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Article

Enantioselective Addition of Allyltin Reagents to Amino Aldehydes Catalyzed with Bis(oxazolinyl)phenylrhodium(III) Aqua Complexes

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Abstract: Bis(oxazolinyl)phenylrhodium(III) aqua complexes, (Phebox)RhX₂(H₂O) [X = Cl, Br], were found to be efficient Lewis acid catalysts for the enantioselective addition of allyl- and methallyltributyltin reagents to amino aldehydes. The reactions proceed smoothly in the presence of 5–10 mol % of (Phebox)RhX₂(H₂O) complex at ambient temperature to give the corresponding amino alcohols with modest to good enantioselectivity (up to 94% ee).

Keywords: allylation; amino aldehydes; Lewis acids; pincer ligands; rhodium

1. Introduction

The development of enantioselective synthesis of chiral homoallylic alcohols containing aminofunctional groups is of great importance to synthetic organic and medicinal chemistry. Despite much effort directed at enantioselective allylation of aldehydes [1-3], there are only a few systems for enantioselective allylation of amino aldehydes as substrates because the high coordination ability of amino groups to the metal species often leads to deactivation of chiral allylmetals or catalysts. Therefore, most of these reactions need a stoichiometric amount of chiral sources. For examples, Brown [4] and Chen [5] reported the utility of allylboron reagents (Figure 1, **A**–**C**) for the reaction with pyridinecarboxaldehydes and 1-methyl-2-pyrrolecarboxaldehyde [4,5]. Denmark and co-workers developed a new reaction system for the allylation of aldehydes, but the enantioselectivity of the reaction with 4-dimethylaminobenzaldehyde was not so high (Figure 1, **D**) [6]. The other one is a catalytic reaction using 20 mol % of BINOL-derived chiral titanium complex/allyltributyltin *via* transmetalation mechanism reported by Umani-Ronchi (Figure 1, **E**) [7].

Figure 1. Chiral allylmetal reagents for the asymmetric allylation of amino aldehydes.



We have previously developed a meridional tridentate ligand, 2,6-bis(oxazolinyl)phenyl derivative (abbreviated to Phebox) as a chiral N–C–N pincer type ligand with one central covalent bond to a metal [8-14], and have demonstrated that rhodium(III) aqua complexes bearing the Phebox ligand, (Phebox)RhX₂(H₂O) [1: X = Cl, 2: X = Br], acted as recoverable chiral Lewis acid catalysts for the enantioselective addition of allylic tributyltin reagents to aldehydes [15-17] and the asymmetric hetero Diels-Alder reactions of Danishefsky's dienes and glyoxylates [18]. During the course of our studies on the Phebox-Rh(III) system as a chiral transition metal Lewis acid, we have found that tertiary amines such as *N*,*N*-diisopropylethylamine or triethylamine cannot bind to the rhodium atom [19]. This discovery encouraged us to use these air-stable and water-tolerant complexes **1** and **2** for the allylation of amino aldehydes as substrates. We wish to report herein the Lewis acid-catalyzed enantioselective addition of allyl- and methallyltributyltin reagents to aldehydes containing amino-functional groups (Scheme 1).

Scheme 1. Enantioselective addition of allyltin reagents to amino aldehydes catalyzed with $(Phebox)RhX_2(H_2O)$ complexes (X = Cl, Br).



2. Results and Discussion

2.1. NMR Studies, Isolation, and X-ray Analysis of Phebox-Rh(III)-Amino Aldehyde Complexes

First, we checked the complexation between Phebox-Rh(III) complex *i*-Pr-1 and amino aldehydes **3a-e** by ¹H and ¹³C-NMR. Selected ¹H and ¹³C-NMR data in CDCl₃ are listed in Table 1. Although rigid complexation was not clearly observed between *i*-Pr-1 and 6-methyl-2-pyridinecarboxaldehyde (3c) (Entry 3), ¹H- and ¹³C-NMR spectra of the other cases showed formation of new complexes. From the NMR spectra of a mixture of *i*-Pr-1 and 3a, the pyridine's nitrogen atom exclusively forms σ -complexes with the (*i*-Pr-Phebox)RhCl₂ fragment; the signals of the protons for 2- and 6-positions of the pyridine ring (H₂ and H₆) appeared at lower field than those of the uncomplexed (free) **3a** (from δ 9.08 to 10.30 ppm for H_2 , and from δ 8.85 to 10.08 ppm for H_6 , respectively) (Entry 1). This amine complex was stable enough to be purified by silica gel chromatography and was eventually characterized by a single-crystal X-ray diffraction (Figure 2, Table 2). In the case of the reaction of *i*-Pr-1 and 3b, H_6 and the formyl proton (H_f) both appeared as broad signals at lower field (δ 9.11 for H_6 and 10.34 ppm for H_f) than those of free **3b** (δ 8.77 for H_6 and 10.07 ppm for H_f) (Entry 2). These results indicate that the coordination of **3b** to the (*i*-Pr-Phebox)RhCl₂ fragment is an equilibrium between the pyridinic nitrogen and carbonyl oxygen. In contrast to the pyridinecarboxaldehydes, solutions of 4-dimethylaminobenzaldehyde (3d) and 4-dimethylaminocinnamaldehyde (3e) in the presence of *i*-Pr-1 showed rigid formation of C=O/ σ type aldehyde complexes. For example, the signals assignable to the dimethylamino group were not changed, but the signals of the formyl proton (H_f) and carbon (C_f) of coordinated **3d** appeared at lower field than the uncomplexed (free) **3d** (from δ 9.74 to 9.92 ppm for H_f , and from δ 190.4 to 207.2 ppm for C_f , respectively) (Entry 4). Similar lower field shifts of H_f and C_f along with the olefinic protons H_{α} and H_{β} (H_{α} = α -proton, H_{β} = β -proton) were also observed for the mixture of *i*-Pr-1 and 3e in ¹H and ¹³C NMR ($\Delta H_{\alpha} = +0.27$ ppm, $\Delta H_{\beta} = +0.14$ ppm, $\Delta H_{f} = +0.77$ ppm, $\Delta C_{f} = +2.9$ ppm, respectively) (Entry 5). It is widely known that chemical shifts of vinylic protons (H_{α} and H_{β}), formyl proton (H_{f}) and carbon (C_{f}) of enals bound

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to Lewis acids by the carbonyl oxygen appear at lower field than those of free enals [20-22]. These lower-field shifts of H_{α} , H_{β} , H_{f} and C_{f} are also observed in the reaction of *i*-Pr-1 and (*E*)-cinnamaldehyde [16]. The above NMR and X-ray studies thus indicated that the (Phebox)RhCl₂ fragment, generated by releasing H₂O from (Phebox)RhCl₂(H₂O), captures amino aldehydes **3b–e** at the carbonyl oxygen to form aldehyde complexes (Scheme 2).

