

Effect of Genotype and Environment on *Salvia miltiorrhiza* Roots Using LC/MS-Based Metabolomics

Qi Zhao, Zhenqiao Song, Xinsheng Fang, Yuling Pan, Linlin Guo, Tian Liu and Jianhua Wang

Table S1. The correlation coefficients from OPLS-DA of *S. miltiorrhiza* extracts from paired locations for each genotype (ZY, Zhuyang; CQ, Changqing; TA, Taian).

Putative Annotation	Genotype 1			Genotype 2			Genotype 3			Genotype 4		
	ZY/CQ	ZY/TA	CQ/TA	ZY/CQ	ZY/TA	CQ/TA	ZY/CQ	ZY/TA	CQ/TA	ZY/CQ	ZY/TA	CQ/TA
Salvianolic acid F	-0.998 ^a	0.991	1.000	0.365	0.369	-0.111	0.830	-0.987	-0.995	-0.997	-0.999	-0.911
Salvianolic acid I	-0.998	-0.999	0.987	-1.000	-0.984	0.985	-0.990	-0.999	-0.971	0.149	-0.998	-0.999
Salvianolic acid E	-0.999	-0.998	0.996	-0.993	-0.994	-0.964	-1.000	-1.000	-1.000	-0.991	-0.991	-0.985
Rosmarinic acid	0.790	0.999	0.997	-0.979	-0.910	-0.866	-0.816	-0.983	-0.975	-0.999	-0.984	-0.826
Lithospermic acid	-0.967	0.987	0.989	-0.999	-0.999	-0.979	-0.864	-0.998	-0.976	-0.999	-1.000	-0.992
Prolithospermic acid derivative	-0.956	0.998	0.995	-0.999	-0.998	-0.980	0.956	-0.999	-0.992	-0.494	-0.438	0.749
Salvianolic acid B	0.946	0.975	0.966	-0.998	-0.998	-0.884	-0.987	-0.983	-0.999	-0.960	-0.998	-0.960
Tanshinone IIB	-0.931	-0.941	-0.980	-0.962	-0.967	-0.994	-0.926	-0.932	0.342	-0.963	-0.974	-0.924
Trijukanone C	-0.915	-0.923	-0.658	-0.978	-0.978	-0.867	-0.997	-0.999	-0.955	-0.799	-0.939	-0.996
15,16'-dihydrotanshinone I	-0.872	-0.901	-0.971	-0.997	-0.997	-0.994	-0.961	-0.971	-0.949	-0.994	-0.998	-0.997
Methyltanshinonate	-0.924	-0.942	-0.995	-1.000	-0.999	-0.999	-0.998	-0.999	-0.986	-0.998	-0.997	-0.998
Trijukanone B	-0.420	-0.919	-0.627	-0.988	-0.991	-0.999	-0.965	-0.973	-0.996	-0.988	-0.990	-0.069
Cryptotanshinone	-0.886	-0.928	-0.942	-0.978	-0.981	-1.000	-0.962	-0.967	-0.953	-0.951	-0.936	0.841
Tanshinone I	-0.806	-0.845	-0.811	-0.992	-0.992	1.000	-0.839	-0.906	-0.981	-0.998	-0.998	-0.996
1,2'-dihydrotanshinone I	-0.594	-0.769	-0.997	-1.000	-0.999	-0.996	-0.998	-1.000	-0.995	-0.993	-0.990	-0.740
Tanshinone IIA	-0.860	-0.922	-0.997	-1.000	-1.000	-0.996	-0.997	-0.999	-0.991	-0.993	-0.990	-0.226

^a Positive and negative signs indicate positive and negative correlation in the concentration, respectively. The coefficient of 0.90 was used as the cutoff value based on the significant difference evaluation ($p < 0.001$).

Table S2. *p*-Value of *S. miltiorrhiza* extracts from paired locations for each genotype (ZY, Zhuyang; CQ, Changqing; TA, Taian).

Putative Annotation	Genotype 1				Genotype 2				Genotype 3				Genotype 4			
	ZY/CQ	ZY/TA	CQ/TA	ZY/CQ	ZY/TA	CQ/TA	ZY/CQ	ZY/TA	CQ/TA	ZY/CQ	ZY/TA	CQ/TA	ZY/CQ	ZY/TA	CQ/TA	
Salvianolic acid F	1.4×10^{-17} a,*	5.0×10^{-11} *	7.4×10^{-19} *	5.5×10^{-1}	7.7×10^{-1}	1.1×10^{-1}	7.6×10^{-4}	2.4×10^{-9} *	1.1×10^{-10} *	3.7×10^{-14} *	5.7×10^{-15} *	2.3×10^{-5} *				
Salvianolic acid I	7.5×10^{-22} *	1.4×10^{-21} *	8.8×10^{-7} *	1.2×10^{-13} *	5.8×10^{-10} *	3.3×10^{-10} *	3.2×10^{-9} *	2.4×10^{-13} *	7.3×10^{-8} *	2.6×10^{-1}	2.6×10^{-14} *	2.3×10^{-14} *				
Salvianolic acid E	1.1×10^{-18} *	2.2×10^{-16} *	2.2×10^{-13} *	5.6×10^{-12} *	1.1×10^{-12} *	1.8×10^{-4} *	1.5×10^{-17} *	2.3×10^{-20} *	8.1×10^{-16} *	7.1×10^{-11} *	2.8×10^{-11} *	2.5×10^{-4} *				
Rosmarinic acid	1.2×10^{-2}	8.1×10^{-15} *	1.9×10^{-14} *	2.7×10^{-7} *	3.1×10^{-4} *	3.1×10^{-4} *	5.1×10^{-3}	1.4×10^{-8} *	4.5×10^{-7} *	5.0×10^{-11} *	7.6×10^{-10} *	9.5×10^{-4}				
Lithospermic acid	8.7×10^{-5} *	4.0×10^{-10} *	5.4×10^{-11} *	2.4×10^{-17} *	3.0×10^{-18} *	8.8×10^{-9} *	2.6×10^{-4} *	5.5×10^{-10} *	3.1×10^{-8} *	5.7×10^{-18} *	3.6×10^{-19} *	5.3×10^{-11} *				
Prolithospermic acid derivative	2.5×10^{-6} *	5.3×10^{-12} *	2.0×10^{-13} *	2.7×10^{-13} *	3.4×10^{-15} *	3.8×10^{-9} *	2.3×10^{-7} *	1.7×10^{-8} *	1.9×10^{-11} *	2.5×10^{-1}	3.6×10^{-1}	2.6×10^{-1}				
Salvianolic acid B	1.1×10^{-4} *	1.6×10^{-8} *	1.1×10^{-7} *	2.0×10^{-12} *	2.2×10^{-13} *	8.9×10^{-5} *	6.8×10^{-11} *	2.9×10^{-9} *	1.1×10^{-13} *	5.2×10^{-7} *	8.7×10^{-11} *	1.2×10^{-7} *				
Tanshinone IIB	4.2×10^{-6} *	1.7×10^{-6} *	1.1×10^{-5} *	1.2×10^{-7} *	7.0×10^{-8} *	1.9×10^{-4} *	4.4×10^{-6} *	4.6×10^{-6} *	7.3×10^{-1}	7.3×10^{-8} *	2.9×10^{-8} *	8.7×10^{-5} *				
Trijugaranone C	1.1×10^{-5} *	7.2×10^{-6} *	1.2×10^{-1}	3.4×10^{-9} *	2.5×10^{-9} *	1.1×10^{-1}	4.9×10^{-14} *	4.1×10^{-14} *	1.2×10^{-6} *	1.8×10^{-3}	3.2×10^{-6} *	2.6×10^{-4} *				
15,16'-dihydrotanshinone I	1.9×10^{-4} *	4.3×10^{-5} *	9.6×10^{-5} *	1.3×10^{-14} *	7.9×10^{-15} *	1.3×10^{-4} *	1.6×10^{-7} *	4.7×10^{-8} *	6.3×10^{-4} *	5.2×10^{-13} *	6.8×10^{-14} *	8.0×10^{-5} *				
Methyltanshinonate	9.7×10^{-6} *	1.8×10^{-6} *	5.5×10^{-4} *	5.9×10^{-20} *	2.0×10^{-20} *	8.7×10^{-8} *	9.7×10^{-17} *	8.7×10^{-18} *	5.1×10^{-9} *	1.6×10^{-13} *	4.9×10^{-15} *	4.9×10^{-8} *				
Trijugaranone B	4.0×10^{-1}	5.1×10^{-4} *	9.8×10^{-2}	1.0×10^{-10} *	3.1×10^{-11} *	8.3×10^{-5} *	7.5×10^{-8} *	2.9×10^{-8} *	8.5×10^{-5} *	3.2×10^{-10} *	3.0×10^{-10} *	1.7×10^{-1}				
Cryptotanshinone	6.0×10^{-5} *	5.2×10^{-6} *	2.6×10^{-4} *	8.2×10^{-9} *	2.2×10^{-9} *	2.6×10^{-4} *	1.5×10^{-7} *	4.2×10^{-8} *	5.3×10^{-4} *	8.9×10^{-7} *	1.8×10^{-6} *	1.5×10^{-1}				
Tanshinone I	1.1×10^{-3}	3.6×10^{-4} *	1.1×10^{-1}	7.8×10^{-12} *	1.1×10^{-11} *	2.7×10^{-4} *	2.6×10^{-4} *	2.5×10^{-5} *	4.1×10^{-4} *	1.9×10^{-14} *	1.1×10^{-15} *	7.0×10^{-8} *				
1,2'-dihydrotanshinone I	5.8×10^{-2}	2.9×10^{-3}	3.9×10^{-4} *	2.3×10^{-18} *	5.8×10^{-19} *	1.2×10^{-7} *	3.7×10^{-17} *	7.3×10^{-19} *	2.9×10^{-12} *	9.8×10^{-11} *	3.4×10^{-11} *	1.7×10^{-1}				
Tanshinone IIA	2.2×10^{-4} *	8.2×10^{-6} *	1.0×10^{-4} *	2.6×10^{-18} *	4.8×10^{-19} *	1.2×10^{-8} *	4.6×10^{-15} *	1.2×10^{-16} *	9.0×10^{-10} *	6.7×10^{-11} *	5.5×10^{-11} *	1.5×10^{-1}				

