.; Kolakshyapati, M.; Hilliar, M.; Sibanda, T.Z.; Andrew, N.; Swick, R.; Wilkinson, S.J.; Ruhnke, I. Black soldier fly larvae in broiler diets improve broiler performance and modulate the immune system. *Anim. Nutr.* **2021**, *7*, 695–706. [\[CrossRef\]](#) [\[PubMed\]](#)
24. Murawska, D.; Daszkiewicz, T.; Sobotka, W.; Gesek, M.; Witkowska, D.; Matusievičius, P.; Bakula, T. Partial and total replacement of soybean meal with full-fat black soldier fly (*Hermetia illucens* L.) larvae meal in broiler chicken diets: Impact on growth performance, carcass quality and meat quality. *Animals* **2021**, *11*, 2715. [\[CrossRef\]](#) [\[PubMed\]](#)
25. Gariglio, M.; Dabbou, S.; Crispo, M.; Biasato, I.; Gai, F.; Gasco, L.; Piacente, F.; Odetti, P.; Bergagna, S.; Plachà, I.; et al. Effects of the dietary inclusion of partially defatted black soldier fly (*Hermetia illucens*) meal on the blood chemistry and tissue (spleen, liver, thymus, and bursa of fabricius) histology of muscovy ducks (*Cairina moschata domestica*). *Animals* **2019**, *9*, 307. [\[CrossRef\]](#) [\[PubMed\]](#)
26. Biasato, I.; Bellezza Oddon, S.; Chemello, G.; Gariglio, M.; Fiorilla, E.; Dabbou, S.; Pipan, M.; Dekleva, D.; Macchi, E.; Gasco, L.; et al. Welfare implications for broiler chickens reared in an insect larvae-enriched environment: Focus on bird behaviour, plumage status, leg health, and excreta corticosterone. *Front. Physiol.* **2022**, *13*, 930158. [\[CrossRef\]](#)
27. Ipema, A.; Bokkers, E.A.M.; Gerrits, W.J.J.; Kemp, B.; Bolhuis, J.E. Long-term access to live black soldier fly larvae (*Hermetia illucens*) stimulates activity and reduces fearfulness of broilers, without affecting health. *Sci. Rep.* **2020**, *10*, 17428. [\[CrossRef\]](#)
28. Erensoy, K.; Sarica, M. Fast-growing broiler production from genetically different pure lines in Turkey. 2. Broiler traits: Growth, feed intake, feed efficiency, livability, body defects and some heterotic effects. *Trop. Anim. Health Prod.* **2023**, *55*, 61. [\[CrossRef\]](#)
29. Archer, G.S. Sex, genetics, and test type affect the responses of chickens to fear testing. *Int. J. Poult. Sci.* **2018**, *17*, 320–326. [\[CrossRef\]](#)
30. Jones, R.B. The tonic immobility reaction of the domestic fowl: A review. *World's Poult. Sci. J.* **1986**, *4*, 82–96. [\[CrossRef\]](#)
31. Welfare Quality®. *Welfare Quality® Assessment Protocol for Poultry (Broilers, Laying Hens)*; Welfare Quality® Consortium: Lelystad, The Netherlands, 2009.
32. Ekstrand, C.; Carpenter, T.E.; Anderson, I.; Algers, B. Prevalence and control of foot-pad dermatitis in broilers in Sweden. *Br. Poult. Sci.* **1998**, *39*, 318–324. [\[CrossRef\]](#) [\[PubMed\]](#)
33. Association of Official Analytical Chemistry (AOAC). *Standard Compendium of Laboratory Methods for Analysing Foods and Related Substances*, 16th ed.; AOAC Publishing: Washington, DC, USA, 1995.
34. Anonymous. *Animal Feeds-Metabolizable Energy Method (Chemical Analyses)*; TS 9610/December; TSE: Ankara, Türkiye, 1991.
