

Article

Secondary Metabolites from Two Species of *Tolpis* and Their Biological Activities

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Abstract: Phytochemical research of two *Tolpis* species, *T. proustii* and *T. lagopoda*, led to the isolation of three new compounds: 30-chloro-3 β -acetoxy-22 α -hydroxyl-20(21)-taraxastene (**1**), 3 β ,22 α -diacetoxy-30-ethoxy-20(21)-taraxastene (**2**) and 3 β ,28-dihydroxy-11 α -hydroperoxy-12-ursene (**3**). The structures of the new compounds were elucidated by means of extensive IR, NMR, and MS data and by comparison of data reported in the literature. The *in vitro* antioxidant activities of the extracts were assessed by the DPPH and ABTS scavenging methods. The cytotoxicity of several known compounds and its derivatives was also assessed against human myeloid leukemia K-562 and K-562/ADR cell lines.

Keywords: Asteraceae; *Tolpis proustii*; *Tolpis lagopoda*; ursane-triterpenes; taraxastane-triterpenes

1. Introduction

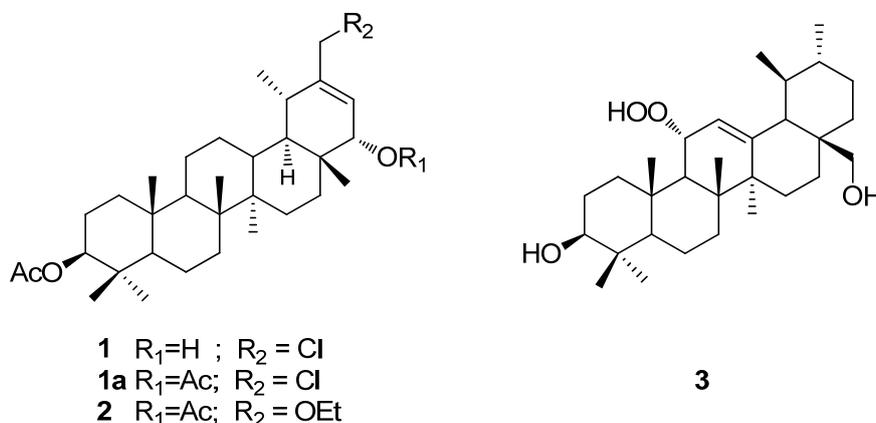
The *Tolpis* genus (Asteraceae: Cichorioideae, Cichoriinae) consists of some 20 species distributed throughout Europe, North Africa, Canary Islands, Cape Verde and Asia, this genus being represented in the Canary Islands by around ten species [1].

Although a wide-ranging study of different species of *Tolpis* from the Canary Islands and from other Macaronesian archipelagos has been carried out from the botanical point of view [2,3], only one phytochemical report has previously appeared on the isolation and structural elucidation of aromatic compounds, triterpenes, and sterols from *T. webbi* and *T. spp* [4].

As a part of our continuing search for novel, plant-derived biological agents and our systematic investigation of the composition of Canarian endemic plants, the present work describes the isolation and structural elucidation of the constituents of the ethanolic extracts of the aerial parts of *T. proustii* Pitard in Pitard and Proust and *T. lagopoda* C.Sm. ex Buch. The constituents of these extracts were purified by CC, MPLC and preparative TLC. The structures of the known compounds were confirmed by comparison of their spectroscopic data with those reported in the literature.

From *T. proustii* seventeen compounds were isolated including two taraxan-triterpenoid 30-chloro-3 β -acetoxy-22 α -hydroxyl-20(21)-taraxastene **1**, and 3 β ,22 α -diacetoxy-30-ethoxy-20(21)-taraxastene **2** a new ursan-triterpenoid 3 β ,28-dihydroxy-11 α -hydroperoxy-12-ursene **3**. From *T. lagopoda*, eight compounds were isolated including a 3 β ,22 α -diacetoxy-30-ethoxy-20(21)-taraxastene **2** (Figure 1).

Figure 1. Compounds **1–3** isolated of the *T. proustii* and *T. lagopoda*.



The high level of solar radiation and the high temperature prevailing in the region of the Canary Islands (opposite the northwest coast of Africa) force plants to develop defence mechanisms against ultraviolet radiation and excessive production of free radicals through the accumulation of antioxidant metabolites [5]. This prompted us to evaluate the antioxidant activity of the crude extract of *T. proustii* and *T. lagopoda*, and that of some of the isolated compounds. Furthermore, one of the most important mechanisms by which tumor cells resist to cytotoxic effects of a variety of chemotherapeutic drugs is overexpression of the human multidrug resistance (MDR1) gene and its product, P-glycoprotein [6]. Here we have evaluated the effects of natural compounds and derivatives on the growth of the human leukemia K-562 and the P-glycoprotein-overexpressing K-562/ADR cell lines.