Entury		δ (μ	$ \Delta (ppm)^{b}$	
Entry		3 <i>i</i> -Pr-1 and 2		
	Her a	H ₂ : 9.08	H ₂ : 10.30	+1.22
1		H ₆ : 8.85	H ₆ : 10.08	+1.23
1	N C _f H _f	H _f : 10.12	H _f : 10.27	+0.15
	H ₂ Ö 3a	C _f : 190.8	C _f : 189.8	-1.0
		H.: 8 77	$H_{c} = 0.11$ (br)	+0.34
2		H_{6} : 0.77	$H_6: 10.34$ (br)	+0.34
	⊓ ₆ N C _f Ö 3b	11 ₁ . 10.07	$\Pi_{\rm I}$. 10.34 (01)	10.27
	~	H _{Me} : 2.66	H _{Me} : 2.66	0.00
3		H _f : 10.04	H _f : 10.06	+0.02
		C _{Me} : 24.5	C _{Me} : 24.5	0.0
	^{ги} е Н _{Ме} Ö Зс	C _f : 193.1	C _f : 194.1	+1.0
	Me	H _{Me} : 3.09	H _{Me} : 3.11	+0.02
	H _{Mex} , N _x	H _f : 9.74	H _f : 9.92	+0.18
4		C _N : 40.2	C _N : 40.2	0.0
	H _{Me} C _f H _f	C _f : 190.4	C _f : 207.2	+16.8
	Ö 3d			
		H _α : 6.54	H _α : 6.81	+0.27
5	Ме	H _β : 7.38	H _β : 7.52	+0.14
		H _{Me} : 3.05	H _{Me} : 3.07	+0.02
	$H_{Me} \stackrel{\Theta N}{\vdash} \qquad \qquad$	H _f : 9.09	H _f : 9.86	+0.77
		C _N : 40.2	C _N : 40.2	0.0
	Η _β Ο 3e	C _f : 193.8	C _f : 196.7	+2.9

Table 1. Selected spectroscopic data for free amino aldehydes 3 and mixtures of *i*-Pr-1 and 3.

^a Observed at 400 MHz for ¹H-NMR and 100 MHz for ¹³C-NMR in CDCl₃ at ambient temperature; ^b Calculated by δ (*i*-Pr-1 and 3) – δ (3).

Scheme 2. Complexation between (Phebox)RhCl₂ fragment ([Rh]) and amino aldehydes.



Figure 2. Molecular structure of (i-Pr-Phebox)RhCl₂(κ -**3a**): there are two independent molecules and one H₂O in the unit cell.



Table 2. Selected bond distances (Å) and angles (deg) for (*i*-Pr-Phebox)RhCl₂(κ-3a).^a

Rh–C1	1.93(1) [1.89(1)]	Rh–N1	2.05(1) [2.06(1)]
Rh-Cl1	2.340(4) [2.334(4)]	Rh–N2	2.05(1) [2.09(1)]
Rh-Cl2	2.334(4) [2.351(4)]	Rh–N3	2.21(1) [2.27(1)]
		C4–O3	1.25(3) [1.27(4)]
C1-Rh-N3	175.2(6) [178.0(5)]	N1-Rh-N3-C2	54(1) [90(1)]
Cl1-Rh-Cl2	178.0(2) [177.2(2)]	O3-C4-C3-C2	-175(2) [19(3)]
N1-Rh-N2	158.4(5) [157.6(5)]		

^a Bond distances and angles of the second molecule are given in brackets.

2.2. Phebox-Rh(III)-Catalyzed Enantioselective Addition of Allyltributyltin to Amino Aldehydes

We also examined the Phebox-Rh(III)-catalyzed reaction of amino aldehydes and allyltributyltin. Allyltributyltin (4a) was added to a suspension of 4Å molecular sieves (MS 4A), amino aldehydes **3b–e** and 5–10 mol % of (S,S)-(Phebox)RhX₂(H₂O) complexes (1 or 2) in dichloromethane at 25 °C for 24 h. These results are summarized in Table 3. First, the reaction of pyridine-2-carboxaldehyde (3b) proceeded smoothly, but the isolated yield of the allylated product 5b was only 14% after purification of the crude material by silica gel chromatography (Entry 1). This result indicates that the alkoxystananne **5b-Sn** formed in the reaction mixture is stable and hardly hydrolyzed under the usual workup process (see Experimental section). Consequently, we adopted a new procedure for conversion of 5b-Sn to the acetate derivative 5b-Ac by treatment with acetic anhydride (Scheme 3). In this manner, **5b-Ac** was obtained in good to high yields and with moderate enantioselectivity (Entries 2, 3, 5, and 6). The absolute configuration of **5b-Ac** obtained by (S,S)-Phebox-derived Rh(III) complexes was determined to be S by comparison of the optical rotation value with literature data [23]. In the case of the reaction using (S,S)-Ph-1, however, ca. 50% of 3b was recovered after silica gel chromatography and (R)-5b-Ac was formed as a major enantiomer (Entry 4). Finally, an enantioselectivity of up to 59% ee was achieved using 10 mol % of the *i*-Pr- and Me-Phebox-derived dibromide complexes (Entries 5 and 6). In the cases of the other aldehydes 3c-e, the products were obtained as a homoallylic alcohol in good to high yields. The reactions of 6-methyl-2-pyridinecarboxaldehyde (3c) and 4-dimethylaminocinnamaldehyde (3e) afforded the corresponding amino alcohols 5c and 5e with good enantioselectivity by using the dibromide complexes (**5c:** 84% ee with Ph-2, and **5e:** 88% ee with Bn-2, respectively) (entries 9 and 15). In sharp contrast, the dichloride complexes showed higher enantioselectivity (84% ee for Me-1, and 81% ee for *s*-Bu-1, respectively) than the parent dibromide one (72% ee for Me-2) in the allylation reaction of 4-dimethylamino- benzaldehyde (**3d**) (Entries 10–12). We also examined the reaction of 3-pyridinecarboxaldehyde (**3a**), however, no allylated product was obtained and the amine complex, (*i*-Pr-Phebox)RhCl₂(κ -**3a**), free **3a**, and allyltributyltin (**4a**) were detected by ¹H-NMR of the crude material.

