OPEN ACCESS **MOLECULES** ISSN 1420-3049 www.mdpi.com/journal/molecules

Article

Design, Synthesis and Anti-HIV Integrase Evaluation of 4-Oxo-4*H*-quinolizine-3-carboxylic Acid Derivatives

Yi-Sheng Xu^{1,2}, Cheng-Chu Zeng^{1,*}, Zi-Guo Jiao¹, Li-Ming Hu¹ and Ru-gang Zhong¹

- ¹ College of Life Science and Bio-Engineering, Beijing University of Technology, Beijing 100124,
 P. R. China
- ² Atmospheric Chemistry & Aerosol Research Department, Chinese Research Academy of Environmental Sciences, Beijing 100012, P. R. China; E-mails: xuys@craes.org.cn (Y-S.X)
- * Author to whom correspondence should be addressed; E-Mail: zengcc@bjut.edu.cn; Tel.: +86-10-67396211; Fax: +86-10-67392001.

Received: 30 December 2008; in revised form: 16 January 2009 / Accepted: 6 February 2009 / Published: 19 January 2009

Abstract: 4-Oxo-4*H*-quinolizine-3-carboxylic acid derivatives bearing sulfamido, carboxylamido, benzimidazole and benzothiazole substituents have been designed and synthesized. The structures of these new compounds were confirmed by ¹H-NMR, ¹³C-NMR, IR and ESI (or HRMS) spectra. Compounds were screened for possible HIV integrase inhibitory activity.

Keywords: 4-Oxo-4*H*-quinolizine-3-carboxylic acid derivatives; HIV-1 integrase; Mg^{2+} binding; Aryl diketo acids.

Introduction

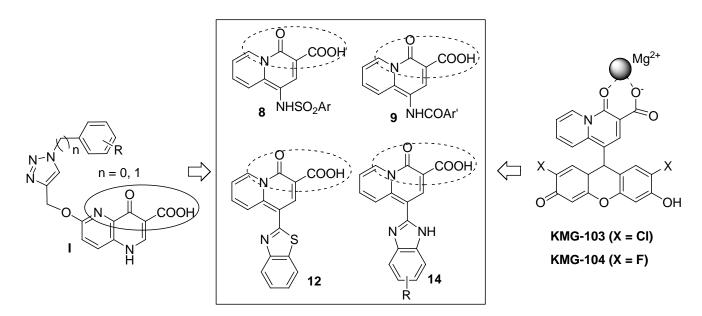
Human immunodeficiency virus type 1 (HIV-1) is the major pathogen of the infection of acquired immunodeficiency syndrome (AIDS) [1]. In the past two and half decades, various compounds have been developed as medicinal candidates for the treatment of HIV infections aiming at one or several steps of the HIV-1 life cycle [2-5] such as absorption, entry, fusion, un-coating, reverse transcription, integration, transcription and maturation. Owing to the facts that HIV integrase (IN) is the indispensable enzyme in the replication of the life cycle for the HIV virus and no cellular homologues

are found in humans [6-9], inhibitors targeted selectively at HIV IN are expected to have low cytotoxicity and thus development of HIV IN inhibitors has attracted more and more attention.

To this end, aryl diketoacids compounds have been reported to be the most promising HIV IN inhibitors [10-12]. Structurally, this type of HIV IN inhibitors consists of two main domains, namely, a diketoacid subunit and an aromatic moiety with one or two arylalkyl substituents. The former serves as the pharmaphore and is proposed to sequester the divalent cofactors (Mg^{2+}) in the IN active site and thus block access of a host DNA to the integrase [13,14]. Meanwhile, the aromatic moiety as hydrophobic domain, typically an indole or benzene moiety, is responsible for the selectivity to the strand transfer (ST) reaction [15,16].

As part of our continuing efforts directed towards the development of potential HIV-1 integrase inhibitors, we have investigated the chemical and electrochemical synthesis of styrylquinolines [17], quinoxalones [18,19] and polyhydroxylated aromatics [20,21]. Based on the structural characteristics of diketo acid-type compounds and their molecular mechanism as HIV IN inhibitors, very recently, a series of 4-oxo-1,4-dihydro-1,5-naphthyridine-3-carboxylic acids have been designed and synthesized by our group (see I, Figure 1) [22]. In view of the fact that the 4-oxo-quinolizine-3-carboxylic acid moiety displays high selectivity and binding ability to Mg(II) (KMG-103 and KMG-104, Figure 1) [23,24] and other derivatives show antibacterial and antimicrobial activities [25-27], in the present work, we further modified the main core via addition of a bulky group, such as an aryl sulfonamido, aryl carboxylamido, benzimidazole or benzothiazole chosen as the side hydrophobic chain and placed at the C-1 position of the 4-oxo-quinolizine-3-carboxylic acid scaffold. Thus, a series of 4-oxoquinolizine-3-carboxylic acid derivatives **8**, **9**, **12** and **14** were synthesized. Results show that, although these compounds could bind Mg²⁺ in CH₃CN solution, no obvious anti-HIV IN activity is observed.

Figure 1. The design of 4-oxo-quinolizine-3-carboxylic acid derivatives as potential HIV IN inhibitors.

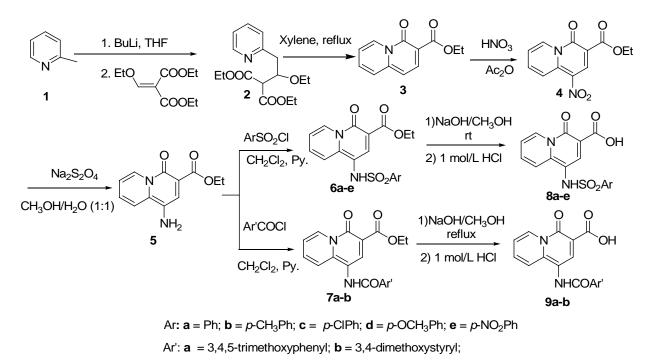


Results and Discussion

Synthesis of 4-oxo-4H-quinolizine-3-carboxylic acid derivatives 8, 9, 12 and 14

The synthesis of 4-oxo-4*H*-quinolizine-3-carboxylic acid derivatives **8** and **9** is described in Scheme 1. Ethyl 4-oxo-4*H*-quinolizine-3-carboxylate (**3**) was synthesized in good yield starting from 2-methylpyridine according to a literature procedure [24]. Nitration of **3** afforded compound **4**, which was reduced by $Na_2S_2O_4$ to generate the key intermediate ethyl 1-amino-4-oxo-4*H*-quinolizine-3carboxylate (**5**). In the presence of pyridine, compound **5** was treated with sulfonyl chloride or acyl chloride to give compounds **6** and **7**, respectively. The target 4-oxo-4*H*-quinolizine-3-carboxylic acids **8** and **9** were finally obtained after hydrolysis in NaOH/methanol followed by neutralization. During the nitration procedure, the speed of addition of nitric acid should be carefully controlled, otherwise, the reaction mixture would become red and the amount of by-products increased. In addition, $Na_2S_2O_4$ was used to reduce nitro group and generated key intermediate **5**, due to the benign reaction conditions (usually at room temperature using water as solvent) and simple workup.

Scheme 1. The synthesis of 4-oxo-4H-quinolizine-3-carboxylic acid derivatives 8 and 9.

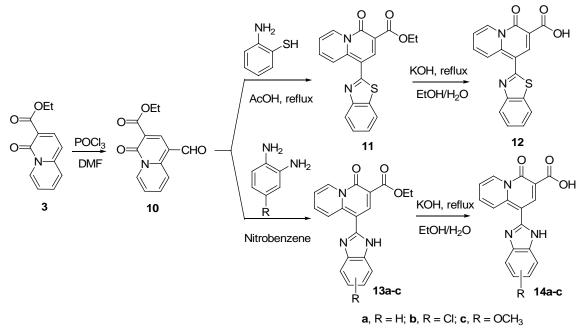


With **3** in hand, we then synthesized benzothiazole- and benzimidazole-substituted 4-oxo-4*H*-quinolizine-3-carboxylic acid derivatives **12** and **14**. As shown in Scheme 2, reaction of compound **3** with DMF and POCl₃ generated intermediate **10** [24], which reacted with 2-aminothiophenol [28, 29] followed by subsequent hydrolysis to generate 1-(benzothiazole-2-yl)-4-oxo-4*H*-quinolizine-3-carboxylic acid (**12**). Similarly, compound **10** underwent oxidative condensation with *o*-phenylene-diamine derivatives in nitrobenzene [30] yielded compounds **13a-c**, which were hydrolyzed [31] to obtain 1-(benzimidazole-2-yl)-4-oxo-4*H*-quinolizine-3-carboxylic acids **14a-c**. It should be pointed out

that two benzimidazole isomers were formed for each of **13b-c** and **14b-c**, that could not be separated each other through conventional procedures.