2. Results and Discussion

2.1. Structure Elucidation of Compounds 1–3

The positive EIMS spectrum of compound **1** showed a molecular ion at m/z 518/520 (3:1) in agreement with the presence of a chlorine atom in the structure, and with the formula $C_{32}H_{51}O_3Cl$ for this compound. In the same spectrum, the ions at m/z 500/502 $[M-H_2O]^+$, 458/460 $[M-CH_3COOH]^+$ suggested that this compound contained a hydroxyl and an acetyl group, respectively. Confirmed by the IR spectrum with absorptions of a hydroxyl 3446 cm^{-1} and a carbonyl 1730 cm^{-1} group. Its HREIMS experiment indicated the molecular formula $C_{32}H_{51}O_3^{37}Cl$ (calcd. for $[M]^+$ 520.3497; found 520.3497) and $C_{32}H_{51}O_3^{35}Cl$ (calcd. for $[M]^+$ 518.3527; found 518.3517). All spectral data suggested that **1** was a pentacyclic triterpene with a trisubstituted double bond in the E ring [7] with a 20(21)-taraxastane structure.

The 1H and ^{13}C -NMR (Table 1) spectra of **1** showed the presence of the oxygenated methine proton at δ_H 3.35 (1H, d, $J = 6.6\text{ Hz}$) and an unusual chloro atom at C-30 at δ_C 47.6 (Table 1). The structure elucidation and NMR assignments were therefore based primarily on the results of COSY, HSQC, HMBC, and NOESY experiments (Figure 2).

Table 1. 1H - and ^{13}C -NMR data for compounds **1**, **1a** and **2** ^a.

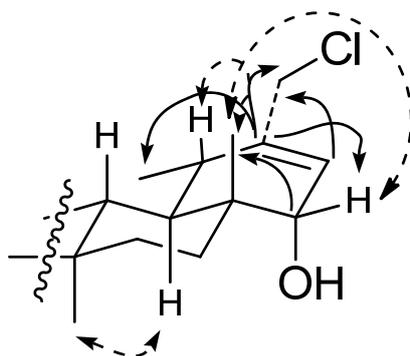
Position	1		1a		2	
	δ_H	δ_C	δ_H	δ_C	δ_H	δ_C
1	1.65 *	38.5	1.65 *	38.7	1.60 *	38.5
2	1.50 *	23.7	1.54 *	21.7	1.55 *	23.7
3	4.41 dd (6.1,10.8)	81.0	4.43 dd (6.4,10.0)	81.1	4.43 dd (5.1 10.4)	81.0
4	-	38.3	-	38.0	-	37.8
5	0.75 *	55.4	0.74 *	55.6	0.75 *	55.4
6	1.45 *	18.2	1.44 *	18.0	1.30 *	18.2
	1.35 *		1.34 *			
7	1.35 *	34.2	1.34 *	34.4	1.35 *	34.2
8	-	41.2	-	41.3	-	41.1
9	1.25 *	50.3	1.26 *	50.6	1.25 *	50.4
10	-	37.1	-	37.3	-	37.1
11	1.20 *	21.6	1.21 *	21.4	1.20 *	21.6
12	1.60 *	27.6	1.60 *	27.7	1.58 *	27.5
	1.20 *					
13	0.95 *	38.6	0.95 *		1.66 *	38.6
14	-	42.3	-	42.4	-	42.3
15	1.72 *	26.7	1.71 *	29.9	1.50 *	26.6
	1.05ddd (2.5,4.0,13.05)		1.02 *			
16	0.95 *	29.7	0.95 *	29.9	1.60 *	29.9
	1.85 dt (9.0,13.0)					
17	-	37.8	-	37.4	-	37.2
18	1.45 *	40.5	1.44 *	41.5	1.45 *	41.5

Table 1. Cont.

Position	1		1a		2	
	δ_H	δ_C	δ_H	δ_C	δ_H	δ_C
19	2.00 t (7.0)	31.6	2.03 tbr (7.0)	31.7	1.79 q (6.6)	32.3
20	-	144.9	-	146.5	-	147.7
21	5.89 d (6.5)	126.3	5.87 d (6.4)	123.1	5.75 d (6.3)	119.2
22	3.35 dbr (6.6)	73.3	4.51 d (6.4)	75.3	4.55 d (6.3)	75.3
23	0.78 s	28.0	0.79 s	28.1	0.81 s	28.0
24	0.77 s	16.6	0.78 s	16.7	0.80 s	16.5
25	0.82 s	16.4	0.82 s	16.7	0.81 s	16.4
26	0.99 s	16.1	0.98 s	16.2	0.96 s	16.1
27	0.93 s	14.7	0.91 s	14.7	0.92 s	14.6
28	0.63 s	17.8	0.70 s	17.9	0.70 s	18.2
29	0.99 d (6.5)	22.2	0.97 d (7.6)	22.1	0.96 d (6.6)	22.6
30	4.15 d (11.2)	47.6	4.14 d (11.2)	47.5	3.96 d (12.6)	72.3
	3.89 d (11.2)		3.89 d (11.6)		3.72 d (12.6)	
OH	3.14 m					
OAc	1.97 s	21.4	1.97 s	21.3	1.97 s	21.3
		171.4	1.98 s	21.3	1.98 s	21.3
				171.1		171.1
				171.1		171.1
OEt					1.13 t (6.9)	15.2
					3.35 m	65.9

^a δ in ppm and J (in Hz) are in parentheses. Recorded in CDCl₃ at 400 MHz and 125 MHz for ¹H and ¹³C-NMR, respectively. * overlapped.

Figure 2. Selected correlations of **1**. Double-ended arrows indicate NOESY, and single arrows indicate HMBC (C to H) correlations.



