

Article

A Facile Synthesis of Arylazonicotinates for Dyeing Polyester Fabrics under Microwave Irradiation and Their Biological Activity Profiles

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Abstract: A as textile dyes and the fastness properties of the dyed samples were measured. Most of the dyed fabrics tested displayed very good washing and perspiration fastness and series of 2-hydroxy- and 2-amino-6-substituted-5-arylazonicotinate monoazo compounds **7a–e** and **9a–c** were prepared via condensation of 3-oxo-3-substituted-2-arylhydrazonals **2a–e** with active methylene nitriles **3a–d** using microwave irradiation as an energy source. These substances were then tested moderate light fastness. Finally, the biological activity of the synthesized compounds against Gram positive bacteria, Gram negative bacteria and yeast were evaluated.

Keywords: polyester fabrics; arylazonicotinates; disperse dyes; microwave irradiation; biological activity

1. Introduction

Disperse dyes are very popular and are important class of dyes for dyeing polyester fabrics because of their brilliancy, wide range of hues, excellent fastness properties, besides environmental and economic reasons [1].

Disperse dyes are organic colors with less water solubility, so they are applied in colloidal aqueous dispersions to hydrophobic textile fibers in which the dyes literally dissolve and produce the desired coloration. The development of disperse dyes is due to significant increase in the World production of polyester fibers as compared to other fibers. Over 95% of disperse dyes usage is for the coloration of polyester and its blends. A monoazo dye with a heterocyclic system would be considered as a useful class of disperse dyes [2,3].

Derivatives of nicotinates, nicotinonitrile, and nicotinamide have a long history of use as heterocyclic components for various disperse dyes [4–7]. Moreover, they have been proven to constitute the active part of several biologically active compounds [8–12].

In view of these findings and in continuation of our previous studies [10,13] on the synthesis of a variety of 2-hydroxy- and 2-amino-6-substituted-5-arylazonicotinate compounds using traditional heating methods, in this article, a new synthesis of these compounds was synthesized under microwave irradiation and their application for dyeing polyester fabrics studied. In addition the biological activity of the synthesized dyes against *Escherichia coli* and *Pseudomonas aeruginosa*. (Gram negative bacteria), *Bacillus subtilis* and *Staphylococcus aureus* (Gram positive bacteria) and *Candida albicans* (yeast) was also studied.

2. Results and Discussion

2.1. Synthesis

Recently we have reported the synthesis of some 2-hydroxy- and 2-amino-6-substituted-5-arylazonicotinates dyes [13]. Herein, in an attempt to improve and facilitate the synthesis of these compounds, we report a new strategy for the preparation of these compounds in better yields by condensing of 3-oxo-3-substituted-2-arylhydrazonals with active methylene nitriles using microwave irradiation as an energy source.

We observed that reactions of **2a-d** with active methylenes **3a-d** in the presence of a catalytic amount of ammonium acetate and a few drops of acetic acid leads gives the corresponding 2-hydroxy-6-substituted-5-aryl azonicotinates **7a-e** (Scheme 1) after heating in a focused microwave oven at 170 °C for 1 min.

It is believed that the pathways for these processes involve initial reaction of 2a-d with active methylenes 3a-d to yield the hydrazono-enones 4 that then cyclize to generate the pyran-imines 5. In the absence of ammonium ion, compounds 5 undergo a Dimroth type rearrangement to yield 7a-e.

Although not directly related to the processes described above, it is of value to note that earlier studies showed that hydrazono-3-oxo-propanal **2d** exists as an equilibrium mixture of *syn* and *anti*-forms [14] in dimethyl sulfoxide solution, with the latter stereoisomer being predominant. In contrast, the results of our current X-ray crystallographic studies demonstrate that in the solid state this substance only exists in the *anti*-form (Figure 1) [15].

Scheme 1. Synthesis of 2-hydroxy-6-substituted-5-arylazonicotinate derivatives 7a–e.