Scheme 3. Conversion of 5b-Sn to 5b-Ac by treatment with Ac₂O.



Entry	Aldehyde	Catalyst	Product	% Yield	% ee ^b (config.) ^c
1		<i>i</i> -Pr- 1	5b	14	42 (<i>S</i>)
2 ^d		<i>i</i> -Pr- 1	5b-Ac	99	53 (<i>S</i>)
3 ^d		Me-1	5b-Ac	99	56 (<i>S</i>)
4 ^d		Ph- 1	5b-Ac	45	21 (<i>R</i>)
5 ^d		<i>i</i> -Pr- 2	5b-Ac	81	59 (<i>S</i>)
6 ^d	N CHO 3b	Me-2	5b-Ac	85	59 (S)
7		<i>i</i> -Pr- 1	5c	94	$69(S)^{e}$
8		Ph -1	5c	89	$75(S)^{e}$
9		Ph-2	5c	97	84 $(S)^{e}$
10		Me-1	5d	80	84 (S)
11		<i>s</i> -Bu- 1	5d	67	81 (S)
12	Me ₂ N	Me-2	5d	42	72 (<i>S</i>)
	CHO 3d				
13		Bn-1	5e	80	$81(S)^{e}$
14		<i>s</i> -Bu- 1	5e	61	$80(S)^{e}$
15	Me ₂ N	Bn-2	5e	44	88 $(S)^{e}$
	CHO 3e				

Table 3. Enantioselective addition of allyltributyltin 4a to amino aldehydes 3b-e.^a

^a All reactions were carried out using 0.5 mmol of **3**, 0.75 mmol of **4**, and 0.025 mmol (5 mol %) of **1** or 0.05 mmol (10 mol %) of **2** in 2 mL of dichloromethane in the presence of MS 4A (250 mg) at 25 °C for 24 h; ^b Determined by chiral HPLC analysis using Daicel CHIRALCEL OD; ^c Assignment by comparison of the sign of optically rotation with reported value; ^d 0.6 mmol of acetic anhydride was added; ^e Assignment by analogy.

2.3. Phebox-Rh(III)-Catalyzed Enantioselective Addition of Methallyltributyltin to Amino Aldehydes

Table 4 summarizes the results obtained for the methallylation of amino aldehydes **3b**–e catalyzed with Phebox-Rh(III) complexes in dichloromethane in the presence of MS 4A at 25 °C for 24 h. In the reactions of pyridinecarboxaldehydes **3b** and **3c**, the enantiomeric excesses of the methallylated products **6b**-Ac and **6c** were moderate (51% ee for **6b**-Ac, and 45% ee for **6c**, respectively) (Entries 1–10). Similar to the reaction of pyridine-2-carboxaldehyde **3b** and allyltributyltin (Table 2, Entry 4), (*S*,*S*)-Ph-**1** afforded the opposite (*R*)-**6b**-Ac as a major enantiomer (Entry 3). Compared to the reactions with pyridinecarboxaldehydes **3b** and **3c**, the (*S*,*S*)-Phebox-Rh-catalyzed reactions of 4-dimethyl-aminobenzaldehyde (**3d**) and 4-dimethylaminocinnamaldehyde (**3e**) with methallyltributyltin (**4b**) afforded the corresponding (*S*)-products with good to high enantioselectivity (90% ee for **6d** and 94% ee for **6e**, respectively) (Entries 11–17). Incidentally, the Phebox-Rh(III) aqua complexes **1** and **2** can be recovered almost quantitatively from the reaction media by silica gel column chromatography.

Entry	Aldehyde	Catalyst	Product	% Yield	% ee ^b (config.) ^c
1 ^d		Bn- 1	6b-Ac	79	15 (<i>S</i>)
2 ^d	N CHO 3b	Me-1	6b-Ac	76	41 (<i>S</i>)
3 ^d		Ph-1	6b-Ac	18	24 (<i>R</i>)
4 ^d		<i>s</i> -Bu- 1	6b-Ac	52	<2 (-)
5 ^d		Me-2	6b-Ac	48	45 (<i>S</i>)
6 ^d		<i>s</i> -Bu- 2	6b-Ac	22	51 (<i>S</i>)
7		Me-1	6c	60	$45(S)^{e}$
8	N CHO 3c	<i>s</i> -Bu- 1	6с	36	$11(S)^{e}$
9		Me-2	6с	21	$10(S)^{e}$
10		<i>s</i> -Bu- 2	6с	26	$26(S)^{e}$
11	Me ₂ N	<i>i</i> -Pr- 1	6d	84	$85(S)^{e}$
12	CHO 3d	Bn- 1	6d	79	90 $(S)^{e}$
13		<i>s</i> -Bu- 1	6d	68	87 $(S)^{e}$
14		Bn- 2	6d	52	$63(S)^{e}$
15	Me ₂ N	<i>i</i> -Pr- 1	6e	52	$80(S)^{e}$
16		<i>s</i> -Bu- 1	6e	74	84 $(S)^{e}$
17	🏹 🏏 `CHO 3e	<i>s</i> -Bu- 2	6e	20	94 $(S)^{e}$

Table 4. Enantioselective addition of methallyltributyltin 4b to amino aldehydes 3b-e.^a

^a All reactions were carried out using 0.5 mmol of **3**, 0.75 mmol of **4**, and 0.025 mmol (5 mol %) of **1** or 0.05 mmol (10 mol %) of **2** in 2 mL of dichloromethane in the presence of MS 4A (250 mg) at 25 °C for 24 h; ^b Determined by chiral HPLC analysis using Daicel CHIRALCEL OD; ^c Assignment by comparison of the sign of optically rotation with reported value; ^d 0.6 mmol of acetic anhydride was added; ^e Assignment by analogy.

