



Article

Enzymatic Browning in Banana Blossoms and Techniques for Its Reduction

Geerada Kaewjumpol ¹, Surasak Srisamlee ¹, Diane M. Beckles ² and Kietsuda Luengwilai ^{1,3,*}

- ¹ Department of Horticulture, Faculty of Agriculture at Kamphaeng Saen, Kamphaeng Saen Campus, Kasetsart University, Nakhon Pathom 73140, Thailand; geerada20@gmail.com (G.K.); surasak.sri@ku.th (S.S.)
- ² Department of Plant Sciences, Mail Stop 3, University of California, Davis, CA 95616, USA; dmbeckles@ucdavis.edu
- ³ Postharvest Technology Innovation Center, Ministry of Higher Education, Science, Research and Innovation, Bangkok 10400, Thailand
- * Correspondence: kietsuda.l@ku.th

Abstract: Banana blossoms are rich in fiber and nutrients and are a popular plant-based, vegan alternative to fish. However undesirable browning, usually visible at the peduncle cut-end, negatively impacts consumer acceptability of banana blossoms. The aim of this work was to develop safe alternatives to prevent browning in banana blossoms. First, the activities of primary enzymes associated with tissue browning, i.e., polyphenol oxidase (PPO), peroxidase (POD) and phenylalanine ammonia lyase (PAL), were assayed. Our data showed that PPO and POD were the key enzymes responsible for blossom browning as they increased in activity, reaching a maximum at pH 7, as browning developed. In contrast, PAL activity decreased, and total phenolic content did not change as browning progressed, indicating PAL was not induced by cutting and may not be involved in blossom browning. Second, to find antibrowning agents for banana blossoms that can substitute for the use of sodium metabisulfite (SMS), different organic acids of varying concentrations were tested. Among organic acids studied, treatment with 3% (*w/v*) oxalic acid was the most effective method and thus could be a safe substitution for SMS to prevent browning in banana blossoms.

Keywords: banana buds; antibrowning techniques; PPO; POD; PAL



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1. Introduction

Banana blossoms (widely known as flower buds) are purple-skinned bracts that develop at the end of the banana fruit cluster. In Southeast Asia, especially Sri Lanka, Malaysia, Indonesia and the Philippines, banana blossoms are a popular ingredient for cooking, and these tissues are rich in many beneficial nutrients, including fiber; protein; vitamins A, C and E; phosphorus; calcium; iron; magnesium; and antioxidants [1,2]. Banana blossoms are also used as a therapeutic agent to lower the risk of anemia [3], increase milk production of breastfeeding mothers [4] and ameliorate the effects of diabetes [5]. However, there is new interest in banana blossoms because they have a texture similar to cooked fish, and they are set to become widely used in vegan cooking [6]. The current price-point for banana blossoms is 14.00 USD per kilogram as a vegan food [6].

After cutting, an undesirable brown color develops at the cut-end surface of banana blossoms. This reduces the nutritional value of the product and is unacceptable to consumers [7–9]. Three key enzymes, polyphenol oxidase (PPO), peroxidase (POD) and phenylalanine ammonia lyase (PAL), are involved in the browning mechanism in plant tissues [10–13]. PPO (EC 1.10.3.1) is widely distributed in the cytoplasm, while its phenolic substrates are in the vacuoles [10]. When cellular organelles are damaged, e.g., when tissues are cut, PPO mixes with phenolic compounds, catalyzing the oxidation of monophenols into *o*-diphenols. A polymerization reaction then converts *o*-diphenols to quinones, which are the precursors to the visible brown pigment seen at the cut surface of

tissues [10]. POD (EC 1.11.1.7) is a heme-containing enzyme that catalyzes single-electron oxidation of phenolic compounds in the presence of hydrogen peroxide [10]. PAL (EC 4.3.1.5) is the key enzyme in the phenylpropanoid metabolism; it is activated by wounding which leads to the accumulation of phenolic compounds [10]. The relative contribution of these browning enzymes varies according to the species. For example, PAL is the major enzyme responsible for browning in cut lettuce [14], PPO and POD are both important in strawberry [15], and all three enzymes are responsible for browning in peach [16]. In banana, PPO was characterized [17]. However, the relative contribution of PPO, POD and PAL to the browning mechanism in banana blossoms is not known.

Vital factors that limit the rate of the browning reaction include the concentration of related enzymes and phenolic compounds, pH, temperature, and the availability of oxygen to the tissues. Tamarind juice and sodium metabisulfite (SMS) are commonly employed as antibrowning agents to suppress undesirable browning in fresh-cut banana blossoms [18]. However, tamarind juice availability is seasonal and hence unreliable [19] and is thus unsuitable for large-scale production. SMS, while effective, has been restricted for use on most fresh fruit and vegetables in many countries [20]. Therefore, alternative compounds efficient in preventing browning in banana blossoms are required.

Organic acids are considered safe and are widely used as postharvest antibrowning agents. Oxalic acid at 5% (*w/v*) reduced browning in longan fruit stored at 25 °C [21]. Citric acid at 3% (*w/v*) suppressed PPO, POD and PAL activities and delayed browning in longkong fruit [22]. Tartaric, citric and ascorbic acids at 3% (*w/v*) inhibited POD activity and reduced browning in minimally processed yacon tubers after dipping [23]. Malic acid at 1% (*v/v*) suppressed PPO and POD activities and reduced browning of banana skin [24]. Although it has been demonstrated that organic acid treatment significantly reduces browning in a variety of tissues, there are no reports about the use of organic acid treatment to minimize browning in banana blossoms. Here, mechanisms associated with enzymatic browning and the effects of organic acids as antibrowning agents of banana blossoms after cutting were investigated.