The most important HMBC and NOESY correlations are shown in Figure 1. Treatment of **1** with Ac₂O-pyridine gave a diacetyl derivative **1a** for which its HREIMS experiment indicated the molecular formula C₃₄H₅₃O₄³⁷Cl (calcd. for [M]⁺ 562.3603; found 562.3624) and C₃₄H₅₄O₄³⁵Cl (calcd. for [M]⁺ 560.3654; found 560.3644). The data implied the presence of a double bond between C-20 and C-21, the chloro atom at C-30, and the acetyl group at C-3 and hydroxyl group at C-22. H-22 was assigned in β -orientation on the basis of the coupling constant with the vinylic proton at C-21 and cross-peak in the NOESY experiment with the CH₃-28. From the above findings, the structure of

30-chloro-3 β -acetoxo-22 α -hydroxyl-20(21)-taraxastene was assigned to **1**, and it was named chlorotolpidiol. To the best of our knowledge, this is the first example of a pentacyclic triterpene of the taraxastane-ursane series with a chloro functionality.

Compound **2** was obtained as a colourless amorphous solid and its molecular formula was determined by a HRESIMS experiment as C₃₆H₅₈O₅ (calcd. for [M+Na]⁺ 593.4182; found 593.4176). The IR spectrum revealed the absorption bands for a carbonyl group (1732 cm⁻¹) and double bond (2872 cm⁻¹). The ¹H-NMR spectrum (Table 1) exhibited six singlet methyl groups at δ_H 0.70, 0.80, 0.81 (6H), 0.92 and 0.96, a secondary methyl group at δ_H 0.96 (3H, d, J = 6.6 Hz) attributed to C-29, a vinyl proton at δ_H 5.75 (1H, d, J = 6.3 Hz), two acetyl groups at δ_H 1.97 s and 1.98 s, two oxymethine signals at δ_H 4.43 (1H, dd, J = 5.1, 10.4 Hz) and 4.55 (1H, d, J = 6.3 Hz), an oxymethylene signal at δ_H 3.96 (1H, d, J = 12.6 Hz) and 3.72 (1H, d, J = 12.6 Hz) and an ethoxy group at δ_H 3.35 (2H, m) and 1.13 (3H, t, J = 6.9 Hz). The extra acetyl signal at C-22 was observed, since the oxygenated methine proton of **1** at δ_H 3.35 (1H, d, J = 6.6 Hz) was displaced to δ_H 4.55 (1H, d, J = 6.3 Hz) in **2**. Moreover, the halogenated group was replaced by a ethoxyl group at C-30 since the carbon in **2** was displaced to low field at δ_C 72.3 (Table 1). The structure of **2** was determined by a combination of COSY, DEPT, HSQC, HMBC, and NOESY experiments. Based on the above data, the new compound tolpidiol A **2** was identified as 3 β ,22 α -diacetoxo-30-ethoxy-20(21)-taraxastene.

Compound **3** was purified as its diacetate **3a** by treatment with acetic anhydride (Ac₂O) in pyridine, **3a** was isolated as a colourless amorphous solid and its HRESIMS experiment indicated the molecular formula C₃₄H₅₄O₆ (calcd. for [M+Na]⁺ 581.3818; found 581.3801). The IR spectrum revealed the absorption bands for carbonyl 1728 cm⁻¹, and hydroxyl 3391 cm⁻¹ groups, the presence of these groups was confirmed by the ¹H and ¹³C-NMR spectra (Table 2). The ¹H-NMR spectrum of **3a** showed signals for five tertiary methyl groups at δ_H 0.81(6H, br, s), 1.00, 1.02 and 1.11, and two secondary methyl groups at δ_H 0.86 (3H, d, J = 6.4 Hz) and δ_H 0.88 (3H, d, J = 7.3 Hz) suggesting a pentacyclic triterpene with an ursane skeleton. An olefinic proton at δ_H 5.30 (1H, d, J = 3.1 Hz) was assigned to H-12, two oxygenated methines at δ_H 4.45 (1H, dd, J = 3.0, 9.6 Hz) and 4.46 (1H, dd, J = 5.0, 9.5 Hz) corresponding to H-3 and H-11 respectively, the latter showing vicinal correlation in the COSY experiment with the olefinic proton H-12, while the proton H-9 δ_H 1.81 (1H, d, J = 9.5 Hz) indicated the presence of a hydroperoxide at C-11. The presence of an oxygenated methylene was confirmed by the signals at δ_H 3.56 (1H, d, J = 11.0 Hz) and 3.93 (1H, d, J = 11.0 Hz). The ¹³C-NMR (Table 2) and DEPT data indicated the presence of two ester carbonyl groups, nine methyl carbons, nine aliphatic methylenes, two olefinic carbons, and seven methine carbons. Thus, the position of acetyl groups in compound **3a** was assigned by a HMBC correlation between the signal at δ_C 171.0 and that at δ_H 4.45; and the signals at δ_C 171.3 and δ_H 3.93. The coupling constant between H-9 and H-11 (J = 9.5 Hz) established the α -orientation of the hydroperoxide at C-11. The EI-MS data of **3a** showed direct loss of H₂O m/z 540 and H₂O₂ m/z 524, confirming the presence of the hydroperoxide.