3. Experimental

3.1. General

Anhydrous dichloromethane was purchased from Kanto Chemical Co. Carbon tetrabromide, all aldehydes and allyltributyltin were purchased from Tokyo Chemical Industry Co., Ltd. ¹H and ¹³C NMR spectra were measured on a VARIAN Inova-400 (400 MHz) spectrometer. ¹H-NMR (400 MHz) chemical shifts were described in parts per million downfield from tetramethylsilane used as an internal standard ($\delta = 0$) in CDCl₃, unless otherwise noted. ¹³C-NMR (100 MHz) chemical shifts were expressed in parts per million with reference to the residual solvent peak as an internal standard $(\delta = 77.1 \text{ for CDCl}_3)$, unless otherwise noted. IR spectra were measured on a JASCO FT/IR-230 spectrometer. Melting points were measured on a Yanaco MP-J3 apparatus. Elemental analyses were measured on a Yanaco CHN CORDER MT-6 instrument. High performance liquid chromatography (HPLC) analyses were performed with a JASCO PU-980 HPLC pump, UV-975 and 980 UV/VIS detector, and CO-966 column thermostat (at 25 °C) using a Daicel CHIRALCEL OD column. Optical rotations were measured on a JASCO DIP-140 polarimeter. Column chromatography was performed with silica gel (Merck, Art. No. 7734). Analytical thin-layer chromatography (TLC) was performed on glass plates and aluminum sheets pre-coated with silica gel (Merck, Kieselgel 60 F-254, layer thicknesses 0.25 and 0.2 mm, respectively). Visualization was accomplished by UV light (254 nm), anisaldehyde, and phosphomolybdic acid. All reactions were carried out under a nitrogen or argon atmosphere. (Phebox)SnMe₃, (Phebox)RhCl₂(H₂O) **1** and (Bn-Phebox)RhBr₂(H₂O) (Bn-**2**) were prepared by our method [16,18,24]. [(c-octene)₂RhCl]₂ [25] and methallyltributyltin [26] were prepared by the literature methods.

3.2. General Procedure for the Synthesis of (Phebox)RhBr₂(H₂O) Complexes

To a stirred solution of (R-Phebox)SnMe₃ and $[(c\text{-octene})_2RhCl]_2$ in dichloromethane was added tetrabromomethane [*ca*. 8 equivalents with respect to (R-Phebox)SnMe₃] at ambient temperature for 24 h. Then the reaction mixture was concentrated under reduced pressure. Purification of the residue by silica gel chromatography gave (R-Phebox)RhBr₂(H₂O) (R-**2**).

 $(i-Pr-Phebox)RhBr_2(H_2O)$ (i-Pr-2). 51% yield. Pale yellow solid. mp. 167 °C (decomp); IR (KBr) v 3474, 2959, 1622, 1485, 1391, 961, 745 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.94 (d, J = 6.5 Hz, 6H), 0.96 (d, J = 7.0 Hz, 6H), 2.42 (qqd, J = 7.0, 6.5, 2.5 Hz, 2H), 3.43 (bs, 2H), 4.30 (td, J = 8.3, 2.5 Hz, 2H), 4.71 (d, J = 8.3 Hz, 4H), 7.25 (t, J = 7.6 Hz, 1H), 7.58 (d, J = 7.6 Hz, 2H); ¹³C-NMR (CDCl₃) δ 15.4, 19.6, 29.1, 67.5, 71.3, 123.1, 128.1, 131.5, 170.9 (d, $J_{Rh-C} = 4.1$ Hz), 178.8 (d, $J_{Rh-C} = 24.2$ Hz); Anal. C₁₈H₂₅N₂O₃Br₂Rh: Found C 37.31, H 4.33, N 4.74%; Calcd C 37.27, H 4.34, N 4.83%.

 $(Me-Phebox)RhBr_2(H_2O)$ (Me-2). 58% yield. Orange solid; mp. >300 °C (decomp); IR (KBr) v 3397, 3009, 2822, 1617, 1485, 1397, 1148, 958, 739 cm⁻¹; ¹H-NMR (CD₃OD) δ 1.50 (d, J = 6.7 Hz, 6H), 3.32 (bs, 2H), 4.34 (ddq, J = 8.8, 7.7, 6.7 Hz, 2H), 4.45 (dd, J = 8.5, 7.7 Hz, 2H), 5.02 (dd, J = 8.8, 8.5 Hz, 2H), 7.24 (t, J = 7.7 Hz, 1H), 7.58 (d, J = 7.7 Hz, 2H); ¹³C-NMR (CD₃OD) δ 19.3, 58.6, 77.7,

122.3, 126.9, 132.4, 171.1 (d, $J_{Rh-C} = 3.7$ Hz), 182.2 (d, $J_{Rh-C} = 20.0$ Hz); Anal. $C_{14}H_{17}N_2O_3Br_2Rh$: Found C 32.04, H 3.39, N 5.27%; Calcd C 32.09, H 3.27, N 5.35%.

