

# Supplementary Materials: Chemical Constituents from the Roots and Rhizomes of *Asarum heterotropoides* var. *mandshuricum* and the In Vitro Anti-Inflammatory Activity

Yu Jing, Yi-Fan Zhang, Ming-Ying Shang, Guang-Xue Liu, Yao-Li Li, Xuan Wang and Shao-Qing Cai

## 1. Spectra of compounds 1–11

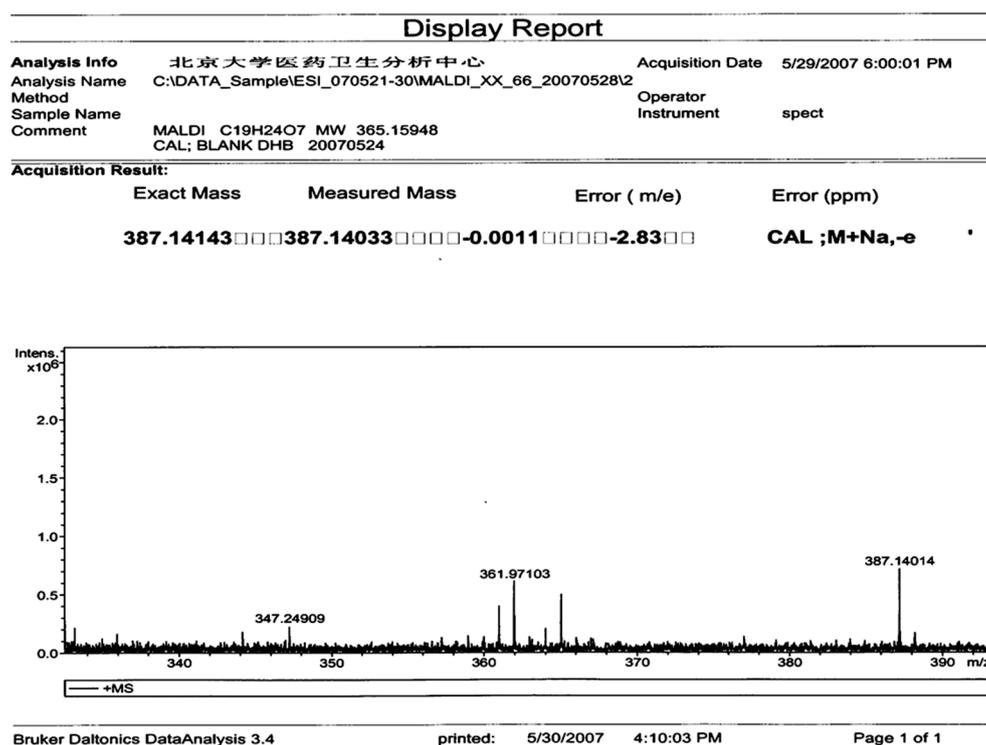