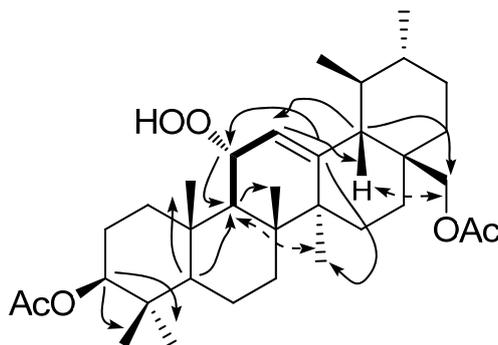
The structure elucidation and NMR assignments were based primarily on the results of HSQC, HMBC, and COSY experiments which allowed the complete assignment of all H- and C-atoms, and the NOESY (Figure 3) data provided the configuration of compound **3a**. Therefore, the structure of **3** was established as 3 β ,28-dihydroxy-11 α -hydroperoxy-12-ursene. To the best of our knowledge, compound **3** is a novel triterpenoid, which we named tolpidiol B.

Table 2. ^1H and ^{13}C -NMR data for compound **3a**^a.

Position	δH	δC
1	0.85 m 2.08 dt (3.5, 7.0)	39.4
2	1.58 *	23.7
3	4.45 dd (3.0, 9.6)	80.6
4	-	38.0
5	0.88 m	55.3
6	1.45 * 1.31 *	18.1
7	1.45 m 1.25 m	33.3
8	-	43.2
9	1.81 d (9.5)	48.8
10	-	37.8
11	4.46 dd (5.0, 9.5)	81.6
12	5.30 d (3.1)	125.8
13	-	144.5
14	-	42.0
15	1.60 * 0.90 m	26.2
16	1.16 * 1.92 td (3.5, 9.0)	23.3
17	-	36.9
18	1.45 m	53.7
19	1.35 m	39.0
20	1.28 m	39.3
21	1.40 m	30.4
22	1.31 m 1.52 dt (3.0, 6.5)	35.5
23	0.81 s	28.2
24	0.81 s	16.7
25	1.02 s	16.8
26	1.00 s	18.0
27	1.11 s	22.2
28	3.56 d (11.0) 3.93 d (11.0)	71.0
29	0.86 d (6.4)	17.4
30	0.88 d (7.3)	21.3
	1.97 s	21.0
OAc	1.97 s	21.3
		171.0
		171.3

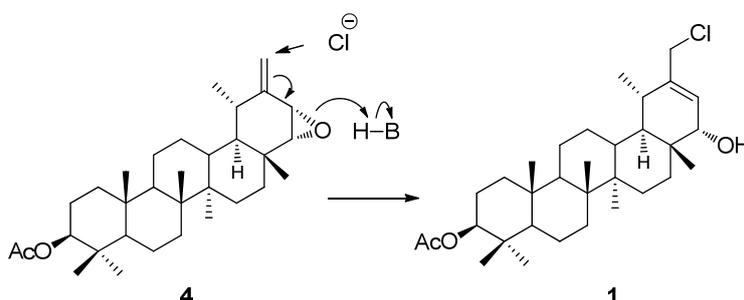
^a δ in ppm and J (in Hz) are in parentheses. Recorded in CDCl_3 at 400 MHz and 125 MHz for ^1H - and ^{13}C -NMR, respectively. * overlapped.

Figure 3. Selected correlations of **3a**. Bond bolded indicate COSY, Double-ended arrows indicate NOESY, and single arrows indicate HMBC (C to H) correlations.



Compound **1** could derive from the known triterpene acetyl-ptiloepoxide **4** [8,9] which was identified by us from *T. proustii* as an inseparable mixture. Triterpenes containing an epoxide at the Δ^{21-22} position are known and have been isolated before from a *Tolpis* species [4]. Based on this, we envisioned the formation of compound **1** by chlorination of the double bond, followed by isomerization and β opening of the epoxide, and protonation, maintaining the α -orientation at the C-22 observed in the precursor compound acetyl-ptiloepoxide (Figure 4).

Figure 4. Tentative forming process of compound **1**.



Although the number is relatively small, several halogenated triterpenes and other higher terpenes have been described mainly from marine sources [10]. However, the presence of chlorinated triterpenoids in terrestrial plants is very rare and just few cases have been reported [11–14]. Initially, compound **1** seemed to be an artifact of the isolation process, but Chen *et al.* [14] proved that to obtain chlorinated compounds, a chlorine source such as CHCl_3 with HCl is necessary. During the isolation process, no chlorinated solvents were used (see Experimental). In the chromatographic separation, dichloromethane was used in the preparative TLC, which was not enough to interact with the possible precursor (ptiloepoxide) of **1**. Compound **2** was isolated as a presumed artifact; this compound was probably obtained from **1**, by a nucleophilic substitution reaction due to the use of hot EtOH during the extraction process.

Additionally, from *T. proustii* 15 known compounds were isolated, including aromatic compounds: scopoletin [15] aesculetin [15] and apigenin [16]; the diterpene phytene-1,2-diol [17] and the triterpenoids stigmasterol [18], ergosterol peroxide [19], ursolic acid [20], lupan-20(29)-ene-3 β ,30-diol [21], 21 α -hydroxytaraxasterol [22], 11-oxo- β -amyrin [23], 3 β -acetoxy-urs-12-ene-1 β ,11 α -diol [24], 21 α ,22 α -epoxy-20 α -hydroxy-20(30)-dihydrotaraxasterol [4], 3 β -acetoxy-21 α ,22 α -epoxytaraxastan-20 α -ol [25], 22-oxo-20-taraxastan-3 β -ol [9], β -amyrin [26]. From *T. lagopoda* seven known

compounds were isolated, including aromatic compounds: 2,4'-dihydroxy-4-methoxybenzophenone [4] and triterpenoids: stigmasterol [18], ergosterol peroxide [19], a mixture of 7-oxo- β -sitosterol and 7-oxo-stigmasterol [26,27], ursolic acid [20], and α -amyrin [28]. Their structures were confirmed by comparison of their spectral data with those reported in the literature.