^a Multiple testing was corrected by Bonferroni correction. * $p < 7.57 \times 10^{-4}$ (0.05/66).

Table S3. Morphological characters and disease resistance of four genotypes.

Morphological characters and disease resistance	Genotype 1	Genotype 2	Genotype 3	Genotype 4	Significance
Cultivar or strain	cultivar	strain	cultivar	strain	
Accession name	Shandong Danshen NO.1	-	Shandong Danshen NO.2	-	
Leaf color	dark green	light green	dark green	light green	
Cauline basal petiole color	purple	purple	purple	green	
Plant types	erect type	spreading type	erect type	central type	
Average blade length (cm)	9 a	6 b	5 b	6 b	***
Average width of blade (cm)	7 a	3 c	4 b	5 b	**
Average plant height (cm)	51 a	40 b	44 b	63 a	*
Average crown width (cm ²)	2400 a	2000 b	1600 c	1700 c	***
Largest stem diameter (mm)	11 a	6 b	3 c	7 b	***
Cauline basal ramification number	8	6	10	7	NS
Longest root length (cm)	29 a	27 b	26 b	16 c	**
Largest root diameter (mm)	10 b	13 a	8 c	5 d	**
Average number of roots ^a	14 c	17 b	6 d	20 a	**
Root dry matter (g.plant ⁻¹)	275 a	298 a	84 b	69 b	***
Root rot	2% b	4% a	4% a	4% a	**
Root knot nematode	2% c	7% a	6% b	5% b	**

^aDiameter > 2mm. ^bValues in each row having different lowercase letters (a, b, c, and d) were significantly different at $p < 0.05$. NS, not significant ($p > 0.05$);

* $p < 0.05$; ** $p < 0.01$; *** $p < 0.001$.

Table S4. The correlation coefficients from OPLS-DA of *S. miltiorrhiza* extracts from different genotypes for each locations (1, Genotype 1; 2, Genotype 2; 3, Genotype 3; 4, Genotype 4).

Putative Annotation	Zhuyang			Changiqng			Taian		
	1/2	2/3	3/4	1/2	2/3	3/4	1/2	2/3	3/4
Procatechuic acid	0.997^a	0.955	-0.990	–	–	–	-0.984	-0.985	0.997
Salvianolic acid F	-0.991	0.951	0.954	0.993	0.987	-0.992	-0.999	-0.973	-0.977
Salvianolic acid I	-0.998	0.996	-0.983	–	0.998	0.529	0.936	0.865	-0.999
Salvianolic acid E	-0.989	-0.748	-0.933	0.940	0.990	-0.996	-0.995	-0.997	0.996
Rosmarinic acid	-0.997	0.932	-0.994	-0.986	0.978	-0.983	-0.999	0.642	-0.733
Lithospermic acid	0.989	0.975	-0.997	-0.987	0.993	-0.988	-0.994	0.999	-0.997
Prolithospermic acid derivative	-0.997	0.992	-0.618	-0.986	0.978	-0.769	-0.999	-0.970	0.997
Salvianolic acid B	0.952	0.918	-0.982	-0.995	0.998	-0.973	-0.995	0.950	-0.921
7 α -hydroxyallyl-royleanone	-0.725	0.623	0.509	-0.997	0.964	0.971	-0.959	0.994	-0.998
Tanshinone IIB	0.140	-0.340	0.759	-0.969	0.970	0.949	-0.997	0.998	0.968
15,16'-dihydrotanshinone I	-0.423	0.768	0.942	-0.973	0.973	0.989	-0.941	0.994	0.991
Methyltanshinonate	0.018	0.986	0.975	-0.980	0.984	0.989	0.136	0.995	0.999
1,2'-dihydrotanshinone I	0.769	-0.980	0.611	-0.968	0.983	0.969	0.991	0.995	0.994
Tanshinone IIA	0.217	0.046	0.909	-0.981	0.982	0.932	0.894	0.991	0.991

^a Positive and negative signs indicate positive and negative correlation in the concentration, respectively. The coefficient of 0.90 was used as the cutoff value based on the significant difference evaluation ($p < 0.001$).

Table S5. *P*-value of *S. miltiorrhiza* extracts from different genotypes for each locations (1, Genotype 1; 2, Genotype 2; 3, Genotype 3; 4, Genotype 4).