2. Materials and Methods

2.1. Chemicals

L-Ascorbic acid and sodium metabisulfite (SMS) were purchased from Ajax Finechem Pty Ltd. (Taren Point, NSW, Australia). Citric acid monohydrate, sodium dihydrogen phosphate monohydrate and disodium hydrogen phosphate dihydrate were purchased from Merck (Darmstadt, Germany). Oxalic acid, boric acid, sodium carbonate and sodium tetraborate were purchased from KemAus (Cherrybrook, NSW, Australia). Malic acid, tartaric acid, guaiacol, L-phenylalanine catechol, polyvinylpyrrolidone (PVP), gallic acid and 6-hydroxy-2,5,7,8-tetramethylchroman-2-carboxylic acid (Trolox) were purchased from Sigma-Aldrich (Shanghai, China). Bovine serum albumin (BSA) was purchased from Sigma-Aldrich (St. Louis, MO, USA), Folin–Ciocalteu phenol reagent was purchased from Merck (Darmstadt, Germany) and 2,2-diphenyl-1-picrylhydrazyl (DPPH) was purchased from Sigma-Aldrich (Steinheim, Germany). All chemicals were of analytical grade.

2.2. Banana Blossom Sampling and Postharvest Treatments

Banana blossoms (*Musa* ABB ‘Kluai Namwa’) were collected from the agricultural fields in Kamphaeng Saen district, Nakhon Pathom Province, Thailand. Blossoms were harvested using a fruit picker tool, transported to the laboratory within 5 h and sorted for uniform size (weight ~500 g). The purple-skinned bracts were removed, and only the white-skinned bracts remained. The peduncle of each banana blossom was recut and air-dried before being stored in polyethylene (PE) bags under partial vacuum. To determine the enzymatic browning that occurs in association with wounding and to exclude browning associated with chilling injury, banana blossoms were kept at ambient temperature and humidity (30 ± 5 °C, 72 ± 7% RH) for 0, 12 and 24 h to observe browning at the cut-end surface. All experiments were carried out with six replicates, and the whole

experiments were repeated three times. Similar results were observed; thus, the data from one experiment are presented.

2.3. Browning Score

Browning score evaluation was adapted from Lichanporn et al. [25] with some modification. Degree of browning was expressed as the average of six replicates, scored as a visual determination of the peduncle cut-end surface color, where 0 = completely white (no browning), 1 = light brown, 3 = brown and 5 = dark brown or black.

2.4. Color Assessment

Colors on cut-end surfaces were assessed using a Chroma Meter (CR-400, Konica Minolta, Tokyo, Japan) calibrated with a white porcelain reference plate. Color parameters were quantified using the CIE system where L^* corresponds to lightness, a^* corresponds to redness ($+a^*$) or green ($-a^*$) and b^* corresponds to yellowness ($+b^*$) or blue ($-b^*$). A lower value of L^* indicates darker brown color, while a higher value of $+a^*$ refers to increased red color [26]. Total color change (ΔE^*) was computed by the formula $\Delta E^* = \sqrt{\Delta L^{*2} + \Delta a^{*2} + \Delta b^{*2}}$, where ΔL^* , Δa^* and Δb^* represent differences in lightness, redness and yellowness, respectively, before and after the specified period of storage duration [27]. Differences in perceivable color were analytically classified as very distinct ($\Delta E > 3$), distinct ($1.5 < \Delta E < 3$) and small difference ($1.5 < \Delta E$) [28].

2.5. Total Phenolic Compounds

Crude extracts were extracted from the peduncle cut-end of banana blossom and analyzed for phenolic content and DPPH scavenging activity. Banana blossom peduncles (24 g) were homogenized in 100 mL of 80% methanol (v/v) and centrifuged at $19,230 \times g$ at 4°C for 10 min. The supernatant was collected and total phenolic content (TPC) was determined by the Folin–Ciocalteu method [29]. A mixture of 0.3 mL of crude extract and 2.25 mL of Folin–Ciocalteu phenol reagent (Folin:DI water; 1:10 v/v) was left for 5 min at $25 \pm 3^\circ\text{C}$. Then, 2.25 mL of 7% sodium carbonate was added to the mixture, which was vortexed and left in the dark at $25 \pm 3^\circ\text{C}$ for 90 min. The reaction was then measured at 725 nm by a spectrophotometer. The standard curve was established by gallic acid equivalent (GAE) and expressed as $\mu\text{g GAE/g FW}$.

2.6. DPPH Radical Scavenging Assay

Radical scavenging activity of the banana blossom peduncle was determined by DPPH assay following the method of Havananda and Luengwilai [29]. A 0.1 mL aliquot of crude extract was mixed with 1.9 mL of 0.004% DPPH solution in methanol, and the mixture was incubated at $25 \pm 3^\circ\text{C}$ under dark condition for 30 min. Absorbance was read at 515 nm. Percentage DPPH inhibition was calculated by Equation (1) [30].

$$\text{DPPH inhibition (\%)} = A_0 - A_1 / A_0 \times 100 \quad (1)$$

where A_0 is absorbance of the control (using methanol instead of crude extracts) and A_1 is absorbance of the reaction mixture.

