

Article

Synthesis and Biological Activity Evaluation of Novel β -Substituted Nitromethylene Neonicotinoid Analogues

Baozhu Wang, Jiagao Cheng, Zhiping Xu, Xiaoyong Xu, Xusheng Shao and Zhong Li *

Shanghai Key Laboratory of Chemical Biology, School of Pharmacy, East China University of Science and Technology, P.O. Box 544, 130 Meilong Road, Shanghai 200237, China

* Author to whom correspondence should be addressed; E-Mail: lizhong@ecust.edu.cn; Tel.: +86-021-6425-2603; Fax: +86-021-6425-3540.

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Abstract: The structure-based design and synthesis of a series of novel neonicotinoid analogues are described. The novel neonicotinoid analogues were designed based upon the reaction of enamine derivatives with electron-withdrawing β -substituents with electrophilic thiocyanogen reagents. These compounds were characterized by spectroscopic methods. Bioassays indicated that some of the synthesized compounds exhibited excellent bioactivity against cowpea aphids (*Aphis craccivora*). The LC₅₀ values of compounds **7**, **9**, **12**, **13**, **15**, **17**, **19**, **20** and commercial imidacloprid were 0.01567, 0.00974, 0.02494, 0.01893, 0.02677, 0.01778, 0.0220, 0.02447 and 0.03502 mmol L⁻¹, respectively, which suggested that they could be used as leads for future development of new insecticides.

Keywords: neonicotinoid analogues; new design strategy; thiocyanogen; insecticidal activities

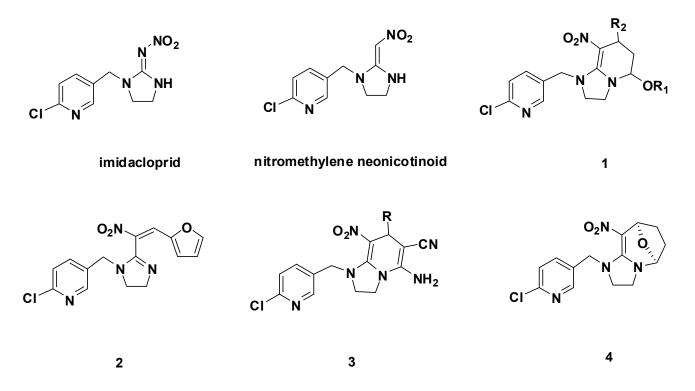
1. Introduction

In recent years, neonicotinoid insecticides have been the fastest growing class of insecticides in modern crop protection, with widespread use against a broad spectrum of sucking and certain chewing pests [1–5]. Significant progress had been obtained on the chemistry of the insecticidal nicotinic agonists and the chemical biology of the nicotinic acetylcholine receptors (nAChRs), that is, chemorational approaches including physicochemical considerations, metabolism, resistance

mechanisms, and chemical and structural biology aspects potentially expediting receptor structure-guided insecticide design [6–12].

Our past endeavors in this field have focused on systematic pharmacophore modifications of nitromethylene neonicotinoids that appeared to capture the "bioactive" conformation (Figure 1). Several types of nitromethylene neonicotinoids with the *cis*-configuration were presented; that is, neonicotinoids **1** with a tetrahydropyridine fixed *cis*-configurations; neonicotinoids **2** with bulky group fixed *cis*-configurations, neonicotinoids **3** with *cis*-configuration constructed by aza-Diels-Alder reactions, and divalent and oxa-bridged neonicotinoids **4** constructed from dialdehydes (Figure 1). Bioassays indicated that the synthesized *cis* compounds **1**–**4** exhibited excellent bioactivity against cowpea aphids [13–16]. According to our knowledge, the nitro groups in all commercialized neonicotinoids have a *trans* configuration, on which three proposals for mode of action are based [17]. The high bioactivity of compounds **1**–**4** implies that neonicotinoids in the *cis* configuration might bind to the receptor in a different way [13]. These efforts accordingly encourage the discovery of "superneonicotinoids" effective for imidacloprid-resistant pests and lepidoptera species.