It is noteworthy that the conditions for the hydrolysis of **11** and **13** were not similar to those used for the synthesis of **8** and **9**. The reaction proceeded very slowly in the NaOH/MeOH system. For example, after 48 hours stirring at room temperature, the reaction mixtures were still slurries and most of the starting materials were present in the reaction system. Consequently, KOH in 80% aqueous ethanol was used for the synthesis of **12** and **14**. As the reaction proceeded the slurry of the reaction mixture turned clear and the products precipitated when 1 mol·L⁻¹aqueous HCl was added.

Scheme 2 Synthesis of 4-oxo-4H-quinolizine-3-carboxylic acid derivatives 12 and 14.



Structural analysis

The structures of all new 4-oxo-4*H*-quinolizine-3-carboxylic acid derivatives **6-14** were confirmed by ¹H-NMR, ¹³C-NMR, IR and ESI-MS (or HRMS) spectra. In the ¹H-NMR spectra of ethyl 4-oxo-4*H*-quinolizine-3-carboxylates **6**, **7**, **11**, **13**, the 4-oxo-4*H*-quinolizine ring proton signals appeared at lower field compared with those of the benzene ring. For example, in the ¹H-NMR spectra of **6b**, the two doublets at 9.43 ppm and 8.28 ppm were ascribed to the protons at C-5 and C-8 of the 4-oxo-4*H*-quinolizine ring, respectively, while the protons at C-6 and C-7 exhibited two doublets at 7.76 ppm and 7.29 ppm. The proton at C-2 resided at 7.63 ppm. A high field AB system (7.31 ppm and 7.64 ppm) corresponded to the signals of the four benzene ring protons.

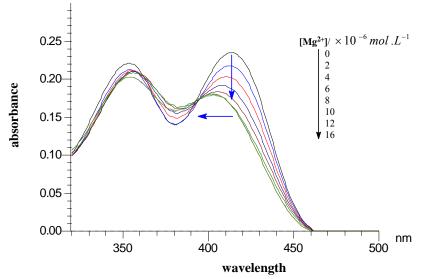
In the ¹H-NMR of target compounds **8**, **9**, **12**, **14a**, one sharp singlet in the δ 13.7-14.1 range was observed, which disappeared after the D₂O exchange and was consequently attributed to the COOH proton. For example, the proton signals of the COOH groups of compounds **8a** and **14a** were located at 13.7 ppm and 13.8 ppm, respectively. In addition, compound **14a** has a strong singlet at δ 13.7 ppm, which demonstrated the existence of a N-H proton in the benzimidazole ring. In the ¹³C-NMR spectra of these compounds one C=O peak at about 166 ppm and one around 159 ppm corresponded to the signals of the 3-carboxylic acid and carbonyl groups in the quinolizine rings. The IR spectra of these

compounds further support their postulated structures. There were several medium broad bands at $3,200 \sim 3,400 \text{ cm}^{-1}$ corresponding to O-H and N-H bond and strong bands at $1,690 \sim 1,730 \text{ cm}^{-1}$ due to C=O bonds. The mass spectra of compounds **8c**, **13b** and **14b** exhibited a M+2 signal, whose intensity was 1/3 of the molecular ion signal, and corresponding to the Cl isotopes in these compounds.

Preliminary investigation of Mg^{2+} binding

To elucidate a two-metal inhibition model, very recently, Kawasuji *et al* [32] performed metal titration studies of 2-hydroxy-3-heteroarylacrylic acid derivatives and observed a two-step shift of the UV-Vis spectra upon the addition of Mg^{2+} if these compounds exhibited HIV-IN inhibitory activity. On the contrary, simple change of UV-Vis absorption curves or no shift was found for their mutated compounds which do not inhibit HIV integrase. Inspirited by this assumption, preliminary ion binding experiments were also carried out by UV-Vis spectroscopy, using compound **14a** as an example. Due to the low solubility of **14a** in water, CH₃CN was used to prepare the stock solution for the UV-Vis measurements. As shown in Figure 2, for the free compound **14a**, two intense absorption bands at 352 nm and 414 nm were observed in the 320 – 550 nm range. As the Mg²⁺ concentration (as perchlorate salt) increased, the absorption intensity of the peak at 414 nm first decreased and then the wavelength shifted blue to around 402 nm. Figure 3 further demonstrated the selectivity of compound **14a** to Mg²⁺. In the presence of a 100-fold equivalent amount of Na⁺ or K⁺, no obvious change was observed. However, the Mg²⁺ induced a blue shift of the absorption curve of compound **14a**. Similar behaviors were also observed for other divalent metals, including Mn²⁺, Co²⁺, Cu²⁺, Ca²⁺ and Ni²⁺ (not shown in Figure 3).

Figure 2. Changes of UV-vis spectra with **14a** in CH₃CN on addition of $0-16 \times 10^{-6}$ mol·L⁻¹ of [Mg²⁺] at 25 °C.

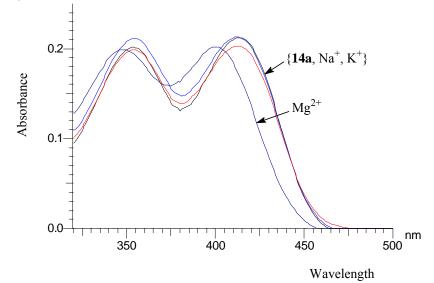


Conditions: $[Mg^{2+}] = 2 \times 10^{-3} \text{ M}$. $[14a] = 2 \times 10^{-5} \text{ MBlack}$: $0 \times 10^{-6} \text{ mol} \cdot \text{L}^{-1}$, blue: $2 \times 10^{-6} \text{ mol} \cdot \text{L}^{-1}$, red: $4 \times 10^{-6} \text{ mol} \cdot \text{L}^{-1}$, navy: $6 \times 10^{-6} \text{ mol} \cdot \text{L}^{-1}$, brown: $8 \times 10^{-6} \text{ mol} \cdot \text{L}^{-1}$, green: $10 \times 10^{-6} \text{ mol} \cdot \text{L}^{-1}$, dark cyan: $12 \times 10^{-6} \text{ mol} \cdot \text{L}^{-1}$, brass: $16 \times 10^{-6} \text{ mol} \cdot \text{L}^{-1}$.

Molecules 2009, 14

The titration experiment of **14a** with Mg^{2+} further demonstrated that the binding ratio is 1:1, which is consistent with that of KMG-103 and KMG-104 to Mg^{2+} , indicating that the introduction of benzimidazole substituent on the C-1 position of the 4-oxo-4*H*-quinolizine-3-carboxylic acid scaffold does not affect its Mg^{2+} binding ability. In a word, a two-step shift of the absorption curve of compound **14a** upon the addition of Mg^{2+} was observed, similar to 2-hydroxy-3-heteroaryl acrylic acid derivatives reported by Kawasuji *et al* [32]. These UV-Vis results indicated that our compounds might interact with metal ions (Mg^{2+}) in the active site of HIV IN and thus offer a piece of *in vitro* indication of possible HIV IN inhibitory activity of these new compounds, in spite of no strict demonstration of correction between the inhibitory activity and the metal affinity.

Figure 3. UV-Vis absorption spectra of free **14a** and **14a** in the presence of K^+ , Na^+ or Mg^{2+} ion in CH₃CN solution.



Conditions: $[14a] = 2 \times 10^{-5} \text{ M}; [Mg^{2+}] = [K^+] = [Na^+] = 2 \times 10^{-3} \text{ M};$ Black: 14a, blue: Na⁺ complex, red: K⁺ complex, navy: Mg^{2+} complex.