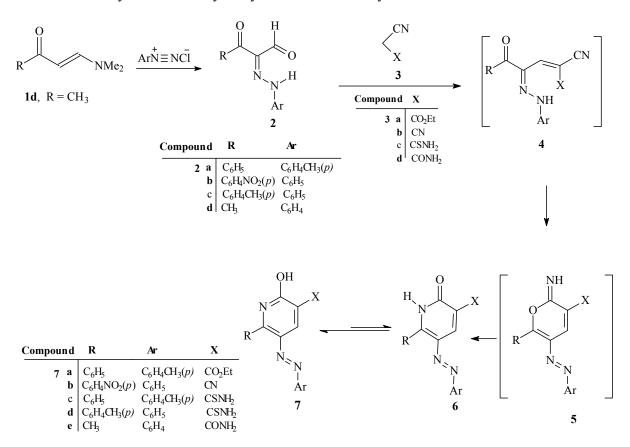
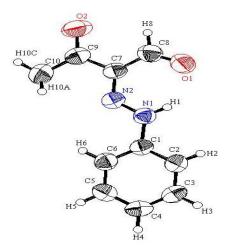


Figure 1. ORTEP drawing of 2d.



In contrast, when **2a,b,e** are condensed with ethyl cyanoacetate in the presence of excess ammonium acetate in a focused microwave oven at 170 °C for 1 min, ethyl 2-amino-6-substituted-5-arylazonicotinate derivatives **9a–c** are produced. It is believed that the pathways for these processes involve initial reaction of **2a,b,e** with ethyl cyanoacetate to yield the hydrazono-enones **4** that then cyclize to generate the pyran-imines **5**. In the presence of a high concentration of ammonium acetate, pyran-imines **5** participate undergo a ring opening to yield amidines **8** that then cyclize followed by water elimination to yield **9a–c** (Scheme 2).

Scheme 2. Synthesis of 2-amino-6-substituted-5-arylazonicotinate derivatives 9a-c.

2.2. Dyeing

The functionalized compounds 7a-e and 9a-c were applied to polyester fabrics at 3% weight of fibre using the high temperature dyeing method at 130 °C. Color shades varying from yellow to brownish-green were obtained. The dyeing on the polyester fabrics was evaluated in terms of their fastness properties (e.g., fastnesses to washing, perspiration, and light). The optical measurements and fastness property data for the dyed fabrics are listed in Tables 1 and 2. The color of dyeing on polyester fabrics is expressed in terms of CIELAB values (Table 1), and the following CIELAB coordinates were measured: lightness (L *); chroma (C *); hue angle (H) from 0 to 360°; A *, whose value represents the degree of redness (positive) and greenness (negative); and B^* , whose value represents the degree of yellowness (positive) and blueness (negative). A reflectance spectrophotometer was used for the colorimetric measurements of the dyed samples. The K/S values (where K is the absorbance coefficient and S is the scattering coefficient) given by the reflectance spectrometer were calculated at λ_{max} (wavelength of maximum absorption) and were directly correlated with the dye concentration on the dye substrate according to the Kubelka-Munk equation. In general, the color hues of the arylazonicotinates 7b on the polyester fabric shifted to the greenish directions; this was indicated by the low value of $A^* = 0.87$ (red-green axis). The positive values of B^* (yellow-blue axis) indicated that the color hues of the arylazonicotinates 7a,d and 9a-c on the polyester fabric shifted in the yellowish direction.

Dye No.	UV, [λ _{max} (DMF)/nm]	Molar extinction coefficient (mol ⁻¹ cm ⁻¹)	K/S	<i>L</i> *	A *	В *	C *	Н
7a	371	16,484	19.30	51.58	14.63	39.13	41.78	69.50
7 b	368	9,237	20.48	34.08	0.87	12.74	12.77	86.11
7c	349	6,813	17.10	40.76	9.15	25.31	26.92	70.13
7 d	347	11,762	16.89	58.82	8.74	36.37	37.41	76.49
7e	360	10,769	10.27	50.60	4.65	22.63	23.10	78.40
9a	374	16,524	22.02	54.95	18.01	48.60	51.83	69.67
9b	373	9,499	21.22	52.53	13.85	41.29	43.56	71.44
9c	379	25,924	25.50	56.06	21.78	57.05	61.07	69.10

Table 1. Optical measurements of the synthesized monoazo compounds on the polyester fabrics.