2.7. Phenylalanine Ammonia Lyase (PAL) Activity Assay

PAL was extracted from the peduncle of banana blossom. One gram of the cut-end peduncle surface was extracted with 10 mL of 0.1 M borate–boric acid buffer (pH 8.8) solution containing 0.2 g of insoluble PVP and then homogenized for 1 min on ice using a Polytron PT2100 (Kinematica, Luzern, Switzerland). The homogenate was filtered through layers of cheesecloth before centrifugation at $12,000 \times g$ for 30 min at 4°C (Model 6500, Kubota Corporation, Tokyo, Japan). The supernatant was collected for PAL activity assay, measured according to Faragher and Chalmers [31] with some modifications. The reaction mixture consisted of 0.1 M borate–boric acid buffer (pH 8.8, 2 mL), 60 mM

L-phenylalanine (0.7 mL) and PAL extract (0.3 mL). The mixture was incubated for 1 h at 30 °C, then the reaction was stopped with 5 N HCl (0.5 mL). During incubation, the reaction of transforming L-phenylalanine into trans-cinnamic acid was catalyzed by PAL activity and assayed by measuring the increase in trans-cinnamic acid at 280 nm using a UV-vis spectrophotometer (Thermo Spectronic, Rochester, NY, USA). The standard curve was created by L-phenylalanine, related to a rise in the amount of trans-cinnamic acid, with PAL activity recorded as mmol trans-cinnamic acid/h/mg protein.

2.8. Polyphenol Oxidase (PPO) and Peroxidase (POD) Activity Assays and Their Optimal pH Condition for Their Activities

A 2.5 g cut-end surface of banana peduncle was homogenized for 1 min at 4 °C in 0.1 M sodium phosphate buffer (pH 7.0, 5 mL) containing 0.2 g of an insoluble PVP. The homogenate was filtered through layers of cheesecloth before centrifugation at 12,000 × g for 10 min at 4 °C. The supernatant was collected for PPO and POD assays following [32,33]. The reaction mixture of PPO activity consisted of 0.1 M sodium phosphate buffer (pH 7.0, 0.4 mL), 0.1 M catechol (0.5 mL) and PPO extract (0.1 mL). Absorbance was measured at 420 nm. The reaction mixture of POD activity contained 0.1 M sodium phosphate buffer (pH 7.0, 0.8 mL), 8 mM guaiacol (0.05 mL), POD extract (0.1 mL) and 24 mM hydrogen peroxide (H₂O₂, 0.05 mL). Absorbance was measured at 470 nm. PPO and POD specific enzyme activities were defined as increase in absorbance per min in 1 mg protein in enzyme of the reaction mixture (A₄₂₀/min/mg protein for PPO activity and A₄₇₀/min/mg protein for POD activity). Protein contents in PAL, PPO and POD extract solutions were analyzed using bovine serum albumin (BSA) as a standard [34].

To determine their optimal pH conditions for enzyme activities, PPO and POD activities were monitored at different pH values ranging from 1 to 7. All procedures followed those described above, except that the reaction buffer, i.e., 0.1 M sodium phosphate buffer (pH 7.0) was replaced by 0.1 M citric acid–0.15 M HCl (pH 1 and 2) or 0.3 M citrate–phosphate buffer (pH 3 to 7).

2.9. Inhibition of PPO and POD Activities Using Different Organic Acids as Inhibitors

The inhibitory effects of different organic acids on PPO and POD activities were tested. Tartaric (TA), citric (CA), malic (MA), oxalic (OA) and ascorbic (AA) acids at 1, 3 and 5% (*w/v*) dissolved in 0.1 M phosphate buffer (pH 7.0) were used. Crude enzymes were extracted following the same procedure described in Section 2.7. The PPO activity assay consisted of 0.1 mL crude enzyme extract, 0.1 mL of 0.1 M sodium phosphate buffer (pH 7.0), 0.5 mL of 0.1 M catechol and 0.3 mL of inhibitor (organic acid), while the POD activity assay employed 0.5 mL of 0.1 M sodium phosphate buffer (pH 7.0), 0.05 mL of 8 mM guaiacol, 0.3 mL of inhibitor, 0.05 mL of 24 mM H₂O₂ and 0.1 mL of crude enzyme extract. The positive control was the reaction mixture without inhibitor, while the negative control was the reaction mixture without crude extract. Percentage inhibitions of PPO and POD activities were calculated using Equation (2) [35].

$$\text{Inhibition of enzyme activity (\%)} = \frac{A_c - A_i}{A_c} \times 100 \quad (2)$$

where A_c is initial PPO or POD activity (without inhibitor) and A_i is PPO or POD activity with inhibitor.

2.10. Antibrowning of Banana Blossoms by Different Organic Acids

The blossoms were harvested and prepared as described in Section 2.2. The peduncle of banana blossom was recut and dipped for 5 min in 5% (*w/v*) TA, 3% (*w/v*) OA or 3% (*w/v*) AA (the optimal concentrations were selected based on preliminary experiments). DI water was used as the negative control, and 1% (*w/v*) SMS and 2.5% (*w/v*) tamarind juice (TJ) were used as positive controls. Samples were dried with a paper towel before being packed into PE bags under partial vacuum and stored at 15 ± 2 °C and 90 ± 5% RH in the dark for 0, 5 and 10 days. These storage conditions, i.e., temperature, humidity, and

duration, were found to be optimal for banana blossom shelf-life based on preliminary testing. Cut-end surfaces of the peduncle were monitored for weight loss and color changes before (0 days) and after storage (5 and 10 days). All experiments were carried out in five replicates and the entire experiment was repeated three times. Similar results were observed; thus, the data from one experiment are presented.

2.11. Determination of Weight Loss Rate

The difference between tissue mass before and after storage, in relation to mass before storage, was calculated and expressed as a percentage of the initial fresh weight.

2.12. Statistical Analysis

All experiments were conducted using a completely randomized design (CRD) with six replicates, unless otherwise stated. Results were analyzed using the SPSS software program version 19 for Windows (SPSS Inc., IBM Company, Chicago, IL, USA). SPSS was used for analysis of variance (ANOVA) and to estimate significant differences between the means, using Tukey's multiple range test. Acceptable significant difference was set at $p < 0.05$. Correlations between enzyme activities, color parameters, phenolic contents and DPPH scavenging activities were also tested by Pearson's correlation coefficient using SPSS software. The principal component analysis (PCA) was analyzed using PAST4.0 software [36].