HIV integrase inhibitory activities of 4-oxo-4H-quinolizine-3-carboxylic acid derivatives

The target compounds **8**, **9**, **12**, **14** were screened against purified HIV IN to determine any potential inhibitory activity. Thus, a synthesized 30 oligonucleotide and a 20 oligonucleotide were used as donor DNA and target DNA, respectively, which were purchased from Shanghai Sangon Biological Engineering Technology & Services Co. Ltd. The donor DNA was biotinylated at the 5' end (5' bio-DNA) and immobilized on streptavidin-coated 96-well microtiter plates. To the donor DNA streptavidin-coated 96-well microtiter plate were added test sample, 3' digoxygenin-labeled target DNA (3' dig-DNA) and recombinant HIV integrase and subjected to incubation in optimized condition.

The plate was further washed with phosphate-buffered saline (PBS) containing 0.1% Tween[®]-20 and anti-digoxigenin-peroxidase (POD, 100 μ L) was added to each well. After incubation at 37°C for 60 min, the plate was washed with PBS containing 0.1% Tween[®]-20. The digoxygenin-labeled products were visualized by adding a POD substrate, tetramethylbenzidine (TMB). The colorimetric

reaction was stopped by addition of 0.5 mol/L H_2SO_4 (100 µL), and the absorbance was measured at 450 nm on a microplate reader. The total inhibitory effect (including inhibiting 3' process reaction and strand transfer reaction) were obtained by comparing the absorbance of drug group with control group (without test compound). As shown in Table 1, the values of IC₅₀ of these compounds are more than 100 µg/mL, which mean a low inhibitory activity to HIV integrase.

The low HIV IN inhibitory activity of these designed 4-oxo-4*H*-quinolizine-3-carboxylic acid derivatives might stem from various cofactors. Structurally, compounds **8**, **9**, **12** and **14** possess a chelating scaffold suitable for binding only one ion. However, a two-metal chelating scaffold to form respectively a five-membered and a six-membered system is required according to the two-metal coordinating model for diketoacid-type HIV IN inhibitors. Therefore, the selective binding to Mg^{2+} is not enough, the two-metal binding scaffold is also necessary in the design of diketoacid-type HIV IN inhibitors. In addition, an appropriate substituent at appropriate position of the chelating scaffold is also essential to ensure potential inhibitory activity.

Sample	Initial concentration (µg/mL)	IC ₅₀ (μg/mL)	Sample	Initial concentration (µg/mL)	IC ₅₀ (µg/mL)
$S-y^1$	12	0.56	9a	100	_2
8a	100	_2	9b	100	_2
8b	100	_2	12	100	_2
8c	100	_2	14a	100	_2
8d	100	_2	14b	100	_2
8e	100	_2	14c	100	_2

Table 1. Integrase inhibitory activity data of 4-oxo-4H-quinolizine-3-carboxylic acid derivatives.

¹ S-y provided by the Shanghai Institute of Organic Chemistry was used as positive contrast.

² "-" indicates that the HIV-IN inhibitory effect was less than 50% at the initial concentration.

Conclusions

Aryl diketoacid-containing compounds were demonstrated to be the most promising HIV IN inhibitors. Based on the molecular mechanism of aryl diketoacids as HIV IN inhibitors and the selective binding to Mg^{2+} of 4-oxo-4*H*-quinolizine-3-carboxylic acid unit, a series of 4-oxo-4*H*-quinolizine-3-carboxylic acid derivatives, substituted by sulfonoamido, carboxylamido, benzimidazole and benzothiazole groups, were designed and synthesized as diketoacid bioisosters. Their anti-HIV IN activity was evaluated and no obvious inhibitory activities were observed. Based on the present results, in the course of our coming works on development of HIV IN inhibitors, we would have a concept that new types of HIV IN inhibitors would incorporate a two-metal chelating scaffold and a appreciate hydrophobic domain. Moreover, the two-metal chelating scaffold could have an obvious binding affinity to Mg^{2+} .

Experimental

General

All melting points (m.p.) were measured with a XT4A electrothermal apparatus equipped with a microscope and are uncorrected. Infrared spectra (IR) were recorded as thin films on KBr plates with a Bruker IR spectrophotometer and are expressed in v (cm⁻¹). ¹H- and ¹³C-NMR spectra were recorded with an AV 400 M Bruker spectrometer (400 MHz ¹H frequency, 100 MHz ¹³C frequency). Chemical shifts were measured in CDCl₃ or DMSO- d_6 with TMS as internal reference. The MS spectra (ESI) were recorded on a Bruker Esquire 6000 mass spectrometer. HRMS spectra were recorded in the negative ion mode using APEX II FT-ICRMS of Bruker Daltonics Inc. All solvents were of commercial quality and were dried and purified by standard procedures. Compounds **2**, **3**, **4**, **10** were synthesized according to the literature procedure [24].

General procedure for the synthesis of ethyl 1-(substituted phenylsulfonamido)-4-oxo-4H-quinolizine-3-carboxylates **6**

Compound 4 (1.97 g, 7.5 mmol) was suspended in a 1:1 mixture of methanol and water (100 mL) in a 250 mL flask. Na₂S₂O₄ (7.8 g, 45 mmol) was then added in one portion at room temperature and the mixture was stirred for about 2 hours. After the reaction, the organic compound was extracted three times using CH₂Cl₂ and dried over anhydrous Na₂SO₄. After filtration of the Na₂SO₄, the solvent was removed under reduced pressure to give 1.0 g (57%) of **5** as a red solid, which was used for the following reactions without further purification. The appropriate substituted sulfonyl chloride (3.44 mmol) in CH₂Cl₂ (30 mL) was added dropwise to a CH₂Cl₂ solution (30 mL) containing **5** (3.44 mmol) and dry pyridine (0.3 mL). The solution changed from red to dark and TLC showed the formation of new compounds with red fluorescence emission. After the reaction, the solution was washed with water and the organic layer was dried over anhydrous MgSO₄. After filtration of the MgSO₄, the solvent was removed under reduced pressure and the residue was recrystallized by ethyl acetate to give the desired products **6** as yellow solids.

Ethyl 1-phenylsulfonamido-4-oxo-4H-quinolizine-3-carboxylate (**6a**): Yield 72%. m.p.: 207-209 °C; ¹H-NMR (CDCl₃): δ 1.34 (t, 3H, J = 7.1 Hz, CH₃), 4.32 (q, 2H, J = 7.1 Hz, OCH₂), 7.25 (dd, 1H, J = 7.7 Hz, J = 5.9 Hz), 7.48~7.91 (m, 8H, Ar-H), 9.42 (d, 1H, J = 7.1 Hz,); ¹³C-NMR (CDCl₃): δ 14.4, 60.9, 105.6, 106.9, 117.3, 122.0, 128.9, 129.3, 130.1, 134.6, 135.5, 138.4, 143.9, 147.2, 154.7, 164.5; IR: v 3491, 3139, 2981, 1734, 1698, 1493 cm⁻¹; MS (ESI): m/z 370.9 [M - H]⁻.

Ethyl 1-p-methylphenylsulfonamido-4-oxo-4H-quinolizine-3-carboxylate (**6b**): Yield 60%. m.p.: 115-117 °C; ¹H-NMR (CDCl₃): δ 1.31 (t, 3H, J = 7.2 Hz, CH₃), 2.45 (s, 3H, PhCH₃), 4.30 (q, 2H, J = 7.2 Hz, CH₂), 6.38 (s, 1H, NH), 7.29~7.33 (m, 3H, Ar-H), 7.64(s, 1H, Ar-H), 7.65(d, 2H, J = 7.6 Hz, Ar), 7.77 (dd, 1H, J = 7.5 Hz, J = 8.0 Hz, Ar-H), 8.28 (d, 1H, J = 8.8 Hz, Ar-H), 9.43 (d, 1H, J = 7.1 Hz); ¹³C-NMR (CDCl₃): δ 14.6, 21.8, 61.1, 104.9, 108.3, 117.7, 122.9, 128.0, 130.1, 130.1, 135.4, 135.5,

141.5, 144.7, 146.9, 155.1, 165.3; IR: v 3452, 3164, 2972, 1718, 1629, 1490 cm⁻¹; MS (ESI): m/z 387.0 [M + H]⁺.