The physical data for the dyed fabrics, given in Table 2, shows that these compounds displayed very good washing and perspiration fastness. The light fastness is moderate with respect to most of the tested compounds, except for **7d** and **9b** which showed good results.

I able 2. Fastness	properties of m	nonoazo comp	ounds on	polyester fabrics	٠.

Deva	Color shade on polyester	Wash fastness			Perspiration fastness					T !-1.4	
Dye No					Alkaline		Acidic			Light	
No.		Alt	SC	SW	Alt	SC	SW	Alt	SC	SW	fastness
7a	Yellow	5	5	5	5	5	5	5	4–5	5	2–3
7b	Brownish-green	5	5	5	5	5	5	5	4–5	5	2–3
7c	Yellowish-brown	4–5	4–5	4–5	5	5	5	5	4–5	5	2–3
7 d	Yellow	5	4–5	5	5	5	5	5	4–5	5	4–5
7e	Yellowish-brown	5	5	5	5	5	5	5	4–5	5	3–4
9a	Dark yellow	5	4–5	5	5	5	5	5	4–5	5	2–3
9b	Yellowish-brown	5	5	5	5	5	5	5	4–5	5	4–5
9c	Yellowish-orange	5	5	5	5	5	5	5	4–5	5	3–4

2.3. Antimicrobial Activities

The antimicrobial activities of the synthesized compounds were screened against selected bacteria and fungi by the agar well diffusion method and their inhibition zones diameters, given in Table 3, reveal that all of the tested arylazonicotinates compounds showed positive antimicrobial activities against at least one of the tested microorganisms. All of them showed strong activities (>10 mm inhibition zone) against *S. auerus*. Two of the compounds **7a** and **9a**, showed medium activities against Gram negative bacteria. Where most of the compounds showed no activities against the two strains of Gram negative bacteria used in the study. Also the majority of compounds showed weak to no activities at all with *B. subtilis*. Only dye **7d** showed significant inhibition zone > 10 mm, against *Candida albican*. The other compound that showed medium activities against yeast is **7c** while all the other compounds failed to affect the yeast growth.

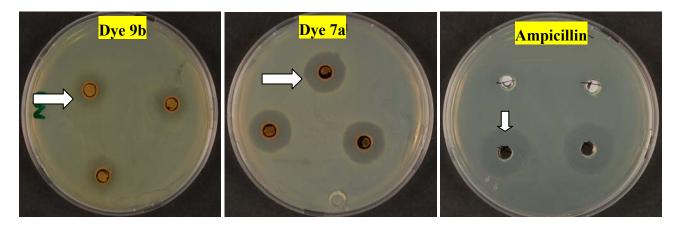
	Inhibition zone diameter (Nearest mm)								
Compound No	B. subtilis	S. aureus	E. coli	P. aeruginosa	C. albicans				
	$Mean \pm SD$	$Mean \pm SD$	$Mean \pm SD$	$Mean \pm SD$	$Mean \pm SD$				
7a	0.1 ± 0.08	15 ± 0.02	7 ± 0	-	-				
7 b	-	11 ± 0.87	-	-	-				
7c	0.7 ± 0.13	16 ± 0	-	-	7 ± 0.07				
7 d	0.1 ± 0	16 ± 0.08	-	-	12 ± 0.1				
9a	-	13 ± 0	7 ± 0	7 ± 0.05	-				
9b	-	10 ± 0.2	-	-	-				
Ampicillin *	30 ± 0.05	46 ± 0.7	31 ± 0.14	17 ± 0.07					

Table 3. Diameter of the zones of inhibition of the tested compounds that showed weak to strong antimicrobial against microorganisms.