35. Van Soest, P.J.; Robertson, J.B.; Lewis, B.A. Methods for dietary fibre, neutral detergent fibre, and non-starch polysaccharides in relation to animal nutrition. *J. Dairy Sci.* **1991**, *74*, 3583–3597. [\[CrossRef\]](#) [\[PubMed\]](#)
36. Marono, S.; Piccolo, G.; Loponte, R.; Di Meo, C.; Attia, Y.A.; Nizza, A.; Bovera, F. In vitro crude protein digestibility of *Tenebrio molitor* and *Hermetia illucens* insect meals and its correlation with chemical composition traits. *Ital. J. Anim. Sci.* **2015**, *14*, 3889. [\[CrossRef\]](#)
37. Folch, J.; Lees, M.; Sloane Stanley, G.H. A simple method for the isolation and purification of total lipids from animal tissues. *J. Biol. Chem.* **1957**, *226*, 497–509. [\[CrossRef\]](#)
38. Onsongo, V.; Osuga, I.; Gachui, C.; Wachira, A.; Miano, D.; Tanga, C.; Ekesi, S.; Nakimbugwe, D.; Fiaboe, K. Insects for income generation through animal feed: Effect of dietary replacement of soybean and fish meal with black soldier fly meal on broiler growth and economic performance. *J. Econ. Entomol.* **2018**, *111*, 1966–1973. [\[CrossRef\]](#)
39. Sharpe, D. Chi-square test is statistically significant: Now what? *Pract. Assess. Res. Eval.* **2015**, *20*, 1–10.
40. Da Silva, V.P., Jr.; van der Werf, H.; Spies, A.; Soares, S.R. Variability in environmental impacts of Brazilian soybean according to crop production and transport scenarios. *J. Environ. Manag.* **2010**, *91*, 1831–1839. [\[CrossRef\]](#)
41. Song, X.P.; Hansen, M.C.; Potapov, P.; Adusei, B.; Pickering, J.; Adami, M.; Lima, A.; Zalles, V.; Stemhan, S.V.; Di Bella, C.M.; et al. Massive soybean expansion in South America since 2000 and implications for conservation. *Nat. Sustain.* **2021**, *4*, 784–792. [\[CrossRef\]](#)
42. Villamide, M.J.; San Juan, L.D. Effect of chemical composition of sunflower seed meal on its true metabolizable energy and amino acid digestibility. *Poult. Sci.* **1998**, *77*, 1884–1892. [\[CrossRef\]](#)
43. McCarthy, A.; O'Callaghan, Y.; Piggott, C.; FitzGerald, R.; O'Brien, N. Brewers' spent grain, bioactivity of phenolic component, its role in animal nutrition and potential for incorporation in functional foods: A review. *Proc. Nutr. Soc.* **2013**, *72*, 117–125. [\[CrossRef\]](#)

44. Cobb500 Performance and Nutrient Supplement. 2022. Available online: www.cobb-vantress.com (accessed on 1 January 2022).
45. Pires Filho, I.C.; Broch, J.; Eyng, C.; Silva, I.M.; Souza, C.; Avila, A.S.; Castilha, L.D.; Cirilo, E.H.; Tesser, G.L.S.; Nunes, R.V. Effects of feeding dried brewers grains to slow-growing broiler chickens. *Livest. Sci.* **2021**, *250*, 104561. [[CrossRef](#)]
46. Cullere, M.; Schiavone, A.; Dabbou, S.; Gasco, L.; Dalle Zotte, A. Meat quality and sensory traits of finisher broiler chickens fed with black soldier fly (*Hermetia illucens* L.) larvae fat as alternative fat source. *Animals* **2019**, *9*, 140. [[CrossRef](#)] [[PubMed](#)]
47. Seyedalmoosavi, M.M.; Mielenz, M.; Görs, S.; Wolf, P.; Daş, G.; Metges, C.C. Effects of increasing levels of whole Black Soldier Fly (*Hermetia illucens*) larvae in broiler rations on acceptance, nutrient and energy intakes and utilization, and growth performance of broilers. *Poult. Sci.* **2022**, *101*, 102202. [[CrossRef](#)] [[PubMed](#)]