Putative Annotation	Zhuyang			Changiqng			Taian		
	1/2	2/3	3/4	1/2	2/3	3/4	1/2	2/3	3/4
Procatechuic acid	4.3×10^{-11} a,*	1.8×10^{-8} *	1.6×10^{-14} *	—	—	—	2.1×10^{-10} *	1.4×10^{-4} *	4.5×10^{-16} *
Salvianolic acid F	2.9×10^{-13} *	1.7×10^{-8} *	3.7×10^{-7} *	4.1×10^{-12} *	4.1×10^{-12} *	6.7×10^{-15} *	1.0×10^{-20} *	2.5×10^{-8} *	9.9×10^{-10} *
Salvianolic acid I	4.4×10^{-26} *	1.1×10^{-10} *	3.5×10^{-11} *	—	4.1×10^{-18} *	1.2×10^{-1}	2.0×10^{-7} *	6.3×10^{-5} *	4.5×10^{-14} *
Salvianolic acid E	1.6×10^{-8} *	3.1×10^{-1}	4.0×10^{-6} *	3.8×10^{-7} *	2.9×10^{-11} *	8.0×10^{-16} *	3.0×10^{-17} *	1.1×10^{-11} *	1.3×10^{-8} *
Rosmarinic acid	1.7×10^{-11} *	9.5×10^{-7} *	2.3×10^{-19} *	1.0×10^{-12} *	2.1×10^{-7} *	2.3×10^{-7} *	3.0×10^{-17} *	1.1×10^{-3}	8.0×10^{-3}
Lithospermic acid	2.1×10^{-12} *	2.1×10^{-8} *	1.4×10^{-18} *	5.3×10^{-10} *	2.7×10^{-14} *	1.3×10^{-12} *	5.1×10^{-16} *	2.7×10^{-6} *	3.1×10^{-7} *
Prolithospermic acid derivative	8.4×10^{-5} *	5.4×10^{-4} *	8.3×10^{-2}	7.3×10^{-10} *	1.2×10^{-8} *	2.8×10^{-2}	3.1×10^{-21} *	3.1×10^{-4} *	2.3×10^{-13} *
Salvianolic acid B	1.5×10^{-4} *	4.8×10^{-7} *	1.8×10^{-12} *	4.1×10^{-11} *	3.2×10^{-14} *	1.1×10^{-10} *	5.7×10^{-15} *	1.4×10^{-4} *	2.5×10^{-4} *
7 α -hydroxyallyl-royleanone	2.5×10^{-2}	1.4×10^{-2}	1.1×10^{-3}	1.3×10^{-12} *	5.2×10^{-6} *	5.7×10^{-10} *	3.6×10^{-9} *	2.1×10^{-4} *	7.7×10^{-5} *
Tanshinone IIB	1.5×10^{-2}	1.5×10^{-1}	4.1×10^{-2}	4.3×10^{-4} *	1.0×10^{-4} *	4.2×10^{-8} *	6.6×10^{-13} *	1.1×10^{-19} *	2.3×10^{-9} *
15,16'-dihydrotanshinone I	8.6×10^{-1}	2.6×10^{-1}	1.8×10^{-4} *	9.4×10^{-8} *	2.1×10^{-7} *	7.5×10^{-12} *	5.3×10^{-6} *	7.7×10^{-11} *	6.9×10^{-13} *
Methyltanshinonate	1.3×10^{-2}	2.0×10^{-4} *	5.9×10^{-4} *	3.1×10^{-4} *	4.7×10^{-11} *	6.8×10^{-15} *	2.3×10^{-1}	3.4×10^{-14} *	5.5×10^{-21} *
1,2'-dihydrotanshinone I	2.2×10^{-3}	4.1×10^{-4} *	2.1×10^{-1}	1.8×10^{-5} *	1.1×10^{-10} *	3.8×10^{-10} *	2.1×10^{-4} *	6.3×10^{-4} *	4.6×10^{-16} *
Tanshinone IIA	2.3×10^{-1}	1.6×10^{-1}	3.4×10^{-4} *	4.4×10^{-7} *	2.4×10^{-9} *	3.0×10^{-7} *	1.5×10^{-2}	2.3×10^{-4} *	2.2×10^{-15} *

^a Multiple testing was corrected by Bonferroni correction. * $p < 7.57 \times 10^{-4}$ (0.05/66).

Table S6. Unidentified compounds in *S. miltiorrhiza* samples.

Peak No.	Unidentified Metabolites	Molecular Weight	λ_{max} (nm)	RT (min)	Negative Ion Mode		Positive Ion Mode	
					[M-H] ⁻	Major Fragments	[M+Na] ⁺	[M+H] ⁺
3	U1	226	294–310	5.20			249	227
8	U2	418	316–345–358–371	17.40	417		419	397
9	U3	339	319	17.60	338		362	340
11	U4	452	251–288–317	19.20			475	453
12	U5	162	212–282–324	19.60				163
19	U6	396	212–255–280	26.60			419	397
22	U7	718	285–310	28.49	717			719
25	U8	301	285–310–330	32.51			324	302
27	U9	296	260–310	35.62	295			
28	U10	298	285–320	36.35	297			299
31	U11	298	285–315–380	42.24	297			
35	U12	340	255–270–380	45.37			363	341
38	U13	318	250–380	45.92			341	319
41	U14	488	250(sh)–380	46.69	487			
43	U15	308	255–350–380	47.35			331	309
45	U16	358	245(sh)	48.12			381	359
51	U17	314	250–280–360	50.31	313			
54	U18	314	250–350–360	53.82	313			
56	U19	280	250	56.65				281
57	U20	331	255–270–360–380	56.97			354	332
58	U21	279	250–320–330–360	57.72			310	280
61	U22	273	255–335–360	59.44			296	274
63	U23	317	255–280–410	60.76			318	340
64	U24	314	275–410–440–255	61.26	313	269, 213		
65	U25	358	255–280	62.70			381	359
66	U26	477	255	63.10			500	478

Table S7. Percentage of paired locations (12) and paired genotypes (18) with relative abundances that were statistically significant ($p < 7.57 \times 10^{-4}$).

Metabolites	Percentage of Paired Locations ^a	Percentage of Paired Genotypes ^b	Metabolites	Percentage of Paired Locations	Percentage of Paired Genotypes	Metabolites	Percentage of Paired Locations	Percentage of Paired Genotypes
Danshensu	67%	56%	Tormentic acid	58%	44%	Dehydromiltirone	8%	6%
Procatechuic acid	67%	61%	Trijukanone C	75%	33%	Procatechu aldehyde	25%	33%
Salvianolic acid F	67% ^c	100%	Danshenxinkun A	75%	33%	Caffeic acid	33%	33%
Salvianolic acid I	92%	78%	Przewa tanshinone A	50%	39%	Isosalvianolic acid B	33%	33%
Salvianolic acid E	100%	83%	Trijukanone B	75%	39%	Salvianolic acid L	42%	33%
Rosmarinic acid	75%	72%	Cryptotanshinone	92%	33%	Salvianolic acid A	33%	39%
Luteolin	75%	50%	Tanshinone I	83%	44%	Tanshindiol C	42%	44%
Lithospermic acid	100%	100%	Vanillic acid	42%	50%	Royleanone-4	42%	44%
Prolithospermic acid derivative	75%	72%	Ferulic acid	25%	50%	1-Ketoisocryptotanshinone	42%	28%
Salvianolic acid B	100%	100%	7 α -Hydroxyallyl-royleanone	25%	78%	Neocryptotanshinone	33%	22%
Tanshinone IIB	92%	61%	Trijukanone A	17%	39%	Miltipolone	42%	39%
β -Sitosterol	67%	50%	Miltirone	17%	33%			
Tanshinonealdehyde	75%	50%						
15,16-Dihydrotanshinone I	100%	78%						
Methyltanshinonate	100%	78%						
1,2-Dihydrotanshinone I	75%	72%						
Tanshinone IIA	92%	56%						

^a Multiple testing was corrected by Bonferroni correction. Bonferroni-corrected p value for significance: 7.57×10^{-4} (0.05/66). ^a Paired locations (12) were ZY vs. CQ, ZY vs. TA, CQ vs. TA in four genotypes. ^b Paired genotypes (18) were Genotype 1 vs. Genotype 2, Genotype 1 vs. Genotype 3, Genotype 1 vs. Genotype 4, Genotype 2 vs. Genotype 3, Genotype 2 vs. Genotype 4, Genotype 3 vs. Genotype 4 at three locations. ^c The locations-specific and genotypes-specific metabolites were labeled by bold fonts.