(-) no inhibition; * Ampicillin: antibacterial (100 $\text{mg}\cdot\text{mL}^{-1}$); ** Cycloheximide: antifungal (100 $\text{mg}\cdot\text{mL}^{-1}$), SD = Standard Deviation.

It is of value to mention here that as Figure 2 shows, after six days the inhibition zone did not show any difference in the size, yet the zone is not clear which indicates that the compound number **9b** did not kill the microorganisms, but rather had weakened their growth only, this is comparison to compound number **7a** or to ampicillin as reference.

Figure 2. Staphylococcus aureus treated with 10 mg·mL⁻¹ of compounds **9b** and **7a** with comparison to Ampicillin after six days of incubation.



Therefore, the new synthesized classes showed promising results regarding the potential to be utilized for medicinal purposes. Currently, we are inspecting the biological activity of the polyester fabrics dyed with the arylazonicotinate compounds against Gram positive bacteria, Gram negative bacteria and yeast.

3. Experimental

Cyloheximide **

3.1. General

Melting points were recorded on a Gallenkamp apparatus. IR spectra were recorded using KBr pellets on a JASCO FTIR-6300 FT-IR spectrophotometer. ¹H- and ¹³C-NMR spectra were recorded on

Bruker DPX 400 MHz or AvanceII 600 MHz super-conducting NMR spectrometers with proton spectra measured at 400, 600 MHz and carbon spectra at 100 and 150 MHz, respectively. Mass spectra were measured on a high resolution GC/MS DFS-Thermo. Microanalyses were performed on Elementar-Vario Micro cube Analyzer. X-Ray analyses were performed using a Rigaku Rapid II diffractometer. The microwave oven used is a single mode cavity Explorer Microwave (CEM Corporation, Matthews, NC, USA). The dyeing of polyester fabrics were conducted using LINITEST + Laboratory High Temperature Dyeing and Fastness System (ATLAS MTT GmbH, Vogelbergstrasse 22, Altenhasslau, Germany). The colorimetric parameters of the dyed polyester fabrics were determined on a reflectance spectrophotometer (UltraScan PRO D65, HunterLab, Virginia, VA, USA).

Synthesis of 3-oxo-2-(phenylhydrazono)-butyraldehyde (2d). A cold solution of the diazonium salt (10 mmol), prepared by adding a cold solution of sodium nitrite (0.7 g) in water (5 mL) to a solution of the corresponding aniline (10 mmol) in conc. HCl (5 mL), was added to a cold solution of enaminone 1d (10 mmol) in EtOH (10 mL) containing NaOH (1.6 g). The resulting mixture was stirred at room temperature for 1 h. The precipitate that formed was collected by using filtration and crystallized from EtOH to give 2d. This compound was obtained as buff crystals (82%); mp. 124 °C (Lit. 122 °C) [16]; IR (KBr): = 3137 (NH), 1669 (CO) cm⁻¹; ¹H-NMR (DMSO- d_6): δ = 2.50 (s, H, CH₃), 7.25 (t, 1H, J = 7.2 Hz phenyl-H), 7.46 (t, 2H, J = 8.4 Hz phenyl-H), 7.64 (d, 2H, J = 8.4 Hz phenyl-H); 9.93 (s, 1H, CHO, D₂O exchangeable), 14.14 (s, 1H, NH, D₂O exchangeable). ¹³C-NMR (DMSO- d_6): δ = 196.3 (CO), 189.7 (CHO), 141.3, 133.5, 129.6, 126.2, 116.8, 24.3 (CH₃); MS, m/z (%), 190 (M⁺, 100), 148.1 (80), 120.1 (58), 77.0 (71). HRMS: m/z (EI) for C₁₀H₁₀N₂O₂; calcd. 190.0736; Found: 190.0736.

General Procedure for the Preparation of Compounds 7a-e

Method A: Independent mixtures of 2a-e (0.01 mol), active methylene nitrile derivatives 3a-d (0.01 mol), and ammonium acetate (0.5 g) in acetic acid (2 mL) were irradiated by focused microwaves at 170 °C for 1 min. Completion of the reactions was monitored by TLC. The pressure build-up in the closed reaction vessel was carefully monitored. After the irradiation, the reaction tube was cooled with high-pressure air through an inbuilt system in the instrument until the temperature had fallen below 50 °C. The mixtures were cooled and then poured into ice-water. The solids that formed were collected by using filtration and crystallized from proper solvents to give 7a-e.