Ethyl 1-p-chloro-phenylsulfonamido-4-oxo-4H-quinolizine-3-carboxylate (**6c**): Yield 73%. m.p.: 210-212°C; ¹H-NMR (DMSO-*d*₆): δ 1.24 (t, 3H, *J* = 7.6 Hz, CH₃), 4.18 (q, 2H, *J* = 7.6 Hz, CH₂), 7.54 (s, 1H, Ar-H), 7.55 (dd, 1H, *J* = 7.2 Hz, *J* = 6.4 Hz, Ar-H), 7.68 (d, 2H, *J* = 8.8 Hz, Ar-H), 7.73 (d, 2H, *J* = 8.8 Hz, Ar-H), 7.97-8.05 (m, 2H, Ar-H), 9.29 (d, 1H, *J* = 7.2 Hz, Ar-H); ¹³C-NMR (DMSO-*d*₆): δ 14.6, 60.3, 103.1, 108.5, 118.7, 122.4, 129.7, 129.9, 130.0, 136.6, 138.0, 138.6, 140.1, 146.0, 154.0, 164.5; IR: v 3445, 3155, 2972, 1715, 1494 cm⁻¹; MS (ESI): *m/z* 404.9 [M -1]⁻, 428.8 [M + Na]⁺.

Ethyl 1-p-methyloxyphenylsulfonamido-4-oxo-4H-quinolizine-3-carboxylate (**6d**): Yield 92%. m.p.: 236-238°C; ¹H NMR (DMSO-*d*₆): δ 1.19 (t, 3H, *J* = 6.9 Hz, CH₃), 3.82 (s, 3H, OCH₃), 4.11 (q, 2H, *J* = 7.0, CH₂), 7.06 (d, 2H, *J* = 8.6, Ar-H), 7.49-7.51 (m, 2H, Ar-H), 7.60 (d, 2H, *J* = 8.6 Hz, Ar-H), 7.90-8.01 (m, 2H, Ar-H), 9.24 (d, 1H, *J* = 7.2 Hz, Ar-H), 9.58 (s, 1H, NH); ¹³C NMR (DMSO-*d*₆): δ 14.5, 56.1, 60.4, 103.2, 109.1, 114.9, 118.7, 122.5, 129.8, 129.9, 130.6, 136.3, 140.3, 146.0, 154.1, 163.2, 164.6; IR: *v* 3442, 3170, 2927, 1716, 1650, 1494 cm⁻¹; MS (ESI): *m/z* 424.9 [M+Na]⁺.

Ethyl 1-p-nitrophenylsulfonamido-4-oxo-4H-quinolizine-3-carboxylate (**6e**): Yield: 76%. m.p.: 265-267°C; ¹H-NMR (DMSO-*d*₆): δ 1.15 (t, 3H, *J* = 7.2 Hz, CH₃), 4.1 (q, 2H, *J* = 7.2 Hz, CH₂), 7.51 (s, 1H, Ar-H), 7.49-7.53 (m, 1H, Ar-H), 7.94-7.98 (m, 4H, Ar-H), 8.39 (d, 2H, *J* = 8.8 Hz, Ar-H), 9.27 (d, 1H, *J* = 7.2 Hz, Ar-H), 10.20 (s, 1H, NH); ¹³C-NMR (DMSO-*d*₆): δ 14.5, 60.4, 103.1, 108.0, 118.8, 122.2, 125.0, 129.4, 130.1, 136.8, 140.1, 144.8, 145.8, 150.4, 154.1, 164.6; IR: *v* 3446, 3204, 2981, 1727, 1627, 1497, 1233 cm⁻¹; MS (ESI): *m/z* 439.9 [M + Na]⁺.

General procedure for the synthesis of 1-(substituted phenylsulfonamido)-4-oxo-4H-quinolizine-3carboxylic acids **8**

To a suspension of 6 (1.2 mmol) in methanol (15 mL) was added 3 mol/L of NaOH (0.9 mL). The reaction mixture was stirred overnight, followed by evaporation of the solvent under reduced pressure. The remaining solid was dissolved in water (20 mL) and the clear solution was acidified with 1 mol/L HCl to pH 1. The resulting suspension was filtered and dried under vacuum over P_2O_5 to give yellowish solids.

1-Phenylsulfonamido-4-oxo-4H-quinolizine-3-carboxylic acid (**8a**): Yield 82%. m.p.: 245-247°C; ¹H-NMR (DMSO-*d*₆): δ 7.51-7.55 (m, 2H, Ar-H), 7.63-7.66 (m, 5H, Ar-H), 8.00-8.09 (m, 2H, Ar-H), 9.29 (d, 1H, *J* = 7.2 Hz, Ar-H), 10.05 (s, 1H, NH), 13.71 (s, 1H, COOH); ¹³C-NMR (DMSO-*d*₆): δ 103.7, 113.0, 120.2, 122.9, 127.6, 129.8, 133.8, 136.9, 138.6, 138.9, 145.0, 158.9, 165.6; IR: *v* 3441, 3293, 3116, 1727, 1616, 1495, 1446, 1365, 1327, 1159 cm⁻¹; MS (ESI): *m/z* 344.9 [M + H]⁺, 366.9 [M + Na]⁺.

1-p-Methylphenylsulfonamido-4-oxo-4H-quinolizine-3-carboxylic acid (**8b**): Yield 75%. m.p.: > 300 °C; ¹H-NMR (DMSO-*d*₆): δ 2.35 (s, 3H, CH₃), 7.33 (d, 2H, *J* = 8.1 Hz, Ar-H), 7.54 (d, 2H, *J* = 8.1 Hz,

Ar-H), 7.64 (dd, 1H, J = 7.0 Hz, Ar-H), 7.67 (s, 1H, Ar-H), 8.03 (dd, 1H, J = 7.2 Hz, J = 8.4 Hz, Ar-H), 8.11 (d, 1H, J = 8.8 Hz, Ar-H), 9.29 (d, 1H, J = 7.1 Hz, Ar-H); ¹³C-NMR (DMSO- d_6): δ 21.5, 103.7, 113.3, 120.2, 123.0, 127.6, 129.8, 130.2, 136.2, 136.8, 138.5, 144.1, 145.0, 158.9, 165.7; IR: v 3265, 3071, 1725, 1619, 1492, 1448, 1292, 1160, 816 cm⁻¹; HRMS: m/z calcd for C₁₇H₁₃ N₂O₅S: 357.0551; found: 357.0550.

1-p-Chlorophenylsulfonamido-4-oxo-4H-quinolizine-3-carboxylic acid (**8c**): Yield 79%. mp > 300°C; ¹H-NMR (DMSO-*d*₆): δ 7.65-7.72 (m, 6H, Ar-H), 8.09-8.15 (m, 2H, Ar-H), 9.36 (d, 1H, *J* = 7.1 Hz, Ar-H), 10.22 (s, 1H, NH), 13.70 (s, 1H, OH); ¹³C-NMR (DMSO-*d*₆): δ 103.7, 112.7, 120.3, 122.9, 129.6, 129.9, 129.9, 137.0, 137.8, 138.5, 138.6, 145.0, 158.9, 165.7; IR: *v* 3446, 3265, 2924, 1720, 1489, 1163 cm⁻¹; MS (ESI): *m/z* 376.9 [M-H]⁻.

1-p-Methoxyphenylsulfonamido-4-oxo-4H-quinolizine-3-carboxylic acid (**8d**): Yield 86%. mp > 300°C; ¹H-NMR (DMSO-*d*₆): δ 3.79 (s, 3H, CH₃), 7.03 (d, 2H, *J* = 8.8 Hz, Ar-H), 7.57 (d, 2H, *J* = 8.8 Hz, Ar-H), 7.65 (dd, 1H, *J* = 10.0 Hz, *J* = 7.2 Hz, Ar-H), 7.68 (s, 1H, Ar-H), 8.03 (dd, 1H, *J* = 8.6 Hz, *J* = 7.0, Ar-H), 8.10 (d, 1H, *J* = 8.8Hz, Ar-H), 9.29 (d, 1H, *J* = 7.2Hz, Ar-H), 9.86 (s, 1H, NH); ¹³C-NMR (DMSO-*d*₆): δ 56.2, 103.7, 113.3, 114.9, 120.2, 123.0, 129.7, 129.8, 130.5, 136.8, 138.6, 145.1, 158.9, 163.3, 165.7; IR: *v* 3248, 2918, 2849, 1724, 1618, 1597, 1496, 1450, 1294, 1157, 836 cm⁻¹; MS (ESI): *m/z* 375.0 [M+H]⁺.