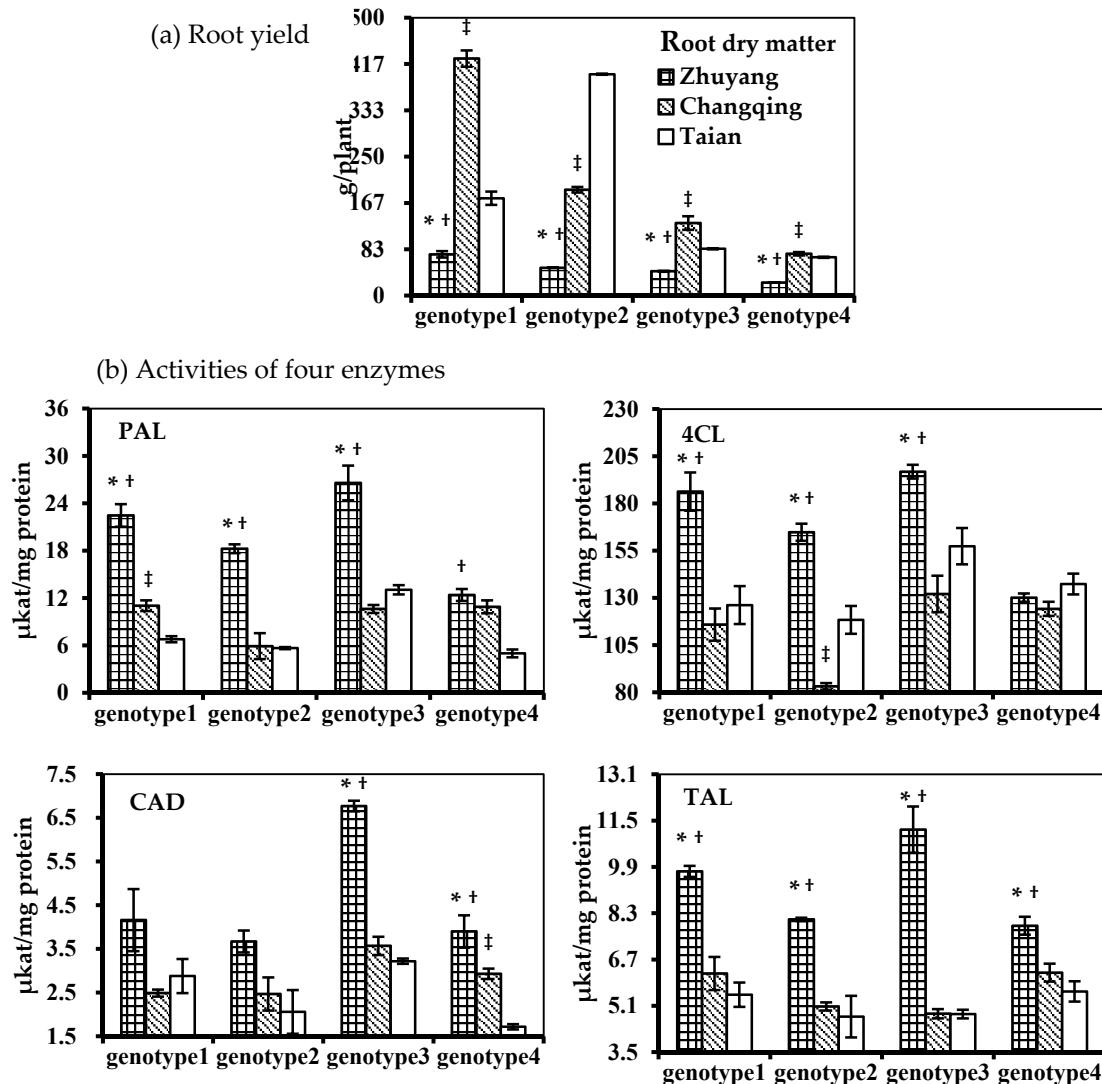


Figure S1. Root yield (a), and activities of four enzymes (b) in *S. miltiorrhiza* in different locations. Four enzymes include PAL, 4CL, CAD, TAL. * Significant difference according to Bonferroni correction ($p < 0.00033$) Zhuyang vs. Changqng; † Significant difference according to Bonferroni correction ($p < 0.00033$) Zhuyang vs. Taian; ‡ Significant difference according to Bonferroni correction ($p < 0.00033$) Changqing vs. Taian; PAL, phenylalanine ammonia-lyase; 4CL, 4-coumarate-CoA ligase; CAD, cinnamyl alcohol-NADPH dehydrogenase; TAL, tyrosine ammonic-lyase.

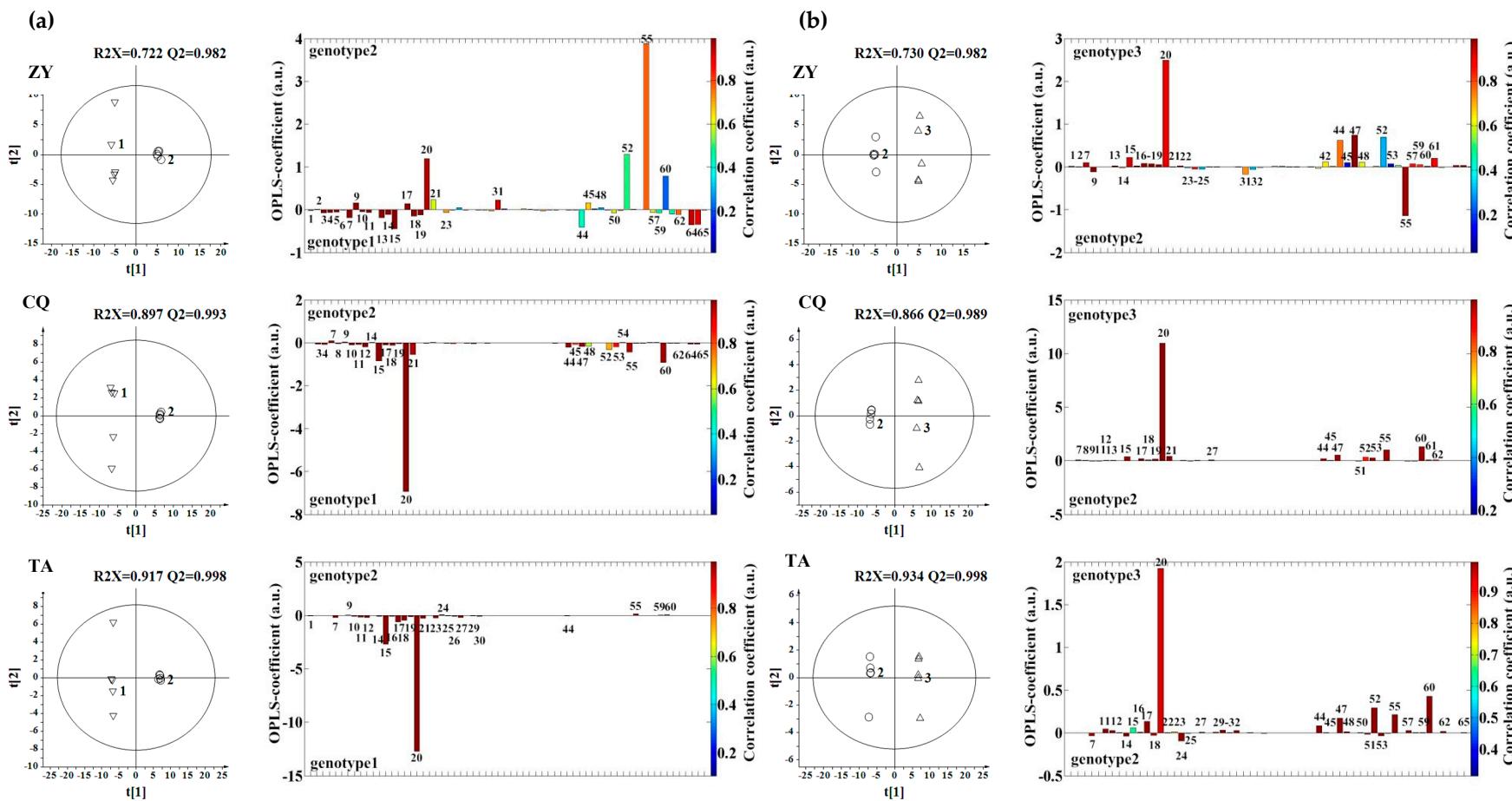


Figure S2. Cont.