2-Hydroxy-6-phenyl-5-p-tolylazonicotinic acid ethyl ester (**7a**). This compound was obtained as yellowish-brown crystals (84%); mp. 180–182 °C; IR (KBr): = 3401 (OH), 1692 (CO) cm⁻¹; ¹H-NMR (DMSO- d_6): δ = 1.35 (t, 3H, J = 7.2 CH₃), 2.50 (s, 3H, CH₃), 4.36 (q, 2H, J = 7.2 CH₂), 7.32 (d, 2H, J = 8.0 Hz arom-H), 7.48–7.50 (m, 3H, arom-H), 7.60 (d, 2H, J = 8.0 Hz arom-H), 7.78–7.79 (m, 2H, arom-H), 7.98 (s, 1H, OH, D₂O exchangeable). 8.54 (s, 1H, arom-H); ¹³C-NMR (DMSO- d_6): δ = 166.5, 161.0, 158.5, 154.5, 150.6, 142.9, 138.3, 136.5, 131.0, 130.8, 129.8, 127.4, 122.3, 105.2, 61.0, 20.9, 14.1; MS, m/z (%), 361 ([M]⁺, 54), 314 (25), 289 (44), 213 (6), 182 (7), 105 (100), 91 (42), 77 (48). λ_{max} (DMF)/nm 371. HRMS: m/z (EI) for C₂₁H₁₉N₃O₃; calcd. 361.1418; Found: 361.1418.

2-Hydroxy-6-(4-nitrophenyl)-5-phenylazonicotinonitrile (**7b**). This compound was obtained as brownish-green powder (85%); mp. 276–278 °C; IR (KBr): = 3446 (OH), 2201 (CN) cm⁻¹; ¹H-NMR

(DMSO- d_6): 7.25–7.62 (m, 6H, arom-H, OH), δ = 7.27 (d, 1H, J = 8.8 Hz arom-H), 8.01–8.08 (m, 1H, arom-H), 8.20 (d, 1H, J = 8.8 Hz arom-H), 8.27–8.36 (m, 2H, arom-H), ¹³C-NMR (DMSO- d_6): δ = 159.8, 152.2, 151.1, 147.6, 141.7, 139.6, 139.0, 131.1, 129.7, 129.2, 126.3, 123.4, 123.1, 114.0, MS, m/z (%), 345 ([M]⁺, 6), 268 (10), 239 (100), 210 (12), 194 (42), 150 (10), 93 (10), 57 (20). λ_{max} (DMF)/nm 368. Anal. Calcd for C₁₈H₁₁N₅O₃: C, 62.61; H, 3.21; N, 20.28. Found: C, 62.78; H, 3.28; N, 20.20.

2-Hydroxy-6-phenyl-5-p-tolylazothionicotinamide (**7c**). This compound was obtained as brown powder (84%); mp. 276–278 °C; IR (KBr): = 3441 (OH), 3385, 3291 (NH₂), 1660 (CO) cm⁻¹; ¹H-NMR (DMSO- d_6): δ = 2.26 (s, 3H, CH₃), 7.02–8.01 (m, 9H, arom-H), 8.57 (s, 1H, OH), 8.76 (s, 1H, pyridyl-H), 12.38 (s, 2H, NH₂). ¹³C-NMR (DMSO- d_6): δ = 206.0, 162.2, 158.0, 152.7, 141.7, 138.8, 137.3, 135.8, 131.7, 130.6, 129.4, 129.0, 124.5, 94.0, 20.8; MS, m/z (%), 348 ([M]⁺, 5), 315 (100), 300 (28), 279 (8). λ_{max} (DMF)/nm 349. Anal. Calcd for C₁₉H₁₆N₄OS: C, 65.50; H, 4.63; N, 16.08; S, 9.20. Found: C, 65.60; H, 4.30; N, 16.43; S, 8.83.