48. Bongiorno, V.; Gariglio, M.; Zambotto, V.; Cappone, E.E.; Biasato, I.; Renna, M.; Forte, C.; Coudron, C.; Bergagna, S.; Gai, F.; et al. Black soldier fly larvae used for environmental enrichment purposes: Can they affect the growth, slaughter performance, and blood chemistry of medium-growing chickens? *Front. Vet. Sci.* **2022**, *9*, 1064017. [[CrossRef](#)] [[PubMed](#)]
49. Dabbou, S.; Lauwaerts, A.; Ferrocino, I.; Biasato, I.; Sirri, F.; Zampiga, M.; Bergagna, S.; Pagliasso, G.; Gariglio, M.; Colombino, E.; et al. Modified black soldier fly larva fat in broiler diet: Effects on performance, carcass traits, blood parameters, histomorphological features and gut microbiota. *Animals* **2021**, *11*, 1837. [[CrossRef](#)] [[PubMed](#)]
50. Cullere, M.; Woods, M.J.; van Emmenes, L.; Pieterse, E.; Hoffman, L.C.; Dalle Zotte, A. *Hermetia illucens* larvae reared on different substrates in broiler quail diets: Effect on physicochemical and sensory quality of the quail meat. *Animals* **2019**, *9*, 525. [[CrossRef](#)] [[PubMed](#)]
51. Huang, T.; Han, J.; Liu, Y.; Fei, M.; Du, X.; He, K.; Zhao, A. Dynamic distribution of gut microbiota in posthatching chicks and its relationship with average daily gain. *Poult. Sci.* **2023**, *e102*, 103008. [[CrossRef](#)]
52. Mench, J.A. *Behaviour of Fowl and Other Domesticated Birds*; Jensen, P., Ed.; The Ethology of Domestic Animals; CABI Publishing: Surrey, UK, 2009.
53. Heuel, M.; Sandrock, C.; Leiber, F.; Mathys, A.; Gold, M.; Zurbrüegg, C.; Gangnat, I.D.M.; Kreuzer, M.; Terranova, M. Black soldier fly larvae meal and fat as a replacement for soybeans in organic broiler diets: Effects on performance, body N retention, carcass and meat quality. *Br. Poult. Sci.* **2022**, *63*, 650–661. [[CrossRef](#)]
54. Araújo WAG de Albino, L.F.T.; Rostagno, H.S.; Hannas, M.I.; Pessoa, G.B.S.; Messias, R.K.G.; Lelis, G.R.; Ribeiro, V., Jr. Sunflower meal and enzyme supplementation of the diet of 21- to 42-d-old broilers. *Rev. Bras. De Ciência Avícola* **2014**, *16*, 17–24. [[CrossRef](#)]
55. Schiavone, A.; Dabbou, S.; Marco, M.D.; Cullere, M.; Biasato, I.; Biasibetti, E.; Capucchio, M.T.; Bergagna, S.; Dezzutto, D.; Meneguz, M.; et al. Black soldier fly larva fat inclusion in finisher broiler chicken diet as an alternative fat source. *Animal* **2018**, *12*, 2032–2039. [[CrossRef](#)]
56. Zaefarian, F.; Abdollahi, M.R.; Cowieson, A.; Ravindran, V. Avian liver: The forgotten organ. *Animals* **2019**, *9*, 63. [[CrossRef](#)]
57. Guo, X.; Li, H.; Xu, H.; Woo, S.L.; Dong, H.; Lu, F.; Lange, A.; Wu, C. Glycolysis in the control of blood glucose homeostasis. *Acta Pharm. Sin. B* **2012**, *2*, 358–367. [[CrossRef](#)]
58. Chen, Y.; Chi, S.; Zhang, S.; Dong, X.; Yang, Q.; Liu, H.; Tan, B.; Xie, S. Effect of black soldier fly (*Hermetia illucens*) larvae meal on lipid and glucose metabolism of pacific white shrimp *litopenaeus vannamei*. *Br. J. Nutr.* **2021**, *128*, 1–39. [[CrossRef](#)]
59. Parpinelli, W.; Cella, P.S.; Eyng, C.; Broch, J.; Savaris, V.D.L.; Santos, E.C.; Avila, A.S.; Nunes, R.V. Impact of dried brewers' grains supplementation on performance, metabolism and meat quality of broiler chickens. *S. Afr. J. Anim. Sci.* **2020**, *2020*, 50. [[CrossRef](#)]