2.2. Antioxidant Activities

Natural antioxidants that are present in plants are responsible for inhibiting or preventing the deleterious consequences of oxidative stress. In Table 3 the relative antioxidant efficiency of both *Tolpis* extracts against the DPPH radical is shown. Antioxidants suppress the absorbance at 515 nm on a time scale dependent on the antioxidant activity of extracts. The RSA of the crude extract of *T. proustii* (59.6%) was higher than that of *T. lagopoda* (41.4%). FRAP assay was used to study the ability of the antioxidants in the extracts to reduce ferric iron to the ferrous form. The same behaviour as for the DPPH assay was observed, *T. lagopoda* being less active than *T. proustii* (4.1 and 18.1 μmol of Fe(III) reduced to Fe(II) per gram of dry plant respectively) (Table 3). On the other hand, the free radical scavenging and ferric reducing power assays revealed that aesculetin (isolated from *T. proustii*) showed the highest antioxidant activities as compared with those of α -tocopherol and BHA (Table 4). Aesculetin gave a RSA value of 100% with a $t_{1/2}$ (time required for 50% scavenging of DPPH radical in the specified concentration of antioxidant) of 22.5 seconds, while BHA and α -tocopherol showed RSA of 21.9 and 17.7% respectively after 20 min. Aesculetin (at concentration 0.1 mg mL^{-1}) showed also higher antioxidant activities than both extracts (at concentration 10 mg mL^{-1}), because the extracts are complex mixtures that include active components at lower levels. Moreover, the crude extracts tend to have more interfering substances that may interact with the antioxidants, decreasing their effectiveness. The antioxidant activities found in this study indicated that aesculetin, as well as both extracts, are ideal for use in the health food industry. Because of the high content of aesculetin found in the *T. proustii* extract (566.8 mg), this extract may be considered to be a natural source of aesculetin with diverse potential therapeutic uses.

Table 3. Antioxidant activity of crude extracts derived from *T. proustii* and *T. lagopoda*.

Assays	<i>T. proustii</i>	<i>T. lagopoda</i>
RSA ^a	59.6 \pm 0.4	41.4 \pm 0.1
FRAP ^b	18.1 \pm 0.4	4.1 \pm 0.2
FRAP ^c	93 \pm 2	41 \pm 1

^a % inhibition \pm standard deviation of three measurements. ^b μmol of Fe(III) reduced to Fe(II) per gram of dry plant \pm standard deviation of three measurements. ^c μmol of Fe(III) reduced to Fe(II) per gram of ethanolic residue \pm standard deviation of three measurements.

Table 4. Antioxidant activity of aesculetin, α -tocopherol and butylated hydroxyanisole (BHA).

Assays	Aesculetin 0.1 mg mL^{-1}	α -Tocopherol 0.1 mg mL^{-1}	BHA 0.1 mg mL^{-1}
DPPH ^a	100 \pm 0	17.7 \pm 0.1	21.9 \pm 0.6
FRAP ^b	9.4 \pm 0.7	0.97 \pm 0.03	3.13 \pm 0.05

^a % inhibition \pm standard deviation of three measurements. ^b μmol of Fe(III) reduced to Fe(II) per mg of compound \pm standard deviation of three measurements.

2.3. Cytotoxic Activity

K-562 and K-562/ADR cells which are sensitive or resistant to doxorubicin, respectively, were incubated with the compounds shown in Table 5 to evaluate their potential cytotoxicity. After 72 h, cell survival was determined by the MTT assay and the IC₅₀ values are summarized in Table 5. Among the different compounds ursolic acid and 22-oxo-20-taraxasten-3β-ol exhibit the strongest effects in mitochondrial reduction of tetrazolium salts to formazan, while the ursolic derivatives and the 1,2-diacetoxiphytene exhibit the weakest effects. Furthermore, K-562 and K-562/ADR cells exhibit comparable sensitivity to compounds ursolic acid and 22-oxo-20-taraxasten-3β-ol (Table 5). These results suggest that the overexpression of the drug efflux protein, P-glycoprotein does not confer resistance against these compounds.

Table 5. Effects of some compounds and derivatives isolated from *T. proustii* and *T. lagopoda* on the growth of the human leukemia cell lines.

Compound	IC ₅₀ (μM)	
	K562	K562/ADR
Ursolic acid	40.6 ± 3.6	49.2 ± 3.1
Ursolic acid methyl ester	59.3 ± 15.5	64.0 ± 14.5
Acetyl ursolic acid	99.5 ± 20.5	>100
Acetyl ursolic acid methyl ester	>100	>100
Aesculetin	63.2 ± 3.2	77.0 ± 5.1
Aesculetin acetyl	68.6 ± 17.1	70.3 ± 19.2
Aesculetin diacetyl	62.3 ± 6.6	59.5 ± 4.5
11-Oxo-β-amyrin	>100	>100
22-Oxo-20-taraxasten-3β-ol	30.0 ± 10.0	43.0 ± 7.0
1,2-Diacetoxiphytene	>100	>100

Cells were cultured for 72 h and the IC₅₀ values were calculated as described in the Experimental section. The data shown represent the means ± SEM of 3–5 independent experiments with three determinations in each.