(*Ph-Phebox*)*RhBr*₂(*H*₂*O*) (Ph-**2**). 37% yield. Pale yellow solid. mp. 189 °C (decomp); IR (KBr) v 3452, 2980, 2825, 1613, 1485, 1326, 1149, 968 cm⁻¹; ¹H-NMR (CDCl₃) δ 1.79 (bs, 2H), 4.58 (dd, J = 10.4, 8.6 Hz, 2H), 5.18 (dd, J = 10.2, 8.6 Hz, 2H), 5.31 (dd, J = 10.4, 10.2 Hz, 2H), 7.35-7.46 (m, 7H), 7.46-7.74 (m, 4H), 7.72 (d, J = 7.7 Hz, 2H); ¹³C-NMR (CDCl₃) δ 31.7, 67.1, 76.8, 123.3, 128.5, 128.8, 128.9, 131.4, 137.4, 172.5 (d, $J_{Rh-C} = 4.2$ Hz), 180.4 (d, $J_{Rh-C} = 21.1$ Hz); Anal. C₂₄H₂₁N₂O₃Br₂Rh: Found C 44.48, H 3.19, N 4.30%; Calcd C 44.47, H 3.27, N 4.32%.

(*s*-*Bu*-*Phebox*)*RhBr*₂(*H*₂*O*) (*s*-Bu-**2**). 21% yield. Pale yellow solid. mp. 119 °C (decomp); IR (KBr) v 3448, 2968, 2822, 1617, 1484, 1394, 1145, 963 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.96 (d, *J* = 6.8 Hz, 6H), 1.00 (t, *J* = 7.3 Hz, 6H), 1.24 (m, 2H), 1.39 (m, 2H), 2.17 (m, 2H), 3.43 (bs, 2H), 4.34 (ddd, *J* = 9.9, 6.7, 3.2 Hz, 2H), 4.69 (dd, *J* = 8.8, 6.7 Hz, 2H), 4.74 (dd, *J* = 9.9, 8.8 Hz, 2H), 7.26 (t, *J* = 7.7 Hz, 1H), 7.59 (d, *J* = 7.7 Hz, 2H); ³C-NMR (CDCl₃) δ 12.0, 12.8, 35.8, 66.4, 71.4, 77.4, 123.4, 128.2, 131.4, 170.7 (d, *J*_{Rh-C} = 4.2 Hz), 176.9 (d, *J*_{Rh-C} = 24.5 Hz); Anal. C₂₀H₂₉N₂O₃Br₂Rh: Found C 39.51, H 4.73, N 4.63%; Calcd C 39.50, H 4.81, N 4.61%.

3.3. General Procedure for the Catalytic Enantioselective Addition of Allyl- or Methallyltributyltin to Aldehydes Catalyzed with (Phebox)RhX₂(H₂O) Complexes

To a suspension of MS 4A (250 mg) in dichloromethane (2 mL) was added (Phebox)RhX₂(H₂O) complex (0.025-0.05 mmol, 5-10 mol %), amino aldehyde (0.5 mmol) and allyl- or methallyltributyltin (0.75 mmol) at 25 °C. After it was stirred for 24 h at that temperature, the reaction mixture was concentrated under reduced pressure. Purification of the residue by silica gel chromatography gave homoallylic alcohol: the enantioselectivity was determined by chiral HPLC analysis.

I-(*2*-*Pyridyl*)-*3*-buten-*I*-ol (**5b**) [4,5]. $[\alpha]_D^{20}$ −27.1° (c 1.22, CHCl₃) for 42% ee: lit. 4 $[\alpha]_D^{23}$ −32.5° (c 3.5, EtOH) for ≥99% ee, 1*S*; IR (neat) v 3267, 2922, 1733, 1699, 164, 1562, 1474, 1066, 702 cm⁻¹; ¹H- NMR (CDCl₃) δ 2.48 (ddddd, *J* = 14.5, 7.3, 6.9, 1.3, 1.1 Hz, 1H), 2.63 (ddddd, *J* = 14.5, 6.9, 4.7, 1.4, 1.2 Hz, 1H), 4.07 (bs, 1H), 4.81 (bs, 1H), 5.09 (dddd, *J* = 10.1, 2.0, 1.2, 1.1 Hz, 1H), 5.11 (dddd, *J* = 17.2, 2.0, 1.4, 1.3 Hz, 1H), 5.83 (dddd, *J* = 17.2, 10.1, 7.3, 6.9 Hz, 1H), 7.20 (ddd, *J* = 7.5, 4.9, 1.2 Hz, 1H), 7.28 (ddd, *J* = 7.8, 1.2, 1.0 Hz, 1H), 7.68 (ddd, *J* = 7.8, 7.5, 1.5 Hz, 1H), 8.54 (ddd, *J* = 4.9, 1.5, 1.0 Hz, 1H); ¹³C-NMR (CDCl₃) δ 43.0, 72.3, 118.1, 120.5, 122.4, 134.2, 136.7, 148.4, 161.4. Enantiomeric excess was determined by after conversion to the corresponding acetate **5b**-Ac.

1-Acetoxy-1-(2-pyridyl)-3-butene (**5b-Ac**) [23]. IR (neat) v 1738, 1592, 1372, 1235, 1047, 921 cm⁻¹; ¹H-NMR (CDCl₃) δ 2.13 (s, 3H), 2.69 (ddddd, J = 14.3, 7.5, 6.9, 1.3, 1.1 Hz, 1H), 2.76 (ddddd, J = 14.3, 7.1, 5.8, 1.4, 1.2 Hz, 1H), 5.04 (dddd, J = 10.2, 1.9, 1.2, 1.1 Hz, 1H), 5.08 (dddd, J = 17.2, 1.9, 1.4, 1.3 Hz, 1H), 5.74 (dddd, J = 17.2, 10.2, 7.1, 6.9 Hz, 1H), 5.86 (dd, J = 7.5, 5.8 Hz, 1H), 7.20 (ddd, J = 7.6, 4.8, 1.3 Hz, 1H), 7.30 (ddd, J = 7.9, 1.3, 0.9 Hz, 1H), 7.67 (ddd, J = 7.9, 7.6, 1.8 Hz, 1H), 8.60 (ddd, J = 4.8, 1.8, 0.9 Hz, 1H); ¹³C-NMR (CDCl₃) δ 21.2, 39.2, 75.7, 118.2, 121.3, 122.8, 133.2, 136.7, 149.5, 158.9, 170.4; [α]_D²⁰ -40.8° (c 1.10, CHCl₃) for 59% ee: lit. [α]_D²⁵ +75° (c 2.01, 10.2) [α]_D²⁰ -40.8° (c 1.10, CHCl₃) [α]_D²⁵ +75° (c 2.01, 10.2) [α]_D²⁵ +75° (c 2.01) [α]_D²⁵ +75° (c CHCl₃) for 92% ee, 1*R* [23]; Daicel CHIRALCEL OD, UV Detector 254 nm, hexane/*i*-PrOH = 9:1, flow rate 0.5 mL/min. $t_{\rm R}$ = 10.2 min (*R*), 12.8 min (*S*).