3. Results

3.1. Browning of Banana Blossom Cut-End Surfaces

Browning scores increased sharply from 1.2, when assayed at harvest, to 4.2, after 12 h storage at 30 °C. Color components (L^* , a^* and b^*) measured by a colorimeter at the cut-end surfaces of the banana blossoms were also determined. At 12 h after harvest, the L^* value significantly decreased from 77.5 to 55.7, while the a^* value increased from -2.7 to 4.2 giving a total color change of $\Delta E^* \sim 21$. In contrast, b^* value was not significantly changed during 24 h. In addition, browning and color components of the cut-end surface after 24 h were not significantly different from those at 12 h (Table 1).

3.2. Total Phenolic Content (TPC) and DPPH

At harvest, banana blossoms had a TPC of 46.5 $\mu\text{g GAE/g FW}$, and this amount did not change after 24 h (Figure 1A). The antioxidant activity of the banana blossoms was evaluated using the DPPH scavenging assay. Similar to TPC, antioxidant activity was not statistically different at 24 h after harvest. DPPH values ranged from 48.3 to 60.9% inhibition (Figure 1A).

3.3. PAL, PPO and POD Activities

The activities of PAL, PPO and POD in the banana blossoms were low after cutting. POD and PPO activities linearly increased from 1.25 to 6.14 and 0.5 to 1.8 unit/min/mg protein, respectively, 12 h after cutting, remaining high after storage for 24 h. In contrast, PAL activity linearly decreased after storage for 24 h from 1.7 to 1.2 mmol trans-cinnamic acid/h/mg protein (Figure 1B).

3.4. TPC, DPPH, PPO, POD and PAL Activities Related to Browning of Banana Blossoms after Cutting

Negative correlations were shown between L^* (indicative of lightness) and PPO activity ($r = -0.957$) and between L^* and POD activity ($r = -0.752$), while positive correlations were found between browning score and PPO activity ($r = 0.954$) and between browning score and POD activity ($r = 0.782$). In contrast, L^* was positively correlated with PAL activity ($r = 0.828$), which was negatively correlated with browning score ($r = -0.840$). Phenolic content and DPPH scavenging activity were stable throughout the 24 h period

after cutting, with a weak relationship found between L* and the browning score, phenolic content and DPPH scavenging activity ($r < 0.54$) (Table 2).

Table 1. Browning of banana blossom cut-end surfaces after cutting. Lightness (L*), redness (a*), yellowness (b*), total color change (ΔE^*) and browning score of the cut-end peduncle surface of banana blossoms stored at 30 °C for 0, 12 and 24 h.

Storage Time (h)		L*	a*	b*	ΔE^*	Browning Score
0		75.5 ± 0.6 a	-2.7 ± 0.2 b	20.9 ± 1.7 NS	-	1.2 ± 0.4 b
12		55.7 ± 8.6 b	4.2 ± 3.2 a	22.9 ± 1.8 NS	21.2 ± 2.8 NS	4.2 ± 0.8 a
24		55.9 ± 10.1 b	4.8 ± 3.5 a	23.1 ± 1.1 NS	21.6 ± 3.2 NS	4.3 ± 0.6 a

Data are means ± SD ($n = 6$). Means followed by different letters in the same columns indicate significant differences ($p < 0.05$) between storage times. NS indicates nonsignificant differences ($p \geq 0.05$) between storage times.

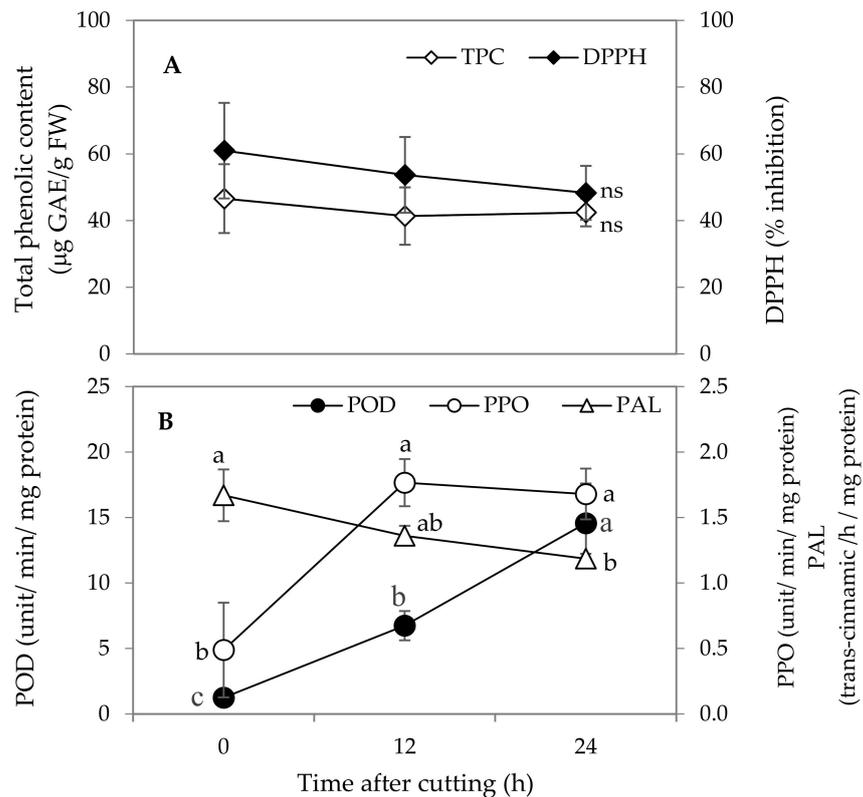


Figure 1. Substrates and enzyme activities associated with browning in cut banana blossom peduncles. Total phenolic content (TPC) and DPPH scavenging activity (A); activities of polyphenol oxidase (PPO), peroxidase (POD) and phenylalanine ammonia lyase (PAL) (B). Activity and content were assayed in banana blossoms stored at 30 °C for 0, 12 and 24 h. Data are shown as mean ± SD. Different letters (a–c) on the line graph of PPO, POD and PAL activities indicate significant differences ($p < 0.05$) between storage times. ns on the line graph of TPC and DPPH indicates nonsignificant differences ($p \geq 0.05$) between storage times.