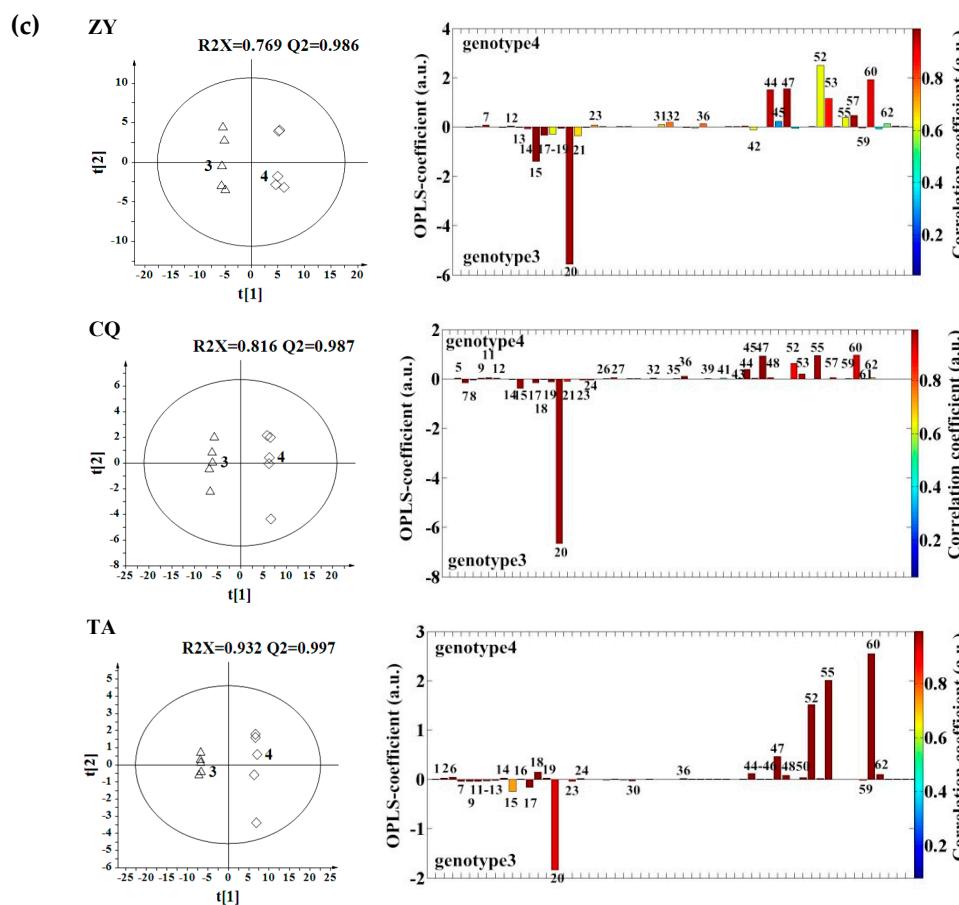


Figure S2. OPLS-DA scores plots (left) and coefficient-coded loadings plots (right) derived from LC-MS data for *S. miltiorrhiza* extracts obtained from four different genotypes (1, Genotype 1; 2, Genotype 2; 3, Genotype 3; 4, Genotype 4) for each location (ZY, Zhuyang; CQ, Changqing; TA, Taian); 2, Procatechuic acid; 7, Salvianolic acid F; 13, Salvianolic acid I; 14, Salvianolic acid E; 15, Rosmarinic acid; 17, Lithospermic acid; 18, Prolithospermic acid derivative; 20, Salvianolic acid B; 30, 7 α -hydroxyallyl-royleanone; 32, Tanshinone II B; 44, 15,16'-dihydrotanshinone I; 47, Methyltanshinonate; 55, 1,2'-dihydrotanshinone I; 60, Tanshinone II A. (a), *S. miltiorrhiza* extracts obtained from Genotype 1 vs. Genotype 2 for each location; (b), *S. miltiorrhiza* extracts obtained from Genotype 2 vs. Genotype 3 for each location; (c), *S. miltiorrhiza* extracts obtained from Genotype 3 vs. Genotype 4 for each location.

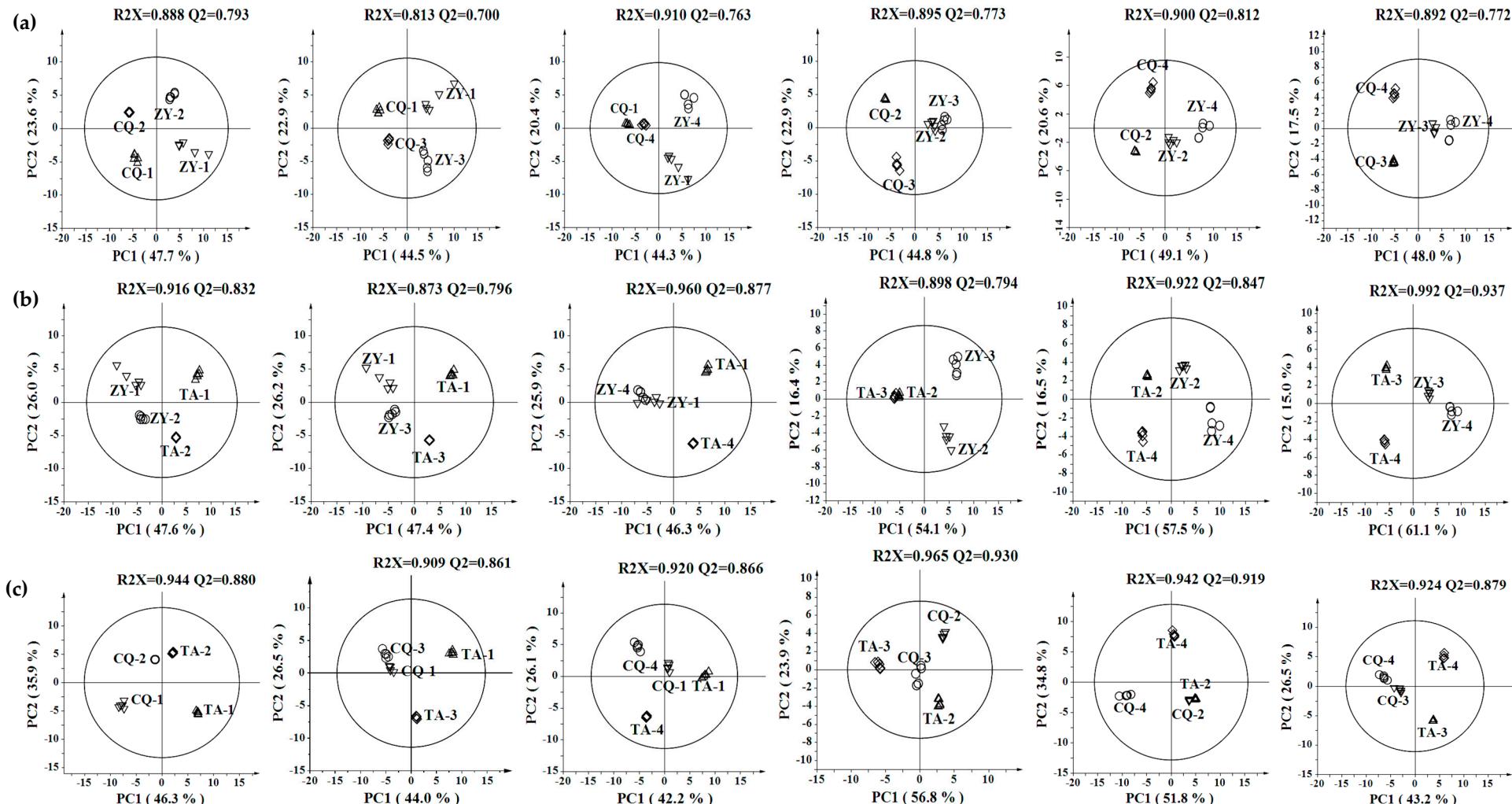


Figure S3. PCA scores plots derived from LC-MS data for *S. miltiorrhiza* extracts obtained from paired locations and genotypes. (ZY, Zhuyang; CQ, Changqing; TA, Taian. 1, Genotype 1; 2, Genotype 2; 3, Genotype 3; 4, Genotype 4). (a), *S. miltiorrhiza* extracts obtained from ZY vs. CQ; (b), *S. miltiorrhiza* extracts obtained from ZY vs. TA; (c), *S. miltiorrhiza* extracts obtained from CQ vs. TA.



