

Review

# Chemopreventive Potential of Oils Extracted from Seeds of Three *Annona* Species

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**Abstract:** *Annona* fruit, leaves, seeds, roots, and bark have been conventionally used in many countries for medical treatments as they are considered ideal sources of pharmacologically active compounds, but *Annona* remains an underutilized fruit in many countries. The fruit of these plants is delicately flavored and is used in industrial products such as ready-to-serve beverages, wine, jellies, jam, and fruit-butter preserve, while the seeds generally go to waste. *Annona* seed oil contains numerous health-benefiting factors such as vitamins, minerals, bioactive compounds, fatty acids, antioxidants, and phenolic compounds, which are responsible for various biological activities, including antibacterial, antioxidant, and antitumor activities. Cancer is a worldwide major health problem that remains unresolved. Even though the current treatments can manage to reduce tumor growth, there is an urgent need to investigate more efficient but less expensive novel techniques to overcome some of the restrictions in treating tumors. *Annona* might offer an indispensable choice besides chemotherapy and radiotherapy, especially for terminally ill patients, as the *Annona* genus contains secondary metabolites in nearly every component of *Annona* plants. Research has shown that many *Annona* species contain promising components that could potentially exhibit anticancer activity, but the information available is scarce and inconsistent. *Annona muricata* (Soursop, “Katuanoda”), *Annona squamosa* (Sweetsop, “Seenianoda”), and *Annona reticulata* (Custard apple, “Welianoda”) are three commonly cultivated edible *Annona* species in Sri Lanka. The main objective of the review was to present an updated comprehensive literature analysis of the putative chemopreventive functions against cancer cell lines/the anticancer effect on cancers, phytochemical properties, and antioxidant properties possessed by the seed oils of three selected common *Annona* species. Although there are some in vitro and in vivo experimental investigations supporting the benefits of *Annona* seed oils, clinical investigations are still needed to explore concealed areas, determine the effects on the human body, determine the safest concentration, and determine health-contributing benefits before they are submitted to clinical trials.

**Keywords:** *Annona muricata*; *Annona squamosa*; *Annona reticulata*; *Annona* seed oil; antioxidants; chemopreventive potential; phytochemicals



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## 1. Introduction

*Annona* is considered an important yet underutilized fruit with high nutritional and medicinal value [1]. These different plant-based medicinal compounds are prepared using a variety of different procedures, including infusions, pastes, and decoctions, in traditional remedies [2–4]. In particular, *Annona reticulata* bark, root bark, and root decoction are used for toothache, fever, and, as an antipyretic and immature fruit, for the treatment of dysentery and diarrhea. In addition, *Annona squamosa* and *Annona muricata* have been recorded for their antidiarrheal effects, and their leaves are used to make tea to treat colic; leaf decoction has been used to treat worms, boils, and abscesses; and seed paste is used to treat cancer and heal cattle wounds [5–8]. Regarding *A. muricata*, its dried leaves are used to treat spasms and dysentery and taken orally for analgesic effects in certain regions of Indonesia, and its fruit juice is used as a diuretic and to treat liver disorders [8–11]. According to ethnobotanical studies, *A. muricata* seeds are toxic, yet the powder made from toasted seeds has been employed in traditional Mexican medicine [10]. In tropical America and India, the leaves of *A. squamosa* have been used systemically to treat dysentery as a tonic and cold medication [11]. *Annona cherimola*, *A. squamosa*, and *A. reticulata* have been documented for their antiparasitic properties. *A. muricata* and *Annona glabra* have also historically been utilized for curing rheumatism [12] while also being used to treat ulcers [13]. The leaves of *Annona diversifolia* have been utilized as analgesic, anticonvulsant, and anti-inflammatory medications [14].

Various studies have reported a variety of pharmacological activities, including antibacterial, anticancer, antidiarrhea, antidiabetic, anti-inflammatory, antioxidant, and antiulcer, that are attributed to the most abundant component vitamins, minerals, bioactive compounds, fatty acids (FAs), antioxidants, etc., of either crude extracts or isolated compounds from various parts of *Annona* species, especially leaves and seeds [2,15–23]. Among the 30 reviewed *Annona* species, six species, *A. squamosa*, *A. muricata*, *A. cherimola*, *A. senegalensis*, *A. reticulata*, and *A. coriacea*, are the most widely studied for their pharmacological activities and the phytochemical profiles of their bark, leaves, fruits, and seeds. For example, the leaf extracts of both *A. squamosa* and *A. reticulata* exhibited potent antiproliferative effects [24], and the ethanolic extract of *A. muricata* leaves was reported for its cytotoxicity against promonocytic leukemic cells [25]. Isocoreximine isolated from *A. cherimola* demonstrated cytotoxicity against multiple cancer cell lines, including the breast cancer cell line, the human colorectal carcinoma cell line, the human prostate tumor cell line, and the human leukemia cell line [26]. Additionally, various parts of *A. muricata*, including its bark, leaves, and stems, were shown to exhibit antioxidant activity using a DPPH assay [22], and the ethanolic extract of *A. squamosa* leaves was also reported to have antioxidant activity [27]. With regard to the phytochemistry of *Annona* species, alkaloids are mainly present in the leaves, whereas acetogenins are present in the seeds and found in smaller quantities in the pulp and leaves [28,29].

*Annona* fruit flesh is edible and is currently used in industrial products such as ready-to-serve beverages, wine, jellies, jam, and fruit butter preserve. These industries usually generate a considerable number of seeds as waste, and a renewed focus on the utilization of these wastes in food processing could minimize the accumulation of waste while concurrently contributing to minimizing the waste disposal problem and generating extra revenue [30]. Similarly, seed oils of edible fruits are important common food ingredients, and evidence has suggested that some compounds available in seeds may play different roles in human health, especially in acute and chronic disease management, such as cancer, cardiovascular diseases, and diabetes [31].

Cancer is a worldwide major health problem that remains unresolved, and the incidence of cancer is rising each year [32,33]. According to data from Cancer Research UK, there are more than 8 million cancer-related deaths every year in the world, and it is predicted that this figure will rise to 14.6 million by 2035 [34,35]. Cancer is a hereditary condition that develops when our bodies' genes, which regulate cell functions, growth, reproduction, and death, experience alterations [36]. The right cancer treatment, includ-

ing surgery, radiotherapy, chemotherapy, hormone treatments, and targeted biological therapies, depends on a proper cancer diagnosis. Concerns about the effectiveness of the current treatment options have been raised due to the prevalence and severity of cancer, which has led patients to search for alternatives that can complement or substitute the current treatment options [36,37]. There is an urgent need to investigate new techniques to overcome some of the restrictions in treating tumors, even though the current treatments can manage to reduce tumor growth [38].

Research has shown that many *Annona* species contain promising components that could potentially exhibit anticancer activity [39]. It has been decided to focus on three common *Annona* species, *A. muricata*, *A. squamosa*, and *A. reticulata*, since available evidence suggests that their uses and importance may be expanded, but the information available is scarce and inconsistent. Consequently, academics, researchers, extension workers, and farmers need better information [40]. A relationship between *Annona*'s potential as an industrial pharmaceutical and a food supplement was established in the current investigation. As this notion has not yet been much revealed, an updated summary of the putative chemopreventive functions of *Annona* seed oils is provided in this review, along with a critical evaluation of the cytotoxicity activity against cancer cell lines, phytochemical properties, and antioxidant properties possessed by the seed oils of the three selected common *Annona* species.

## 2. *Annona* Genus

The family Annonaceae, which includes a variety of aromatic trees, bushes, shrubs, and climbers or lianas, includes the genus *Annona*. A few of these plants grow in temperate zones, although they are mostly found in tropical and subtropical areas [40,41]. *Annona* is the most significant genus in the Annonaceae family due to its edible fruits and therapeutic benefits [42]. The Latin word “anon,” which means “yearly produce” and refers to the production of fruits from approximately 119 species globally, is the source of the genus name “*Annona*” [43,44]. Despite *Annona* having many species, only limited species of this family are economically important and commonly found in Sri Lanka, such as *A. squamosa* (sugar apple); *A. cherimola* (cherimoya); *A. muricata* (soursop); Atemoya, a hybrid between *A. cherimola* and *A. squamosa*; *A. reticulata* (Custard apple); and *A. glabra* (pond apple) [40,45]. There are many shared traits, and the species can be distinguished based on the traits of the plant, such as fruit size, shape, color, and quality, among others [46].

According to the Department of Agriculture, Sri Lanka, six edible *Annona* species can be commonly found in Sri Lanka, namely *A. muricata*, *A. squamosa*, *A. reticulata*, *A. glabra*, *A. cherimola*, and Atemoya (Figure 1) (Table 1) [40,45,47].

Among these, three species are commercially cultivated in Sri Lanka: *A. muricata*, *A. reticulata*, and *A. squamosa* [48].



**Figure 1.** Common *Annona* spp. grown in Sri Lanka.

**Table 1.** Commonly found *Annona* species in Sri Lanka.

Botanical Name	Vernacular Name	Common Name
<i>Annona muricata</i> L.	Katuanoda	Soursop
<i>Annona reticulata</i> L.	Welianoda	Bullock's heart, custard apple
<i>Annona squamosa</i> L.	Sinianoda	Sweetsop/sugar apple
<i>Annona cherimola</i> Miller	Cherimoya	Cherimoya
A cross of <i>A. cherimola</i> and <i>A. squamosa</i>	Atemoya	Pineapple sugar apple
<i>Annona glabra</i> L.	Welatha	Pond apple, alligator apple

### 2.1. *Annona muricata* (Soursop)

*A. muricata* is called soursop in English, 'Katu anoda' in Sinhalese, and 'Sitha palam' in Tamil. It is probably indigenous to the West Indies and is grown in Sri Lanka's lowlands in wet zones [49]. It is a small tree, about 4–10 m tall, and its fruit are ovoid or irregularly ovoid, conical heart-shaped, dark green in color, and covered with soft spines (Figure 1). The fruit resembles white, less sweet, more acidic, cottony/fibrous, and juicy flesh and is used commercially for the production of candy, juice, and sherbets. It contains 127–170 blackish-brown seeds, which later turn brown during storage and are about 1–2 cm in size (Figure 2) [50–52]. This is the widely available *Annona* species in Sri Lanka, and several *A. muricata* accessions have been identified within the different agro-climatic zones in Sri Lanka. Most of the plant's parts have historically been used to treat a variety of illnesses, such as fever, rheumatism, and cancer [53]. Presently, individuals with cancer frequently utilize this plant, and numerous studies concentrate on the dietary, therapeutic, and bioactive components of *A. muricata* leaves [5].

**Figure 2.** General appearance of *A. muricata*, *A. squamosa*, and *A. reticulata* seeds (Scale 4:3).

### 2.2. *Annona reticulata* (Bullock's Heart/Custard Apple)

*A. reticulata* is named bullock's heart/custard apple in English, 'Welianoda' in Sinhalese, and 'Ramsitha' in Tamil. It originated in the West Indies and is grown as a fruit in Sri Lankan wet-zone home gardens. It has a small tree height of about 5–10 m, and the fruit is spherical, ovoid or heart-shaped, reddish-yellow, and possesses impressed lines above the carpels (Figure 1) [54]. For the treatment of epilepsy, diarrhea, cardiac issues, constipation, bleeding, bacterial infections, parasite and worm infestations, fever, ulcers, and as a pesticide, this plant's many parts are used in traditional medicines. There are frequently more than 40 black-brown, compacted, and glossy seeds about 1 cm in size (Figure 2) in its thick, cream-white flesh, and it has a somewhat granular body [55,56].