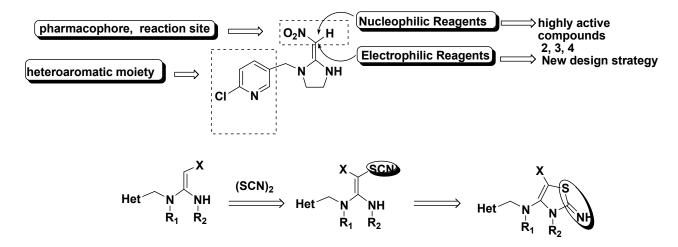
Figure 1. Structures of imidacloprid neonicotinoids and structurally related analogues.



The structure derivative method through the organic reaction is another important tool for systematic pharmacophore modifications [18]. It had been theorized that the nitroenamine entities in these high active compounds play important roles for specific binding to the desired insect nAChRs site [19–22]. Encouraged by these reports, the modifications of nitroenamine entities to fix its configuration through organic reactions has attracted our attention. The chemistry of enamines, including push–pull enamines bearing an additional electron-acceptor group at the β -position, has been extensively developed, mainly as a result of electrophilic reactions that proceed predominantly at the β -position [23–25]. Electrophilic modification, followed by a set of transformations, have made enamines an integral part of synthetic organic chemistry methodology [26]. Additionally, electron-acceptor

groups can participate in chemical transformations. According to this knowledge, a series of prototype compounds with extended N-substituted (thiocyanogen) substituents were designed in this paper (Figure 2). The choice of the thiocyanogen was predominantly governed by its traits as electrophilic reagent, and mainly by its ability to easily cyclize with the NH group in the imidazoline ring to fix the electron-withdrawing group in the *cis* position (Figure 2) [27].

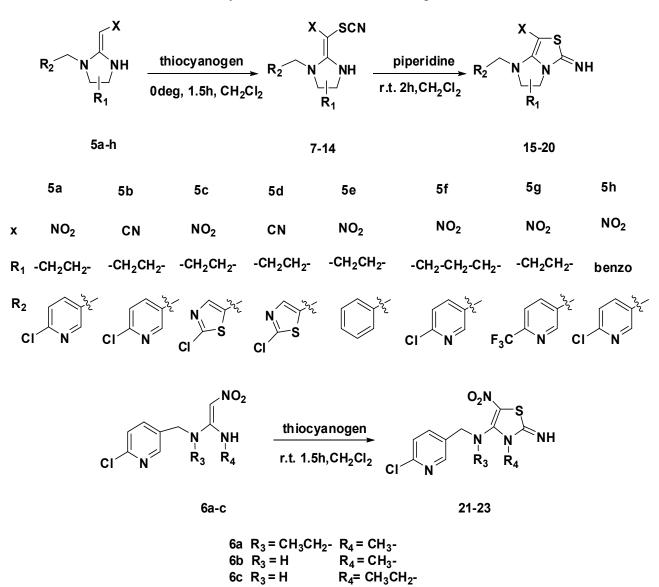
Figure 2. Structures of imidacloprid neonicotinoids and its structurally related analogues.