1-p-Nitrophenylsulfonamido-4-oxo-4H-quinolizine-3-carboxylic acid (**8e**): Yield 82%; mp > 300°C; ¹H-NMR (DMSO-*d*₆): δ 7.60-7.80 (m, 1H, Ar-H), 7.73 (s, 1H, Ar-H), 7.94 (d, 2H, *J* = 8.0 Hz, Ar-H), 8.02-8.05 (m, 2H, Ar-H), 8.35 (d, 2H, *J* = 8.0 Hz, Ar-H), 9.32 (d, 1H, *J* = 6.4 Hz, Ar-H), 10.49 (s, 1H, NH), 13.68 (s, 1H, OH); ¹³C-NMR (DMSO-*d*₆): δ 103.8, 112.3, 120.3, 122.6, 125.1, 129.3, 129.9, 137.2, 138.6, 144.7, 144.8, 150.4, 158.9, 165.7; IR: *v* 3454, 3234, 3082, 1715, 1488, 1167, 786 cm⁻¹; MS (ESI): *m/z* 387.8 [M-H]⁻.

General procedure for the synthesis of 7a,b

To a solution of 3,4,5-trimethoxylbenzoic acid or 3,4-dimethoxycinnamic acid (4 mmol) in ether (20 mL) was added PCl₅ (20 mL) in one portion. The reaction mixture was heated to reflux for about 4 hours and then cooled to room temperature. After maintaining in a refrigerator for about 1 hour, a white precipitate formed which was filtered and recrystallized from cold ether to give the corresponding carboxylic chloride. The newly-synthesized acyl chloride dissolved in CH₂Cl₂ (20 mL) was added dropwise to a mixture of compound **5** (3 mmol) and triethylamine (3 mmol) in CH₂Cl₂ (20 mL). The reaction solution was stirred overnight at room temperature. After the reaction, the solution was washed with water and the organic layer was dried over anhydrous MgSO₄. After filtration of the MgSO₄, the solvent was removed under reduced pressure and the residue was purified by recrystallization from ethyl acetate to give **7a** or **7b** as a yellow solid.

Ethyl 1-(3,4,5-trimethoxyphenylcarbonylamino)-4-oxo-4H-quinolizine-3-carboxylate (**7a**): Yield 40%; mp 93°C; ¹H-NMR (CDCl₃): δ 1.29 (t, 3H, J = 7.2 Hz, CH₃), 3.92 (s, 3H, OCH₃), 3.95 (s, 6H, OCH₃),

4.26 (q, 2H, J = 7.2 Hz, OCH₂), 7.11 (dd, 1H, J = 6.8 Hz, J = 7.2 Hz, Ar-H), 7.39 (s, 2H, Ph-H), 7.57 (dd, 1H, J = 8.8 Hz, J = 6.8 Hz, Ar-H), 7.66 (d, 1H, J = 8.8 Hz, Ar-H), 8.08 (s, 1H, Ar-H), 8.66 (bs, 1H, NH), 9.17 (d, 1H, J = 7.2 Hz, Ar-H); ¹³C-NMR (CDCl₃): $\delta = 14.4$, 56.4, 60.9, 61.0, 105.1, 105.1, 110.6, 116.9, 122.0, 128.4, 129.4, 134.2, 140.2, 141.5, 143.7, 153.3, 154.9, 165.3, 166.9; IR: v 3462, 3328, 2939, 1720, 1678, 1587, 1487, 1336, 1216, 1126, 1009, 772 cm⁻¹; MS (ESI): m/z 449.0 [M + Na]⁺.

Ethyl 1-[3-(3,4-dimethoxyphenyl)acrylamido]-4-oxo-4H-quinolizine-3-carboxylate (**7b**): Yield 85%; m.p.: 105-107°C; ¹H-NMR (DMSO-*d*₆): δ 1.30 (t, 3H, *J* = 7.2 Hz, CH₃), 3.81 (s, 3H, OCH₃), 3.83 (s, 3H, OCH₃), 4.26 (q, 2H, J = 7.2 Hz, CH₂), 6.81 (d, 1H, *J* = 15.6 Hz, CH=CH), 7.03 (d, 1H, *J* = 8.4 Hz, Ar-H), 7.21 (d, 1H, *J* = 8.4 Hz, Ar-H), 7.26 (s, 1H, Ar-H), 7.51 (dd, 1H, *J* = 6.4 Hz, *J* = 7.2 Hz, Ar-H), 7.55 (d, 1H, *J* = 15.6 Hz, Ar-H), 7.86 (d, 1H, J = 8.8 Hz, Ar-H), 7.96 (dd, 1H, *J* = 7.2 Hz, *J* = 8.0 Hz, Ar-H), 8.26 (s, 1H, Ar-H), 9.30 (d, 1H, *J* = 7.2 Hz, Ar-H), 9.89 (s, 1H, NH); ¹³C-NMR (DMSO-*d*₆): δ 14.8, 55.9, 56.0, 60.5, 103.7, 110.6, 111.1, 112.2, 118.2, 119.4, 122.3, 122.4, 127.9, 129.8, 135.8, 139.0, 141.1, 143.1, 149.4, 150.9, 154.1, 165.3, 165.9; IR: *v* 3436, 3226, 2935, 1702, 1677, 1595, 1491, 1377, 1236, 1114, 1026, 768 cm⁻¹; MS (ESI): *m/z* 445.0 [M + Na]⁺.

General procedure for the synthesis of 9a,b

To a suspension of **7a** or **7b** (0.5 mmol) in 80% methanol (20 mL) was added NaOH (5 mmol). The reaction mixture was heated to reflux till the starting **7a** or **7b** disappeared according to monitoring by TLC. After evaporation of the solvent under reduced pressure, the remaining solid was dissolved in water (20 mL) and the clear solution was acidified with 1 mol/L HCl to pH 1 and the resulting suspension extracted with CH_2Cl_2 . The combined organic layer was dried over anhydrous MgSO₄ followed by filtration. The desired **9a** or **9b** were finally obtained as solids by column chromatography (eluents: $CH_3OH/CHCl_3 = 1:30$)

1-(3,4,5-Trimethoxyphenylcarbonylamino)-4-oxo-4H-quinolizine-3-carboxylic acid (**9a**): Yield 92%; m.p.: 280-282°C; ¹H-NMR (DMSO-*d*₆): δ 3.67 (s, 3H, OCH₃), 3.80 (s, 6H, OCH₃), 7.33 (s, 2H, Ar-H), 7.64 (dd, 1H, *J* = 7.2 Hz, *J* = 6.8 Hz, Ar-H), 7.94 (d, 1H, *J* = 8.8 Hz, Ar-H), 8.02 (dd, 1H, *J* = 6.8 Hz, *J* =8.8 Hz, Ar-H), 8.29 (s, 1H, Ar-H), 9.30 (d, 1H, *J* = 7.2 Hz, Ar-H), 10.29 (s, 1H, NH), 13.96 (s, 1H, COOH); ¹³C-NMR (DMSO-*d*₆): δ 56.6, 60.6, 103.9, 105.9, 115.2, 105.9, 115.2, 120.2, 123.3, 129.1, 129.7, 136.6, 138.2, 141.0, 143.0, 153.2, 159.0, 166.1, 166.5; IR: *v* 3440, 3115, 2924, 2853, 1696, 1657, 1615, 1587, 1496, 1453, 1364, 1292, 1123 cm⁻¹; MS (ESI): *m/z* 397.0 [M - H]⁻.