1-(6-Methyl-2-pyridyl)-3-buten-1-ol (**5c**). IR (neat) v 3417, 2907, 1642, 1594, 1459, 1066, 799 cm⁻¹; ¹H-NMR (CDCl₃) δ 2.45 (dtt, *J* = 14.2, 7.0, 1.1 Hz, 1H), 2.55 (s, 3H), 2.61 (dddt, *J* = 14.2, 6.9, 4.8, 1.1 Hz, 1H), 4.40 (d, *J* = 4.8 Hz, 1H), 4.76 (dt, *J* = 7.0, 4.8 Hz, 1H), 5.01 (ddt, *J* = 10.1, 2.0, 1.2 Hz, 1H), 5.12 (ddt, *J* = 17.1, 2.0, 1.5 Hz, 1H), 5.85 (ddt, *J* = 17.1, 10.1, 7.0 Hz, 1H), 7.04 (d, *J* = 7.7 Hz, 1H), 7.06 (d, *J* = 7.7 Hz, 1H), 7.56 (t, *J* = 7.7 Hz, 1H); ¹³C-NMR (CDCl₃) δ 24.4, 43.1, 71.9, 117.3, 117.8, 121.9, 134.5, 136.9, 157.1, 160.5; Anal. C₁₀H₁₃NO: Found C 73.69, H 8.15, N 8.49%; Calcd C 73.59, H 8.03, N 8.58%; [α]_D²⁵ –14.7° (c 0.95, CHCl₃) for 84% ee; Daicel CHIRALCEL OD, UV Detector 254 nm, hexane/*i*-PrOH = 30:1, flow rate 0.5 mL/min. *t*_R = 12.0 min (*R*), 13.8 min (*S*).

1-(p-Dimethylaminophenyl)-3-buten-1-ol (**5d**) [7]. IR (neat) v 3435, 2802, 1614, 1522, 1348, 1162, 1052, 915, 819 cm⁻¹; ¹H-NMR (CDCl₃) δ 1.89 (d, *J* = 2.6 Hz, 1H), 2.45-2.58 (m, 2H), 2.95 (s, 6H), 4.65 (ddd, *J* = 7.1, 6.2, 2.6 Hz, 1H), 5.11 (dm, *J* = 10.2 Hz, 1H), 5.15 (dm, *J* = 17.2 Hz, 1H), 5.82 (dddd, *J* = 17.2, 10.2, 7.3, 6.9 Hz, 1H), 6.72 (d, *J* = 8.8 Hz, 2H), 7.24 (d, *J* = 8.8 Hz, 2H); ¹³C-NMR (CDCl₃) δ 40.7, 73.4, 112.6, 117.9, 126.9, 131.9, 135.1, 150.3; [α]_D²⁴ –51.8° (c 0.35, CHCl₃) for 84% ee; Daicel CHIRALCEL OD, UV Detector 254 nm, hexane/*i*-PrOH = 9:1, flow rate 0.5 mL/min. *t*_R = 15.9 min (minor), 18.5 min (major).

(*E*)-1-(*p*-Dimethylaminophenyl)-1,5-hexadien-3-ol (**5e**). IR (neat) v 3674, 1730, 1610, 1522, 1437, 1352, 968, 806 cm⁻¹; ¹H-NMR (CDCl₃) δ 1.75 (d, *J* = 3.7 Hz, 1H), 2.36 (ddddd, *J* = 14.0, 7.4, 6.9, 1.1, 1.0 Hz, 1H), 2.43 (ddddd, *J* = 14.0, 6.8, 5.4, 1.4, 1.2 Hz, 1H), 2.95 (s, 6H), 4.31 (dddd, *J* = 7.2, 6.9, 5.4, 3.7 Hz, 1H), 5.14 (dddd, *J* = 10.2, 2.1, 1.2, 1.0 Hz, 1H), 5.17 (dddd, *J* = 17.1, 2.1, 1.4, 1.1 Hz, 1H), 5.86 (dddd, *J* = 17.1, 10.2, 7.4, 6.8 Hz, 1H), 6.03 (dd, *J* = 15.8, 6.9 Hz, 1H), 6.50 (d, *J* = 15.8 Hz, 1H), 6.67 (d, *J* = 8.9 Hz, 2H), 7.27 (d, *J* = 8.9 Hz, 2H); ¹³C-NMR (CDCl₃) δ 40.6, 42.2, 72.4, 112.5, 118.2, 125.1, 127.2, 127.5, 130.8, 134.5, 150.3; Anal. C₁₄H₁₉NO: Found C 77.30, H 8.89, N 6.44%; Calcd C 77.38, H 8.81, N 6.45%; [α]_D²⁵ -21.5° (c 1.08, CHCl₃) for 88% ee; Daicel CHIRALCEL OD, UV Detector 254 nm, hexane/*i*-PrOH = 9:1, flow rate 0.5 mL/min. *t*_R = 20.0 min (minor), 21.1 min (major).