2. Results and Discussion

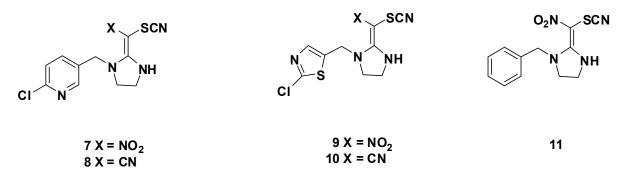
2.1. Synthetic Chemistry

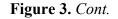
The synthetic route of the target compounds is summarized in Scheme 1. The chemical structures of the novel neonicotinoid analogues under study are depicted in Figure 3. The preparation of the designed compounds began with the common intermediates 5a-h and 6a-c which were synthesized according to the method of Kagabu et al. [19]. In the reaction of 5a-h with thiocyanogen at 0 °C for 1.5 h, the corresponding β -thiocyanated products 7–14 were produced in 63%–82% yields. In this procedure, 5a appears to produce a mixture of 7 and cyclization product 15. Similarly, 5c and 5f also gave mixtures of aminothiazole and the relevant cyclization product. Compounds 7, 9 and 12 were synthesized by reducing the reaction time and reaction temperature without forming cyclization products. Then, these compounds 7–9 and 11–13 were converted quantitatively into the corresponding cyclization products, 15–20, by stirring at room temperature in dichloromethane solution containing 0.1% piperidine as catalyst. Unexpectedly, 10 and 14 could not be converted into the corresponding cyclization products. The substrates 6a-c yielded only the corresponding cyclization products 21-23 respectively under the same conditions. Whereas configuration of compounds 7-14 and 21-23 cannot be determined easily by standard ¹H- and ¹³C-NMR techniques, the compounds 15-20 obtained from the reaction system are considered to be produced with retention of the configuration of the preformed enamines with electron-withdrawing β -substituents, followed by catalyzed cyclization. The structures of the title compounds were well characterized by ¹H-NMR, ¹³C-NMR and HRMS.

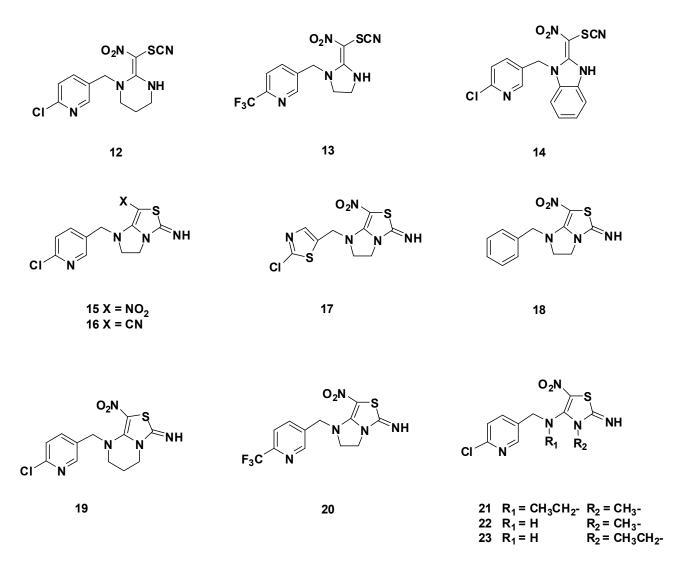


Scheme 1. Synthetic route for the title compounds 7–23.

Figure 3. Chemical structures of novel neonicotinoid analogues under study.







2.2. Biological Activities

The insecticidal activities of the designed compounds against cowpea aphids are shown in Table 1. Most of the title compounds showed excellent insecticidal activities against cowpea aphids. For example, the LC₅₀ values of compounds **7**, **9**, **12**, **13**, **15**, **17**, **19**, and **20** were 0.01567, 0.00974, 0.02494, 0.01893, 0.02677, 0.01778, 0.0220 and 0.02447 mmol L⁻¹, respectively, whereas that of imidacloprid was only 0.03502 mmol L⁻¹ [16]. Activities varied significantly depending upon the patterns of substituents on the enamine and imidazole ring. It was observed that compounds with NO₂ substituents (**7**, **15**, **9** and **17**) demonstrated better activities than that with CN substituents (**8**, **10** and **16**). Generally, the imidazolidine substituents appear to be the important factor for insecticidal activities. A benzene substituent (compound **14**) resulted in complete inactivity. Moreover, the cyclic addition of SCN substituents with the imidazolidine ring is unfavourable for high activities from the data analysis present of compounds **7**, **9**, **15** and **17**.

