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Article

Platycoside N: A New Oleanane-Type Triterpenoid Saponin from the Roots of *Platycodon grandiflorum*

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Abstract: A new oleanane-type triterpenoid saponin, named platycoside N (1), together with six known saponins, was isolated from the roots of *Platycodon grandiflorum*. On the basis of acid hydrolysis, comprehensive spectroscopic data analyses and comparison with the spectral data of the known compounds, its structure was elucidated as 3-*O*- β -Dglucopyranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl-2 β ,3 β ,1 6α ,23-tetrahydroxyolean-12-en-28-oic acid 28-*O*- β -L-rhamnopyranosyl-(1 \rightarrow 2)- α -L-arabinopyranoside. The six known compounds were platycodin D (2), deapioplatycodin D (3), platycodin D₃ (4), deapioplatycodin D₃ (5), platycoside E (6) and deapioplatycoside E (7).

Keywords: platycoside N; Platycodon grandiflorum; triterpenoid saponin

1. Introduction

8.

The roots of *Platycodon grandiflorum* A. DC (Campanulaceae), Platycodi Radix, are traditionally used as food and a herbal medicine in the treatment of a wide range of diseases, including bronchial asthma, hepatic fibrosis, bone disorders [1-4], hypercholesterolemia and hyperlipidemia [5]. The principal bioactive constituents of this herb are triterpenoid saponins (platycosides), which exhibit a variety of pharmacological activities, such as anti-inflammatory [6,7], anti-obesity [8-11], anti-cancer [12-15] and hypoglycemic effects [16,17]. To date, more than 30 saponins have been isolated from this plant [18-25]. In order to find more bioactive compounds, we have now studied the chemical constituents of *P. platycodiflorum*, and in this paper, we report the presence in this species of a new oleanane-type triterpenoid saponin, named platycoside N, together with six known compounds, from the roots of *Platycodon grandiflorum*. (Figure 1)



Figure 1. Chemical structures of compounds 1-7.

2. Results and Discussion

Platycoside N (1) was a white amorphous powder, and its molecular formula $C_{53}H_{86}O_{24}$ was determined based on the HR-ESI-MS spectra. The oleanane-type triterpenoid saponin nature of compound 1 was revealed through analysis of its spectral features. The IR spectrum exhibited absorptions at 3,425 cm⁻¹ (OH), 1,647 cm⁻¹ (ester carbonyl), and 1,616 cm⁻¹ (double bond). Six methyl groups (δ 0.89 × 2, 0.98, 1.17, and 1.58 × 2) and one olefinic proton (δ 5.46, br s) of the aglycon were observed in the ¹H-NMR spectrum. The ¹³C-NMR spectrum showed that the aglycon had six methyl carbons at δ 16.0, 18.6, 17.7, 27.2, 33.3, and 24.8, two olefinic carbons at δ 123.1 (CH) and 144.5 (C), one oxymethylene and three oxymethine carbons at δ 66.6, and 70.1, 74.2 and 83.1, respectively, and one carbonyl carbon at δ 176.0 (Table 1). The information of the ¹H-NMR spectrum

coupled with the ¹³C-NMR spectrum indicated that **1** had $2\beta_{3}\beta_{1}6\alpha_{2}$ -tetrahydroxyolean-12-en-28oic acid (polygalacic acid) as an aglycon [20]. The ¹³C-NMR spectrum showed 53 signals, of which 30 were assigned to a triterpenoid moiety and 23 to the saccharide portion. The downfield shift of C-3 (\delta 83.1) and for the upfield shift of C-28 (δ 176.0), revealed that the sugar moieties were attached to the aglycon at these two positions. The ¹H and ¹³C-NMR spectra of **1** exhibited four anomeric protons at δ 5.10×2 (2H, d, J = 7.5 Hz), 6.27(1H, d, J = 2.5 Hz), 5.68 (1H, br s) ppm and carbons at $\delta 106.7 \times 2$, 93.9, 101.5 (Table 1). In the ¹H-NMR spectrum, one methyl signal at δ 1.59 (3H, d, J = 5.5 Hz) belonging to rhamnose was observed. In addition, the monosaccharides were identified as glucose, rhamnose and arabinose by TLC and a combination of DEPT, HMQC and HMBC experiments. Acid hydrolysis of 1 also gave glucose, arabinose and rhamnose in a ratio of 2:1:1 respectively, as confirmed by GC analysis of the respective trimethylsilyl derivatives [20]. The ¹H- and ¹³C-NMR and 2D-NMR analysis indicated that all the monosaccharides of **1** were in pyranose forms. The β -anomeric configurations of the D-gulucose units were determined by its ${}^{3}J_{H1 H2}$ coupling constants (7.5 Hz). The α-anomeric configurations of the L-arabinose and L-rhamnose were determined by the broad singlet of their anomeric protons [24]. The linkages between sugar moieties and C-3 of the aglycon were corroborated through HMBC experiments, i.e., H-1 (8 5.10) of the terminal glucose correlated with C-6 (δ 70.8) of the inner glucose, and H-1 (δ 5.10) of the inner glucose correlated with C-3 (δ 83.1) of the sapogenin. The linkages of sugar moieties at C-28 were established based on HMBC correlations between H-1 (& 5.68) of rhamnose and C-2 (& 75.3) of arabinose, and H-1 (& 6.27) of arabinose and C-28 (δ 176.0) of aglycone (Figure 2). On the basis of all the above evidence, platycoside N (1) was identified as 3-O- β -D-glucopyranosyl-(1 \rightarrow 6)- β -D-glucopyranosyl-2 β ,3 β ,16 α ,23-tertahydroxyolean-12en-28-oic acid 28-*O*- β -L-rhamnopyranosyl-(1 \rightarrow 2)- α -L-arabinopyranoside.