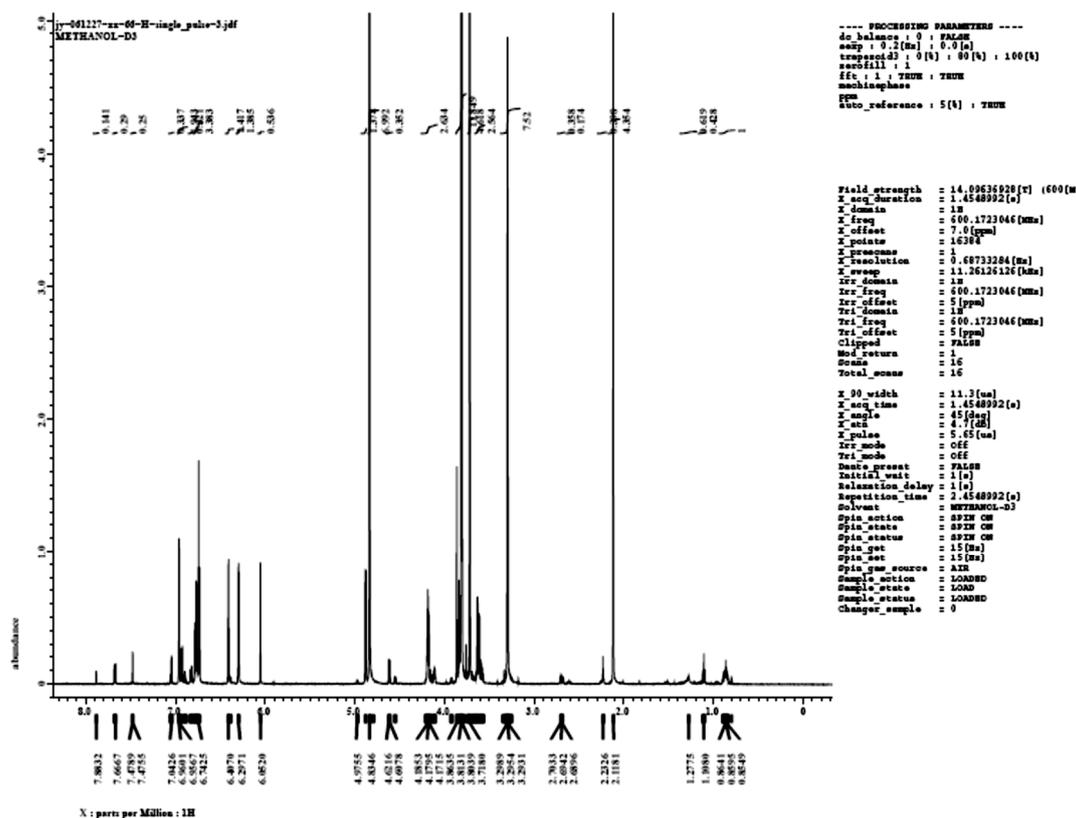
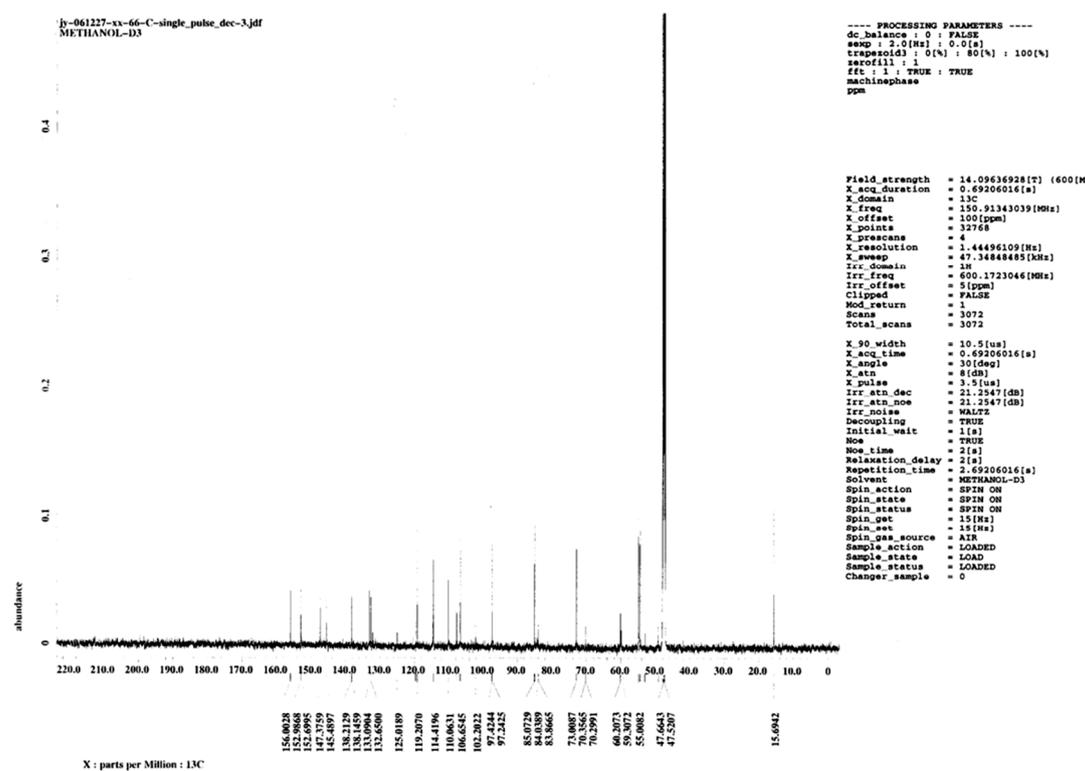
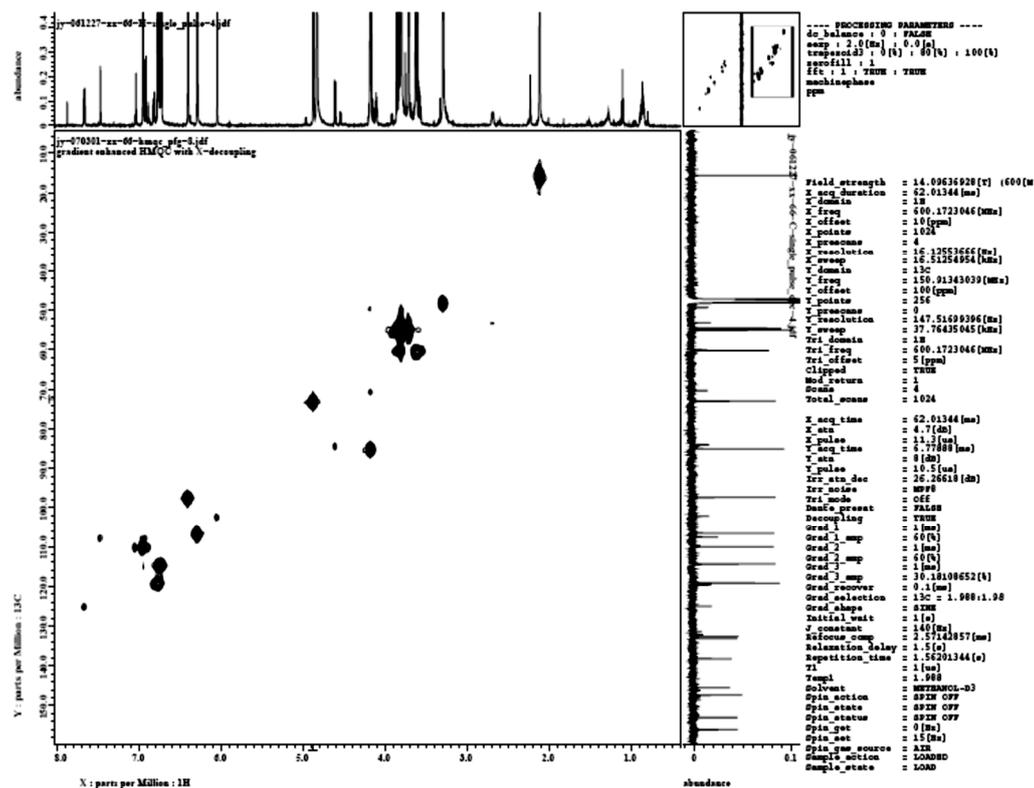
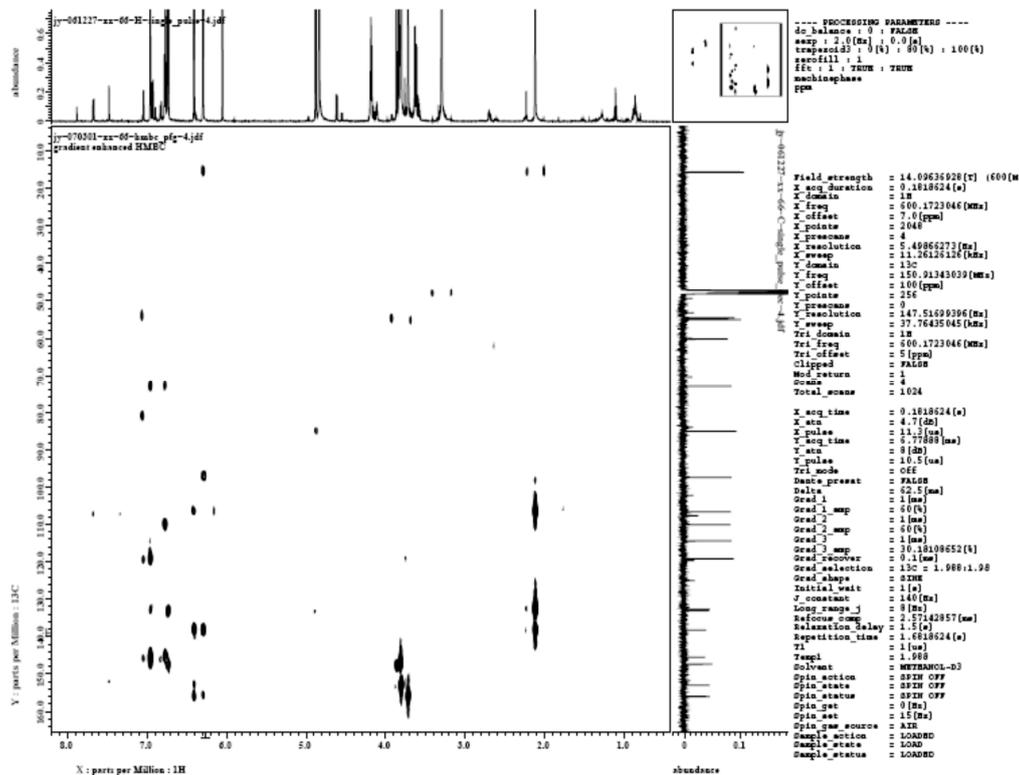