Compounds	Mortality (%, 500mg L^{-1})	$LC_{50} (mmol L^{-1})$
7	100	0.01567
8	34.6	nt ^a
9	100	0.00974
10	0	nt
11	23.98	nt
12	100	0.02494
13	100	0.01893
14	87.82	nt
15	100	0.02677
16	32.4	nt
17	100	0.01778
18	16.19	nt
19	100	0.02200
20	100	0.02447
21	57.6	nt
22	100	0.11839
23	50.79	nt

Table 1. Insecticidal activities of compounds 7–23 against cowpea aphids (*Aphis craccivora*).

3. Experimental

3.1. General

Melting points (mp) were recorded on a Büchi B540 apparatus (Büchi Labortechnik AG, Flawil, Switzerland) and are uncorrected. ¹H-NMR and ¹³C-NMR spectra were recorded on a Bruker AM-400 (400 MHz) spectrometer with CDCl₃ or DMSO- d_6 as the solvent and TMS as the internal standard. Chemical shifts are reported in δ (parts per million) values. High-resolution mass spectra were recorded under electron impact (70 eV) condition using a MicroMass GCT CA 055 instrument. Analytical thin-layer chromatography (TLC) was carried out on precoated plates (silica gel 60 F₂₅₄), and spots were visualized with ultraviolet (UV) light. The compounds **5a–h** and **6a–c** were synthesized according to the method of Kagabu *et al.* [19]. Unless otherwise noted, reagents and solvents were used as received from commercial suppliers. Yields were not optimized. All reactions were carried out under a protective atmosphere of drying nitrogen or utilizing a calcium chloride drying tube.

3.1.1. General Synthetic Procedure for 7-14

To a solution of 5a-g (10 mmol) in dichloromethane (100 mL), a thiocyanogen (10 mmol) solution in dichloromethane (50 mL), which was previously prepared from lead thiocyanate and bromine at -15 °C, was added dropwise with stirring at -15 °C and then the mixture was stirred for 1.5 h at 0 °C. The reaction mixture was washed with cold water (3 × 50 mL) and brine (50 mL), and dried with sodium sulfate. The solvent was removed to give a crude solid, which was recrystallized from a mixture of dichloromethane/ether (2:1, v/v) to give 7–14. 2-Chloro-5-((2-(nitro(thiocyanato)methylene)imidazolidin-1-yl)methyl)pyridine (7). Yield: 92%, m.p.: 130.6–132.1 °C; ¹H-NMR (DMSO- d_6): δ 3.76 (br.s, 4H), 4.77 (s, 2H), 7.55 (d, J = 8.4 Hz, 1H), 7.86 (dd, $J_1 = 2.6$ Hz, $J_2 = 8.4$ Hz, 1H), 8.42 (d, J = 2.0 Hz, 1H), 9.43 (s, 1H) ppm; ¹³C-NMR (DMSO- d_6): δ 162.6, 149.9, 149.1, 139.1, 131.8, 124.7, 112.3, 87.5, 50.7, 50.2, 42.7; HRMS (ES+) calcd for C₁₁H₁₀³⁵ClN₅O₂S (M+H)⁺: 312.0244. Found: 312.0330. Calcd for C₁₁H₁₀³⁷ClN₅O₂S (M+H)⁺: 314.0214 Found: 314.0284.

2-(1-((6-Chloropyridin-3-yl)methyl)imidazolidin-2-ylidene)-2-thiocyanatoacetonitrile (**8**). Yield: 63%, m.p.: 118.2–119.7 °C; ¹H-NMR (DMSO- d_6): δ 3.51–3.56 (m, 2H), 3.66–3.70 (m, 2H), 4.79 (s, 2H), 7.54 (d, J = 8.4 Hz, 1H), 7.78–7.80 (m, 1H), 8.11 (s, 1H), 8.36 (d, J = 2.4 Hz, 1H) ppm; ¹³C-NMR (DMSO- d_6): δ 164.9, 150.0, 149.2, 139.2, 132.1, 124.8, 122.1, 113.8, 51.6, 47.4, 41.5, 31.2; HRMS (EI+) calcd for C₁₂H₁₀³⁵ClN₅S (M⁺): 291.0345. Found: 291.0344. Calcd for C₁₂H₁₀³⁷ClN₅S (M⁺): 293.0316 Found: 293.0318.