Position	δ_{C}	Position	δ _C
1	46.8	3-O-Glc	
2	70.1	1	106.7
3	83.1	2	75.3
4	42.4	3	78.7
5	47.5	4	72.1
6	20.5	5	76.0
7	33.6	6	70.8
8	40.2	Glc	
9	47.5	1'	106.7
10	37.2	2'	74.9
11	24.4	3'	78.7
12	123.1	4′	71.6
13	144.5	5'	78.7
14	42.4	6'	62.3
15	36.2	Ara	
16	74.2	1	93.9
17	48.6	2	75.3
18	41.5	3	70.8

Table 1. ¹³C-NMR data of compound **1** in pyridine– d_5 (δ ppm).

19	47.2	4	66.6
20	31.0	5	63.7
21	36.2	Rha	
22	32.2	1	101.5
23	66.6	2	71.6
24	16.0	3	72.7
25	18.6	4	73.1
26	17.7	5	68.7
27	27.2	6	18.6
28	176.0		
29	33.3		
30	24.8		

 Table 1. Cont.

Figure 2. The key mulde conclutions of compound 1 (nom 11 to e	Figure 2. The key HMBC correlations of co	mpound 1 (from H to C
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The six known saponins were identified as platycodin D (2), deapio platycodin D (3), platycodin D₃ (4), deapio platycodin D₃ (5), platycoside E (6) and deapio platycoside E (7) through comparison of their UV, IR, NMR and MS data with literature values [25,26].

3. Experimental

3.1. General

ESI-MS (negative mode) measurements were carried out on an Agilent 1100 series LC/MSD Trap SL mass spectrometer. HR-ESI-MS (positive and negative modes) was analyzed on a Bruker FT-ICRMS spectrometer. IR spectra were recorded on an IR-47 spectrometer. NMR spectra were recorded on a Bruker Avance DRX 400 NMR spectrometer using TMS as internal standard, and chemical shifts δ were given in ppm. Silica gel (200–300 mesh) for column chromatography and silica

gel G for TLC were purchased from Qingdao Marine Chemical Factory, Qingdao, China. AB-8 macroporous resin was purchased from Tianjin Nankai factory. Preparative HPLC was performed on a Waters 600 liquid chromatography instrument with a UV detector, monitored at 210 nm using a C18 column (Zorbax Eclipse XDB, 250 mm \times 9 mm; 10 µm)

3.2. Plant material

The roots of *P. grandiflorum* were purchased at Changchun Guangfulu market in Changchun-city of Jilin province, China and identified by Prof. Yi-Nan Zheng, College of Chinese Material Medicine, Jilin Agricultural University. A voucher specimen (No.20050116) has been deposited in the herbarium of the same college.

3.3. Extraction and isolation

Dry and powdered roots of *P. grandiflorum* (2.0 Kg) were refluxed three times with 30 L of 70% methanol, 3 h each time. Extracts were concentrated, suspended in water and sequentially partitioned with ethyl acetate and *n*-butanol. The *n*-butanol fraction was subjected to macroporous resin AB-8 column and eluted sequentially with water, 30% ethanol and 70% ethanol. The 30% ethanol elution was repeatedly chromatographed on a reverse-phase column, and eluted with aqueous methanol, affording three fractions A-C. Fraction A was purified by HPLC to afford compounds **1** (23 mg), **2** (15 mg) and **3** (40 mg). Fraction B and C gave **4** (32 mg), **5** (22 mg), **6** (16 mg) and **7** (15 mg). Platycoside N (**1**): White amorphous powder; IR (KBr) cm⁻¹: 3425, 2947, 1647, 1616, 1114; ESI-MS *m/z*: 1105 [M-H]⁻, HR-ESI-MS *m/z* 1105.5408 [M-H]⁻ (Calcd for C₅₃H₈₅O₂₄, 1105.5431). ¹H-NMR (400 MHz, pyridine-*d*₅) δ : 0.89 × 2, 0.98, 1.17, 1.58 × 2 (each 3H, s, CH₃ of C-26, C-29, C-30, C-24, C-25, C-27), 1.59 (3H, d, CH₃ of rhamnose), 4.36 (1H, d, *J* = 3.0Hz, H-3), 3.82, 4.60 (each 1H, d, H-23), 4.72(1H, m, H-2), 5.10 × 2 (each 1H, d, *J* = 7.5 Hz, H-1' and H-10f glucose), 5.68 (1H, br s, H-1 of rhamnose), 5.46 (1H, br s, H-12) 5.03 (1H, br s, H-16). ¹³C-NMR (100 MHz, pyridine-*d*₅) data: see Table 1.

3.4. Acid hydrolysis of 1

Compound **1** (2.0 mg) was refluxed with 4.0 M HCl (5.0 mL) for 1 h at 95 °C, and the reaction mixture was extracted with ethyl acetate. The aqueous layer was then adjusted to pH 7.0 with NaHCO₃. After evaporating to dryness, the sugar mixture was dissolved in pyridine and developed on silica gel TLC [CHCl₃-MeOH-H₂O (7:3:0.5, lower phase), *n*-BuOH-AcOH-H₂O (4:1:5, upper phase). Three spots were seen on the TLC after spraying with 4% α -naphthol-EtOH-5% H₂SO₄. Through comparison with authentic sugar standards (purchased from Sigma), it was found that compound **1** possessed D-glucose, L-rhamnose and L-arabinose units.

4. Conclusions

In summary, we have isolated a new oleanane-type triterpenoid saponin, named platycoside N (1), together with six known saponins from the roots of *Platycodon grandiflorum*.

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Sample Availability: Samples of the compounds are available from the authors (liwei7727@126.com).

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