### 2.3. *Annona squamosa* (Sugar Apple/Sweetsop)

*A. squamosa* is called sugar apple/sweetsop in English, 'Sinianoda' in Sinhalese, and 'Sittapan' in Tamil and originated in tropical America and the West Indies. It is cultivated in Sri Lankan home gardens in low- and mid-country wet zones. It has a small tree, about 3–8 m high, with globular, cordate-ovoid, conical, or heart-shaped, yellowish-green fruit, which contains white pulp, which is pleasant, sweet-sour-flavored (Figure 1). The seeds are brownish black, about 1 cm in size (Figure 2), and each fruit contains about 35–45 [55,57]. This *Annona* species remains one of the most researched in the world and is well known for

having antidiabetic effects [58]. It has a variety of pharmacological characteristics, primarily antitumor properties, in its seeds, bark, and leaves [59].

### 3. Proximate Composition of Annona Seeds and Seed Oil Extraction

The impact of seed oil on human health is determined by its chemical composition. The proximate composition of *A. muricata*, *A. squamosa*, and *A. reticulata* seeds on a dry basis (g/100 g dry weight (DW) (%)) is summarized in Table 2.

Generally, Annona seeds are mainly composed of about 32.4% seed coat and 67.7% seed kernel. A proximate composition analysis of *A. muricata* seeds demonstrated the presence of 40% lipids, where *A. squamosa* seeds contain about 26.8% lipids, and *A. reticulata* seeds reveal the presence of 30.34% lipids g/100 g dry weight (DW) (%) [60–65]. These seeds have special potential for industrial use because of their high oil contents. The oil yield from seeds varies depending on the variety, seed size and shape, stage of fruit ripening, timing of seed harvest, soil and environmental conditions around the oil-bearing plant, pre-treatment procedure, and the particular extraction method used [60].

**Table 2.** The proximate analysis of seeds of *A. muricata*, *A. squamosa*, and *A. reticulata*.

Parameter (% g/100 g Dry Weight)	<i>A. muricata</i> *	<i>A. squamosa</i> **	<i>A. reticulata</i> ***
Moisture	7.7 ± 0.24	6.7 ± 0.2	15.2 ± 0.02
Lipid	40 ± 0.82	26.8 ± 0.4	30.34 ± 0.04
Protein	8.5 ± 0.52	17.5 ± 0.2	17.12 ± 0.01
Ash	9.7 ± 0.12	2.2 ± 0.1	4.5 ± 0.00
Fiber	5.2 ± 0.26	16.8 ± 0.2	32.0 ± 0.01
Carbohydrate (By Difference)	34.1	30.0	37.91

(\* Onimawo [63]; \*\* Abdalbasit et al. [64]; \*\*\* Ezekiel et al. [65]).

Palm and olive oils are extracted from their fruits, and most vegetable oils such as cotton, sesame, ground nut, rape, corn, soya bean, and sunflower are extracted from seeds. According to the findings, seed oils can be extracted either by conventional extraction methods, such as solvent extraction, steam distillation, and mechanical pressing, or by non-conventional extraction methods, such as supercritical fluid extraction, pressurized liquid extraction, microwave-assisted extraction, ultrasound extraction, and pulsed electric field extraction. In fact, conventional methods are technically and scientifically challenging to handle and non-environmentally friendly [66–68], and until recently, the majority of reported seed oil extractions have been performed by Soxhlet extraction or cold pressing. The difference in oil content between these two extraction methods was not significant. This can be attributed to the possibility that during Soxhlet extraction, sample heating from the high temperature used for solvent evaporation may have caused oil droplets to emerge from the sample more readily; thus, the percentage of oil recovery from the seeds is high [61,69]. Though the majority of fruit seed oils are made by cold pressing the seeds, there are some instances where the oil content of the seeds is too low for cold pressing [70]. Regarding Annona seed oil (ASO), the maximally efficient extraction method has been shown to be Soxhlet extraction [68]. In Soxhlet extraction, the oils from the powdered seeds are extracted exhaustively with petroleum analytical-grade ether (boiling point: 60–80 °C) as the extraction solvent in the Soxhlet apparatus for 16 h, and the petroleum ether is separated from the oil by distillation at 100 °C in an air oven for 1 h [71].

Methyl esters of ASO have been studied using the GC/MS (gas chromatography–mass spectroscopy) method to determine their chemical composition. According to Table 3, oleic acid, linoleic acid, palmitic acid, and stearic acid were present in higher amounts, respectively, in all three ASO varieties, while linolenic acid, palmitoleic acid, myristic acid, and capric acid were found in trace amounts. Oleic acid was the most dominant FA (fatty acid), and linoleic acid was the second most dominant FA [63,72]. Further analyses of the *A. squamosa* seed oil (ASSO) revealed a significant concentration of UFAs (unsaturated fatty acids), and the linoleic and oleic acids provided about 70.3% of the total [63,73]. Even

within the same species, there may be variations in the FA composition of the seed oil. For instance, research revealed that oleic (37.0%), palmitic (25.1%), and linoleic (10.9%) acids were the predominant FAs in ASSO [74–76]. In contrast, 32.0% linoleic and 29.0% oleic contents have been observed [60]. Numerous studies have examined the chemical composition of ASSO, which was mostly found to be made up of oleic acid, linoleic acid, stearic acid, and palmitic acid, with UFAs composing up to approximately 70% of this mixture [77].

**Table 3.** The fatty acid composition of *A. muricata*, *A. squamosa*, and *A. reticulata* seeds, soybean, rapeseed, and sunflower.

Compound (%)	<i>A. muricata</i> seeds *	<i>A. squamosa</i> seeds **	<i>A. reticulata</i> seeds ***	Soybean ****	Rapeseed ****	Sunflower seed *****
Saturated fatty acids (SFA)						
Capric acid (10:0)	Traces	0.17 ± 0.03	Traces	-	-	-
Myristic acid (14:0)	Traces	0.67 ± 0.01	0.35 ± 0.03	0.10 ± 0.001	0.1	0.10 ± 0.003
Palmitic acid (C16:0)	20.41 ± 1.58	15.47 ± 0.17	14.1 ± 0.13	11.33 ± 0.56	3.49	6.94 ± 0.14
Stearic acid (C18:0)	4.13 ± 0.29	8.14 ± 0.04	6.86 ± 0.02	4.55 ± 0.19	0.85	5.90 ± 0.10
Unsaturated fatty acids (UFA)						
Monounsaturated fatty acids (MUFA)						
Palmitoleic acid (C16:1)	1.44 ± 0.45	1.43 ± 0.03	1.41 ± 0.01	0.1	0.2	0.2
Oleic acid (C18: 1)	41.29 ± 0.53	48.54 ± 0.13	47.4 ± 0.01	22.24 ± 0.28	64.40	19.49 ± 0.76
Polyunsaturated fatty acids (PUFA)						
Linoleic acid (C18:2)	30.85 ± 0.34	23.40 ± 0.06	22.9 ± 0.03	54.67 ± 0.98	22.30	64.87 ± 1.94
Linolenic acid (C18:3)	1.88 ± 0.25	2.18 ± 0.03	1.79 ± 0.03	6.07 ± 0.17	8.23	1.90 ± 0.06
Total SFAs	24.54	24.45	24.35			
Total U FAs	75.46	75.55	75.65			

Values are mean ± SD for triplicate determinations. (\* Kimbonguila et al. [78]; \*\* Abdalbasit et al. [63]; \*\*\* Abdalbasit et al. [63], Omkaresh et al. [72]; \*\*\*\* Ma and Hana. [79]; \*\*\*\*\* Talpur et al. [80]; Kalo; and Kemppinen [81]).

*Annona muricata* seed oil (AMSO) contained SFAs (saturated fatty acids) and UFAs: 24.54% and 75.46%, respectively. Since it prevents heart problems, linoleic acid is one of the most significant PUFAs (polyunsaturated fatty acids) in human nutrition [82]. By using the Blye and Dyer method, 2.38% linolenic acid is recorded, and by using the Soxhlet method, 1.88% is recorded. Additionally, the Blye and Dyer method yields 1.20% palmitoleic acid, and the Soxhlet method yields 1.44%. Comparing the fatty acid composition of AMSO to that of vegetable oils reveals that this plant is particularly high in oleic acid, linoleic acid, and palmitic acid [78].

The amount of UFAs in *Annona reticulata* seed oil (ARSO) comprised a substantial amount of the total discovered FAs (75.65%), making the oil nutritionally desirable, while the proportion of SFAs was only 24.35% [72]. Research data on ARSO were rare, though it has been used commonly for indigenous medicinal purposes; thus, there is huge research potential.

#### 4. Chemopreventive Agents of Annona Seed Oils and Their Chemopreventive Potential

Chemoprevention is a method of controlling cancer with which the development of the illness can be completely avoided, slowed down, or reversed by one or more organic or inorganic substances. Annona might offer an indispensable choice besides chemotherapy

and radiotherapy, especially for terminally ill patients, as the *Annona* genus contains secondary metabolites, including phytochemicals, antioxidants, essential oils, FAs, minerals, and vitamins, in nearly every component of *Annona* plants [40,83]. The structure of bioactive substances affects their ability to treat or prevent cancers. The efficiency of cytotoxic activity is determined by the high number of hydroxyl groups supported by the hydroxyl position flanking the  $\gamma$ -lactone ring and the stereo-chemical configuration of the ring [84,85]. Researchers have recently focused on the anticancer effects of *Annona* seeds, including ASSO, AMSO, and ARSO, and unique chemical compounds extracted from *Annona* have shown anticancer activities, which allows for further improvements to the available treatment methods, and this research needs to continue to allow for the development of more efficient but less expensive treatments against cancer [86–88].

#### 4.1. Fatty Acids in *Annona* Seed Oils

FAs are usually found as components of many complex lipid molecules and can be identified as SFAs or UFAs based on the hydrogen atom bonding to the carbon atoms in the molecule. UFAs can be either MUFAs (mono-unsaturated fatty acids) or PUFAs [66]. In terms of composition, unlike animal oils that are made up mainly of SFAs, ASOs contain varying proportions of SFAs and UFAs tied in their TAG (triacylglyceride) molecules, and the percentage of UFAs is higher than that of SFAs (i.e., FAs esterified to a glycerol moiety). Additionally, this makes ASO an interesting source of two PUFAs; linoleic and linolenic acids, which are termed EFAs (essential fatty acids) because humans must obtain them from their diets. TAG is the principal means of storing FAs in biological systems and is considered cytotoxic [89].