1-Acetoxy-1-(2-pyridyl)-3-methyl-3-butene (**6b-Ac**). IR (neat) v 3076, 2933, 1742, 1651, 1591, 1472, 1236, 894 cm⁻¹; ¹H-NMR (CDCl₃) δ 1.77 (s, 3H), 2.11 (s, 3H), 2.65 (d, *J* = 6.8 Hz, 2H), 4.72 (bs, 1H), 4.79 (bs, 1H), 5.98 (t, *J* = 6.8 Hz, 1H), 7.20 (dd, *J* = 7.7, 4.8 Hz, 1H), 7.31 (d, *J* = 7.7 Hz, 1H), 7.67 (td, *J* = 7.7, 1.7 Hz, 1H), 8.60 (dd, J = 4.8, 1.7 Hz, 1H); ¹³C-NMR (CDCl₃) δ 21.1, 22.6, 74.7, 113.7, 121.2, 122.8, 136.7, 141.2, 149.5, 159.3, 170.4; Anal. C₁₂H₁₅NO₂: Found C 70.29, H 7.29, N 6.76%; Calcd C 70.22, H 7.37, N 6.82%; [α]_D²⁶ -37.5° (c 1.08, CHCl₃) for 51% ee; Daicel CHIRALCEL OD, UV Detector 254 nm, hexane/*i*-PrOH = 50:1, flow rate 0.5 mL/min. *t*_R = 17.8 min (*R*), 21.0 min (*S*).

1-(6-Methyl-2-pyridyl)-3-methyl-3-buten-1-ol (**6c**). IR (neat) v 3399, 2924, 1645, 1591, 1458, 1156, 889 cm⁻¹; ¹H-NMR (CDCl₃) δ 1.82 (bs, 3H), 2.37 (dd, *J* = 14.0, 8.8 Hz, 1H), 2.52 (dd, *J* = 14.0, 4.4 Hz, 1H), 2.55 (s, 3H), 4.14 (d, *J* = 4.4 Hz, 1H), 4.81 (bs, 1H), 4.85 (dt, *J* = 8.8, 4.4 Hz, 1H), 4.89 (bs, 1H), 7.04 (d, *J* = 7.8 Hz, 1H), 7.08 (d, *J* = 7.8 Hz, 1H), 7.56 (t, *J* = 7.8 Hz, 1H); ¹³C- NMR

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(CDCl₃) δ 22.7, 24.4, 47.4, 70.9, 113.6, 117.3, 121.9, 136.9, 142.5, 157.2, 161.1; Anal. C₁₁H₁₅NO: Found C 74.58, H 8.53, N 7.80%; Calcd C 74.54, H 8.53, N 7.90%; $[\alpha]_D^{20}$ –29.1° (c 1.37, CHCl₃) for 45% ee; Daicel CHIRALCEL OD, UV Detector 254 nm, hexane/*i*-PrOH = 30:1, flow rate 0.5 mL/min. t_R = 12.5 min (*R*), 13.5 min (*S*).

1-(p-Dimethylaminophenyl)-3-methyl-3-buten-1-ol (**6d**). White solid. mp. 28–30 °C; IR (neat) v 3251, 2886, 2800, 1524, 1442, 1350, 1162, 1054, 816 cm⁻¹; ¹H-NMR (CDCl₃) δ 1.79 (bs, 3H), 1.98 (bs, 1H), 2.39 (dd, *J* = 14.1, 4.4 Hz, 1H), 2.48 (dd, *J* = 14.1, 9.2 Hz, 1H), 2.94 (s, 6H), 4.74 (ddd, *J* = 9.2, 4.4, 1.5 Hz, 1H), 4.85 (bs, 1H), 4.90 (bs, 1H), 6.73 (d, *J* = 8.8 Hz, 2H), 7.26 (d, *J* = 8.8 Hz, 2H); ¹³C-NMR (CDCl₃) δ 22.5, 40.8, 48.0, 71.4, 112.6, 113.7, 126.9, 132.0, 142.9; Anal. C₁₃H₁₉NO: Found C 76.07, H 9.29, N 6.77%; Calcd C 76.06, H 9.33, N 6.82%; [α]_D¹⁹ –55.7° (c 1.41, CHCl₃) for 90% ee; Daicel CHIRALCEL OD, UV Detector 254 nm, hexane/*i*-PrOH = 9:1, flow rate 0.5 mL/min. *t*_R = 15.0 min (*R*), 17.4 min (*S*).

(*E*)-1-(*p*-Dimethylaminophenyl)-3-methyl-1,5-hexadien-3-ol (**6e**). IR (neat) v 3631, 3397, 2926, 2801, 1611, 1447, 1167, 965, 804 cm⁻¹; ¹H-NMR (CDCl₃) δ 1.79 (d, *J* = 2.8 Hz, 1H), 1.80 (bs, 3H), 2.34 (d, *J* = 6.6 Hz, 2H), 2.96 (s, 6H), 4.40 (tdd, *J* = 6.6, 6.6, 2.8 Hz, 1H), 4.85 (bs, 1H), 4.90 (bs, 1H), 6.03 (dd, *J* = 15.7, 6.6 Hz, 1H), 6.54 (d, *J* = 15.7 Hz, 1H),6.68 (d, *J* = 8.8 Hz, 2H), 7.28 (d, *J* = 8.8 Hz, 2H); ¹³C-NMR (CDCl₃) δ 22.7, 40.6, 46.5, 70.6, 112.5, 113.6, 125.3, 127.6, 130.0, 130.5, 142.4, 150.3; Anal. C₁₅H₂₁NO: Found C 76.07, H 9.29, N 6.77%; Calcd C 76.06, H 9.33, N 6.82%; $[\alpha]_D^{20}$ -30.2° (c 1.66, CHCl₃) for 84% ee; Daicel CHIRALCEL OD, UV Detector 254 nm, hexane/*i*-PrOH = 9:1, flow rate 0.5 mL/min. *t*_R = 20.0 min (*R*), 21.1 min (*S*).