3. Experimental

3.1. General Experimental Procedures

Optical rotations: Perkin-Elmer *model 343* polarimeter. IR Spectra: Bruker model *IFS-55* spectrophotometer. ¹H and ¹³C-NMR spectra: Bruker model AMX-500 and AMX-400 spectrometers with standard pulse sequences, operating at 500 and 400 MHz for ¹H-, and 125 MHz for ¹³C-NMR, CDCl₃ was used as solvent and TMS as internal standard. EI-MS: *Micromass model Autospec* (70 eV) spectrometer. The constituents of the ethanolic extracts were separated by gravity column chromatography, medium pressure liquid chromatography (MPLC) and preparative TLC. Column chromatography (CC): silica gel SiO₂; (70–230 mesh, Merck), column fractions were monitored by TLC (silica gel *60 F₂₅₄*), Medium pressure column chromatography (MPLC): silica gel *Merck* (40–63 μm). Prep. TLC: silica gel *60 PF₂₅₄₊₃₆₆* plates (20 × 20 cm, 1-mm thickness).

3.2. Plant Material

The plant material was identified by Dr. Rosa Febles. *T. proustii* was collected at Roque Agando (La Gomera, Canary Islands) in May 2009. A voucher specimen has been deposited at the Herbarium of the Viera y Clavijo Botanical Garden in Las Palmas de Gran Canaria (LPA 24194). *T. lagopoda* was collected at Tenteniguada (Gran Canaria, Canary Islands) in May 2008. A voucher specimen has been deposited at the Herbarium of the Viera y Clavijo Botanical Garden in Las Palmas de Gran Canaria (LPA 23596).

3.3. Extraction and Isolation

The aerial parts of *T. proustii* (3.0 Kg) were extracted with ethanol (4 L) in a Soxhlet until exhaustion. Solvent removal afforded a viscous residue (541.5 g) which was fractionated by means of CC (SiO₂; hexane/AcOEt step gradients). The fractions eluted with hexane-AcOEt (7:3) were submitted to a new MPLC chromatography with hexane-AcOEt (9:1), to give stigmasterol (30.6 mg); 3 β -acetoxy-21 α ,22 α -epoxy-20 α -hydroxy-20(30)-dihydrotaraxasterol (6.3 mg), obtained by preparative TLC (benzene-ethyl acetate 8:2); ergosterol peroxide (obtained as its acetylated derivative upon acetylation of one of the obtained fractions (2.3 mg); 3 β ,30-dihydroxylup-20(29)-ene (31.1 mg); phytene-1,2-diol (7.6 mg); **3a** (2.3 mg), isolated by successive preparative TLCs (benzene-AcOEt, 9.9:0.1, 2 elutions; dichloromethane-hexane 1:1, 3 elutions) and **1** (3.7 mg), obtained by preparative TLC (dichloromethane, 3 elutions). Subsequent chromatography by MPLC with hexane-AcOEt (8.5:1.5) gave ursolic acid (193.8 mg); 21 α -hydroxytaraxasterol (8,9 mg), obtained by preparative TLC (benzene-AcOEt, 8.5:1.5, 4 elutions); 11-oxo- β -amyryn (15.4 mg); 22-oxo-taraxasten-3 β -ol (2.7 mg) isolated by preparative TLC (dichloromethane-AcOEt 9:1, 4 elutions); β -amyryn (5.2 mg); and **2** (2.1 mg), obtained upon crystallization in hexane-AcOEt and subsequent purification by preparative TLC (dichloromethane-AcOEt 8:2).

The fractions eluted with hexane-AcOEt (3:2) were partially rechromatographed by MPLC in hexane-AcOEt (7:3) giving, after preparative TLC (benzene-AcOEt, 8:2, 2 elutions), apigenin (41.2 mg) and scopoletin (6.7 mg). The remainder of these fractions was submitted to subsequent rechromatography by MPLC in hexane-AcOEt (8.5:1.5) to give 3 β -acetoxy-1 β ,11 α -dihydroxy-urs-12-ene (9.7 mg), isolated by preparative TLC (benzene-AcOEt, 8:2, 4 elutions), and 21 α ,22 α -epoxy-20 α -hydroxy-20(30)-dihydrotaraxasterol (3.6 mg), purified by preparative TLC (hexane-AcOEt, 8:2, 2 elutions). Finally, the fractions eluted with hexane-AcOEt (1:1) afforded aesculetin (566.8 mg), upon crystallization in hexane-AcOEt.