The oil and fat available in *Annona* seeds consist of different FA compositions and lipid profiles. By utilizing the GC/MS method to examine the methyl esters of the seed oil's fatty oil, it is possible to ascertain the chemical composition of the oil [90]. In order to be an oil that is nontoxic for humans, the concentration of linoleic acid and oleic acid should not be more than 12% and 1%, respectively, of that of FA methyl esters, according to European guidelines. Despite being toxic, ASO has large levels of UFAs, particularly oleic and linoleic acids. After detoxification, ASO can also be consumed because it contains a lot of UFAs [91]. Consumed oils have a significant impact on human physiology, including lipid metabolism, chronic illness prevention, and general well-being [92–96]. ASO can be widely used to cure cancer because it includes oleic acid, linoleic acid, stearic acid, palmitic acid, and other UFAs [97].

The physio-chemical properties, such as melting point, solid fat index, saponification value, cloud point, flash point, iodine value, color, viscosity, density, specific heat, and the heat of fusion, etc., of oils are largely dependent on the nature of their constituent FA and TAG compositions. Nonetheless, the FA and TAG compositions of different oils from diverse sources mostly determine their qualitative features and applications, and some of them can become source materials for functional lipids [73,98]. In this context, the FA type, the degree of saturation or desaturation, the method of delivery to cancer cells or hosts, and the tumor/cell type must be considered when evaluating the effects of FAs on tumor cell lines [99]. In the literature, the cytotoxic effects of FAs have been individually evaluated in different tumor cell lines with differing results [100–104]. For example, ASSO has been shown to suppress H22 solid tumor development and show selective cytotoxic activity against HepG2 cell lines, which might be attributed to the presence of UFAs and acetogenins [105].

According to the criteria adopted by the American National Cancer Institute and some cytotoxic screening assays, a crude extract oil showing an IC<sub>50</sub> value < 30  $\mu\text{g}\cdot\text{mL}^{-1}$  in tumor cell lines is considered promising for anticancer drug development [106]. Since ASO has a high content of UFAs, a well-known antineoplastic activity, and different cytotoxic effects have been reported for some species of *Annona*, its cytotoxic effects against tumor and non-tumor cell lines should be further studied [107,108].

#### 4.2. Phytochemicals in Annona Seed Oils

Plant species rich in phytochemicals are believed to reduce disease risk and have therapeutic characteristics, thus having potential medicinal value [91]. Phytochemicals are secondary metabolic compounds, namely acetogenins, alkaloids, phenolic compounds, essential oils, flavonoids, terpenoids, cyclopeptides, carotenoids, amino acids, etc. (Figure 3). Some phytochemicals, such as acetogenins, have shown neurotoxicity in *in vitro* and *in vivo* studies. More research is needed to quantify the number, of neurotoxic compounds and to determine the level of human exposure. Metabolic studies are also necessary to determine whether digestive processes decrease or increase the bioactivity and/or neurotoxicity of the active compounds. These studies have been extended to ASOs used in medicinal treatments [109].

Due to their capacity to scavenge potentially damaging free radicals, it has been discovered that the pharmacological effects of ASOs, including anticancer properties, are connected to the availability and content of phytochemicals and are influenced by the structure of these bioactive compounds. *Annona* has attracted the interest of individuals interested in the utilization of natural compounds that play a crucial role in the food and pharmaceutical industries [84,85]. Due to *Annona*'s alleged ethno-medical applications against tumors and cancer, extensive anticancer research has been carried out on these plants [110]. Some studies stated that ASO showed high antioxidant activities and phenolic contents and thus may have therapeutic uses [111,112].

The most prevalent bioactive molecules in the Anonaceae family are Anonaceous acetogenins, a special class of long-chain FA derivatives produced via the polyketide pathway. More than 120 acetogenins have been discovered in earlier phytochemical studies on *Annona*'s leaves, stems, bark, seeds, pulp, and fruit peel [113]. Solamin, annoneticulin-9-one, annononicin, squamone, and rolliniastatin are the principal five anonaceous acetogenins with cytotoxic properties [87]. The mechanism of acetogenin's cytotoxic action is the inhibition of mitochondrial complex I [114] and the inhibition of ubiquinone-linked NADH oxidase in the plasma membranes of cancerous cells, causing apoptosis [115]. A broad class of naturally occurring secondary metabolites is called alkaloids, and they are derived from amino acids or the transamination process [116]. Isolated and reported alkaloids include anomurine, anomuricine, isoquinoleic alkaloids, anonaine, and isolaureline [117]. Since the majority of phenolic compounds are water-soluble and the conventional medicinal extract is an aqueous infusion, they are regarded as the most significant phytochemicals [118].

The important bioactive constituents of ARSO are alkaloids, tannins, flavonoids, cardiac glycosides, phenols, terpenoids, steroids, and saponins [62]. The phytochemical concentrations ranged from 0.16 mg/100 g to 16.15 mg/100 g, with phenol found to be significantly prevalent, whilst tannins, alkaloids, and flavonoids were found to be moderately prevalent, and terpenoids and steroids had the lowest prevalence [119–121]. In general, flavonoids have the potential to significantly reduce the risk of disease through a variety of physiological processes, including cytotoxic and antioxidant actions [7]. Acetogenins have been identified from ARSOs, and some of these substances have demonstrated strong cytotoxic action against four human cancer cell lines (Table 4). The biological activity of annonacin was investigated after further purification, and it was discovered that this substance killed cells in several cancer cell lines. The biological effects of squamocin were also examined, and this research demonstrated that squamocin is a cytotoxic component for almost all cancer cell lines [122–124].

According to preliminary phytochemical analyses, the biological actions of ASSO are primarily brought on by phenolic chemicals such as alkaloids, peptides, amino acids, sterols, tannins, flavonoids, polysaccharides, and tocopherols, which are known to have strong anticancer activities, and they can be used to treat or prevent a variety of illnesses, including cancer [12,125–128]. ASSO has demonstrated substantial antitumor activity *in vitro* and *in vivo* against human hepatoma cells, suggesting the potential for the extract to be developed as a novel anti-liver cancer medication [129,130]. An earlier investigation

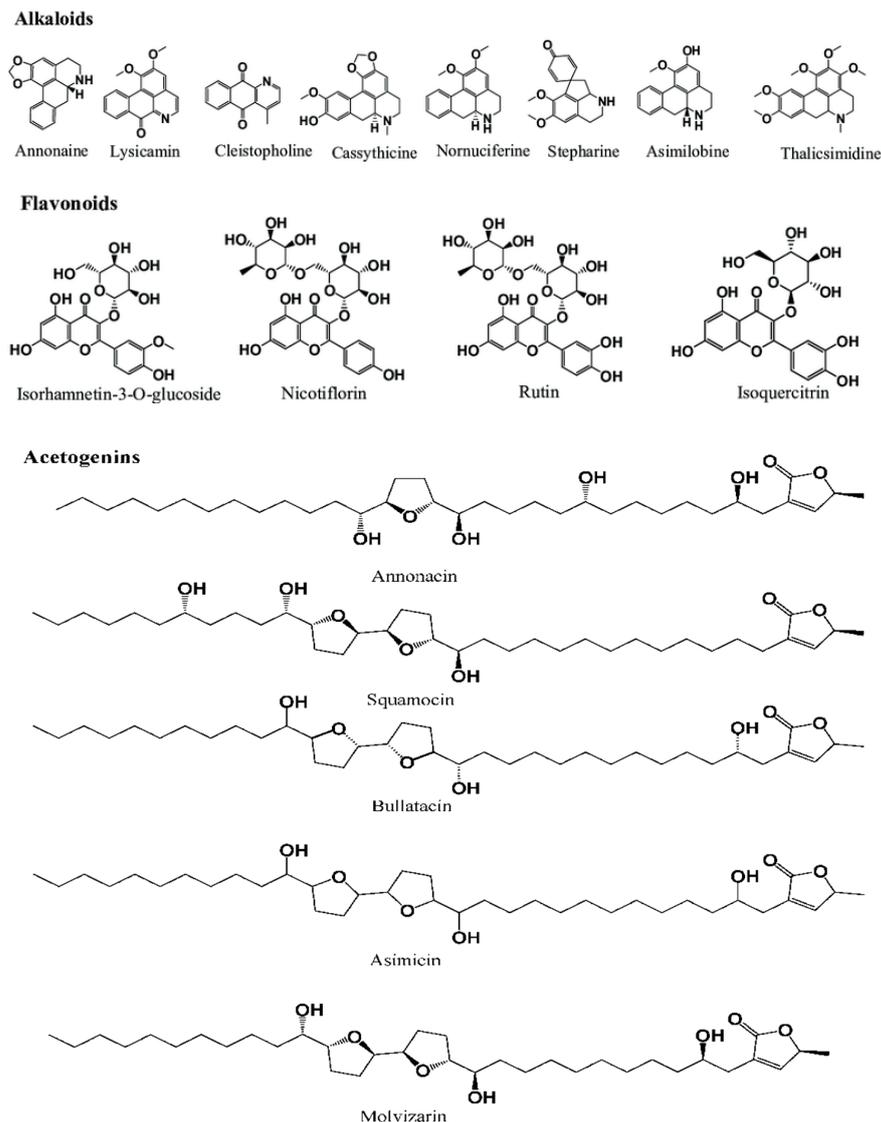
into the pharmacology of ASSO revealed that it has specific cytotoxic effects on the HepG2 cell line [131].

**Table 4.** Reported cytotoxic activities of acetogenins present in ASOs.

Seed Oil	Acetogenins	Reported Cytotoxic Activities
<i>Annona reticulata</i> seed oil *	bullatacin	- strong cytotoxic action against human cancer cell lines, including breast cancer (CCM2), mouth epidermal cancer (Hep.G2 and Hep.2, 3, 15), and liver cancer (Hep.G2 and Hep.2, 3, 15)
	cis-/trans-isomurisolenin	- cytotoxic for other cancer cell lines, including the HeLa and HeLa S3 cervical cancer cell lines, the SKOV3 and PA-1 ovarian cancer cell lines, the BCC-1 skin cancer cell line, and the MCF-7 breast cancer cell line
	cis-/trans-bullatacinone	- apoptosis was induced as a result of encouraging the production of the Bax protein and raising caspase 3 activity
	annoreticulin	- kills cells in several cancer cell lines, including T24 bladder cancer cells
	annoreticulin-9-one	- blocks the c-test
	cis-/trans-murisolenone	- cytotoxic component for almost all cancer cell lines
	squamocin	
<i>Annona squamosa</i> seed oil **	12, 15-cis-squamostatin-A	- significant anticancer capabilities against the cancer cells A-549, Hela, MCF-7, and HepG2, particularly for Hep G2 and MCF-7
	bullatacin	- taken orally to inhibit the growth of hepatic cancer cell (H22) lines in mice
	squadiolin A,	- significant anticancer activity in animals carrying H22 xenografts; its inhibitory rate was 53.54% against the growth of H22 cell lines
	squadiolin B,	- significantly damages MDA-MB-231
	squadfosacin B	- significantly cytotoxic to the breast cancer cells MDA-MB-231
		- extremely hazardous to human Hep G2 and 3B hepatoma cells as well as MCF-7 breast cancer cells, and it demonstrates cytotoxicity against HepG2, Hep 3B, and the MCF-7 cell line
<i>Annona muricata</i> Seed oil ***	muricins	- cytotoxic effects on leukemia cells CCRF-CEM
	muricatetrocin A	- significantly harms the human hepatoma cell lines Hep G2 and 2,2,15 in in vitro experiments
	muricatetrocin B	- in vitro tests using the human hepatoma cell lines Hep G2 and Hep 2.2.15 revealed strong antiproliferative effects
	longifolicin	
	corossolin	
	corossolone	
annotacin A and B		

\* Chang et al. [122]; Yuan et al. [123]; Yuan et al. [124]; \*\* Chen et al. [69]; Chen et al. [129]; Liaw et al. [132]; Yang et al. [133]; \*\*\* Kuete et al. [134]; Chang et al. [135].