2-Hydroxy-5-phenylazo-6-p-tolylthionicotinamide (**7d**). This compound was obtained as yellowish-brown powder (72%); mp. 166–168 °C; IR (KBr): = 3409 (OH), 3283, 3197 (NH₂), 1697 (CO) cm⁻¹; ¹H-NMR (DMSO- d_6): δ = 2.33 (s, 3H, CH₃), 7.33–7.95 (m, 9H, arom-H), 8.32 (s, 1H, OH), 8.77 (s, 1H, pyridyl-H), 10.33 (s, 1H, NH), 10.64 (s, 1H, NH). ¹³C-NMR (DMSO- d_6): δ = 206.5, 162.1, 158.1, 144.4, 141.1, 137.3, 133.2, 132.3, 130.7, 129.2, 129.0, 128.8, 125.6, 115.2, 21.2; MS, m/z (%), 348 ([M]⁺, 5), 315 (100), 300 (32), 290 (6). λ_{max} (DMF)/nm 347. Anal. Calcd for C₁₉H₁₆N₄OS: C, 65.50; H, 4.63; N, 16.08; S, 9.20. Found: C, 65.78; H, 4.39; N, 16.43; S, 9.36.

2-Hydroxy-6-methyl-5-phenylazonicotinamide (7e). This compound was obtained as brown powder (74%); mp. >300 °C; IR (KBr): = 3446 (OH), 3342, 3242 (NH₂), 1657 (CO) cm⁻¹; ¹H-NMR (DMSO- d_6): δ = 2.39 (s, 3H, CH₃), 7.56–7.69 (m, 5H, phenyl-H), 7.95 (s, 1H, OH, D₂O exchangeable), 8.25 (s, 1H, pyridyl-H), 11.30 (s, 2H, NH₂, D₂O exchangeable). ¹³C-NMR (DMSO- d_6): δ = 168.1, 153.9, 149.0, 139.5, 130.8, 130.4, 128.8, 126.7, 125.0, 97.3, 15.7; MS, m/z (%), 256 ([M]⁺, 12), 207 (17), 129 (32), 93 (54), 77 (100). λ_{max} (DMF)/nm 360. Anal. Calcd for C₁₃H₁₂N₄O₂: C, 60.93; H, 4.72; N, 21.86. Found: C, 61.43; H, 4.06; N, 21.81.

Method B: Independent mixtures of 2a-e (0.01 mol), active methylene nitrile derivatives 3a-d (0.01 mol), and ammonium acetate (0.5 g) in acetic acid (10 mL) were stirred at reflux for 30 min. Completion of the reactions was monitored by TLC. The same treatment as described in method A to give 7a-e in (83, 91, 77 [13]), 68 and 70%, respectively.

General Procedure for the Preparation of Compounds 9a-c

Method A: Independent mixtures of 2a,b or 2e (0.01 mol), ethyl cyanoacetate (0.01 mol), and ammonium acetate (2 g) in acetic acid (2 mL) were irradiated by focused microwaves at 170 °C for 1 min. Completion of the reactions was monitored by TLC. The mixtures were cooled and then poured into ice-water. The solids that formed were collected by using filtration and crystallized from proper solvents to give 9a–c.