60. Hossain, S.M.; Blair, R. Chitin utilisation by broilers and its effect on body composition and blood metabolites. *Br. Poult. Sci.* **2007**, *48*, 33–38. [[CrossRef](#)]
61. Attia, Y.A.; Bovera, F.; Asiry, K.A.; Alqurashi, S.; Alrefaei, M.S. Fish and black soldier fly meals as partial replacements for soybean meal can affect sustainability of productive performance, blood constituents, gut microbiota, and nutrient excretion of broiler chickens. *Animals* **2023**, *13*, 2759. [[CrossRef](#)] [[PubMed](#)]
62. Loponte, R.; Nizza, S.; Bovera, F.; De Riu, N.; Fliegerova, K.; Lombardi, P.; Vassalotti, G.; Mastellone, V.; Nizza, A.; Moniello, G. Growth performance, blood profiles and carcass traits of barberry partridge (*Alectoris barbara*) fed two different insect larvae meals (*Tenebrio molitor* and *Hermetia illucens*). *Res. Vet. Sci.* **2017**, *115*, 183–188. [[CrossRef](#)]
63. Bovera, F.; Loponte, R.; Pero, M.E.; Cutrignelli, M.I.; Calabrò, S.; Musco, N.; Vassalotti, G.; Panettieri, V.; Lombardi, P.; Piccolo, G.; et al. Laying performance, blood profiles, nutrient digestibility and inner organs traits of hens fed an insect meal from *Hermetia illucens* larvae. *Res. Vet. Sci.* **2018**, *120*, 86–93. [[CrossRef](#)]
64. Marono, S.; Loponte, R.; Lombardi, P.; Vassalotti, G.; Pero, M.E.; Russo, F.; Gasco, L.; Parisi, G.; Piccolo, G.; Nizza, S.; et al. Productive performance and blood profiles of laying hens fed *Hermetia illucens* larvae meal as total replacement of soybean meal from 24 to 45 weeks of age. *Poult. Sci.* **2017**, *96*, 1783–1790. [[CrossRef](#)]
65. Ipema, A.; Bokkers, E.A.M.; Gerrits, W.J.J.; Kemp, B.; Bolhuis, J.E. Provision of black soldier fly larvae (*Hermetia illucens*) in different ways benefits broiler welfare and performance, with superior effects of scattering live larvae. *Physiol. Behav.* **2022**, *257*, 113999. [[CrossRef](#)]
66. De Jong, I.C.; Gunnink, H.; Harn, J.V. Wet litter not only induces footpad dermatitis but also reduces overall welfare, technical performance, and carcass yield in broiler chickens. *J. Appl. Poult. Res.* **2014**, *23*, 51–58. [[CrossRef](#)]
67. Biasato, I.; Ferrocino, I.; Dabbou, S.; Evangelista, R.; Gai, F.; Gasco, L.; Cocolin, L.; Capucchio, M.T.; Schiavone, A. Black soldier fly and gut health in broiler chickens: Insights into the relationship between cecal microbiota and intestinal mucin composition. *J. Anim. Sci. Biotechnol.* **2020**, *11*, 1–12. [[CrossRef](#)] [[PubMed](#)]

-
68. Ndotono, E.W.; Khamis, F.M.; Bargul, J.L.; Tanga, C.M. Insights into the gut microbial communities of broiler chicken fed black soldier fly larvae-desmodium-based meal as a dietary protein source. *Microorganisms* **2022**, *10*, 1351. [[CrossRef](#)] [[PubMed](#)]
 69. Van der Hoeven-Hangoor, E.; van de Linde, I.B.; Paton, N.D.; Verstegen, M.W.A.; Hendriks, W.H. Effect of different magnesium sources on digesta and excreta moisture content and production performance in broiler chickens. *Poult. Sci.* **2013**, *92*, 382–391. [[CrossRef](#)]

Disclaimer/Publisher’s Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.