The total acetogenin content in ASSO was 41.00 mg/g, and it exhibited remarkable cytotoxic activities against human tumor cell lines in a study and helped treat cancers of the liver, cervix, pancreas, etc. [69,129]. The study showed that various acetogenin types exhibit various inhibitory effects against various cancer cells (Table 4), and they have been shown to be the major bioactive compounds that possess strong antitumor/anticancer activity [112]. Using high-performance liquid chromatography (HPLC), two significant acetogenins have been identified in ASSO: 12, 15-cis-squamostatin-A (47.98 mg/g) and bullatacin (256.18 mg/g). Other constituents with antitumor activities include annosquacin A, B, and C, annosquatin A and B, squamostanin A, B, and D, squamostolide, and uvarigrandin A. Although it has been established that the fundamental mechanism is the stimulation of apoptosis, ASSO shows selectivity for certain cancer cells. Free radical production is also considered to be a key mechanism for the anticancer action of seed oils [129,133].



**Figure 3.** Structures of a few phytochemicals present in ASSO, ARSO, and AMSO [136].

Cancer cells are frequently treated with *A. muricata*, and the entire plant has a promising future as a chemotherapeutic treatment for cancer, according to research. AMSO contains about thirty-seven phenolic compounds, such as acetogenins, alkaloids, phenolic compounds, etc. [137]. In addition to the vitamins and carotenes found in seeds, there have also been shown to be 37 volatile chemicals and 18 essential oils. It improves the capacity of non-cancerous cells in the body. Higher levels of flavonoids are present, which are crucial for reducing cancer cell migration and cellular proliferation, both of which cause a response to reactive oxygen species (ROS) [138]. AMSO-containing acetogenins play a crucial role in preventing cancers (Table 4). Soursop seeds are low in toxins such as tannins, phytates, and cyanides and are high in oil and vitamins [53]. Accordingly, ASOs have huge potential to be used in chemoprevention, which needs to be researched further.

#### 4.3. Antioxidant Activity of Annona Seed Oils

Natural antioxidants include nitrogen compounds such as alkaloids, amino acids, peptides, and amines and derivatives of chlorophyll, carotenoids, and ascorbic acid, as well as phenolic compounds such as flavonoids, phenolic acids, and tannins [139]. Food antioxidants have a significant impact on maintaining good health and are directly linked to the prevention of degenerative diseases like cancer, cardiovascular disease, and neurological disorders [140]. Free radicals such as peroxide, hydroperoxide, or lipid peroxy, which

are harmful to the body, causing cancer, aging, and cellular damage, are scavenged by antioxidants and prevent the oxidative processes that cause degenerative illnesses [141]. Omega-3 (n-3) UFAs have been linked to cancer prevention [142]. Reactive oxygen species (ROS), also known as oxygen free radicals, play a dual role in biological systems, where both beneficial and detrimental effects are possible [143].

Different antioxidant screening methods were used to examine Annona's capacity to scavenge free radicals, and the findings showed that ASO has significant antioxidants. Additional research may result in the development of powerful antioxidant agents from ASOs [110,144]. Antioxidant-based chemoprevention has been proposed to have a promising future in the provision of significant fundamental advantages to public health. Many scientists and medical professionals are now considering this as a crucial tactic for preventing, postponing, or even stopping the development of cancer [145]. Although numerous research studies have found a correlation between plant phenolic content and antioxidant capacity, it was noted that data about the antioxidant potential of ASO are rare to find [146]. To develop products, cosmetics, nutraceuticals, and biopharmaceuticals to combat cancer and to add to the database of highly significant medicinal plants, further investigation is needed to identify the bioactive compounds responsible for antioxidant activity [147].

Using a DPPH assay, a study found that ASSO has antioxidant activity, with an IC50 value of 7.88 g/mL [130]. In a different investigation, the antioxidant activity of oral-administered ASSO was evaluated in rats (150–210 g) with alcohol-induced liver damage. Superoxide dismutase, glutathione, and catalase levels were shown to be significantly raised following treatment with the ethanolic seed extract [77]. As a result, ASSO therapy results in the restoration of normal antioxidant enzymes. Through oral administration, ASSO reduced the development of H22 tumor cells in mice by a maximum inhibitory rate of 53.54% [69].

In addition to the six previously reported cytotoxic acetogenins annoreticuin, annoreticuin-9-one, cis-/trans-bullatacinone, bullatacin, cis-/trans-murisolinone, and squamocin, cis-/trans-isomurisolenin was also discovered [146,148]. There have also been reports of spathenolol, muurolene, copaene, and eudesmol, among other terpenes. Cyclohexapeptide glabrin A was recovered from a methanol extract of ARSO, and novel cyclooctapeptides were discovered by analyzing the sequence and three-dimensional structure of cycloreticulins A and B. N-fatty acyl tryptamines were also obtained [148,149]. AMSOs contain a considerable amount of antioxidants, and the antioxidant activity of soursop seed oil was 77.34% (dry basis) [150].

Mother nature has provided us with many naturally occurring medicinal plants, yet we often use them without realizing their therapeutic value [151]. Several medicinal plants have been examined scientifically, and it has been found that their secondary metabolites and bioactive chemicals have the potential to have an anticancer impact. The best example of this is the *Annona* species. The chosen *A. squamosa*, *A. muricata*, and *A. reticulata* have also grown in favor as a result of more modern research being conducted on the bioactivities and health benefits of various plant parts, such as the seeds, bark, leaves, fruits, etc. Although the seeds of the fruit are usually waste and the fruit itself is edible, literature reviews have reported that ASO primarily consists of fatty acids, phytochemicals, and antioxidants like polyketides, annonaceous acetogenins (neurotoxins), cyclopeptides, carbohydrates, proteins, lipids, oleic acid, and linoleic acid. The phytochemical composition and molecular basis of the bioactivities of Annona seeds have been the subject of a few studies. The demand for innovative treatments is growing as a result of the wide variety of cancer types and their primary causes. A novel cancer medication that can target and prevent apoptosis in cancer cells is the subject of intense research efforts today. Accordingly, it has been noted that plants of the same species collected from various areas have varying levels of secondary metabolites, indicating that the synthesis of a plant's bioactive components may also vary, impacting how effective it is against cancer cells [16,152].

In conventional medicines, Annona is regarded as an ideal source of pharmacologically active compounds. Several prior investigations on the biological functions of the various

*Annona* spp. components have been conducted [153,154]. *Annona* seed oil contains phytochemicals, antioxidants, and fatty acids found in ARSO, AMO, and ASSO, which are employed for the treatment of cancer. In vitro and in vivo studies that show their usefulness in chemoprevention have been explored, as have anticancer agents. Fatty acid-derived lipids serve as the building blocks for cell membrane formation in addition to being crucial for hormone and signal transmission [155]. Due to their rapid proliferation and division, tumor cells require a lot of FAs. FAs make up a large portion of ASO. Tumor cells will undoubtedly absorb more ASO. Additionally, ASO primarily enters cells through passive transport [156].

## 5. Conclusions

ASOs were found to be effective in key bioactivities, including antioxidant activity and antitumor/anticancer activity, and based on in vivo and in vitro investigations, secondary metabolites of ASO cause anticancer and other immune system-associated effects. This review outlined the many pharmacological effects, particularly the aforementioned species' anticancer properties. To undertake thorough investigations and gain a better grasp of *Annona*'s anticancer potential, more studies will eventually need to be conducted. The bioactive substances that contribute to its bioactivities have also not been correctly identified or qualitatively and quantitatively studied as chemical markers for standardization and quality control purposes, and their mechanisms of action need to be adequately established. Therefore, there is a ton of room for further research into the phytochemical and pharmacological properties of ASOs to support their legitimate place in evidence-based chemopreventative medications. Hence, to isolate and identify the active metabolites that contribute to their robust anti-inflammatory and anticancer actions, future research on ASOs should concentrate on broad phytochemical investigations. Before being submitted to clinical trials to determine their safest concentration, their active metabolites should also be subjected to more mechanistic studies, in vivo and in vitro investigations, and toxicity tests. Further improvements to available treatment methods need to continue to allow for the development of more efficient but less expensive treatments against cancer.

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## References

1. Momin, K.C.; Sangma, A.N.; Suresh, S.P.; Singh, Y.S.; Rao, R. A potential nutraceutical and therapeutic fruit plant. *Int. J. Minor Fruit. Med. Aromat. Plants* **2018**, *4*, 44–49.
2. Dahanayake, N. Some neglected and underutilized fruit crops in Sri Lanka. *Int. J. Sci. Res. Pub.* **2015**, *5*, 1–7.
3. Sousa, C.M.M.; Silva, H.R.; Ayres, M.C.C.; Costa, C.L.S.; Araújo, D.S.; Cavalcante, L.C.D.; Barros, E.D.S.; Araújo, P.B.D.M.; Brandão, M.S.; Chaves, M.H. Total phenolics and antioxidant activity of five medicinal plants. *Quím. Nova* **2007**, *30*, 351–355. [[CrossRef](#)]
4. Kessler, P.J.A. Annonaceae. In *Flowering Plants-Dicotyledons; The Families and Genera of Vascular Plants*; Kubitzki, K., Rohwer, J.G., Bittrich, V., Eds.; Springer: Berlin/Heidelberg, Germany, 1993; Volume 2, pp. 93–129. [[CrossRef](#)]
5. Wahab, S.M.A.; Jantan, I.; Haque, M.A.; Arshad, L. Exploring the leaves of *Annona muricata* L. as a source of potential anti-inflammatory and anticancer agents. *Front. Pharm.* **2018**, *9*, 661. [[CrossRef](#)]
6. Jansen, P.; Lemmens, R.; Oyen, L. *Plant Resources South-East Asia: Basic List of Species Commodity Grouping*; Final Version; Pudoc: Wageningen, The Netherlands, 1991.
7. Pandey, N.; Barve, D. Phytochemical and Pharmacological Review on *Annona squamosa* Linn. *Int. J. Res. Pharm. Biomed. Sci.* **2011**, *2*, 1404–1412.