1-(3,4-Dimethoxystyrylcarboxylamino)-4-oxo-4H-quinolizine-3-carboxylic acid (**9b**): Yield 91%; m.p.: 306-308°C; ¹H-NMR (DMSO-*d*₆): δ 3.82 (s, 3H, OCH₃), 3.84 (s, 3H, OCH₃), 6.84 (d, 1H, *J* = 16.0 Hz, CH=CH), 7.03 (d, 1H, *J* = 8.4 Hz, Ar-H), 7.23 (d, 1H, *J* = 8.4 Hz, Ar-H), 7.27 (s, 1H, Ar-H), 7.58 (d, 1H, *J* = 16.0 Hz, Ar-H), 7.70-7.74 (m, 1H, Ar-H), 8.10-8.12 (m, 2H, Ar-H), 8.44 (s, 1H, Ar-H), 9.37 (d, 1H, *J* = 7.2 Hz, Ar-H), 10.11 (s, 1H, NH), 14.13 (s, 1H, OH); ¹³C-NMR (DMSO-*d*₆): δ 56.0, 56.1, 103.9, 110.9, 112.4, 115.5, 119.3, 120.0, 122.3, 122.8, 127.9, 136.3, 136.9, 141.4, 142.1, 149.5, 151.1,

158.8, 165.8, 166.1; IR: v 3251, 1728, 1612, 1541, 1515, 1464, 1303, 1266 cm⁻¹; MS (ESI): *m/z* 393.1 [M - H]⁻.

Synthesis of ethyl 1-benzothiazol-2-yl-4-oxo-4H-quinolizine-3-carboxylate (11)

2-Aminophenthiol (250 mg, 2 mmol) was added to a 50 mL of round flask charged with compound **10** (490 mg, 2 mmol) and acetic acid (15 mL). The reaction mixture was heated to reflux for about one hour till TLC confirmed that the reaction had finished. After removal of the solvent, the residue was recrystallized from ethyl acetate to offer a needle-like yellow solid. Yield: 65%; m.p.: 199-200 °C; ¹H-NMR (DMSO-*d*₆): δ 1.35 (t, 3H, *J* = 7.1 Hz, CH₂CH₃), 4.32 (q, 2H, *J* = 7.1 Hz, CH₂CH₃), 7.48 (t, 1H, *J* = 7.6 Hz, Ar-*H*), 7.57 (t, 1H, *J* = 7,6 Hz, Ar-*H*), 7.70 (t, 1H, *J* = 7.2 Hz), 8.07 (d, 1H, *J* = 8.0 Hz, Ar-*H*), 8.15 (d, 1H, *J* = 8.0 Hz, Ar-*H*), 8.25 (t, 1H, *J* = 7.6 Hz, Ar-*H*), 8.75 (s, 1H, Ar-*H*), 9.47 (d, 1H, *J* = 7.2 Hz, Ar-*H*), 9.59 (d, 1H, *J* = 8.0 Hz, Ar-*H*); ¹³C-NMR (DMSO-*d*₆): δ 15.2, 61.2, 105.0, 106.0, 120.0, 122.8, 123.3, 124.9, 126.1, 127.5, 131.3, 134.2, 139.4, 142.9, 144.6, 154.3, 154.7, 165.4, 165.9; IR: *v* 1736, 1670, 1623, 1583, 1496, 1222, 1133, 780 cm⁻¹; MS (ESI): *m/z* 373 [M + Na]⁺.

Synthesis of 1-benzothiazol-2-yl-4-oxo-4H-quinolizine-3-carboxylic acid (12)

KOH (765 mg, 13.7 mmol) dissolved in water (6 mL) was added to a 100 mL round flask charged with compound **11** (480 mg, 1.37 mmol) and ethanol (24 mL). The reaction mixture was heated to reflux for about four hours till TLC confirmed that the reaction had finished. After removal of the solvent, the residue was re-dissolved in water (30 mL) and then acidified to pH 2. The product was extracted three times with CH₂Cl₂ (50 mL) and the combined organic layer was dried over anhydrous MgSO₄. After filtration of the MgSO₄, the solvent was removed under reduced pressure and the residue was purified by recrystallization from chloroform/petroleum ether to generate **12** as a yellow solid. Yield 68%. m.p.: >300 °C; ¹H-NMR (CDCl₃): δ 7.32 (dd, 1H, *J* = 7.6 Hz, *J* = 7.2 Hz, Ar-*H*), 7.64-7.67 (m, 2H, Ar-*H*), 7.81-7.91(m, 2H, Ar-*H*), 7.95 (d, 1H, *J* = 8.0 Hz, Ar-*H*), 8.78 (d, 1H, *J* = 4.8 Hz), 9.21 (s, 1H, Ar-*H*), 9.43 (d, 1H, *J* = 7.2 Hz, Ar-*H*), 13.95 (s, 1H, COO*H*); ¹³C-NMR (DMSO-*d*₆): δ 110.2, 113.2, 119.9, 120.6, 122.6, 123.3, 127.6, 128.3, 130.7, 137.3, 138.3, 138.7, 147.7, 151.4, 152.4, 163.8, 165.7; IR: *v* 1750, 1611, 1564, 1453 cm⁻¹; MS (ESI): *m/z* 345.0 [M + Na]⁺, 361.0 [M + K]⁺.

General procedure for the synthesis of compounds 13a-c

To a 50 mL of round flask charged with phenylenediamine derivative (1 mmol) and nitrobenzene (10 mL) was added compound **10** (245 mg,1 mmol) and then the solution was slowly heated to 100 °C. After 1 h stirring, the reaction mixture was further heated to 150 °C and stirred till TLC showed the absence of starting **10**. The solution was cooled to room temperature and directly loaded onto a silica column for chromatographic purification. The eluents were first petroleum ether and then ethyl acetate/petroleum ether (1:2). After the nitrobenzene was eluted out, CHCl₃/CH₃OH was used to elute the corresponding **13a-c**.

Ethyl 1-(1H-benzimidazol-2-yl)-4-oxo-4H-quinolizine-3-carboxylate (**13a**): Yield 85 %. m.p.: 194-195 °C; ¹H-NMR (CDCl₃): δ 1.20 (t, 3H, J = 7.2 Hz, OCH₂CH₃), 4.12 (q, 2H, J = 7.2 Hz, OCH₂CH₃), 7.21 (t, 1H, J = 7.2 Hz, Ar-H), 7.31~7.36 (m, 2H, Ar-H), 7.67~7.75 (m, 3H, Ar-H), 8.56 (s, 1H, Ar-H), 8.94 (d, 1H, J = 8.9 Hz, Ar-H), 9.30 (d, 1H, J = 7.2 Hz), 11.5 (br, 1H, NH); ¹³C-NMR (CDCl₃): δ 14.2, 61.1, 104.8, 105.2, 117.7, 122.9, 124.8, 129.6, 135.6, 141.1, 144.6, 148.8, 155.2, 165.3; IR: v 1725, 1678, 1624, 1538 cm⁻¹; MS (ESI): m/z 334 [M+1], 356 [M + Na]⁺.

Ethyl 1-(5(6)-chloro-1H-benzimidazol-2-yl)-4-oxo-4H-quinolizine-3-carboxylate (**13b**): Yield 83 %; m.p.: 216-217 °C; ¹H-NMR (CDCl₃): δ 1.18 (t, 3H, J = 7.2 Hz, OCH₂CH₃), 4.10 (q, 2H, J = 7.2 Hz, OCH₂CH₃), 7.23-7.29 (m, 2H, Ar-*H*), 7.63-7.76 (m, 3H, Ar-*H*), 8.57 (s, 1H, Ar-*H*), 9.04 (d, 1H, J = 8.8 Hz, Ar-*H*), 9.33 (d, 1H, J = 4.8 Hz, Ar-*H*), 11.8 (br, 1H, N*H*); ¹³C-NMR (CDCl₃): δ 14.1, 61.0, 104.4, 104.9, 117.9, 123.3, 124.7, 128.3, 129.6, 135.8, 141.1, 144.5, 150.0, 155.1, 165.1; IR: *v* 3444, 3219, 1730, 1644, 1488, 1106 cm⁻¹; MS (ESI): *m/z* 366 [M-1]⁻.

Ethyl 1-(5(6)-methoxy-1H-benzimidazol-2-yl)-4-oxo-4H-quinolizine-3-carboxylate (**13c**): Yield 88%; m.p.: 234-236 °C; ¹H-NMR (CDCl₃): δ 1.20 (t, 3H, J = 7.2 Hz, OCH₂CH₃), 3.89 (s, 3H, OCH₃), 4.13 (q, 2H, J = 7.2 Hz, OCH₂CH₃), 6.95 (d, 1H, J = 8.8, Ar-H), 7.21-7.71 (m, 3H, Ar-H), 8.24 (d, 1H, J = 8.0 Hz, Ar-H), 8.54 (s, 1H, Ar-H), 8.95 (d, 1H, J = 8.8 Hz, Ar-H), 9.30 (d, 1H, d, J = 7.2 Hz, Ar-H); ¹³C-NMR (CDCl₃): δ 165.36, 156.81, 155.19, 144.55, 140.98, 135.40, 134.56, 129.53, 129.30, 124.83, 123.49, 117.66, 105.13, 105.08, 61.05, 55.85, 14.18; IR: v 3435, 1723, 1615, 1450 cm⁻¹; MS (ESI): m/z 362 [M-1]⁻.