Table 2. Substrates and enzyme activities in relation to browning of banana blossoms after cutting. Pearson's correlation coefficients (r) between activities of polyphenol oxidase (PPO), peroxidase (POD) and phenylalanine ammonia lyase (PAL); browning scores; color parameters; phenolic content; and DPPH scavenging activity.

	PAL	PPO	POD	L* Value	Browning Score	Phenolic Content	DPPH Assay
PAL	1.000						
PPO	−0.829 **	1.000					
POD	−0.870 **	0.660	1.000				
L* value	0.826 **	−0.957 **	−0.752 *	1.000			
Browning score	−0.840 **	0.954 **	0.782 *	−0.977	1.000		
Phenolic content	0.430	−0.547	−0.187	0.317	−0.402	1.000	
DPPH assay	0.638	−0.590	−0.481	0.423	−0.532	0.923 **	1.000

* indicates a significant correlation at $p < 0.05$. ** indicates a significant correlation at $p < 0.01$.

3.5. Optimal pH for PPO and POD Activities

One important aim of this study, apart from identifying antibrowning agents, was to identify the pH optima of PPO and POD. The pH within plant cells and of all organic acids is naturally below 7. Therefore, pHs above pH 7 were not included in this study because they would not be physiologically relevant. By monitoring the PPO and POD activities at pH between 1 and 7, it was found that the optimal pH for PPO was 7, whereas a broad optimal pH range of 4–7 was found for POD. Rapid loss of PPO activity occurred below pH 7. PPO activity was relatively stable between pH 6 and pH 4 before decreasing to minimal activity below pH 3. For POD, the activity remained steady at pH 7–4 and then decreased to minimal activity below pH 2 (Figure 2).

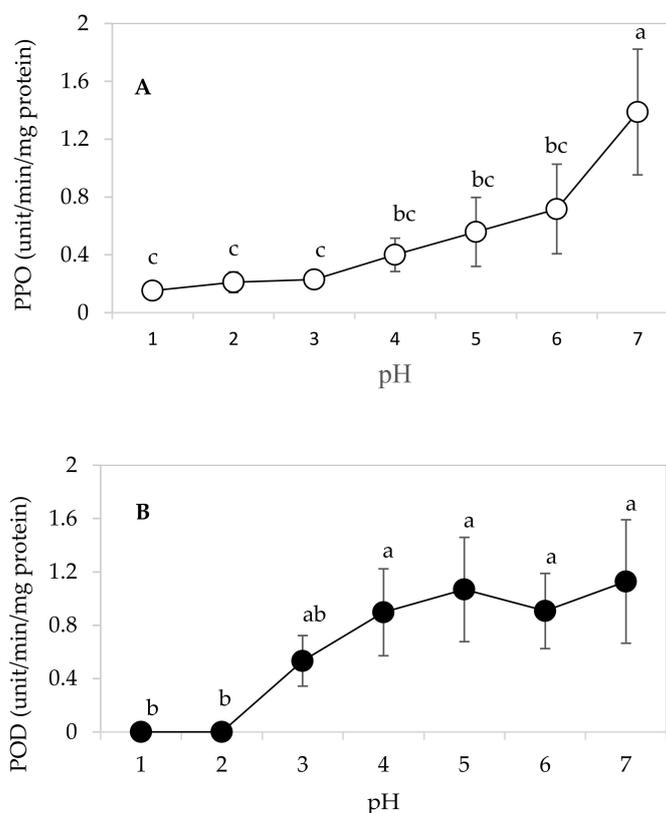


Figure 2. Optimal pH for polyphenol oxidase (PPO) and peroxidase (POD) activities. Activities of PPO (A) and POD (B) at different pH values. Enzymes were assayed at pH 1 and 2 using 0.1 M citric acid–0.15 M HCL and at pH 3 to 7 using 0.3 M using citrate–phosphate buffer. Data are mean \pm SD ($n = 3$). Different letters (a–c) on the line graph of PPO and POD activities indicate significant differences ($p < 0.05$) between different pH conditions.

3.6. Effects of Organic Acids as Inhibitors on PPO and POD Activities of Banana Blossoms

Overall, most of the acid treatments reduced PPO activity by 75–90%, except for 1% citric acid and tartaric acid which led to merely 46% and 68% inhibition, respectively (Table 3). POD activity was 100% inhibited when 5% tartaric acid, 3–5% oxalic acid and ascorbic acid were used. Other acids, i.e., 3% tartaric acid, 5% malic acid and 5% citric acid, also effectively inhibited POD activity by 75–83%. Some acids, i.e., citric, malic, oxalic, and ascorbic acids at the lowest concentrations used, i.e., 1%, did not inhibit POD activity. All organic acids at 3–5% had pH lower than 4, except for 1% ascorbic acid with pH of 6. All organic acids, except for 1% ascorbic acid, inhibited PPO activity. Although control treatment (reaction buffer without organic acid agents) with a pH lower than 3 could inhibit PPO and POD activity by more than 50%, 3% citric acid and malic acid also had pHs lower than 3, but could inhibit POD activity to 0% and 36%, respectively (Table 3).