8. Bele, M.Y.; Focho, D.A.; Egbe, E.A.; Chuyong, B.G. Ethnobotanical survey of the uses Annonaceae around mount Cameroon. *Afr. J. Plant Sci.* **2011**, *5*, 237–247.
9. Attiq, A.; Jalil, J.; Husain, K. Annonaceae: Breaking the wall of inflammation. *Front. Pharmacol.* **2017**, *8*, 752. [[CrossRef](#)]
10. Zubaidi, S.N.; Mohd Nani, H.; Ahmad Kamal, M.S.; Abdul Qayyum, T.; Maarof, S.; Afzan, A.; Mohmad Misnan, N.; Hamezah, H.S.; Baharum, S.N.; Mediani, A. *Annona muricata*: Comprehensive Review on the Ethnomedicinal, Phytochemistry, and Pharmacological Aspects Focusing on Antidiabetic Properties. *Life* **2023**, *13*, 353. [[CrossRef](#)]
11. Morton, J.F.; Dowling, C.F. *Fruits of Warm Climates (Sugar Apple)*; Scientific Research: Miami, FL, USA, 1987; pp. 69–72. Available online: <http://www.hort.purdue.edu/newcrop/mortonne/roselle.html> (accessed on 1 June 2023).
12. Siebra, C.A.; Nardin, J.M.; Florão, A.; Rocha, F.H.; Bastos, D.Z.; Oliveira, B.H.; Weffort-Santos, A.M. Potencial antiinflamatório de *Annona glabra*, Annonaceae. *Rev. Bras. Farmacogn.* **2009**, *19*, 82–88. [[CrossRef](#)]
13. Biba, V.; Amily, A.; Sangeetha, S.; Remani, P. Anticancer, antioxidant and antimicrobial activity of Annonaceae family. *World J. Pharm. Sci.* **2014**, *3*, 1595–1604. [[CrossRef](#)]
14. González-Trujano, M.; Navarrete, A.; Reyes, B.; Hong, E. Some pharmacological effects of the ethanol extract of leaves of *Annona diversifolia* on the central nervous system in mice. *Phytother. Res.* **1998**, *12*, 600–602. [[CrossRef](#)]
15. Oliveira, G.N.D.S.A.; Dutra, L.M.; Paz, W.H.P.; da Silva, F.M.A.; Costa, E.V.; da Silva, A.J.R.G. Chemical constituents from the leaves and branches of *Annona coriacea* Mart. (Annonaceae). *Biochem. Syst. Ecol.* **2021**, *97*, 104297. [[CrossRef](#)]
16. Nugraha, A.S.; Damayanti, Y.D.; Wangchuk, P.; Keller, P.A. Anti-infective and anti-cancer properties of the Annona species: Their ethnomedicinal uses, alkaloid diversity, and pharmacological activities. *Molecules* **2019**, *24*, 4419. [[CrossRef](#)]
17. Takahashi, J.A.; Pereira, C.R.; Pimenta, L.P.; Boaventura, M.A.D.; Silva, L.G.E. Antibacterial activity of eight Brazilian Annonaceae plants. *Nat. Prod. Res.* **2006**, *20*, 21–26. [[CrossRef](#)]
18. Ajaiyeoba, E.; Falade, M.; Ogbale, O.; Okpako, L.; Akinboye, D. In vivo antimalarial and cytotoxic properties of *Annona senegalensis* extract. *J. Tradit. Complement. Med.* **2006**, *3*, 137–141. [[CrossRef](#)]
19. Afroz, N.; Hoq, M.A.; Jahan, S.; Islam, M.M.; Ahmed, F.; Shahid-Ud-Daula, A.; Hasanuzzaman, M. Methanol soluble fraction of fruits of *Annona muricata* possesses significant antiarrhythmic activities. *Heliyon* **2020**, *6*, e03112. [[CrossRef](#)] [[PubMed](#)]
20. Panda, S.; Kar, A. Antidiabetic and antioxidative effects of *Annona squamosa* leaves are possibly mediated through quercetin-3-O-glucoside. *Biofactors* **2007**, *31*, 201–210. [[CrossRef](#)] [[PubMed](#)]
21. Rocha, R.S.; Kassuya, C.A.L.; Formagio, A.S.N.; Mauro, M.D.O.; Andrade-Silva, M.; Monreal, A.C.D.; Cunha-Laura, A.L.; Vieira, M.D.C.; Oliveira, R.J. Analysis of the anti-inflammatory and chemopreventive potential and description of the antimutagenic mode of action of the *Annona crassiflora* methanolic extract. *Pharm. Biol.* **2016**, *54*, 35–47. [[CrossRef](#)] [[PubMed](#)]
22. Essama, S.R.; Nyegue, M.; Foe, C.N.; Silihe, K.K.; Tamo, S.B.; Etoa, F. Antibacterial and antioxidant activities of hydro-ethanol extracts of barks, leaves and stems of *Annona muricata*. *Am. J. Pharmacol. Sci.* **2015**, *3*, 126–131.
23. Nakano, D.; Ishitsuka, K.; Kamikawa, M.; Matsuda, M.; Tsuchihashi, R.; Okawa, M.; Okabe, H.; Tamura, K.; Kinjo, J. Screening of promising chemotherapeutic candidates from plants against human adult T-cell leukemia/lymphoma (III). *J. Nat. Med.* **2013**, *67*, 894–903. [[CrossRef](#)] [[PubMed](#)]
24. Osorio, E.; Arango, G.J.; Jiménez, N.; Alzate, F.; Ruiz, G.; Gutiérrez, D.; Paco, M.A.; Giménez, A.; Robledo, S. Antiprotozoal and cytotoxic activities in vitro of Colombian Annonaceae. *J. Ethnopharmacol.* **2007**, *111*, 630–635. [[CrossRef](#)]
25. Martínez-Vázquez, M.; Diana, G.; Estrada-Reyes, R.; González-Lugo, N.M.; Apan, T.R.; Heinze, G. Bio-guided isolation of the cytotoxic corytenchine and isocoreximine from roots of *Annona cherimolia*. *Fitoterapia* **2005**, *76*, 733–736. [[CrossRef](#)]
26. Castillo-Juárez, I.; González, V.; Jaime-Aguilar, H.; Martínez, G.; Linares, E.; Bye, R.; Romero, I. Anti-helicobacter pylori activity of plants used in Mexican traditional medicine for gastrointestinal disorders. *J. Ethnopharmacol.* **2009**, *122*, 402–405. [[CrossRef](#)] [[PubMed](#)]
27. Baskar, R.; Rajeswari, V.; Kumar, T.S. In vitro antioxidant studies in leaves of *Annona* species. *Indian J. Exp. Biol.* **2007**, *45*, 480–485. [[PubMed](#)]
28. Al Kazman, B.S.M.; Harnett, J.E.; Hanrahan, J.R. Traditional Uses, Phytochemistry and Pharmacological Activities of Annonaceae. *Molecules* **2022**, *27*, 3462. [[CrossRef](#)] [[PubMed](#)]
29. Rabelo, S.V.; Quintans, J.d.S.S.; Costa, E.V.; da Silva Almeida, J.R.G.; Júnior, L.J.Q. *Annona Species (Annonaceae) Oils*; Academic Press: Cambridge, MA, USA, 2015; pp. 221–229.
30. Tamang, A.; Subba, S.K.; Chhetri, S. Wild edible and minor fruits of Odisha. *Pharma Innov. J.* **2021**, *10*, 609–613.
31. Nyam, K.L.; Tan, C.P.; Lai, O.M.; Long, K.; Che Man, Y.B. Physicochemical properties and bioactive compounds of selected seed oils. *LWT—Food Sci. Technol.* **2009**, *42*, 1396–1403. [[CrossRef](#)]
32. Guo, L.; Wang, S.; Zhang, J.; Yang, G.; Zhao, M.; Ma, W.; Zhang, X.; Li, X.; Han, B.; Chen, N.; et al. Effects of ecological factors on secondary metabolites and inorganic elements of *Scutellaria baicalensis* and analysis of geo-herbalism. *Sci. China Life Sci.* **2013**, *56*, 1047–1056. [[CrossRef](#)] [[PubMed](#)]
33. Madhujith, T.; Shahidi, F. Antioxidant and Antiproliferative Potential of Pearled Barley (*Hordeum vulgare*). *Pharm. Biol.* **2008**, *46*, 88–95. [[CrossRef](#)]
34. Ranganathan, P.; Sengar, M.; Chinnaswamy, G.; Agrawal, G.; Arumugham, R.; Bhatt, R. Impact of COVID-19 on cancer care in India: A cohort study. *Lancet Oncol.* **2021**, *22*, 970–976. [[CrossRef](#)]
35. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics. *Cancer J. Clin.* **2018**, *68*, 7–30. [[CrossRef](#)] [[PubMed](#)]