Figure S4. Plants and roots of four genotypes (a, Genotype 1; b, Genotype 2; c, Genotype 3; d, Genotype 4).

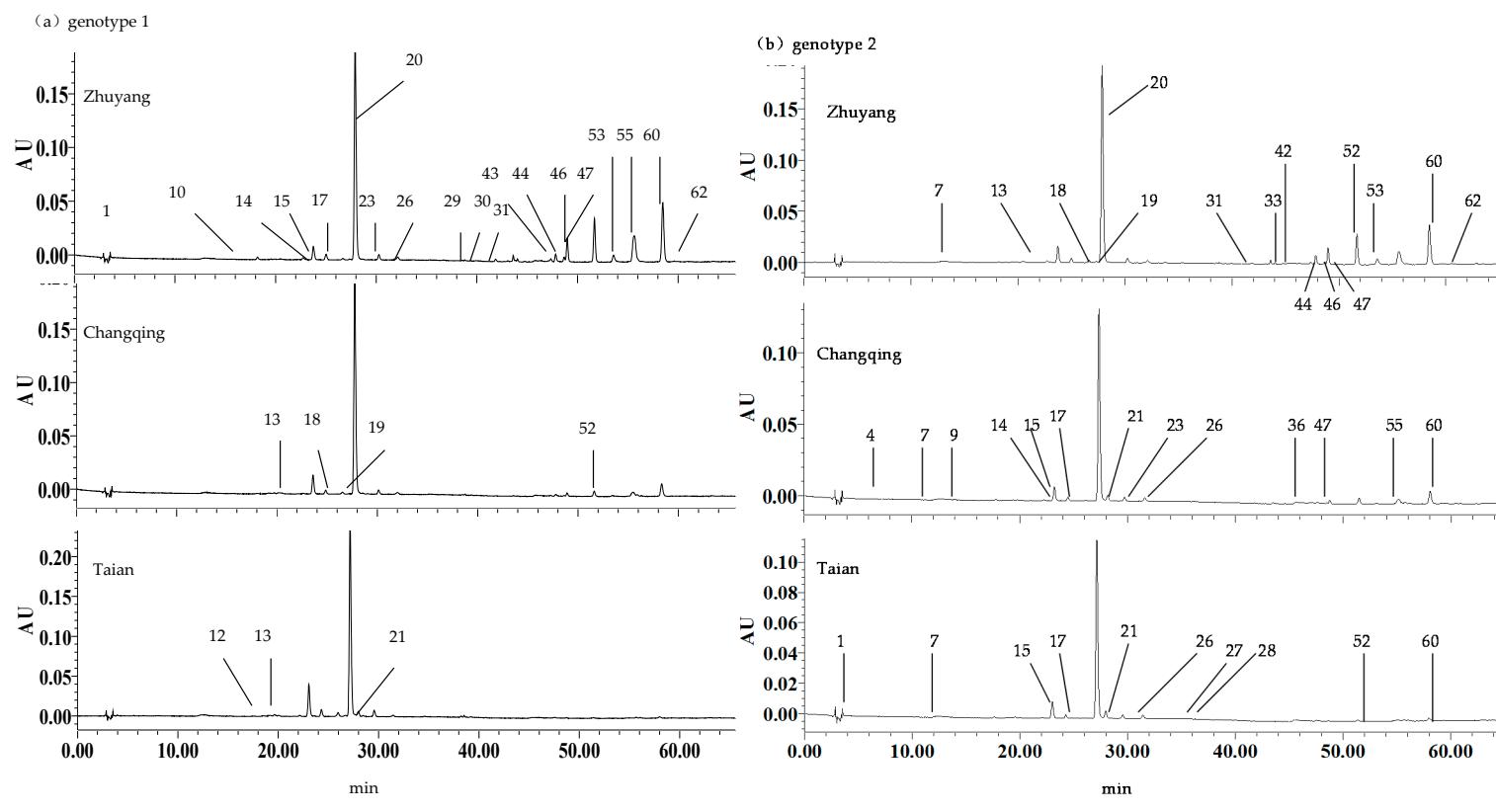


Figure S5. Cont.