1-((2-Chlorothiazol-5-yl)methyl)-2-(nitro(thiocyanato)methylene)imidazolidine (**9**). Yield: 75%, m.p.: 85.1–86.3 °C; ¹H-NMR (DMSO-*d*₆): δ 3.69–3.78 (m, 4H), 4.84 (s, 2H), 7.73 (s, 1H), 9.40 (s, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 162.2, 151.6, 141.8, 135.9, 112.1, 87.3, 50.1, 46.3, 42.6; HRMS (ES+) calcd for C₉H₈³⁵ClN₅O₂S₂ (M+H)⁺: 317.9808. Found: 317.9890. Calcd for C₉H₈³⁷ClN₅O₂S₂ (M+H)⁺: 319.9778 Found: 319.9860.

2-(1-((2-Chlorothiazol-5-yl)methyl)imidazolidin-2-ylidene)-2-thiocyanatoacetonitrile (**10**). Yield: 52%, m.p.: 120.3–122.2 °C; ¹H-NMR (DMSO- d_6): δ 3.30–3.70 (m, 4H), 4.93 (s, 2H), 7.70 (s, 1H), 8.17 (s, 1H) ppm; ¹³C-NMR (DMSO- d_6): δ 164.5, 151.3, 141.6, 135.8, 122.0, 113.5, 51.0, 43.0, 41.5, 31.8; HRMS (ES–) calcd for C₁₀H₈³⁵ClN₅S (M–H)⁺: 295.9910. Found: 295.9829. Calcd for C₁₀H₈³⁷ClN₅S (M–H)⁺: 297.9880 Found: 297.9803.

1-Benzyl-2-(nitro(thiocyanato)methylene)imidazolidine (**11**). Yield: 82%, m.p.: 102.3–104.2 °C; ¹H-NMR (DMSO-*d*₆): δ 3.72 (s, 4H), 4.75 (s, 2H), 7.32–7.42 (m, 5H), 9.40 (s, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 162.3, 136.1, 129.2, 128.1, 127.6, 112.3, 86.9, 52.7, 50.3, 42.6; HRMS (ES+) calcd for C₁₂H₁₂N₄O₂S (M+H)⁺: 277.0681. Found: 277.0750.

1-((6-Chloropyridin-3-yl)methyl)-2-(nitro(thiocyanato)methylene)-hexahydropyrimidine (**12**). Yield: 70%, m.p.: 158.1–160.3 °C; ¹H-NMR (DMSO-*d*₆): δ 2.14 (t, *J* = 5.2, 2H), 3.38 (t, *J* = 5.6 Hz, 2H), 3.85 (t, *J* = 5.6 Hz, 2H), 4.06 (s, 2H), 7.57 (d, *J* = 8.0 Hz, 1H), 7.91 (dd, *J*₁ = 8.4 Hz, *J*₂ = 2.8 Hz, 1H), 8.45 (d, *J* = 2.4 Hz, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 164.7, 158.9, 150.2, 146.2, 140.1, 131.0, 124.5, 106.5, 56.8, 47.2, 45.1, 19.1; HRMS (ES+) calcd for C₁₂H₁₂³⁵ClN₅O₂S (M+H)⁺: 326.0400. Found: 326.0477. Calcd for C₁₂H₁₂³⁷ClN₅ O₂S (M+H)⁺: 328.0371 Found: 328.0465.