3.4. Synthesis and X-ray Analysis of (i-Pr-Phebox)RhCl₂ (к-3а)

 $(i-Pr-Phebox)RhCl_2(\kappa-3a)$. To a stirred solution of *i*-Pr-1 (200 mg, 0.41 mmol) in dichloromethane (5 mL) was added **3a** (39 µL, 0.41 mmol) at ambient temperature. After it was stirred for 2 h, the mixture was concentrated under reduced pressure. Purification of the residue by silica gel chromatography (dichloromethane/ether = 1:1) gave (*i*-Pr-Phebox)RhCl₂(κ -**3a**) in 84% yield (200 mg). Orange solid. mp. 203-205 °C (decomp); IR (KBr) v 2958, 1711, 1620, 1485, 1394, 1214, 963, 739 cm⁻¹; ¹H-NMR (CDCl₃) δ 0.64 (d, J = 6.8 Hz, 6H), 0.73 (d, J = 6.8 Hz, 6H), 1.37 (dsept, J = 2.8, 6.8 Hz, 2H), 4.04 (ddd, J = 10.0, 6.4, 2.8 Hz, 2H), 4.62 (dd, J = 8.8, 6.4 Hz, 2H), 4.74 (dd, J = 10.0, 8.8 Hz, 2H), 7.30 (t, J = 7.6 Hz, 1H), 7.65 (d, J = 7.6 Hz, 2H), 7.79 (dd, J = 7.6, 5.6 Hz, 1H), 8.49 (dd, J = 7.6, 1.6 Hz, 1H), 10.07 (dd, J = 5.6, 1.6 Hz, 1H), 10.27 (s, 1H), 10.30 (d, J = 1.6 Hz, 1H); ¹³C-NMR (CDCl₃) δ 15.1, 19.2, 29.4, 66.9, 71.1, 123.3, 125.2, 128.1, 131.6, 132.6, 136.6, 155.1, 157.2, 172.4 ($J_{Rh-C} = 3.4 \text{ Hz}$), 185.9 ($J_{Rh-C} = 19.7 \text{ Hz}$), 189.8; Anal. $C_{24}H_{28}Cl_2N_3O_3Rh$: Found C 49.62, H 4.86, N 7.22%; Calcd C 49.67, H 4.86, N 7.24%. X-ray-quality crystals of (*i*-Pr-Phebox)RhCl₂(κ-3a) was obtained from benzene-ether-hexane at room temperature and mounted in glass capillary. Diffraction experiments were performed on a Rigaku AFC-7R four-circle diffractometer equipped with graphite-monochromated Mo K \langle radiation; $\downarrow = 0.71069$ Å. The lattice parameters and an orientation matrix were obtained and refined from 25 machine-centered reflections with $29.82 < 2 \leq 29.97^{\circ}$. Intensity data were collected using a $-2\sqrt{\text{scan technique}}$, and three standard reflections were recorded every 150 reflections. The data were corrected for Lorentz and polarization effects. The structure was solved by direct methods [27] and expanded using Fourier techniques [28]. The non-hydrogen atoms were refined anisotropically. Hydrogen atoms were included but not refined. The final cycle of full-matrix least-squares refinement was based on 5306 observed reflections ($I > 3 \int (I)$) and 598 variable parameters. Neutral atom scattering factors were taken from Cromer and Waber [29]. All calculations were performed using the teXsan crystallographic software package [30]. Final refinement details are collected in Table 5 and the numbering scheme employed is shown in Figure 2, which was drawn with ORTEP at 30% probability ellipsoid. Crystallographic data (excluding structure factors) for the structures reported in this paper have been deposited with the Cambridge Crystallographic Data Centre as supplementary publication no. CCDC-826794. Copies of the data can be obtained free of charge on application to CCDC, 12 Union Road, Cambridge CB21EZ, UK (fax: (+44)1223-336-033; e-mail: deposit@ccdc.cam.ac.uk).

Empirical Formula	C48H58N6O7Cl4Rh2	Temperature	23.0 °C
Formula Weight	1178.65	Scan type	ω -2 θ
Crystal Dimensions	$0.15 \times 0.5 \times 0.5 \text{ mm}$	Scan Width	94. 3̃tanθ deg
Crystal System	monoclinic	$2\theta_{\rm max}$	55.0 deg
Lattice Type	C-centered	No. of Reflection Total: 6787	
Lattice Parameters: a	18.307(4) Å	measured	
b	14.886(5) Å	No. of Unique data	6581 ($R_{int} = 0.018$)
С	21.056(4) Å	Structure Solution	Direct methods
eta	106.55(2) deg	Refinement	Full-matrix
Volume	5500(2) Å ³		least squares
Space Group	<i>C</i> 2 (#5)	No. of Observations	5306 (I>3 <i>σ</i> (I))
Z value	4	No. of Variables	598
$D_{ m calcd}$	1.423 g/cm^3	Reflection/Parameter	8.87
<i>F</i> (000)	2408.00	Ratio	
<i>μ</i> (Mo Kα)	8.44 cm^{-1}	Residuals: <i>R</i> ; <i>R</i> _w	0.058; 0.077
λ	0.71069 Å		

Table 5. Crystallographic data and structure refinement for (*i*-Pr-Phebox)RhCl₂(κ-**3a**).

4. Conclusions

In this paper, we have described the catalytic enantioselective addition of allyl- and methallyltributyltin reagents to amino aldehydes catalyzed with air-stable and water tolerant chiral Phebox-Rh(III) aqua complexes. The reactions proceed under mild conditions to afford the corresponding homoallylic alcohols with modest to good enantioselectivity (up to 94% ee), and these aqua complexes can be recovered from the reaction media by column chromatography. We have clarified that the chiral (Phebox)RhX₂ fragments (X = Cl, Br), generated by releasing water molecule from (Phebox)RhX₂(H₂O), capture amino aldehydes at the carbonyl oxygen and the reaction proceeded *via* a Lewis acid mechanism.

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Sample availability: Samples of the complexes 1 and 2 are available from the authors.

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