36. Habibi, M.F.; Megdad, M.M.; Al-Qadi, M.H.; Al Qatrawi, M.J.; Sababa, R.Z.; Abu-Naser, S.S. A Proposed Expert System for Obstetrics & Gynecology Diseases Diagnosis. *Int. J. Acad. Multidiscip. Res.* **2022**, *6*, 305–321.
37. WHO. *Preventing Chronic Diseases: A Vital Investment*; World Health Organization: Geneva, Switzerland, 2020. [CrossRef]
38. Yan, Z.; Zhang, B.; Huang, Y.; Qiu, H.; Chen, P.; Guo, G. Involvement of autophagy inhibition in *Brucea javanica* oil emulsion induced colon cancer cell death. *Oncol. Lett.* **2015**, *9*, 1425–1431. [CrossRef] [PubMed]
39. Dev, A.R.A.; Joseph, S.M. Anticancer potential of *Annona* genus: A detailed review. *J. Indian Chem. Soc.* **2021**, *98*, 100231. [CrossRef]
40. Pinto, A.C.d.Q.; Corderio, M.C.R.; Andrade, S.R.M.; Ferreira, F.R.; Filguriras, H.A.d.C.; Alves, R.E.; Kinpara, D.I. *Annona* Species. In *International Centre for Underutilised Crops*; Williams, J.T., Ed.; University of Southampton: Southampton, UK, 2005; p. 268.
41. Quílez, A.; Fernández-Arche, M.; García-Giménez, M.; De la Puerta, R. Potential therapeutic applications of the genus *Annona*: Local and traditional uses and pharmacology. *J. Ethnopharmacol.* **2018**, *225*, 244–270. [CrossRef] [PubMed]
42. Rabêlo, S.V.; Quintans, J.S.S.; Costa, E.V.; Almeida, J.R.G.S.; Quintans-Júnior, L. *Essential Oils in Food Preservation, Flavor and Safety*; Elsevier: Amsterdam, The Netherlands, 2016; pp. 221–228. Available online: [https://www.researchgate.net/profile/Rosa-Mourao/publication/288807775\\_Amazon\\_Rosewood\\_Aniba\\_rosaeodora\\_Ducke\\_Oils/links/5b192a92aca272021ceedc49/Amazon-Rosewood-Aniba-rosaeodora-Ducke-Oils.pdf](https://www.researchgate.net/profile/Rosa-Mourao/publication/288807775_Amazon_Rosewood_Aniba_rosaeodora_Ducke_Oils/links/5b192a92aca272021ceedc49/Amazon-Rosewood-Aniba-rosaeodora-Ducke-Oils.pdf) (accessed on 13 March 2023).
43. Mia, M.A.B. Digital Herbarium of Crop Plants, Department of Crop Botany, Bangabandhu Sheikh Mujibur Rahman Agricultural University. 2016. Available online: <http://dhcrop.bsmrau.net/1879-2/> (accessed on 12 March 2022).
44. Dahanayake, N.; Geurts, F. *Annonaceous Fruits*; Royal Tropical Institute: Amsterdam, The Netherlands, 1981; p. 16. Available online: [https://assets.publishing.service.gov.uk/media/57a08c6640f0b652dd0012e6/R7187\\_-\\_Annona\\_monograph\\_-\\_revised.pdf](https://assets.publishing.service.gov.uk/media/57a08c6640f0b652dd0012e6/R7187_-_Annona_monograph_-_revised.pdf) (accessed on 14 July 2023).
45. Dilrukshi, M.K.D.T.; Dhamadasa, R.M.; Abeyasinghe, D.C.; Abhayagunasekara, A.V.C. Selection of Superior Quality *Annona* Species by means of Bioactive Compounds and Antioxidant Capacity. *World J. Agric. Res.* **2020**, *8*, 39–44.
46. Heenkenda, H.M.S.; Pushpakumara, D.K.N.G.; Ranil, R.H.G.; Thantirige, M.J.K. Chapter 9: *Annona*, *Annona* species. In *Underutilized Fruit Trees in Sri Lanka*; Pushpakumara, D.K.N.G., Gunasena, H.P.M., Singh, V.P., Eds.; World Agroforestry Centre, South Asia Office: New Delhi, India, 2011; Volume 2, pp. 158–182. [CrossRef]
47. Abdulrahman, H.; Kumar, R.S. Antibacterial activity of *Annona squamosa* and *Annona reticulata* L. against clinical isolates of mutans streptococci the causative agents of dental caries. *Asian J. Pharm. Clin. Res.* **2015**, *8*, 152–155.
48. Encina, C.L.; Martin, E.C.; Lopez, A.A.; Padilla, I.M.G. Biotechnology applied to *Annona* spp.: A review. *Rev. Bras. Fruticult.* **2014**, *36*, 17–21. [CrossRef]
49. Daniel, Q. *Genetic Resources: Our Forgotten Treasure*; Third World Network: Geneva, Switzerland, 1992.
50. Jayaweera, D.M.A. *Medicinal Plants in Ceylon*; Part 1–5; National Science Council of Sri Lanka: Colombo, Sri Lanka, 1981.
51. De Pauda, L.S.; Lugod, G.; Pancho, J.V. *Handbook on Philippine Medicinal Plants*; University of Philippines: Quezon City, Philippines, 1997; Volume 1.
52. Okoro, C.K.; Osunde, Z.D. Physical Properties Of Soursop (*Annona muricata*) Seeds. *Int. J. Eng. Res. Technol.* **2013**, *2*, 1–4.
53. Badrie, N.; Schauss, A. Soursop (*Annona muricata* L.): Composition, nutritional value, medicinal uses, and toxicology. *Bioact. Food Prom. Health* **2010**, *39*, 621–643. [CrossRef]
54. Jamkhande, P.G.; Wattamwar, A.S. *Annona reticulata* Linn. (Bullock’s heart): Plant profile, phytochemistry and pharmacological properties. *J. Tradit. Complement. Med.* **2015**, *5*, 144–152. [CrossRef]
55. Rajapaksha, U. *Traditional Fruit Plants in Sri Lanka*; Hector Kobbekaduwa Agrarian Research and Training Institute: Colombo, Sri Lanka, 1998; pp. 56–64.
56. Handique, K.; Hazarika, D.N.; Langthasa, S.; Khanikar, H.B.; Deori, G.D. Morphological characterization of custard apple (*Annona reticulata*) grown in Brahmaputra valley of Assam. *Pharm. Innov. J.* **2022**, *11*, 684–688.
57. Martínez, M.F.; Miranda, L.D.; Magnitskiy, S. *Anatomy of Sugar Apple (Annona squamosa L.) Seeds (Annonaceae)* *Agronomía Colombiana*; Universidad Nacional de Colombia Bogotá: Bogotá, Colombia, 2013; Volume 31, pp. 279–287. Available online: <https://www.redalyc.org/articulo.oa?id=180329804003> (accessed on 14 June 2023).
58. Atique, A.; Iqbal, M.; Ghouse, A. Use of *Annona squamosa* and *Piper nigrum* against diabetes. *Fitoterapia* **1985**, *56*, 190–192.
59. Ahmed, R.H.A.; Mariod, A.A. *Annona squamosa*: Phytochemical Constituents, Bioactive Compounds, Traditional and Medicinal Uses. In *Wild Fruits: Composition, Nutritional Value and Products*; Springer: Berlin/Heidelberg, Germany, 2019; pp. 143–155. [CrossRef]
60. Onimawo, I.A. Proximate composition and selected physicochemical properties of the seed, pulp, and oil of soursop (*Annona muricata*). *Plant Foods Hum. Nutr.* **2002**, *57*, 165–171. [CrossRef]
61. Mariod, A.A.; Elkheir, S.; Ahmed, Y.M.; Matthäus, B. *Annona squamosa* and *Catunaregam nilotica* Seeds, the effect of the extraction method on the oil composition. *J. Am. Oil Chem. Soc.* **2010**, *87*, 763–769. [CrossRef]
62. Kumar, M.; Changan, S.; Tomar, M.; Prajapati, U.; Saurabh, V.; Hasan, M.; Sasi, M.; Maheshwari, C.; Singh, S.; Dhupal, S.; et al. Custard Apple (*Annona squamosa* L.) Leaves: Nutritional Composition, Phytochemical Profile, and Health-Promoting Biological Activities. *Biomolecules* **2021**, *11*, 614. [CrossRef] [PubMed]
63. Abdalbasit, A.M.; Ramlah, M.I.; Maznah, I.; Norsharina, I. Antioxidant activities of phenolic rich fractions (PRFs) obtained from black mahlab (*Monechma ciliatum*) and white mahlab (*Prunus mahaleb*) seedcakes. *Food Chem.* **2010**, *118*, 120–127. [CrossRef]

64. Williams, E.T.; Luka, K.; Timothy, N. Phytochemical, Elemental, Proximate, and Antinutrient Composition of Custard Apple Seed (*Annona reticulata*) from Maiha Adamawa State, Nigeria. *Glob. J. Med. Res. L Nutr. Food Sci.* **2020**, *20*, 19–24. Available online: <https://medicalresearchjournal.org/index.php/GJMR/article/download/2333/2222/> (accessed on 1 June 2023).
65. WHO; Food and Agriculture Organization of the United Nations; Joint FAO/WHO Expert Committee on Food Additives. *Evaluation of Certain Food Additives and Contaminants: Seventy-Fourth Report of the Joint FAO/WHO Expert Committee on Food Additives*; World Health Organization: Geneva, Switzerland, 2011. Available online: <https://www.who.int/publications/i/item/9789241209601> (accessed on 1 June 2023).
66. Samaram, S.; Mirhosseini, H.; Tan, C.P.; Ghazali, H.M. Ultrasound-assisted extraction (UAE) and solvent extraction of papaya seed oil: Yield, fatty acid composition, and triacylglycerol profile. *Molecules* **2013**, *18*, 12474–12487. [[CrossRef](#)]
67. Liu, N.; Ren, G.; Faiza, M.; Li, D.; Cui, J.; Zhang, K.; Yao, X. Comparison of conventional and green extraction methods on oil yield, physicochemical properties, and lipid compositions of pomegranate seed oil. *J. Food Compos. Anal.* **2022**, *114*, 104747. [[CrossRef](#)]
68. Aremu, M.O.; Ibrahim, H.; Bamidele, T.O. Physicochemical Characteristics of the Oils Extracted from some Nigerian Plant Foods—A Review. *Chem. Process Eng. Res.* **2015**, *32*, 36–52.
69. Dąbrowski, G.; Czaplicki, S.; Konopka, I. Composition and quality of poppy (*Papaver somniferum* L.) seed oil depending on the extraction method. *LWT* **2020**, *134*, 10167. [[CrossRef](#)]
70. Ali, F.M.; Ali, B.E.; Speight, J.G. *Handbook of Industrial Chemistry: Organic Chemicals*; McGraw–Hill Education: New York, NY, USA, 2005.
71. Patili, S.M.; Gaykar, R.D.; Kuumbhar, G.B. Manufacturing of Natural Pesticide from Custard Apple Seeds. *Int. J. Innov. Res. Technol.* **2022**, *8*, 775–779.
72. Omkaresh, B.R.; Veeranna, K.; Yatish, K.V.; Ibham, V.; Pramoda, K. Optimization and kinetics analysis of biodiesel production from *Annona reticulata* seed oil using magnesium phosphate catalyst. *Braz. J. Chem. Engin* **2022**, *40*, 775–787. [[CrossRef](#)]
73. Rana, V.S. Fatty Oil and Fatty Acid Composition of *Annona squamosa* Linn. Seed Kernels. *Int. J. Fruit Sci.* **2014**, *15*, 79–84. [[CrossRef](#)]
74. Ansari, M.H.; Afaque, S.; Ahmad, M. Isoricinoleic acid in *Annona squamosa* seed oil. *J. Am. Oil Chem. Soc.* **1985**, *62*, 1514. [[CrossRef](#)]
75. Ahmad, S.; Naqvi, F.; Sharmin, E.; Verma, K.L. Development of amine acid cured *Annona squamosa* oil epoxy anticorrosive polymeric coatings. *Prog. Org. Coatings.* **2006**, *55*, 268–275. [[CrossRef](#)]
76. Ao, H.; Lu, L.; Li, M.; Han, M.; Guo, Y.; Wang, X. Enhanced Solubility and Antitumor Activity of *Annona Squamosa* Seed Oil via Nanoparticles Stabilized with TPGS: Preparation and In Vitro and In Vivo Evaluation. *Pharmaceutics* **2022**, *14*, 1232. [[CrossRef](#)]
77. Zahid, M.; Arif, M.; Rahman, M.A.; Singh, K.; Mujahid, M. Solvent extraction and gas chromatography–mass spectrometry analysis of *Annona squamosa* L. seeds for determination of bioactive, fatty acid/fatty oil composition, and antioxidant activity. *J. Diet. Suppl.* **2018**, *15*, 613–623. [[CrossRef](#)] [[PubMed](#)]
78. Kimbonguila, A.; Nzikou, J.M.; Matos, L.; Loumouamou, B.; Ndangui, C.B.; Pambou-Tobi, N.P.G.; Abena, A.A.; Sliou, T.; Scher, J.; Desobry, S. Proximate Composition and Physicochemical Properties on the Seeds and Oil of *Annona muricata* grown In Congo-Brazzaville. *Res. J. Environ. Earth Sci.* **2010**, *2*, 13–18.
79. Ma, F.; Hanna, M.A. Biodiesel production: A review. *Bioresour. Technol.* **1999**, *70*, 15. [[CrossRef](#)]
80. Talpur, M.; Sherazi, S.T.; Mahesar, S.; Kandhro, A. Effects of Chicken Frying on Soybean, Sunflower and Canola Oils. *Pak. J. Anal. Environ. Chem.* **2009**, *10*, 59–66.
81. Kalo, P.; Kemppinen, A. TRIGLYCERIDES | Structures and Properties. In *Encyclopedia of Food Sciences and Nutrition*, 2nd ed.; Caballero, B., Ed.; Academic Press: Cambridge, MA, USA, 2003; pp. 5857–5868. [[CrossRef](#)]
82. Boelhouwer, C.; Mol, J.C. Metathesis of fatty acid esters. *J. Am. Oil Chem. Soc.* **1984**, *61*, 425–430. [[CrossRef](#)]
83. Lee, S.C.; Chan, J. Evidence for DNA damage as a biological link between diabetes and cancer. *Chin. Med. J.* **2015**, *128*, 1543–1548. [[CrossRef](#)]
84. Barbalho, S.; de Goulart, R.; Machado, F.V.F.; da Soares de Souza, M.; Santos Bueno, P.; Guiguer, E.; Araujo, A.; Groppo, M. *Annona* sp: Plants with Multiple Applications as Alternative Medicine—A Review. *Curr. Bioact. Compd.* **2012**, *8*, 277–286. [[CrossRef](#)]
85. Asare, G.A.; Afriyie, D.; Ngala, R.A.; Abutiati, H.; Doku, D.; Mahmood, S.A.; Rahman, H. Antiproliferative activity of aqueous leaf extract of *Annona muricata* L. on the prostate, BPH-1 cells, and some target genes. *Integer. Cancer Ther.* **2015**, *14*, 65–74. [[CrossRef](#)] [[PubMed](#)]
86. Taylor, P.; Arsenak, M.; Abad, M.J.; Fernandez, A.; Milano, B.; Gonto, R.; Ruiz, M.C. Screening of Venezuelan medicinal plant extracts for cytostatic and cytotoxic activity against Tumor cell lines. *Phytother. Res.* **2013**, *27*, 530–539. [[CrossRef](#)] [[PubMed](#)]
87. Chavan, S.S.; Shamkuwar, P.B.; Damale, M.G.; Pawar, D.P. A comprehensive review on *Annona reticulata*. *Int. J. Pharm. Sci. Res.* **2014**, *5*, 45–50. [[CrossRef](#)]
88. Mishra, S.; Ahmad, S.; Kumar, N.; Sharma, B.K. *Annona muricata* (the cancer killer): A Review. *Glob. J. Pharm. Res.* **2013**, *2*, 1613–1618. [[CrossRef](#)]
89. Lima, L.A.R.S.; Johann, S.; Cisalpino, P.S.; Pimenta, L.P.S.; Boaventura, M.A.D. In vitro antifungal activity of fatty acid methyl esters of the seeds of *Annona cornifolia* A. St.-Hil. (Annonaceae) against pathogenic fungus *Paracoccidioides brasiliensis*. *J. Braz. Trop. Med.* **2011**, *44*, 777–780. [[CrossRef](#)]
90. Chowdhury, K.; Banu, L.A.; Khan, S.; Latif, A. Studies on the fatty acid composition of edible oils. *Bangladesh J. Sci. Ind. Res.* **2007**, *42*, 311–316. [[CrossRef](#)]