2-Amino-6-phenyl-5-p-tolylazonicotinic acid ethyl ester (**9a**). This compound was obtained as brownish-yellow crystals (81%); mp. 210–212 °C; IR (KBr): = 3402 , 3275 (NH₂) 1693 (CO) cm⁻¹; ¹H-NMR (DMSO- d_6): δ = 1.37 (t, 3H, J = 7.2 CH₃), 2.50 (s, 3H, CH₃), 4.39 (q, 2H, J = 7.2 CH₂), 7.31 (d, 2H, J = 8.4 Hz arom-H), 7.47–7.48 (m, 3H, arom-H), 7.98 (s, 2H, NH₂, D₂O exchangeable).7.59 (d, 2H, J = 8.4 Hz arom-H), 7.80–7.82 (m, 2H, arom-H), 8.54 (s, 1H, arom-H); ¹³C-NMR (DMSO- d_6): δ = 166.2, 161.3, 159.5, 150.4, 140.6, 137.2, 136.5, 130.8, 129.8, 129.2, 127.4, 127.2, 122.3, 104.9, 61.0, 20.9, 14.1; λ_{max} (DMF)/nm 374. Anal. Calcd for C₂₁H₂₀N₄O₂: C, 69.98; H, 5.59; N, 15.55. Found: C, 69.99; H, 5.50; N, 15.25. HRMS: m/z (EI) for C₂₁H₂₀N₄O₂: calcd. 360.1582; Found: 360.1582.

2-Amino-6-(4-nitrophenyl)-5-phenylazonicotinic acid ethyl ester (**9b**). This compound was obtained as yellowish-brown powder (80%); mp. 200–202 °C; IR (KBr): = 3385, 3201 (NH₂), 1656 (CO) cm⁻¹; ¹H-NMR (DMSO- d_6): δ = 1.32 (t, 3H, J = 7.2 CH₃), 4.35 (q, 2H, J = 7.2 CH₂), 7.32–7.62 (m, 3H, arom-H), 7.68 (d, 2H, J = 7.2 Hz arom-H), 8.02 (d, 2H, J = 8.4 Hz arom-H), 8.16–8.41 (m, 4H, arom-H, NH₂), 8.56 (s, 1H, pyridyl-H); ¹³C-NMR (DMSO- d_6): δ = 166.2, 163.2, 159.8, 152.3, 148.6, 143.7, 136.8, 132.3, 131.4, 130.7, 128.6, 124.1, 122.7, 106.3, 61.0, 14.2; MS, m/z (%), 391 ([M]⁺, 100), 360 (28), 286 (22), 256 (64), 213 (13), 152 (12), 111 (21), 97 (42), 77 (72). λ_{max} (DMF)/nm 373. HRMS: m/z (EI) for C₂₀H₁₇N₅O₄; calcd. 391.1275; Found: 391.1275.

2-Amino-5-(4-chlorophenylazo)-6-phenylnicotinic acid ethyl ester (**9c**). This compound was obtained as yellowish-orange powder (68%); mp. 188–190 °C; IR (KBr): = 3275 , 3182 (NH₂), 1693 (CO) cm⁻¹; ¹H-NMR (DMSO- d_6): δ = 1.36 (t, 3H, J = 7.2 CH₃), 4.36 (q, 2H, J = 7.8 CH₂), 7.23 (d, 1H, J = 10.2 Hz arom-H), 7.39 (d, 1H, J = 7.8 Hz arom-H), 7.49–7.82 (m, 7H, arom-H, NH₂), 7..89 ((d, 1H, J = 10.2 Hz arom-H), 7.98 (d, 1H, J = 7.8 Hz arom-H), 8.56 (s, 1H, pyridyl-H); ¹³C-NMR (DMSO- d_6): δ = 166.1, 161.9, 159.7, 150.9, 141.7, 139.8, 137.1, 136.4, 134.8, 129.8, 127.3, 123.9, 105.0, 61.0, 14.1; λ_{max} (DMF)/nm 379. Anal. Calcd for C₂₀H₁₇ClN₄O₂: C, 63.08; H, 4.50; N, 14.71. Found: 63.55; H, 4.37; N, 14.77. HRMS: m/z (EI) for C₂₀H₁₇ClN₄O₂; calcd. 380.1035; Found: 380.1035.

Method B: Independent mixtures of **2a,b** or **2e** (0.01 mol), ethyl cyanoacetate (0.01 mol), and ammonium acetate (3 g) in acetic acid (10 mL) were stirred at reflux for 30 min. Completion of the reactions was monitored by TLC. The same treatment as described in method A gave **9a–c** in 83 [13], 60, and 65%, respectively.