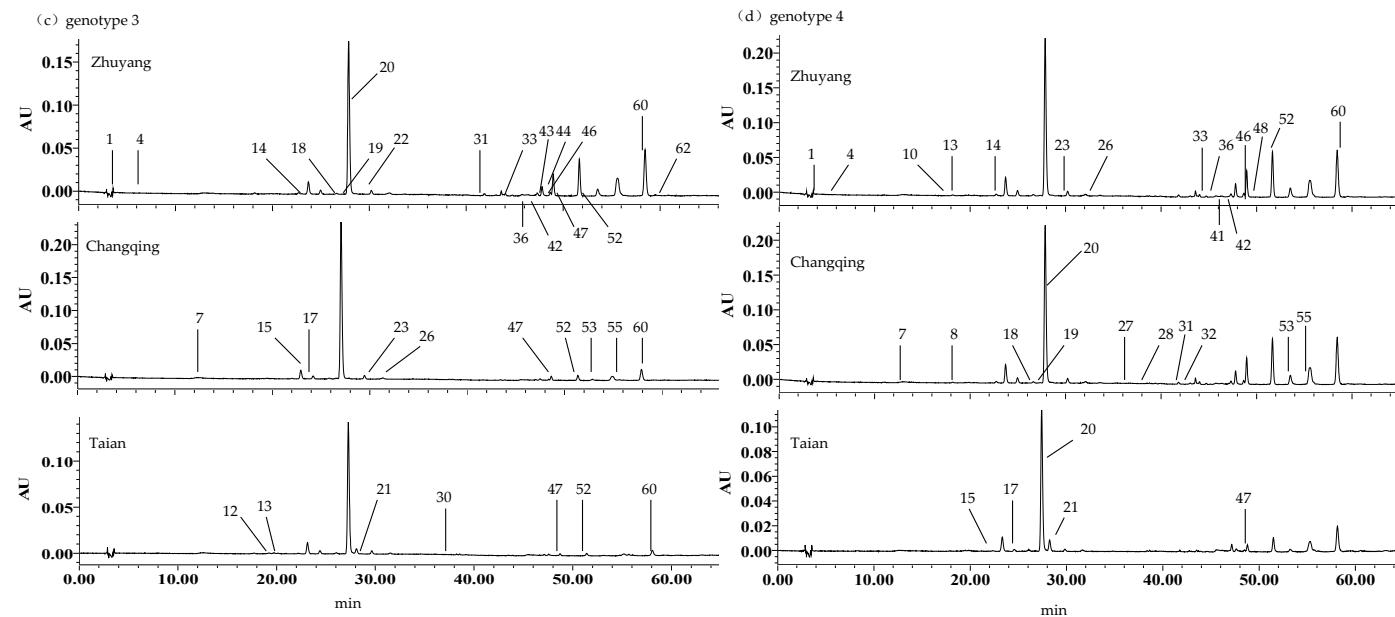
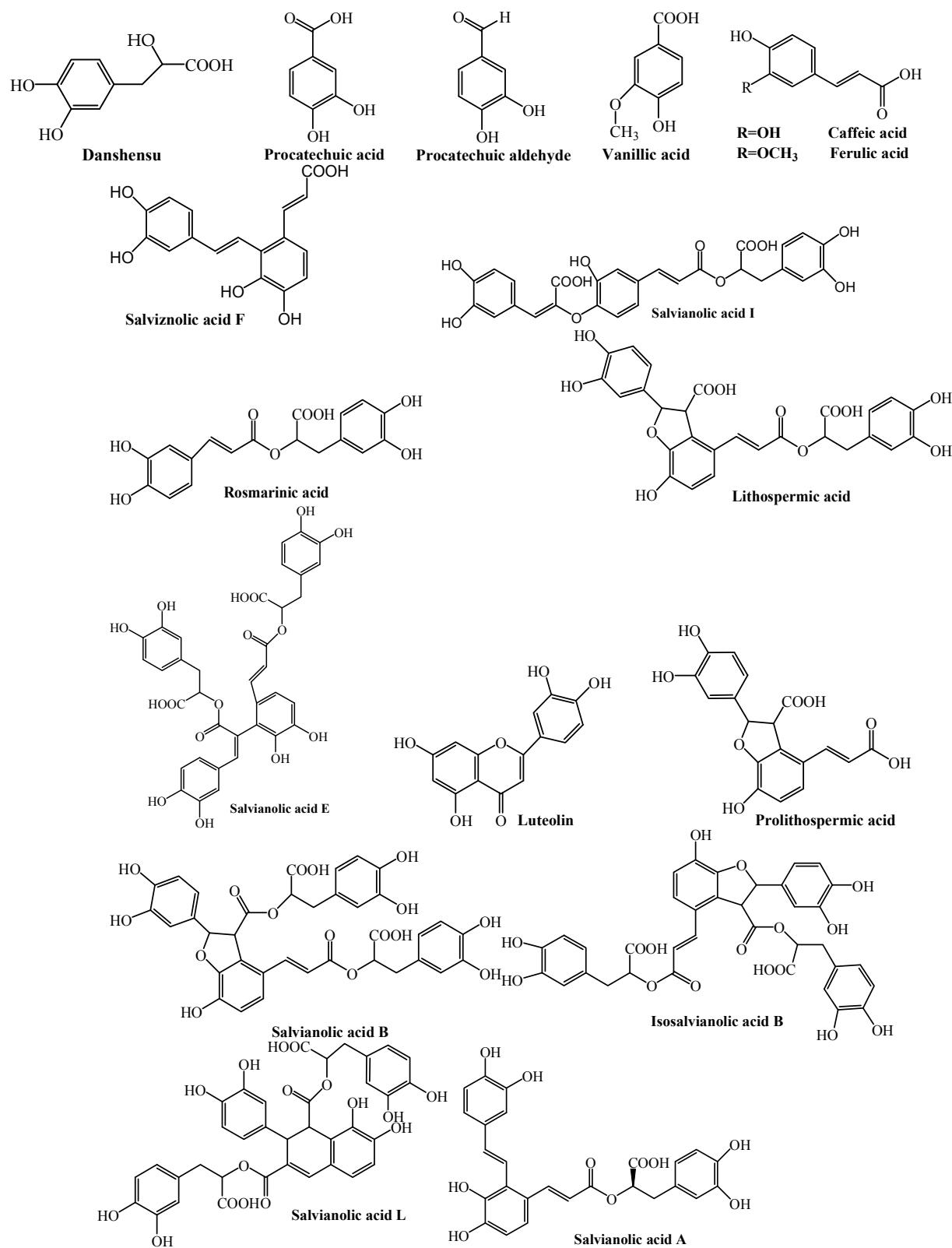


Figure S5. LC-UV chromatograms (280nm) for *S. miltiorrhiza* from three different locations (Zhuyang, Changqing, Taian) for each genotype: 1, Danshensu; 4, Procatechuic aldehyde; 7, Salvianolic acid F; 8, U2; 9, U3; 10, Ferulic acid; 12, U5; 13, Salvianolic acid I; 14, Salvianolic acid E; 15, Rosmarinic acid; 17, Lithospermic acid; 18, Prolithospermic acid derivative; 19, U6; 20, Salvianolic acid B; 21, Isosalvianolic acid B; 22, U7; 23, Salvianolic acid L; 24, Salvianolic acid A; 26, Tanshindiol C; 27, U9; 28, U10; 29, Royleanone-4; 30, 7 α -hydroxyallyl-royleanone; 31, U11; 32, Tanshinone II B; 33, 1-ketoisocryptotanshinone; 36, Trijukanone C; 41, U14; 42, Przewa tanshinone A; 43, U15; 44, 15,16'-dihydrotanshinone I; 46, Neocryptotanshinone; 47, Methyltanshinonate; 48, Trijukanone B; 52, Cryptotanshinone; 53, Tanshinone I; 55, 1,2'-dihydrotanshinone I; 60, Tanshinone II A; 62, Miltirone. (a), LC-UV chromatograms from three different locations for Genotype 1; (b), LC-UV chromatograms from three different locations for Genotype 2; (c), LC-UV chromatograms from three different locations for Genotype 3; (d), LC-UV chromatograms from three different locations for Genotype 4.

**Figure S6. Cont.**

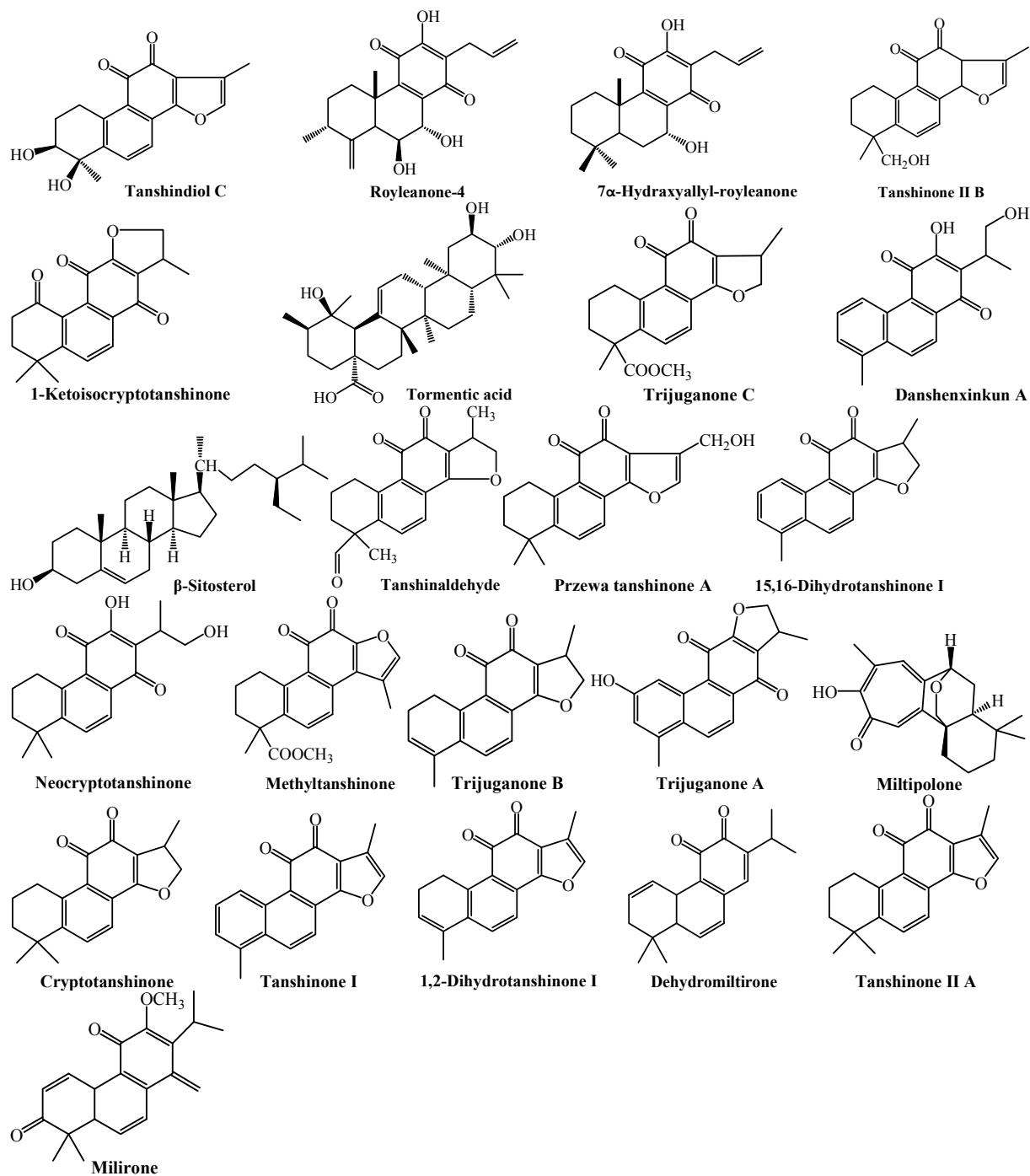


Figure S6. The structures of some secondary metabolites in *S. miltiorrhiza*.