The aerial parts of *T. lagopoda* (2.7 Kg) were extracted with ethanol (4 L) in a Soxhlet until exhaustion. Removal of the solvent afforded a viscous residue (279.8 g), which was fractionated by CC (SiO₂; hexane/AcOEt step gradients). Fractions eluted with hexane-ethyl acetate (4:1) were subsequently chromatographed by MPLC with hexane-ethyl acetate (7.5:2.5) to give five fractions (numbered from fraction 1 to 5). From fraction 1 stigmasterol (14.3 mg) was purified by crystallization in hexane-AcOEt, while fraction 3 afforded 2,4'-dihydroxy-4-methoxybenzophenone, obtained by crystallization with hexane-AcOEt (15.1 mg) and from its mother liquors α -amyryn (2.9 mg) was obtained by preparative TLC with (benzene-AcOEt, 9:1, 5 elutions), as its acetylated derivative, after

acetylation. From fraction 4 ursolic acid (6.1 mg) was purified after a methylation process. Fraction 5 afforded ergosterol peroxide (11 mg) by preparative TLC (hexane-AcOEt, 9:1, 2 elutions), purified after acetylation as its acetyl derivative. Finally, fraction 2 afforded compound **2** (1.8 mg) by preparative TLC (benzene-AcOEt, 9:1). Fractions eluted with hexane-AcOEt (7:3) were subsequently chromatographed by MPLC with hexane-AcOEt (8:2), the most polar fractions affording the mixture of 7-oxo- β -sitosterol and 7-oxo-stigmasterol (18.1 mg) were purified by preparative TLC (hexane-AcOEt, 7:3, 5 elutions).

30-Chloro-3 β -acetoxo-22 α -hydroxyl-20(21)-taraxastene (**1**). Amorphous white solid; $[\alpha]_D^{25} = +6.0$ (CHCl₃, c 0.001); for ¹H and ¹³C-NMR data, see Table 1. IR, ν max (KBr): 3446, 2924, 2853, 1732, 1439, 1244, 1172 cm⁻¹. EIMS m/z 520 (5.7), 518 (15.5), 502 (13.5), 500 (35.2), 483 (7.3), 460 (4.3), 458 (10.5), 464 (13.6), 404 (5.1) 249 (21.0), 190 (36.5) 189 (100.0), 187(40.0) 161 (17.0), 133(29.0) 121 (36.0), 81(31.6). HREIMS m/z 520.3497 [M]⁺ (calcd for C₃₂H₅₁O₃³⁷Cl 520.3497), 518.3517 [M]⁺ (calcd for C₃₂H₅₁O₃³⁵Cl 518.3527), 502.3414 [M-H₂O]⁺ (calcd for C₃₂H₄₉O₂³⁷Cl 502.3392), 500.3436 [M-H₂O]⁺ (calcd for C₃₂H₄₉O₂³⁵Cl 500.3421).

Acetylation of **1**. Compound **1** (2.0 mg) was dissolved in pyridine (1 mL) and acetic anhydride (2 mL), and the reaction was further stirred at room temperature for 12 h and after usual work-up. The product was dried under vacuum to furnish **1a** (1.8 mg). For ¹H and ¹³C-NMR spectroscopic data, see Table 1.

30-Chloro-3 β ,22 α -diacetoxo-20(21)-taraxastene (**1a**). Amorphous white solid; $[\alpha]_D^{25} = +86.7$ (CHCl₃, c 4.5×10^{-3}); for ¹H and ¹³C-NMR data, see Table 1. IR, ν max (KBr): 2920, 2852, 1731, 1652, 1540, 1456, 1372, 1247, 1016, 982 cm⁻¹. EIMS m/z 562 (0.7), 560 (1.7), 525 (34.7), 500 (27.0), 466 (10.4), 404 (6.4), 189 (100), 95 (52.8), 69 (56). HREIMS m/z 562.3624 [M]⁺ (calcd for C₃₄H₅₃O₄³⁷Cl 562.3603), 560.3644 [M]⁺ (calcd for C₃₄H₅₃O₄³⁵Cl 560.3632).

3 β ,22 α -Diacetoxo-30-ethoxy-20(21)-taraxastene (**2**). Colourless amorphous solid; $[\alpha]_D^{25} = +4.8$ (CHCl₃, c 0.015); for ¹H and ¹³C-NMR data, see Table 1. IR, ν max (KBr): 2931, 2872, 1732, 1652, 1464, 1456, 1671, 1244, 1016 cm⁻¹. EIMS m/z 510 (100) [M-OAc]⁺, 450 (40) [M-2OAc]⁺. HRESIMS positive ion m/z 593.4176 [M+Na]⁺ (Calcd. for C₃₆H₅₈O₅Na, 593.4182).

3 β ,28-Diacetoxo-11 α -hydroperoxy-12-ursene **3a**. Colourless amorphous solid; $[\alpha]_D^{25} = +9.5$ (CHCl₃, c 0.014); for ¹H and ¹³C-NMR data, see Table 2. IR, ν max (KBr): 3391, 2951, 2925, 2875, 1728, 1392, 1244, 1032, 983, 903 cm⁻¹. HREIMS m/z 540.3829 [M-H₂O]⁺ (Calcd. for C₃₄H₅₂O₅, 540.3815); m/z 524.3873 [M-H₂O₂]⁺ (Calcd. for C₃₄H₅₂O₄, 524.3866). HRESIMS positive ion m/z 581.3801 [M+Na]⁺ (Calcd. for C₃₄H₅₄O₆Na, 581.3818).