Table 3. Inhibition of PPO and POD activities using different organic acids as inhibitors. Percentage inhibition of PPO and POD activities by five organic acids of varying concentrations and their natural pH.

Organic Acid Agents	Organic Acid Concentration	pH	Inhibition of Enzyme Activity (%)	
			PPO	POD
Control ^{1/}	0%	7.0 ± 0.0	0 ± 0 d	0 ± 0 d
		5.0 ± 0.0	39 ± 2 c	4 ± 7 d
		3.0 ± 0.0	82 ± 7 a	52 ± 7 bc
		1.0 ± 0.0	88 ± 3 a	100 ± 0 a
TA	1%	3.3 ± 0.4 b	68 ± 8 ab	25 ± 18 c
	3%	2.5 ± 0.3 c	86 ± 4 a	77 ± 9 b
	5%	1.7 ± 0.6 c	83 ± 3 a	99 ± 2 a
CA	1%	4.0 ± 0.0 b	46 ± 11 b	0 ± 0 d
	3%	2.8 ± 0.0 c	76 ± 8 a	0 ± 0 d
	5%	2.5 ± 0.0 c	83 ± 4 a	75 ± 6 b
MA	1%	3.6 ± 0.0 b	79 ± 11 a	0 ± 0 d
	3%	2.8 ± 0.0 c	79 ± 1 a	36 ± 7 c
	5%	2.5 ± 0.0 c	78 ± 3 a	83 ± 2 b
OA	1%	2.6 ± 0.0 c	74 ± 14 a	0 ± 0 d
	3%	0.9 ± 0.3 d	78 ± 10 a	100 ± 0 a
	5%	0.7 ± 0.1 d	90 ± 1 a	100 ± 0 a
AA	1%	5.6 ± 0.1 a	83 ± 2 a	0 ± 0 d
	3%	2.7 ± 0.9 c	86 ± 0 a	100 ± 0 a
	5%	2.6 ± 0.8 c	87 ± 7 a	100 ± 0 a

^{1/} The control was the reaction mixture without organic acids but with the pH adjusted with HCl. TA = tartaric acid, CA = citric acid, MA = malic acid, OA = oxalic acid and AA = ascorbic acid. Data are shown as mean ± SD (*n* = 6). Means in columns with different letters indicate significant differences (*p* < 0.05) between different inhibitors at different concentrations.

Three synthetic organic acids at the lowest concentrations of tartaric acid (5%), oxalic acid (3%) and ascorbic acid (3%) provided the greatest inhibition of PPO and POD activity. Therefore, 5% tartaric acid, 3% oxalic acid and 3% ascorbic acid were selected for further study.

3.7. Effect of Antibrowning Agents on Browning and Weight Loss of Banana Blossoms

Among all compounds tested, 3% oxalic acid caused the highest inhibition of browning at the blossom cut-surface after storage at 15 °C for 10 days. The L*, a* and ΔE* values and browning score of the cut-surface of banana blossom treated with 3% oxalic acid were better than that of the control (DI water, 2.5% tamarind juice and 1% SMS). No significant differences in weight loss among inhibitor agents and storage times were observed (Table 4). There was no abnormal odor and no disease incidence in any of the treated banana blossoms after 10 days of storage (data not shown).

Table 4. Browning response of banana blossoms after various organic acid treatments. Color, i.e., lightness (L^*), redness (a^*) and total color change (ΔE^*), was assessed using a colorimeter. Browning score of peduncle cut-end surfaces and weight loss of banana blossoms were calculated as described. All tissues were kept in PE bags at 15 °C for 0, 5 and 10 days.

Inhibitor	L^*		
	0 Day	5 Days	10 Days
DI water	68.6 ± 4.6 B,a	60.6 ± 5.8 B,b	61.5 ± 5.5 BC,b
2.5% TJ	71.4 ± 3.0 B,a	60.2 ± 3.3 B,b	60.1 ± 2.5 BC,b
1% SMS	75.5 ± 0.0 A,a	54.3 ± 6.8 C,b	53.4 ± 6.8 C,b
5% TA	76.9 ± 0.5 A,a	55.4 ± 3.2 BC,b	55.9 ± 3.4 BC,b
3% AA	77.7 ± 0.7 A,a	59.6 ± 2.1 B,b	58.5 ± 2.5 B,b
3% OA	77.3 ± 1.4 A,a	66.8 ± 2.8 A,b	66.4 ± 2.4 A,b
Inhibitor	a^*		
	0 Day	5 Days	10 Days
DI water	−0.3 ± 1.9 AB,b	4.9 ± 2.5 AB,a	5.2 ± 2.4 AB,a
2.5% TJ	−1.0 ± 1.7 AB,b	5.3 ± 1.8 AB,a	6.3 ± 1.6 AB,a
1% SMS	0.8 ± 0.0 A,b	7.6 ± 1.4 A,a	4.3 ± 1.4 A,a
5% TA	−3.4 ± 0.3 B,b	5.2 ± 1.8 B,a	5.0 ± 1.8 B,a
3% AA	−3.5 ± 0.3 B,b	6.3 ± 1.0 AB,a	7.0 ± 0.5 AB,a
3% OA	−3.4 ± 0.6 B,b	0.0 ± 0.7 C,a	0.0 ± 0.7 C,a
Inhibitor	ΔE^*		
	0 Day	5 Days	10 Days
DI water		12.4 ± 4.3 NS	11.8 ± 1.0 NS
2.5% TJ		13.8 ± 2.9 NS	15.5 ± 3.5 NS
1% SMS		23.4 ± 2.5 NS	24.3 ± 5.5 NS
5% TA		23.4 ± 2.6 NS	22.9 ± 2.8 NS
3% AA		22.4 ± 2.0 NS	23.2 ± 2.3 NS
3% OA		11.6 ± 2.2 NS	12.0 ± 1.7 NS
Inhibitor	Browning score		
	0 Day	5 Days	10 Days
DI water	1.0 ± 0.0 A,b	3.6 ± 0.9 B,a	3.6 ± 0.9 B,a
2.5% TJ	1.0 ± 0.0 A,b	3.4 ± 0.5 B,a	3.4 ± 0.5 B,a
1% SMS	1.0 ± 0.5 A,b	2.8 ± 0.8 B,a	3.8 ± 0.8 B,a
5% TA	1.0 ± 0.0 A,b	4.8 ± 0.4 A,a	5.0 ± 0.0 A,a
3% AA	1.0 ± 0.0 A,b	3.8 ± 0.8 B,a	3.8 ± 0.8 B,a
3% OA	1.0 ± 0.0 A,b	2.8 ± 0.8 B,a	3.8 ± 1.3 B,a
Inhibitor	Weight loss (% of initial FW)		
	0 Day	5 Days	10 Days
DI water		0.3 ± 0.4 AB,a	0.8 ± 0.9 A,a
2.5% TJ		0.5 ± 0.3 A,a	0.6 ± 0.3 A,a
1% SMS		0.4 ± 0.2 A,a	0.2 ± 0.0 AB,a
5% TA		0.2 ± 0.1 AB,a	0.3 ± 0.3 AB,a
3% AA		0.3 ± 0.2 AB,a	0.5 ± 0.2 A,a
3% OA		0.2 ± 0.2 AB,a	0.1 ± 0.1 AB,a