3.2. High Temperature Dyeing Method (HT)

3.2.1. Materials

Scoured and bleached polyester 100% (150 130 g/m², 70/2 denier) was obtained from El-Shourbagy Co., Egypt. The fabric was treated before dyeing with a solution containing non-ionic detergent (Sera Wash M-RK, 5 g/L) and sodium carbonate (2 g/L) in a ratio of 50:1 at 60 °C for 30 min, then thoroughly washed with water and air dried at room temperature.

3.2.2. Dyeing

A dispersion of the dye was produced by dissolving the appropriate amount of dye (3% shade w.o.f) in acetone (1 mL) and then added dropwise with stirring to the dyebath (liquor ratio 50:1) in the presence of a 1:1 ratio of Sera Gal P-LP as dispersing agent. The pH value of the bath was adjusted to 4.5–5 with acetic acid (10%) in the presence of a 1:1 ratio of the dispersing agent (Sera Gal P-LP). The temperature was raised to 130 °C at the rate of 7 °C/min, and dyeing continued for 60 min. After dyeing, the fabrics were thoroughly washed and then subjected to a surface reduction cleaning [(2 g NaOH + 2 g sodium hydrosulphite)/L]. The samples were heated in this solution for 30 min. at 70 °C and then thoroughly washed and air-dried.

3.3. Color Measurements and Analyses

3.3.1. Color Measurements

The colorimetric parameters (Table 1) of the dyed polyester fabrics were determined on a reflectance spectrophotometer (UltraScan PRO D65). The color yields of the dyed samples were determined by using the light reflectance technique performed on a Perkin-Elmer (Lambda 3B) UV/VIS Spectrophotometer. The color strengths, expressed as K/S values, were determined by applying the Kubelka-Mink equation as follows:

$$K/S = [(1-R)^2/2R] - [(1-R_o)^2/2R_o]$$

where R = decimal fraction of the reflectance of the dyed fabric; R_o = decimal fraction of the reflectance of the undyed fabric; K = absorption coefficient; S = scattering coefficient.

3.3.2. Fastness Testing

Fastnesses to washing, perspiration, and light were tested according to reported methods [17].

3.4. Antimicrobial Activities Test

The antimicrobial activities of different arylazonicotinates compounds were tested using the agar-well diffusion technique (Isaacson and Kirchbaum, [18]) against five different microbial cultures. Pure cultures of *Escherichia coli* and *pseudomonas aeruginosa* (Gram negative bacteria), *Bacillus subtilis* and *Staphylococcus aureus* (Gram positive bacteria) and *Candida albicans* (yeast) were involved in the test. An aliquot of each bacterial strain (0.1 mL) was inoculated and spread on nutrient agar (NA) while the yeast (0.1 mL) was spread on potato dextrose agar (PDA). The inoculated plates were supplied with 100 μL of each of the tested compounds with a total final concentration of 10 mg mL⁻¹. The compounds were included in 4 mm wells produced by sterile cork borer. The NA plates were incubated at 37 °C for 24 h while PDA plates were incubated at 25 °C for 24–48 h. The zones of inhibition around the wells were determined and the average based on three replica was recorded. cycloheximide and ampicillin, both used as references in the experiment as cycloheximide is known to inhibit eukaryotic organisms while ampicillin inhibits prokaryote organisms. All plates were kept for

six days after inoculation and the changes in the inhibition zone was monitored and documented by photography in order to determine on the cytolytic and cytostatic effect of the tested compounds.

4. Conclusions

A series of arylazonicotinates derivatives were synthesized via condensation of 3-oxo-3-substituted-2-arylhydrazonals with active methylenes using microwave irradiation as an energy source. The new compounds were tested as textile dyes on polyester fabrics, where they display yellow to brownish-green hues, in addition to very good washing and perspiration fastness and moderate light fastness. Finally, the biological activity of the synthesized compounds against Gram positive bacteria, Gram negative bacteria and yeast was determined.

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Sample Availability: Samples of compounds 2d, 7a-e and 9-c are available from the authors.

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