General procedure for the synthesis of compounds 14a-c

To a 100 mL of round flask charged with compound **13** (1 mmol) and ethanol (20 mL) was added KOH (560 mg, 10 mmol) dissolved in water (5 mL). The reaction mixtures were heated to reflux till TLC confirmed that the reactions had finished. After removal of the solvent, the residue was redissolved in water (20 mL) and then acidified to pH 2. The resulting suspensions were filtered, dried and purified by column chromatography (eluents: CHCl₃/CH₃OH) to obtain corresponding **14a-c** as yellow solids.

1-(1H-Benzimidazol-2-yl)-4-oxo-4H-quinolizine-3-carboxylic acid (**14a**): Yield 20 %. m.p.: >300 °C; ¹H-NMR (DMSO-*d*₆): δ7.23 (m, 2H, Ar-*H*), 7.53 (m, 1H, Ar-*H*), 7.70 (m, 1H, Ar-*H*), 7.79 (t, 1H, J = 6.8 Hz, Ar-*H*), 8.28 (t, 1H, J = 5.6 Hz, Ar-*H*), 8.97 (s, 1H, Ar-*H*), 9.47 (d, 1H, J = 6.8 Hz, Ar-*H*), 9.85 (d, 1H, J = 9.2 Hz, Ar-*H*), 13.14 (s, 1H, N*H*), 13.75 (s, 1H, COOH); ¹³C-NMR (DMSO-*d*₆): δ 104.6, 107.0, 111.8, 119.2, 120.6, 122.3, 123.3, 125.8, 130.2, 134.8, 138.1, 139.4, 143.9, 144.2, 149.1, 159.4, 166.1; IR: *v* 1705, 1612, 1497, 1443 cm⁻¹; HRMS: *m/z* calcd for C₁₇H₁₀N₃O₃: 304.0718; found: 304.0727.

1-(5(6)-Chloro-1H-benzimidazol-2-yl)-4-oxo-4H-quinolizine-3-carboxylic acid (**14b**): Yield: 35 %. m.p.>300 °C; ¹H-NMR (DMSO-*d*₆) δ: 7.43 (dd, 1H, *J* = 8.4 Hz, *J* = 1.6 Hz, Ar-*H*), 7.74 (d, 1H, *J* = 8.4 Hz, Ar-*H*), 7.78 (s, 1H, Ar-*H*), 7.79 (t, 1H, *J* = 7.2 Hz, Ar-H), 8.25 (t, 1H, *J* = 7.6 Hz, Ar-*H*), 8.89 (s, 1H, Ar-*H*), 9.09 (d, 1H, J = 8.8 Hz, Ar-*H*), 9.45 (d, 1H, J = 7.2 Hz, Ar-*H*); ¹³C-NMR (DMSO- d_6) δ : 101.9, 104.9, 114.6, 116.3, 120.7, 124.9, 125.2, 129.2, 130.6, 133.9, 136.0, 138.9, 141.7, 144.2, 148.6, 158.6, 165.7; IR: *v* 3434, 1718, 1612, 1499 cm⁻¹; MS (ESI) *m/z*: 338 [M-1]⁻.

l-(5(6)-*Methoxy*-1*H*-benzimidazol-2-yl)-4-oxo-4*H*-quinolizine-3-carboxylic acid (**14c**): Yield 45 %. m.p.: >300 °C; ¹H-NMR (DMSO-d₆): δ 3.86 (s, 3H, OCH₃), 7.06 (dd, 1H, J = 8.4 Hz, J = 2.0 Hz, Ar-*H*), 7.21 (d, 1H, J = 2.0 Hz, Ar-*H*), 7.66 (d, 1H, J = 8.8 Hz, Ar-*H*), 7.80 (t, 1H, J = 6.8 Hz, Ar-*H*), 8.26 (t, 1H, J = 8.4 Hz,Ar-*H*), 8.88 (s, 1H, Ar-*H*), 9.09 (d, 1H, J = 8.8 Hz, Ar-*H*), 9.48 (d, 1H, J = 7.2 Hz, Ar-*H*); ¹³C-NMR (DMSO-d₆): δ 165.76, 158.73, 157.63, 146.81, 144.11, 141.15, 138.64, 136.03, 130.49, 129.85, 125.03, 120.65, 115.89, 114.55, 104.84, 103.86, 97.09, 56.24; IR: v 3415, 1728, 1618 cm⁻¹; MS (ESI): *m/z* 334 [M-1]⁻.

Acknowledgements

This work was supported by Grants from Beijing Novel Project (No. 2005B10), Beijing Natural Science Foundation (No. 2062003) and the National Natural Science Foundation of China (No. 20772010).

References

- 1. Turner, B.G.; Summers, M.F. Structural biology of HIV. J. Mol. Biol. 1999, 285, 1-32.
- 2. Molla, A.; Granneman, G.R.; Sun, E.; Kempf, D.J. Recent developments in HIV protease inhibitor therapy. *Antiviral Res.* **1998**, *39*, 1-23.
- 3. Moyle, G.; Gazzard, B. Current knowledge and future prospects for the use of HIV protease inhibitors. *Drugs* **1996**, *51*, 701-702.
- 4. Pomerantz, R.J.; Horn, D.L. Twenty years of therapy for HIV 1 infection. *Nature Med.* 2003, 9, 867-873.
- 5. De Clercq, E. New developments in anti-HIV chemotherapy. *Biochim. Biophys. Acta* **2002**, *1587*, 258-275.
- 6. Brown, P.O. Integration of retroviral DNA. Curr. Top. Microbiol. Immunol. 1990, 157, 19-48.
- 7. Katz, R.A.; Skalka, A.M. The retroviral enzymes. Annu. Rev. Biochem. 1994, 63, 133-173.
- 8. Farnet, C.M.; Wang, B.; lipford, J.R.; Bushman, F.D. Differential inhibition of HIV-1 preintegration complexes and purified integrase protein by small molecules. *Proc. Natl. Acad. Sci. USA* **1996**, *93*, 9742-9747.
- Molteni, V.; Rhodes, D.; Rubins, K.; Hansen, M.; Bushman, F.D.; Siegel, J.S. A new class of HIV-1 integrase inhibitors: the 3,3,3',3'-tetramethyl-1,1'-spirobi(indan)-5,5',6,6'-tetrol family. J. Med. Chem. 2000, 43, 2031-2039.
- Hazuda, D.J.; Felock, P.; Witmer, M.; Wolfe, A.; Stillmock, K.; Grobler, J.A.; Espeseth, A.; Gabryelski, L.; Schleif, W.; Blau, C.; Miller, M.D. Inhibitors of strand transfer that prevent integration and inhibit HIV-1 replication in cells. *Science* 2000, 287, 646-650.
- 11. Hazuda, D.J.; Yong, S.D.; Guare, J.P.; Anthony, N.J.; Gomez, R.P.; Wai, J.S.; Vacca, J.P.; Handt, L.; Motzel, S.L.; Klein, H.J.; Tussey, L.; Schleif, W.A.; Gabryelski, L.S.; Jin, L.; Miller, M.D.;

Casimiro, D.R.; Emini, E.A.; Shiver, J.W. Integrase inhibitors and cellular immunity suppress retroviral replication in rhesus macaques. *Science* **2004**, *305*, 528-532.

