## **Supplementary Materials**

**Figure S1.** CD spectra for the c-MYC mutant promoter sequence i-motif DNA construct in BPES buffer with added glycerol (20% w/w). Red line (—): the CD spectra of the DNA structure at pH 4.0. Blue line (—): the CD spectra of the DNA structure at pH 4.5. Purple line (—): the CD spectra of the DNA structure at pH 5.0. Yellow line (—): the CD spectra of the DNA structure at pH 5.6. Green line (—): the CD spectra of the DNA structure at pH 6.0. Orange line (—): the CD spectra of the DNA structure at pH 6.5; Black line (—): the CD spectra of the DNA structure at pH 7.0.



Results of the DSC melting experiments for the mutant c-MYC i-motif forming sequence in 0%, 10%, 20% and 30% w/w PEG<sub>12000</sub> containing buffer solutions at pH 5.0 and 6.0 are summarized in Table S1. The DSC results show that at both pH 5.0 and 6.0, the stability of the i-motif, in terms of melting temperature, increases as the concentration of PEG<sub>12000</sub> increases. This observation is consistent with the results of the i-motif stability in PEG<sub>8000</sub> containing buffers with PEG concentrations from 0% to 30%.

**Table S1.** The DSC determined melting temperatures,  $T_ms$  of the i-motif structure formed in 0%, 10%, 20% and 30% w/w PEG<sub>12000</sub> solutions at pH 5.0 and 6.0.

% PEG <sub>12000</sub>	<i>T<sub>m</sub></i> of oligonucleotide in pH 5.0 solutions	<i>T<sub>m</sub></i> of oligonucleotide in pH 6.0 solutions
0	41.5	23.0
10	44.9	26.4
20	48.5	36.8
30	51.0	46.7

If we compare the  $T_ms$  of the c-MYC i-motif in two different PEG containing buffers (PEG<sub>8000</sub> and PEG<sub>12000</sub>), but having the same concentration and the same pH values, we see that at the same concentration (20% or 30% w/w), PEG<sub>8000</sub> exhibits the better stabilization of the i-motif in pH 5.0 solutions, while PEG<sub>12000</sub> exhibits the better stabilization of the i-motif at pH 6.0. The  $T_m$  data in Table 2 in combination with the  $T_m$  data in Table S1 show that at pH = 5.0, the melting of the c-MYC i-motif occurs 1.5 to 2.1°C lower in 20% PEG<sub>12000</sub> solutions than in 20% PEG<sub>8000</sub> solutions. In contrast, at pH = 6.0, the melting of the c-MYC i-motif occurs approximately 4°C higher in 20% PEG<sub>12000</sub> solutions than in 20% PEG<sub>8000</sub> solutions. All of the DSC melting experiments done in the

 $PEG_{12000}$  show the presence of a second higher melting peak in the thermogram that we have attributed to the disassociation or unfolding of a DNA/PEG complex (see Figure 3).

CD spectra for pH titration experiments done on the mutant c-MYC promoter sequence in 0%–30% w/w PEG<sub>12000</sub> containing buffer solutions are shown in Figure S2. The CD results show that in 10%, 20% and 30% PEG<sub>12000</sub> solutions, the highest pH at which a stable i-motif occurs in each of these solutions is approximately pH = 6.1, pH = 6.2, or pH = 6.3. Again, these results in terms of i-motif stabilization as a function of PEG concentration are similar for PEG<sub>8000</sub> and PEG<sub>12000</sub>.

**Figure S2.** CD spectra for pH titration experiments done for the mutant c-MYC promoter sequence in 0-30% w/w PEG<sub>12000</sub> containing buffer solutions.