5-((2-(Nitro(thiocyanato)methylene)imidazolidin-1-yl)methyl)-2-(trifluoromethyl)pyridine (13). Yield: 82%, m.p.: 140.1–142.3 °C; ¹H-NMR (DMSO-*d*₆): δ 3.76 (s, 4H), 3.38 (t, J = 5.6 Hz, 2H), 4.85 (s, 2H), 7.91 (d, J = 8.0 Hz, 1H), 8.04 (s, 1H), 8.75 (s, 1H), 9.45 (s, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 162.7, 149.3, 137.2, 136.6, 121.1, 112.3, 87.7, 50.9, 50.8, 42.8; HRMS (ES+) calcd for C₁₂H₁₀F₃N₅O₂S (M+H)⁺: 345.0507. Found: 345.0617.

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(E-1-((6-Chloropyridin-3-yl)methyl)-2-(nitro(thiocyanato)methylene)-2,3-dihydro-1H-enzo[d]imid-azole (14). Yield: 74%, m.p.: 125.6–127.2 °C; ¹H-NMR (DMSO-*d* $₆): <math>\delta$ 4.79 (s, 2H), 7.39–7.7 (m, 6H), 8.35 (d, *J* = 2.4 Hz, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 150.1, 148.7, 148.6, 138.5, 132.5, 131.3, 130.4, 126.0, 125.4, 124.8, 113.8, 112.2, 112.0, 86.0, 55.4, 47.2; HRMS (ES–) calcd for C₁₀H₈³⁵ClN₅S (M–H)⁺: 358.0244. Found: 358.0130. Calcd for C₁₀H₈³⁷ClN₅S (M–H)⁺: 360.0214. Found: 297.0166.

3.1.2. General Synthetic Procedure for 15–20

The solution of **7–9** and **11–13** (10 mmol) in dichloromethane (50 mL) containing 0.1% piperidine was stirred for 2 h at room temperature. After removal of the solvent, the residue was recrystallized from dichloromethane to give **15–20** in quantitative yields.

1-((6-Chloropyridin-3-yl)methyl)-7-nitro-2,3-dihydroimidazo[1,2-c]thiazol-5(1H)-imine (**15**). Yield: 93%, m.p.: 136.4–138.5 °C; ¹H-NMR (DMSO-*d*₆): δ 3.85 (s, 2H), 4.08 (s, 2H), 5.27 (s, 2H), 7.58 (br.s, 1H), 7.91–7.92 (m, 1H), 8.47 (s, 1H), 8.91 (br.s, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 150.6, 150.1, 149.7, 139.8, 132.0, 124.8, 99.7, 55.7, 49.4, 41.8; HRMS (ES+) calcd for C₁₁H₁₀³⁵ClN₅O₂S (M+H)⁺: 312.0244. Found: 312.0321. Calcd for C₁₁H₁₀³⁷ClN₅O₂S (M+H)⁺: 314.0279. Found: 314.0284.

l-((6-Chloropyridin-3-yl)methyl)-5-imino-1,2,3,5-tetrahydroimidazo[1,2-c]thiazole-7-carbonitrile (**16**). Yield: 82%, m.p.: 201.5–202.3 °C; ¹H-NMR (DMSO-*d*₆): δ 3.87 (s, 4H), 4.61 (s, 2H), 7.41 (d, *J* = 8.0 Hz, 1H), 7.73–7.76 (m, 1H), 8.38 (d, *J* = 2.0 Hz, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 155.2, 153.0, 152.0, 149.3, 138.9, 129.1, 125.0, 114.7, 53.2, 47.8, 47.2, 41.0; HRMS (ES+) calcd for C₁₂H₁₀³⁵ClN₅S (M+H)⁺: 292.0345. Found: 292.0430. Calcd for C₁₂H₁₀³⁷ClN₅S (M+H)⁺: 294.0316. Found: 294.0410.