- Espeseth, A.S.; Felock, P.; Wolfe, A.; Witmer, M.; Grobler, J.; Anthony, N.; Egbertson, M.; Melamed, J.Y.; Young, S.; Hamill, T.; Cole, J.L.; Hazuda, D.J. HIV-1 integrase inhibitors that compete with the target DNA substrate define a unique strand transfer conformation for integrase. *Proc. Natl. Acad. Sci. USA* 2000, *97*, 11244-11249.
- Goldgur, Y.; Craigie, R.; Cohen, G.H.; Fujiwara, T.; Yoshinaga, T.; Fujishita, T.; Sugimoto, H.; Endo, T.; Murai, H.; Davies, D.R. Structure of the HIV-1 integrase catalytic domain complexed with an inhibitor: a platform for antiviral drug design. *Proc. Natl. Acad. Sci. USA* 1999, 96, 13040-13043.
- Grobler, J.A.; Stillmock, K.; Hu, B.; Witmer, M.; Felock, P.; Espeseth, A.; Wolfe, A.; Egbertson, M.S.; Bourgeois, M.; Melamed, J.Y.; Wai, J.S.; Young, S.D.; Vacca, J.P.; Hazuda, D.J. Diketo acid inhibitor mechanism and HIV-1 integrase: implications for metal binding in the active site of phosphotransferase enzymes. *Proc. Natl. Acad. Sci. USA* 2002, *99*, 6661-6666.
- Zhuang, L.H.; Wai, J.S.; Embrey, M.W.; Fisher, T.E.; Egbertson, M.S.; Payne, L.S.; Juare, Jr. J.P.; Vacca, J.P.; Hazuda, D.J.; Felock, P.J.; Wolfe, A.L.; Stillmock, K.A.; Witmer, M.V.; Moyer, G.; Schleif, W.A.; Gabryelsk, L.J.; Leonard, Y.M.; Lynch, Jr. J.J.; Michelson, S.R.; Young, S.D. Design and Synthesis of 8-Hydroxy-[1,6]Naphthyridines as Novel Inhibitors of HIV-1 Integrase in Vitro and in Infected Cells. *J. Med. Chem.* 2003, *46*, 453-456.
- Summa, V.; Petrocchi, A.; Matassa, V.G.; Taliani, M.; Laufer, R.; Francesco, R.D.; Altamnra, S.; Pace, P. HCV NS5b RNA-Dependent RNA Polymerase Inhibitors: From α, γ -Diketoacids to 4,5-Dihydroxypyrimidine- or 3-Methyl-5- hydroxypyrimidinonecarboxylic Acids. Design and Synthesis. J. Med. Chem. 2004, 47, 5336-5339.
- Li, X.-M.; Zeng, C.-C.; Jiao, Z.-G.; Yan, H.; Zhong, R.-G. Synthesis and crystal structure of 7acetamido-5-chloro-8-(p-methyloxybenzenesulfonyloxy)-2-styrylquinoline. *Chinese J. Struct. Chem.* 2007, 26, 669-673.
- 18. Xu, Y.-S.; Zeng, C.-C.; Li, X.-M.; Zhong, R.-G.; Zeng, Y. Design, synthesis and Cu2+ recognition of β -diketoacid and quinoxalone derivatives bearing caffeoyl or galloyl moieties linked by arylamide as potential HIV integrase inhibitors. *Chin. J. Chem.* **2006**, *24*, 1086-1094.
- Zeng, C.-C.; Li, X.-M.; Yan, H.; Zhong, R.-G. Design and synthesis of p/m-[p-(un)substituted phenylsulfonamido]phenyl β -diketo acids and quinoxalone derivatives. *Chin. J. Chem.* 2007, 25, 1174-1182.
- 20. Zeng, C.-C.; Liu, C.-F.; Zeng, J.; Zhong, R.-G. Electrochemical synthesis of 6-arylsulfonyl caffeic acid derivatives in aqueous medium. *J. Electroanal. Chem.* **2007**, *608*, 85-90.
- Zeng, C.-C; Ping, D.-W.; Zhang, S.-C.; Zhong, R.-G.; Becker, J.Y. Electrochemical synthesis of polyhydroxylated aromatic derivatives substituted with mono- and dipyrimidinyl thioethers in aqueous medium. *J. Electroanal. Chem.* 2008, 622, 90-96.
- Zeng, J.; Lv, X.-H.; Zeng, C.-C.; Hu. L.-M, Zhong, R.-G. Design, synthesis and anti-HIV integrase evaluation of 1,2,3-triazol -4-yl-substituted 1,4-dihydro-4-oxo-1,5-napthyridine-3-carboxylic acids. *Chin. J. Chem.* 2009 (In press).
- 23. Komatsu, H.; Iwasawa, N.; Citterio, D.; Suzuki, Y.; Kubota, T.; Tokuno, K.; Kitamura, Y.; Oka, K.; Suzuki, K. Design and synthesis of highly sensitive and selective fluorescein-derived

magnesium fluorescent probes and application to intracellular 3D Mg²⁺ imaging. J. Am. Chem. Soc. **2004**, 126, 16353-16360.

- Otten, P.A.; London, R.E.; Levy, L.A. 4-Oxo-4H-quinolizine-3-carboxylic acids as Mg²⁺ selective, fluorescent indicators. *Bioconjugate Chem.* 2001, *12*, 203-212.
- 25. Li, Q.; Chu, D.T.W.; Claiborne, A.; Cooper, C.S.; Lee, C.M.; Raye, K.; Berst, K.B.; Donner, P.; Wang, W.; Hasvold, L.; Fung, A.; Ma, Z.; Tufano, M.; Flamma, R.; Shen, L.L.; Baranowski, J.; Nilius, A.; Alder, J.; Meulbroek, J.; Marsh, K.; Crowell, D.; Hui, Y.; Seif, L.; Melcher, L.M.; Henry, R.; Spanton, S.; Faghih, R.; Klein, L.L.; Ken Tanaka, S, Plattner, J.J. Synthesis and Structure-Activity Relationships of 2-Pyridones: A Novel Series of Potent DNA Gyrase Inhibitors as Antibacterial Agents. *J. Med. Chem.* **1996**, *39*, 3070-3088.
- Oya, S.; Masuda, N.; Kuroki, Y.; Inoue, T.; Okudo, M.; Iwata, T.; Kokubo, K.; Mizuno, H.; Hagiwara, M. Preparation of 4-oxo-4H-quinolizine-3-carboxylic acid derivatives as antibacterial agents. *Jpn. Kokai Tokkyo Koho* 2003, JP 2003261566 A 20030919
- Fukumoto, R.; Niwano, Y.; Kusakabe, H.; Chu, C.; Kimura, H.; Nagasawa, K.; Yanagihara, S.; Hirosawa, C.; Ishiduka, S. Preparation of 4-oxoquinolizines as antimicrobial agents. *PCT Int. Appl.* 2003, WO 2003029253
- Iwamoto, K.; Araki, K.; Fujishima, H.; Shinkai, S. Fluorogenic calix[4]arene. J. Chem. Soc. Perkin Trans. 1. 1992, 1885-1887.
- 29. Kim, Y.H.; Youk, J.S.; Moon, S.Y.; Choe, J.I.; Chang, S.K. Hg²⁺-selective fluorogenic chemosensor derived from 8-Aminoquinoline. *Chem. Lett.* **2004**, *33*, 702-703.
- 30. Kim, S.J.; Kool, E.T. Sensing Metal Ions with DNA Building Blocks: Fluorescent Pyridobenzimidazole Nucleosides. J. Am. Chem. Soc. 2006, 128, 6164-6171.
- Weatherhead-Kloster, R.A.; Selby, H.D.; Miller III, W.B.; Mash, E.A. Organic Crystal Engineering with 1,4-Piperazine-2,5-diones. 6. Studies of the Hydrogen-Bond Association of Cyclo[(2-methylamino-4,7-dimethoxyindan-2-carboxylicacid)(2-amino-4,7-dimethoxyindan-2carboxylic acid)]. J. Org. Chem. 2005, 70, 8693-8702.
- Kawasuji, T.; Fuji, M.; Yoshinaga, T.; Sato, A.; Fujiwara, T.; Kiyama, R. A platform for designing HIV integrase inhibitors. A two-metal binding model as a potential mechanism of HIV integrase inhibitors. *Bio. & Med. Chem.*, 2006, 14, 8420-8429

Sample Availability: Samples of compounds 6-14 are available from the authors.

© 2009 by the authors; licensee Molecular Diversity Preservation International, Basel, Switzerland. This article is an open-access article distributed under the terms and conditions of the Creative Commons Attribution license (http://creativecommons.org/licenses/by/3.0/).