Data are shown as mean ± SD ($n = 5$). Means in columns with different capital letters indicate significant differences ($p < 0.05$) among inhibitors and means in rows with different small letters indicate significant differences ($p < 0.05$) between storage times. NS indicates nonsignificant differences ($p < 0.05$) among inhibitors and storage times. Inhibitors: DI = deionized water, TA = tartaric acid, OA = oxalic acid, AA = ascorbic acid, TJ = tamarind juice and SMS = sodium metabisulfite.

PCA was performed to provide an overview of the differences caused by the four different antibrowning treatments and storage periods on the banana blossoms. The first two principal components together accounted for 73% of the original variance: PC1 explained 44%, and PC2 explained 29%. Tissues clustered into three distinct groups. In group I, all treated tissues assayed before storage (0 d) clustered together; the only tissues treated with acid

and stored that were in this group were those dipped in 3% oxalic acid. The remaining tissues, all stored for 5 or 10 days, separated into either group II or III. Blossoms treated with DI water, or 2.5% tamarind juice clustered together, while blossoms treated with 1% SMS, 3% ascorbic acid and 5% tartaric acid formed the remaining cluster (Figure 3). Loading analysis indicated that the L^* and b^* values with loading scores of -0.83 and 0.91 on PC1 and PC2, respectively, determined the grouping (data not shown).