91. Akbar, E.; Yaakob, Z.; Kamarudin, S.K.; Smail, M.; Salimon, J. Characteristic and composition of the *Jatropha curcas* oil seed from Malaysia and its potential as biodiesel feedstock. *Eur. J. Sci. Res.* **2009**, *29*, 39–404.
92. Thang, T.D.; Kuo, P.C.; Huang, G.J.; Hung, N.H.; Huang, B.S.; Yang, M.L. Chemical Constituents from the Leaves of *Annona reticulata* and Their Inhibitory Effects on NO Production. *Molecules* **2013**, *18*, 4477–4486. [[CrossRef](#)]
93. Valantina, S.R.; Neelamegan, P. Antioxidant potential in vegetable oil—A review paper. *Res. J. Chem. Environ.* **2012**, *16*, 87–94.
94. Parry, J.; Su, L.; Luther, M.; Zhou, K.; Yurawecz, M.P.; Whittaker, P.; Yu, L. Fatty acid composition and antioxidant properties of cold-pressed marionberry, boysenberry, redraspberry, and blueberry seed oils. *J. Agri. Food Chem.* **2005**, *53*, 566–573. [[CrossRef](#)]
95. Mabaleha, M.B.; Mitei, Y.C.; Yeboah, S.O. A comparative study of the properties of selected melon seed oils as potential candidates for development into commercial edible vegetable oils. *J. Am. Oil Chem. Soc.* **2007**, *84*, 31–36. [[CrossRef](#)]
96. Anhwange, B.A.; Ajibola, V.; Oniye, S.J. Chemical studies of the seeds *Moringa oleifera* (LAM) detarium mirocarpum. *J. Bio. Sci.* **2004**, *4*, 711–715. [[CrossRef](#)]
97. Nie, Y.L.; Liu, K.X.; Mao, X.Y.; Li, Y.L.; Li, J.; Zhang, M.M. Effect of injection of *Brucea javanica* oil emulsion plus chemoradiotherapy for lung cancer: A review of clinical evidence. *J. Evid.-Based Med.* **2012**, *5*, 216–225. [[CrossRef](#)] [[PubMed](#)]
98. Liyanage, T.; Madhujith, T.; Wijesinghe, K.G.G. Comparative study on major chemical constituents in volatile oil of true cinnamon (*Cinnamomum verum*) and five wild cinnamon species grown in Sri Lanka. *Trop. Agri Res.* **2017**, *28*, 270–280. [[CrossRef](#)]
99. Jiang, W.G.; Bryce, R.P.; Horrobin, D.F. Essential fatty acids: Molecular and cellular basis of their anti-cancer action and clinical implications. *Crit. Rev. Oncol. Hematol.* **1998**, *27*, 179–209. [[CrossRef](#)] [[PubMed](#)]
100. Meterissian, S.H.; Forse, R.A.; Steele, G.D.; Thomas, P. Effect of membrane free fatty acid alterations on the adhesion of human colorectal carcinoma cells to liver macrophages and extracellular matrix proteins. *Cancer Lett.* **1995**, *89*, 145–152. [[CrossRef](#)]
101. Du Toit, P.J.; Van Aswegen, C.H.; Du Plessis, D.J. The effect of essential fatty acids on growth and urokinase-type plasminogen activator production in human prostate DU-145 cells, Prostaglandins Leukot. *Essent. Fat. Acids.* **1996**, *55*, 173–177. [[CrossRef](#)]
102. Soto-Guzman, A.; Villegas-Comonfort, S.; Cortes-Reynosa, P.; Perez Salazar, E. Role of arachidonic acid metabolism in Stat5 activation induced by oleic acid in MDA-MB-231 breast cancer cells. Prostaglandins. *Leukot. Essent. Fat. Acids.* **2013**, *88*, 243–249. [[CrossRef](#)]
103. Moon, H.S.; Batirel, S.; Mantzoros, C.S. Alpha linolenic acid and oleic acid additively down-regulate malignant potential and positively cross-regulate AMPK/S6 axis in OE19 and OE33 esophageal cancer cells. *Metabolism* **2014**, *63*, 1447–1454. [[CrossRef](#)]
104. Wang, Z.; Liu, D.; Zhang, Q.; Wang, J.; Zhan, J.; Xian, X.; Du, Z.; Wang, X.; Hao, A. Palmitic acid affects proliferation and differentiation of neural stem cells in vitro. *J. Neurosci. Res.* **2014**, *92*, 574–586. [[CrossRef](#)]
105. Chen, Y.; Chen, Y.; Shi, Y.; Ma, C.; Wang, X.; Li, Y.; Miao, Y.; Chen, J.; Li, X. Antitumor activity of *Annona squamosa* seed oil. *J. Ethnopharmacol.* **2016**, *193*, 362–367. [[CrossRef](#)]
106. Suffness, M.; Pezzuto, J.M. Assays related to cancer drug discovery. In *Methods in Plant Biochemistry: Assays for Bioactivity*; Hostettmann, K., Ed.; Academic Press: London, UK, 1990; pp. 71–133.
107. Fauser, J.K.; Prisciandaro, L.D.; Cummins, A.G.; Howarth, G.S. Fatty acids as potential adjunctive colorectal chemotherapeutic agents. *Cancer Biol. Ther.* **2011**, *11*, 724–731. [[CrossRef](#)]
108. Lima, L.A.R.S.; Alves, T.M.A.; Zani, C.L.; Pimenta, L.P.S.; Boaventura, M.A. Antioxidant and citotoxic potential of fatty acid methyl esters from the seeds of *Annona cornifolia* A. ST-Hil. (Annonaceae). *Food Res. Int.* **2012**, *48*, 873–875. [[CrossRef](#)]
109. Thiviya, P.; Gunawardena, N.; Gamage, A.; Madhujith, T.; Merah, O. Apiaceae Family as a Valuable Source of Biocidal Components and their Potential Uses in Agriculture. *Horticultrae* **2022**, *8*, 614. [[CrossRef](#)]
110. Adewole, S.; Ojewole, J. Protective effects of *Annona muricata* Linn. Leaf aqueous extract on serum lipid profiles and oxidative stress in hepatocytes of streptozotocin-treated diabetic rats. *Afr. J. Tradit. Complement. Altern. Med.* **2009**, *6*, 30–41. [[CrossRef](#)]
111. Kadarani, D.K.; Setyadjit, S.; Seno, D.S.H.; Sakashih, E. Total phenol & antioxidant from seed & peel of ripe and unripe of Indonesian sugar apple (*Annona squamosa* L.) extracted with various Pharmacyts IOSR. *J. Pharm.* **2013**, *5*, 20–25.
112. Eshra, D.H.; Shehata, A.R.; Ahmed, A.N.A.; Saber, J.I. Physicochemical Properties of the Seed Kernels and the Oil of Custard Apple (*Annona squamosa* L.). *Int. J. Food Biotech.* **2019**, *4*, 87–93. [[CrossRef](#)]
113. Sun, S.; Liu, J.; Zhou, N.; Zhu, W.; Dou, Q.P.; Zhou, K. Isolation of three new annonaceous acetogenins from Graviola fruit (*Annona muricata*) and their anti-proliferation on human prostate cancer cell PC-3. *Bioorg. Med. Chem. Lett.* **2016**, *26*, 4382–4385. [[CrossRef](#)]
114. Lannuzel, A.; Michel, P.P.; Höglinger, G.U.; Champy, P.; Jousset, A.; Medja, F.; Ruberg, M. The mitochondrial complex I inhibitor annonacin is toxic to mesencephalic dopaminergic neurons by impairment of energy metabolism. *Neuroscience* **2003**, *121*, 287–296. [[CrossRef](#)]
115. Alali, F.Q.; Xiao-Xi, L.; McLaughlin, J.L. Annonaceous acetogenins: Recent progress. *J. Nat. Prod.* **1999**, *62*, 504–540. [[CrossRef](#)]
116. Dey, P.; Kundu, A.; Kumar, A.; Gupta, M.; Lee, B.M.; Bhakta, T.; Dash, S.; Kim, H.S. Analysis of alkaloids (indole alkaloids, isoquinoline alkaloids, tropane alkaloids). In *Recent Advances in Natural Products Analysis*; Elsevier: Amsterdam, The Netherlands, 2020; pp. 505–567. [[CrossRef](#)]
117. Leboeuf, M.; Cavé, A.; Bhaumik, P.; Mukherjee, B.; Mukherjee, R. The phytochemistry of the Annonaceae. *Phytochemistry* **1980**, *21*, 2783–2813. [[CrossRef](#)]
118. Coria-Téllez, A.V.; Montalvo-González, E.; Yahia, E.M.; Obledo-Vázquez, E.N. *Annona muricata*: A comprehensive review on its traditional medicinal uses, phytochemicals, pharmacological activities, mechanisms of action and toxicity. *Arab. J. Chem.* **2016**, *11*, 662–691. [[CrossRef](#)]