1-((2-Chlorothiazol-5-yl)methyl)-7-nitro-2,3-dihydroimidazo[1,2-c]thiazol-5(1H)-imine (17). Yield: 78%, m.p.: 150.3–151.4 °C; ¹H-NMR (DMSO-*d*₆): δ 3.81–4.16 (m, 4H), 5.34 (s, 2H), 7.77 (s, 1H), 8.96 (s, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 152.3, 150.0, 142.0, 136.0, 99.9, 55.5, 45.2, 41.7; HRMS (EI+) calcd for C₉H₈ClN₅O₂S₂ (M⁺): 316.9808. Found: 316.9807.

1-Benzyl-7-nitro-2,3-dihydroimidazo[1,2-c]thiazol-5(1H)-imine (**18**). Yield: 74%, m.p.: 82.3–84.4 °C; ¹H-NMR (DMSO-*d*₆): δ 3.81–4.06 (m, 4H), 5.26 (s, 2H), 7.32–7.43 (s, 1H), 8.96 (s, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 150.2, 136.4, 129.2, 128.3, 55.5, 52.3, 41.6; HRMS (ES+) calcd for calcd for C₁₂H₁₂N₄O₂S (M+H)⁺: 277.0681. Found: 277.0746.

1-((6-Chloropyridin-3-yl)methyl)-8-nitro-1,2,3,4-tetrahydrothiazolo[3,4-a]pyrimidin-6-imine (19). Yield: 65%, m.p.: 165.3–167.4 °C; ¹H-NMR (DMSO-*d*₆): δ 2.01 (t, *J* = 5.2 Hz, 2H), 3.30 (t, *J* = 5.6 Hz, 2H), 3.73 (t, *J* = 5.6 Hz, 2H), 4.90 (s, 2H), 7.53 (d, *J* = 8.4 Hz, 1H), 7.92 (dd, *J*₁ = 8.4 Hz, *J*₂ = 2.4 Hz, 1H), 8.43 (d, *J* = 2.0, 1H), 9.19 (s, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 153.5, 150.5, 150.0, 149.8, 139.9, 131.9, 124.4, 103.2, 56.5, 47.3, 41.6, 19.3; HRMS (ES+) calcd for C₁₂H₁₂³⁵ClN₅O₂S (M+H)⁺: 326.0400. Found: 326.0476. Calcd for C₁₂H₁₂³⁷ClN₅O₂S (M+H)⁺: 328.0371. Found: 328.0473.

7-*Nitro-1-((6-(trifluoromethyl)pyridin-3-yl)methyl)-2,3-dihydroimidazo[1,2-c]thiazol-5(1H)-imine* (**20**). Yield: 88%, m.p.: 148.2–149.3 °C; ¹H-NMR (DMSO-*d*₆): δ 3.85 (t, *J* = 8.2 Hz, 2H), 4.10 (t, *J* = 9.2 Hz, 2H), 5.36 (s, 2H), 7.94 (d, *J* = 8.4 Hz, 1H), 8.13 (d, *J* = 8.4 Hz, 1H), 8.81(s, 1H), 8.94 (s, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 151.0, 149.8, 137.8, 136.8, 121.1, 56.0, 50.0, 41.8; HRMS (ES+) calcd for C₁₂H₁₀F₃N₅O₂S (M+H)⁺: 346.0507. Found: 346.0634.

3.1.3. General Synthetic Procedure for 21-23

To a solution of **6a–c** (10 mmol) in dichloromethane (50 mL) was added with stirring at -15 °C a dichloromethane solution of thiocyanogen (10 mmol) which had previously prepared from lead thiocyanate and bromine at -15 °C, and then the mixture was stirred for 1.5 h at room temperature. The reaction mixture was washed with cold water (3 × 50 mL) and brine (50 mL) and then dried with sodium sulfate. The solvent was evaporated *in vacuo*, and fractional precipitation of the residue from dichloromethane afforded **21–23**.