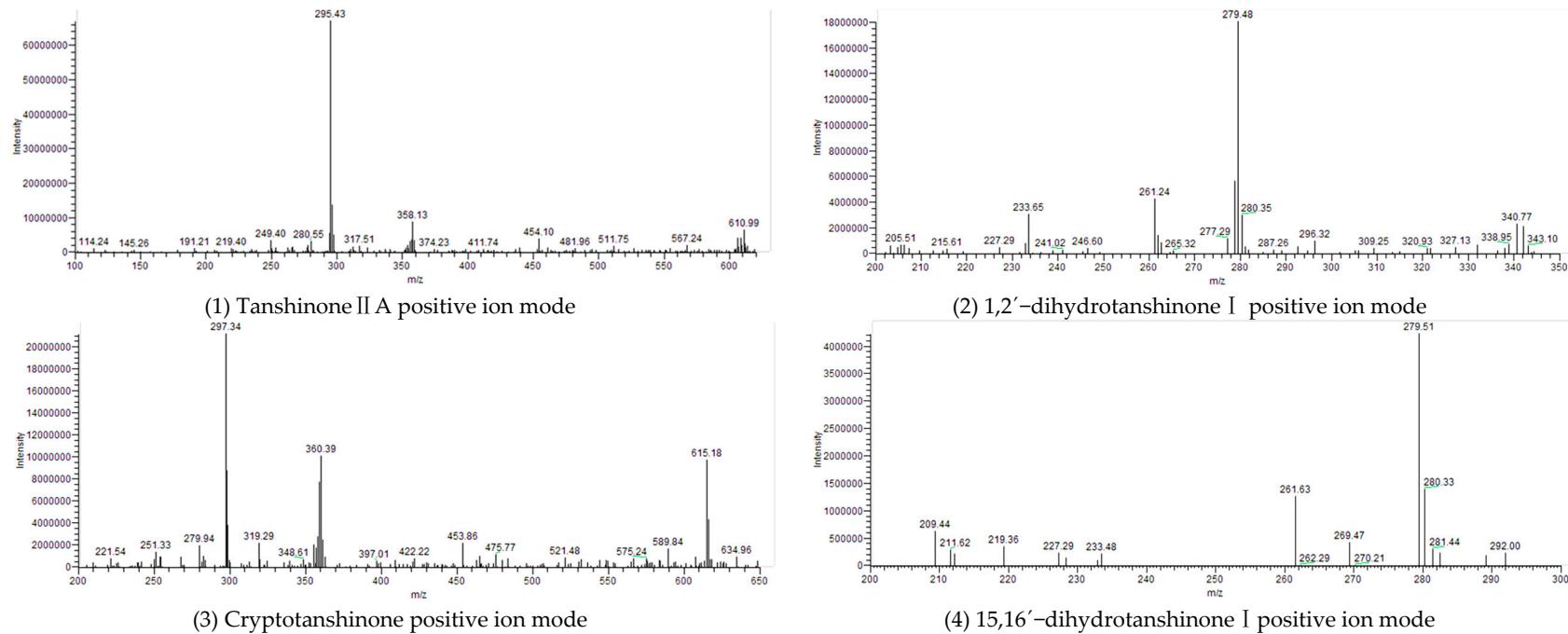


Figure S7. Cont.

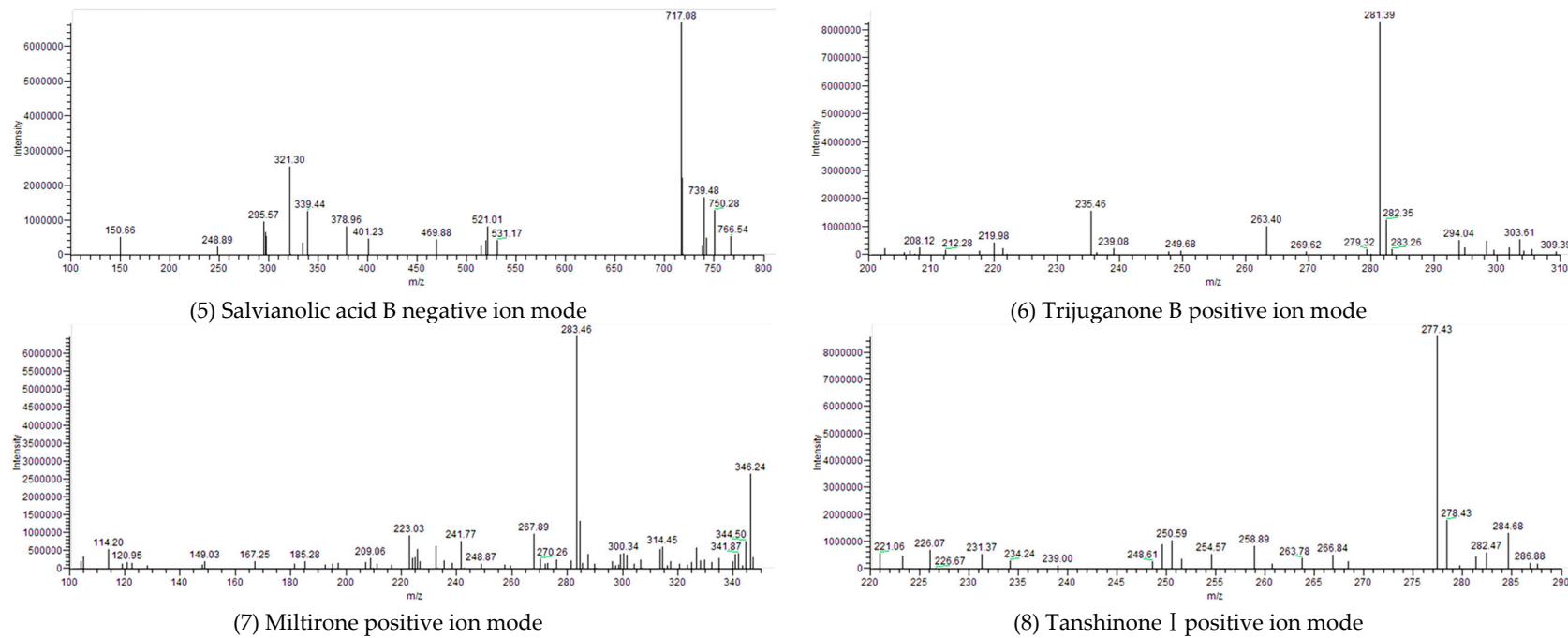


Figure S7. Some spectra of *S. miltiorrhiza* compounds in our studies: (1), tanshinone IIA; (2), 1,2'-dihydrotanshinone I; (3), cryptotanshinone; (4), 15,16'-dihydrotanshinone I; (5), salvianolic acid B; (6), trijukanone B; (7), miltirone; (8), tanshinone I.