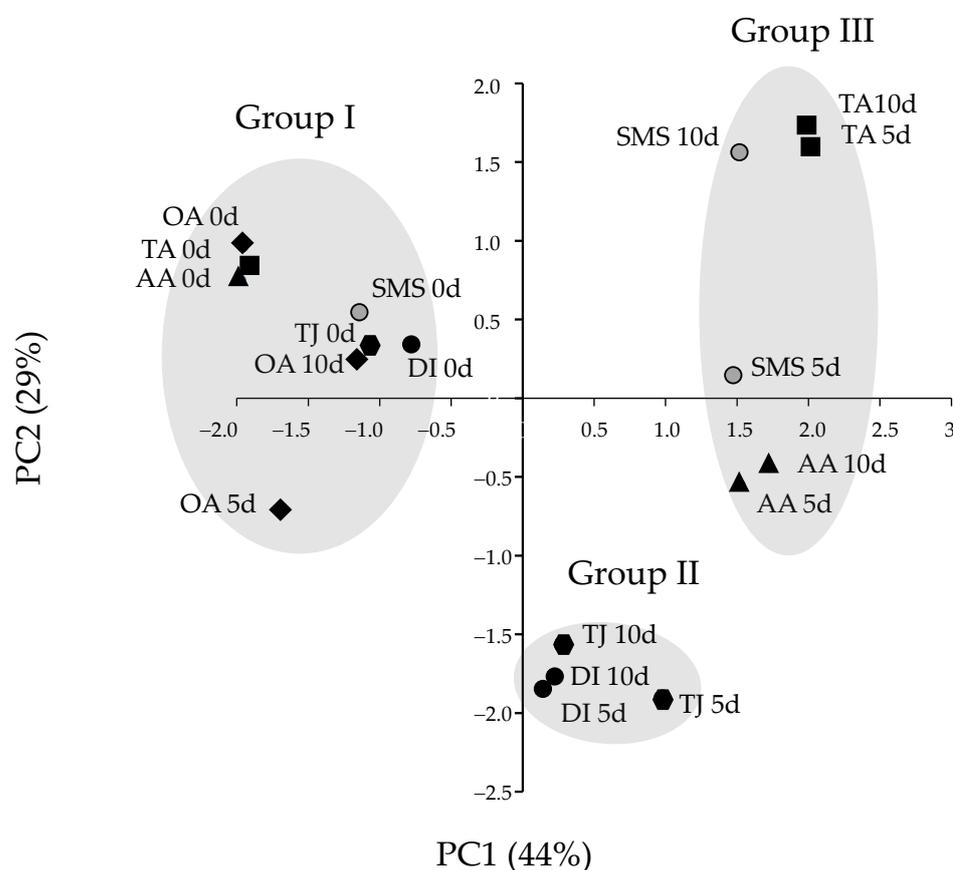


Figure 3. The principal component analysis (PCA) of objective color status and browning of the peduncle cut-end surfaces of banana blossoms after antibrowning treatments. Objective color was assessed by lightness (L^*), redness (a^*), yellowness (b^*) and total color change (ΔE^*); the browning score and weight loss of the peduncle cut-end surfaces of banana blossoms were assessed after storage in polyethylene bags at 15°C for 0, 5 and 10 days. Banana blossoms treated with 3% oxalic acid (OA) and stored for 5 or 10 days were similar to those on the initial day of treatment. Banana blossoms treated with 3% ascorbic acid (AA) and 5% tartaric acid (TA) and stored for 5 or 10 days were similar to those treated with 1% sodium metabisulfite (SMS) and stored for the equivalent period. Banana blossoms treated with 2.5% tamarind juice (TJ) and stored for 5 or 10 days were similar to those treated with DI water.

4. Discussion

Banana blossoms have been traditionally eaten in dishes in SE Asia for hundreds of years, but there is now a growing demand, both locally and globally, for high-quality blossoms with reduced or no browning. This necessitates an investigation of the postharvest browning of fresh-cut banana blossoms. Here, we examined the activity of key browning enzymes and also investigated acid treatments to prevent undesirable browning.

4.1. Browning Mechanism at Cut-End Surface of Banana Blossoms

Browning or blackening from abiotic wounding or cutting occurs by enzymatic oxidation of phenolic compounds [37]. At the site of injury, chemical signals originate

and propagate into adjacent tissue. This induces the de novo synthesis of phenylalanine ammonia lyase (PAL) activity, the first key enzyme that converts L-phenylalanine to trans-cinnamic acid [38,39] resulting in the accumulation of phenolic compounds. In the presence of oxygen, phenolic compounds are oxidized by PPO to quinones that are highly reactive and can combine with other compounds to form brown pigments [40]. POD, on the other hand, acts as an antioxidant enzyme to eliminate excess hydrogen peroxide (H_2O_2) [41] and initiates enzymatic browning of fruit and vegetables [42]. Here, PPO and POD activities increased rapidly 12 h after cutting, in concert with increased browning, while in contrast, PAL activity decreased (Figure 1 and Table 1). Thus, only PPO and POD are induced by wounding, and together they serve as the key enzymes involved in the browning mechanism at the cut-end surface of banana blossoms.

In this study, the phenolic content (TPC) and DPPH scavenging activity of the blossoms were high and did not change as browning progressed (Figure 1). Phenolic compounds contribute to browning by acting as substrates for PPO and POD, and this is true in banana blossoms. The almost constant TPC during browning development indicates that banana blossoms were not limited in the phenolic compounds to supply substrate for PPO and POD.

4.2. Oxalic Acid Treatment as the Most Effective Method to Prevent Browning after Cutting

Different organic acids were tested as effective suppressors of the browning that occurs in the peduncle cut-end surface of banana blossoms. Low pH, the presence of a chelating copper ion and the ability to reduce quinones to their original diphenol are the main properties of organic acids that render them capable of inhibiting PPO and POD activities. Since different types and concentrations of organic acids inhibited the browning of banana blossoms differently, diverse inhibitory mechanisms for PPO and POD by each organic acid were assumed. Both browning scores and browning enzyme activities of banana blossoms were most inhibited by 3% of oxalic acid treatment (Tables 3 and 4). Oxalic acid may have a more effective chelating ability compared to other acids with the same structure [43,44]. Oxalic acid can bind to the PPO copper-containing active sites to form an inactive complex, thus reducing browning [45]. This finding agrees with reports on lettuce where oxalic acid diminished catechol quinone formation and no quinone bleaching was observed [46].

The present study also suggested that pH 2 reduced PPO activity and completely inhibited POD activity in the banana blossoms after tissue wounding by cutting. Similar to our results, Oba et al. [17] found that the activity of PPO from 'Saba' banana blossoms was reduced with decreasing pH from 7.5 to 4. He and Luo [10] reported that environmental pH affected the ionizable groups of the PPO and POD protein structures, which in turn would diminish their ability to catalyze their respective browning reactions. An acidic pH of 3.54 induced the unfolding of the native mushroom PPO with structural changes [47]. Oxalic acid had a low pKa value with a natural pH of 0.9 when 3% concentration was used. Thus, the low pH value of 3% oxalic acid may partially explain why this treatment was the most effective method to prevent banana blossom browning.

In comparison to the commercial antibrowning agent, i.e., 1% SMS, treatment with 3% oxalic acid led to higher browning inhibition at the blossom cut-surface. The 3% oxalic acid treatment is easy to apply, cost-effective and nontoxic. In this study, the organic acid treatments were applied only at the peduncle cut-end surface of the banana blossoms. Possible negative effects on quality attributes that may be caused by the organic acid treatments include unpleasant coloration, browning, weight loss, abnormal odor or disease incidence. However abnormal odor and disease incidence were not detected after organic acid treatment used in this experiment. Hence, this treatment holds great promise for use as a commercial agent for reduced browning of banana blossoms.

5. Conclusions

PPO and POD were identified as the key enzymes involved in browning at the cut-end surface of banana blossoms. Optimal pH values for PPO and POD activity in banana

blossoms were pH 4–6 and pH 7, respectively. A pH value of 2 inhibited PPO and POD activity by as much as 80% and 100%, respectively. Treatment with 3% oxalic acid effectively inhibited browning at the cut-end surface of the banana blossom peduncle for 10 days after 15 °C storage under PE package. It is proposed that 3% oxalic acid should be widely investigated as an effective commercial treatment to reduce browning and enhance the postharvest quality of banana blossoms.

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