3.4. Antioxidant Experiments

3.4.1. Chemicals

Methanol HPLC grade (Panreac, Barcelona, Spain) and Milli-Q water (18MQ, Millipore, Billerica, MA, USA) were always used in this study. Formic acid and sodium acetate (Merck, Darmstadt, Germany) were analytical quality reagents. 1,1-Diphenyl-2-picrylhydrazyl (DPPH) and 2,4,6-tri (2-

pyridyl)-1,3,5-triazine (TPTZ) were purchased from Sigma-Aldrich Chemie (Steinheim, Germany); rutin and gentisic and caffeic acids were supplied by Merck. Ferric chloride ($\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$), ferrous sulphate ($\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$) and glacial acetic acid were obtained from Panreac.

3.4.2. Preparation of Extracts for Antioxidant Assays

The ethanolic extract residue was solved in methanol by stirring at room temperature. After centrifugation at 7,000 rpm for 10 min, the supernatant was collected and the antioxidant activity was measured. Aesculetin, α -tocopherol and BHA (5 mg) were dissolved in methanol (10 mL). The solution was diluted to 0.1 mg mL^{-1} to be used for antioxidant assays.

3.4.3. Free Radical Scavenging Activity on DPPH

The reducing ability of antioxidants towards DPPH radical was evaluated by measuring the loss of 1,1-diphenyl-2-picrylhydrazyl (DPPH) colour at 515 nm after reaction with test extracts [29]. The sample solution (30 μL) was rapidly mixed with 1 mL of a 0.1 mM DPPH solution. After 25 min incubation time in the dark at ambient temperature (23 °C), the decline in absorbance against a methanol blank was recorded. The inhibition percentage values were calculated by equation: Radical Scavenging Activity (RSA) = $100 (1 - \text{Abs in the presence of sample} / \text{Abs in the absence of sample})$.

3.4.4. Ferric Reducing Antioxidant Power Assay (FRAP)

Reducing power was determined according to [30]. This method is based on the reduction of Fe^{3+} to Fe^{2+} , which is recorded by measuring the formation of a blue colored Fe^{2+} -tripirydyltriazine compound from the colourless oxidized Fe^{3+} form by the action of electron donating antioxidants. The FRAP reagent consists of 300 mM acetate buffer (3.1 g sodium acetate + 16 mL glacial acetic acid, made up to 1 litre with distilled water; pH = 3.6), 10 mM TPTZ in 40 mM HCl and 20 mM $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ in the ratio of 10:1:1.

Extract (10 μL) was added to 1.0 mL freshly prepared and prewarmed (37 °C) FRAP reagent. The mixture was incubated at 37 °C for 10 min and the absorbance was measured against a reagent blank (1.0 mL FRAP reagent + 10 μL distilled water) at 593 nm. A standard curve of Fe^{2+} was constructed over the concentration range of $0.2 \mu\text{mol L}^{-1}$ to $1 \mu\text{mol L}^{-1}$. The results were determined by the regression equation of the curve ($y = 0.00221x + 0.00999$, $r = 0.9969$) and expressed as μmol ferric ions reduced to ferrous form per g of dry plant material.

3.5. Cytotoxic Experiments

3.5.1. Cell Culture

Human leukemia K-562 cells (DSMZ No: ACC 10, German Collection of Microorganisms and Cell Cultures, Braunschweig, Germany) were cultured in RPMI 1640 containing 2 mM L-glutamine supplemented with 10% (v/v) heat-inactivated fetal bovine serum. The K-562/ADR cell line was kindly provided by Professor Lisa Oliver (INSERM, Nantes, France) and cultured as above.

3.5.2. Assay for Growth Inhibition and Cell Viability

The cytotoxicity of compounds was assessed using a 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) assay. Briefly, 1×10^4 exponentially growing cells were seeded in 96-well microculture plates with various compounds concentrations (3–100 μM) in a volume of 200 μL . DMSO concentration was the same in all the treatments and did not exceed 0.1% (v/v). After 72 h, surviving cells were detected based on their ability to metabolize 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H-tetrazolium bromide into formazan crystals. Optical density was read with an ELISA reader at a wavelength of 570 nm and was used as a measure of cell viability. The MTT dye reduction assay measures mitochondrial respiratory function and can detect the onset of cell death earlier than dye-exclusion methods. Cell survival was calculated as the fraction of cells alive relative to control for each point: cell survival (%) = mean absorbance in treated cells/mean absorbance in control wells \times 100. Concentrations inducing a 50% inhibition of cell growth (IC_{50}) were determined graphically using the curve fitting algorithm of the computer software Prism 4.0 (GraphPad). Values are means \pm SEM from three independent experiments, each performed in triplicate.

4. Conclusions

We have studied two endemic Canary plants of the *Tolpis* genus and two new compounds tolpidiol A (**2**) and B (**3**) and a novel chloro derivative chlorotolpidiol (**1**) were isolated from *T. proustii* and *T. lagopoda*. The isolation of halogenated triterpenoids from terrestrial sources is very rare, and few cases have been reported. To the best of our knowledge, chlorotolpidiol is the first example of a pentacyclic triterpene of the taraxastane-ursane series chlorinated at the position C-30. Herein we present a tentative explanation of the biogenesis of this class of compounds. The presence of chlorine atoms provides chemically addressable handles for further work in organic-medicinal chemistry. The antioxidant activities of the ethanolic extract were evaluated and it showed relative efficiency. We found weak cytotoxic activity against K562 and K-562/ADR cell lines of some of the isolated and derivative compounds.

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Sample Availability: Samples of the compounds **1a**, **2** and **3** are available from the authors.

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