N-((6-Chloropyridin-3-yl)methyl)-N-ethyl-2-imino-3-methyl-5-nitro-2,3-dihydrothiazol-4-amine (21). Yield: 84%, m.p.: 120.3–121.4 °C; ¹H-NMR (DMSO-*d₆*): δ 1.14 (t, *J* = 7.2 Hz, 3H), 3.17 (d, *J* = 7.2 Hz, 2H), 3.40 (s, 3H), 4.57 (s, 2H), 7.54 (d, *J* = 8.0 Hz, 1H), 7.92–7.95 (m, 1H), 8.49 (d, *J* = 2.0 Hz, 1H) ppm; ¹³C-NMR (DMSO-*d₆*): δ 165.1, 150.7, 150.5, 140.9, 131.8, 124.8, 118.2, 51.9, 46.8, 36.1, 13.9; HRMS (EI+) calcd for C₁₂H₁₄ClN₅O₂S (M⁺): 327.0557. Found: 327.0559.

N-((6-Chloropyridin-3-yl)methyl)-2-imino-3-methyl-5-nitro-2,3-dihydrothiazol-4-amine (22). Yield: 82%, m.p.: 116.4–118.4 °C; ¹H-NMR (DMSO-*d*₆): δ 3.28 (s, 3H), 5.37 (s, 2H), 7.80 (s, 1H) ppm; ¹³C-NMR (DMSO-*d*₆): δ 152.1, 142.5, 141.2, 42.5, 34.6, 33.3; HRMS (ES–) calcd for C₈H₈³⁵ClN₅O₂S₂ (M–H)⁺: 303.9808. Found: 303.9716. calcd for C₈H₈³⁷ClN₅O₂S₂ (M–H)⁺: 305.9778. Found: 305.9703.

N-((6-Chloropyridin-3-yl)methyl)-3-ethyl-2-imino-5-nitro-2,3-dihydrothiazol-4-amine (23). Yield: 78%, m.p.: 115.4–117.2 °C; ¹H-NMR (DMSO-*d*₆): δ 1.23 (t, *J* = 7.2 Hz, 3H), 3.66 (q, *J* = 7.2 Hz, 2H), 5.34 (s, 2H), 7.81 (s, 1H); ¹³C-NMR (DMSO-*d*₆): δ 152.1, 141.2, 134.7, 42.7, 42.1, 15.7; HRMS (ES–) calcd for C₉H₁₀³⁵ClN₅O₂S₂ (M-H)⁺: 317.9964. Found: 317.9870. calcd for C₉H₁₀³⁷ClN₅O₂S₂ (M–H)⁺: 319.9935. Found: 319.9836.

3.2. Biological Assay

All compounds were dissolved in acetone and diluted with water containing Triton X-100 (0.1 mg L⁻¹) to obtain series concentrations of 500.0, 250.0, 125.0 mg L⁻¹ and others for bioassays. Pea aphids (*Aphis craccivora*) were dipped according to our previously reported procedure [13,23]. Tender shoots of soybean with 40-60 healthy apterous adults were dipped in diluted solutions of the chemicals containing Triton X-100 (0.1 mg L⁻¹) for 5 s, the superfluous fluid was removed, and the shoots were placed in the conditioned room ($25 \pm 1 \text{ °C}$, 50% RH). Water containing Triton X-100 (0.1 mg L⁻¹) was used as control. Mortality was assessed after 24 h. Each treatment had three repetitions, and the data were adjusted and subjected to probit analysis as before.

4. Conclusions

In conclusion, a series of a novel class of neonicotinoids, in which the common nitromethylene pharmacophore was substituted by electrophilic thiocyanogen reagents, were designed and synthesized.

Most of the compounds exhibited excellent insecticidal activities against cowpea aphids (*A. craccivora*), which implied that reaction of nitromethylene with the electrophilic system to obtain novel neonicotinoids analogues with high activities was feasible.

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Sample Availability: Samples of the title compounds are available from the authors.

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