119. Galeane, M.C.; Martins, C.H.G.; Massuco, J.; Bauab, T.M.; Sacramento, L.V.S. Phytochemical screening of *Azadirachta indica* A. Juss for antimicrobial activity. *Afr. J. Microbiol. Res.* **2017**, *11*, 117–122. [[CrossRef](#)]
120. Ramadass, N.; Subramanian, N.S. Study of phytochemical screening of neem (*Azadirachta indica*). *Int. J. Zool. Stud.* **2018**, *3*, 209–212.
121. James, J.; Veettil, A.K.T.; Pratyush, K.; Misra, C.S.; Sagadevan, L.D.M.; Thankamani, V. Phytochemical Investigation and Antibacterial Activity of the Fruits of *Alstonia Scholaris*. *Int. J. Phytopharm.* **2012**, *3*, 74–77.
122. Chang, F.R.; Wu, Y.C.; Duh, C.Y.; Wang, S.K. Studies on the acetogenins of Formosan annonaceous plants. II. Cytotoxic acetogenins from *Annona reticulata*. *J. Nat. Prod.* **1993**, *56*, 1688–1694. [[CrossRef](#)]
123. Yuan, S.S.; Chang, H.L.; Chen, H.W.; Yeh, Y.T.; Kao, Y.H.; Lin, K.H.; Wu, Y.C.; Su, J.H. Annonacin, a mono-tetrahydrofuran acetogenin, arrests cancer cells at the G1 phase and causes cytotoxicity in a Bax- and caspase-3-related pathway. *Life Sci.* **2003**, *72*, 2853–2861. [[CrossRef](#)]
124. Yuan, S.S.; Chang, H.L.; Chen, H.W.; Kuo, F.C.; Liaw, C.C.; Su, J.H.; Wu, Y.C. Selective cytotoxicity of squamocin. on T24 bladder cancer cells at the S-phase via Bax-, Bad-, and caspase-3-related pathways. *Life Sci.* **2006**, *78*, 869–874. [[CrossRef](#)]
125. Kintzios, S.E. Terrestrial plant-derived anticancer agents and plant species used in anticancer research. *Crit. Rev. Plant Sci.* **2006**, *25*, 79–113. [[CrossRef](#)]
126. Rahman, M.M.; Parvin, S.; Haque, M.E.; Islam, M.E.; Mosaddik, M.A. Antimicrobial and cytotoxic constituents from the seeds of *Annona squamosa*. *Fitoterapia* **2005**, *76*, 484–489. [[CrossRef](#)]
127. Shirwaikar, A.; Rajendran, K.; Kumar, C.D. Oral antidiabetic activity of *Annona squamosa* leaf alcohol extract in NIDDM rats. *Pharm. Biol.* **2004**, *42*, 30–35. [[CrossRef](#)]
128. Madhuri, S.; Pandey, G. Some anticancer medicinal plants of foreign origin. *Curr. Sci.* **2009**, *96*, 779–783.
129. Chen, Y.; Chen, J.W.; Zhai, J.H.; Wang, Y.; Wang, S.L.; Li, X. Antitumor activity and toxicity relationship of annonaceous acetogenins. *Food Chem. Toxicol.* **2013**, *58*, 394–400. [[CrossRef](#)] [[PubMed](#)]
130. Rakesh, R.; Mahendra, S. Coumarin lignans from the seeds of *Annona squamosa* Linn. *J. Chem.* **2009**, *6*, 518–522. [[CrossRef](#)]
131. Richmond, A.; Su, Y.J. Mouse xenograft models vs GEM models for human cancer therapeutics. *Dis. Models Mech.* **2008**, *1*, 78–82. [[CrossRef](#)] [[PubMed](#)]
132. Liaw, C.C.; Yang, Y.L.; Chen, M.; Chang, F.R.; Chen, S.L.; Wu, S.H.; Wu, Y.C. Mono-tetrahydrofuran annonaceous acetogenins from *Annona squamosa* as cytotoxic agents and calcium ion chelators. *J. Nat. Prod.* **2008**, *71*, 764–771. [[CrossRef](#)]
133. Yang, H.J.; Zhang, N.; Chen, J.W.; Wang, M.Y. Two new cytotoxic acetogenins from *Annona squamosa*. *J. Asian Nat. Prod. Res.* **2009**, *11*, 250–256. [[CrossRef](#)] [[PubMed](#)]
134. Kuete, V.; Dzutam, J.K.; Voukeng, I.K.; Fankam, A.G.; Efferth, T. Cytotoxicity of methanol extracts of *Annona muricata*, *Passiflora edulis*, and nine other Cameroonian medicinal plants towards multi-factorial drug-resistant cancer cell lines. *SpringerPlus* **2016**, *5*, 1666. [[CrossRef](#)]
135. Chang, F.R.; Wu, Y.C. Novel cytotoxic annonaceous acetogenins from *Annona muricata*. *J. Nat. Prod.* **2001**, *64*, 925–931. [[CrossRef](#)] [[PubMed](#)]
136. Kazman, B.S.M.; Al, J.E.; Harnett, A.; Jane, R.H. The Phytochemical Constituents and Pharmacological Activities of *Annona atemoya*: A Systematic Review. *Pharmaceuticals* **2020**, *13*, 269. [[CrossRef](#)] [[PubMed](#)]
137. Yang, C.; Gundala, S.R.; Mukkavilli, R.; Vangala, S.; Reid, M.D.; Aneja, R. Synergistic interactions among flavonoids and acetogenins in *Graviola* (*Annona muricata*) leaves confer protection against prostate cancer. *Carcinogenesis* **2015**, *36*, 656–665. [[CrossRef](#)]
138. Moghadamtousi, S.Z.; Fadaeinasab, M.; Nikzad, S.; Mohan, G.; Ali, H.M.; Kadir, H.A. *Annona muricata* (Annonaceae): A review of its traditional uses, isolated acetogenins, and biological activities. *Int. J. Mol. Sci.* **2015**, *16*, 15625. [[CrossRef](#)]
139. Hassimotto, N.M.A.; Genovese, M.E.; Lajolo, F.M. Antioxidant capacity of Brazilian fruit, vegetables and commercially-frozen fruit pulps. *J. Food Comp. Anal.* **2009**, *22*, 394–396. [[CrossRef](#)]
140. Diplock, A.T.; Miller, N.J.; Rice-Evans, C.A. Evaluation of the total antioxidant activity as a marker of the deterioration of apple juice on storage. *J. Agri Food Chem.* **1995**, *43*, 1794–1801. [[CrossRef](#)]
141. Halliwell, B. Free radicals and antioxidants. *Nutr. Rev.* **1994**, *52*, 253–265. [[CrossRef](#)]
142. Morris-Stiff, G.J.; Bowrey, D.J.; Oleesky, D.; Davies, M.; Clark, G.W.; Puntis, M.C. The antioxidant profiles of patients with recurrent acute and chronic pancreatitis. *Am. J. Gastroenterol.* **1999**, *94*, 2135–2140. [[CrossRef](#)]
143. Bailly, C.; El-Maarouf-Bouteau, H.; Corbineau, F. From intracellular signaling networks to cell death: The dual role of reactive oxygen species in seed physiology. *Comptes R. Biol.* **2008**, *331*, 806–814. [[CrossRef](#)]
144. Vikas, B.; Akhil, B.S.; Remani, P.; Sujathan, K. Free Radical Scavenging Properties of *Annona squamosa*. *Asian Pac. J. Cancer Prev.* **2017**, *18*, 2725–2731. [[CrossRef](#)] [[PubMed](#)]
145. Shureiqi, I.; Chen, D.; Lotan, R.; Yang, P.; Newman, R.A.; Fischer, S.M.; Lippman, S.M. 15-Lipoxygenase-1 mediates nonsteroidal anti-inflammatory drug-induced apoptosis independently of cyclooxygenase-2 in colon cancer cells. *Cancer Res.* **2000**, *60*, 6846–6850. [[PubMed](#)]
146. Shehata, M.G.; Abu-Serie, M.M.; El-Aziz, A.; Mohammad, N.; El-Sohaimy, S.A. Nutritional, phytochemical, and in vitro anticancer potential of sugar apple (*Annona squamosa*) fruits. *Sci. Rep.* **2021**, *11*, 6224. [[CrossRef](#)] [[PubMed](#)]
147. Ma, C.; Chen, Y.; Chen, J.; Li, X.; Chen, Y. A Review on *Annona squamosa* L.: Phytochemicals and Biological Activities. *Am. J. Chin. Med.* **2017**, *45*, 933–964. [[CrossRef](#)] [[PubMed](#)]

148. Chang, F.R.; Chen, J.L.; Chiu, H.F.; Wu, M.J.; Wu, Y.C. Acetogenins from seeds of *Annona reticulata*. *Phytochemistry* **1998**, *47*, 1057–1061. [[CrossRef](#)] [[PubMed](#)]
149. Meada, U.; Hara, N.; Fujimoto, Y.; Srivastava, A.; Gupta, Y.K.; Sahai, M. 'N-fatty Acyl Tryptamines from *Annona reticulata*. *Phytochemistry* **1993**, *34*, 1633–1635. [[CrossRef](#)]
150. Silva, A.C.; Jorge, N. Bioactive compounds of the liquid fractions of agro industrial waste. *Food Res. Int.* **2014**, *66*, 493–500. [[CrossRef](#)]
151. Pathirana, C.K.; Madhujith, T.; Eeswara, J. Bael (*Aegle marmelos* L. Corrêa), a Medicinal Tree with Immense Economic Potentials. *Adv. Agric.* **2020**, *2020*, 8814018. [[CrossRef](#)]
152. Shital, P.; Rujuta, A.; Sanjay, M. Transbronchial needle aspiration cytology (TBNA) in endobronchial lesions: A valuable technique during bronchoscopy in diagnosing lung cancer and it will decrease repeat bronchoscopy. *J. Cancer Res. Clin. Oncol.* **2014**, *140*, 809–815. [[CrossRef](#)] [[PubMed](#)]
153. Fridlender, M.; Kapulnik, Y.; Koltai, H. Plant-derived substances with anti-cancer activity: From folklore to practice. *Front. Plant Sci.* **2015**, *6*, 799. [[CrossRef](#)]
154. Weerasinghe, M.G.W.K.; Dahanayake, N. A review of *Annona* species in Sri Lanka. *Int. J. Minor. Fruits Med. Aromat. Plants* **2022**, *8*, 1–6. [[CrossRef](#)]
155. Santos, C.R.; Schulze, A. Lipid metabolism in cancer. *FEBS J.* **2012**, *279*, 2610–2623. [[CrossRef](#)] [[PubMed](#)]
156. Dallavalle, S.; Dobricic, V.; Lazzarato, L.; Gazzano, E.; Machuqueiro, M.; Pajeva, I.; Tsakovska, I.; Zidar, N.; Fruttero, R. Improvement of conventional anti-cancer drugs as new tools against multidrug resistant tumors. *Drug Resist. Updates* **2020**, *50*, 100682. [[CrossRef](#)] [[PubMed](#)]

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