Figure S1. HRMALDIMS spectrum of compound 1.

Figure S2. <sup>1</sup>H-NMR spectrum of compound 1 in CD<sub>3</sub>OD.Figure S3. <sup>13</sup>C-NMR spectrum of compound 1 in CD<sub>3</sub>OD.

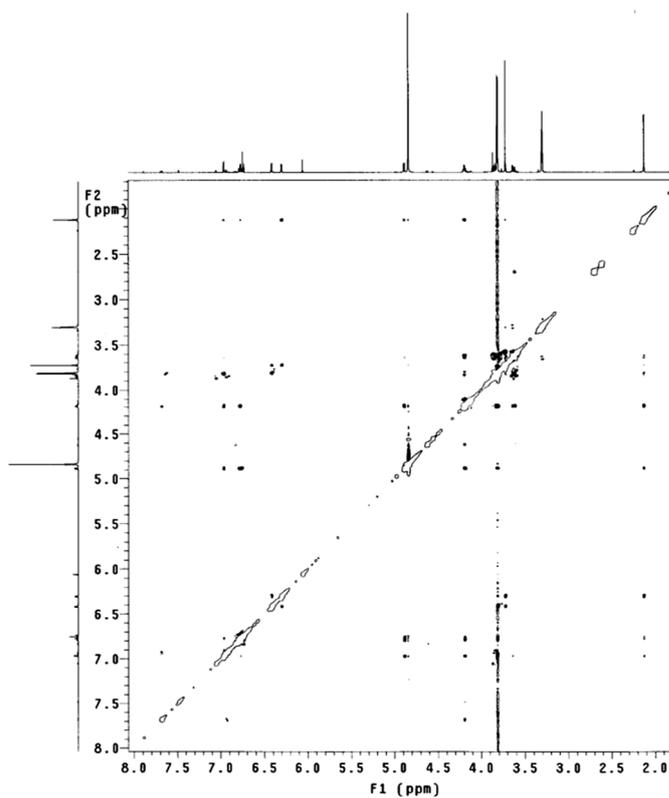


Figure S6. HMQC-NMR spectrum of compound 1 in CD<sub>3</sub>OD.Figure S7. HMBC-NMR spectrum of compound 1 in CD<sub>3</sub>OD.

XX-66

Pulse Sequence: NOESY  
 Solvent: cd3od  
 Temp: 25.0 C / 298.1 K  
 File: jingyu-XX-66-noesy  
 INOVA-500 "gNUS00"

Relax. delay 1.000 sec  
 Mixing 0.600 sec  
 Acq. time 0.228 sec  
 Width 4485.9 Hz  
 2D Width 4485.9 Hz  
 48 repetitions  
 2 x 256 increments  
 OBSERVE H1, 499.9038718 MHz  
 DATA PROCESSING  
 Gauss apodization 0.105 sec  
 F1 DATA PROCESSING  
 Gauss apodization 0.030 sec  
 FT size 4096 x 4096  
 Total time 12 hr, 45 min, 3 sec

Figure S8. NOESY NMR spectrum of compound 1 in CD<sub>3</sub>OD.

## Display Report

<b>Analysis Info</b>	北京大学医药卫生分析中心	Acquisition Date	5/29/2007 7:22:21 PM
Analysis Name	C:\DATA_Sample\ESI_070521-30\MALDI_XX_89_20070528\3	Operator	
Method		Instrument	spect
Sample Name			
Comment	MALDI C20H24O6 MW 360.15729 CAL; DHB 20070524		

## Acquisition Result:

Exact Mass	Measured Mass	Error ( m/e)	Error (ppm)
383.14651	383.14576	-0.0008	-1.96

CAL ; M+Na, e

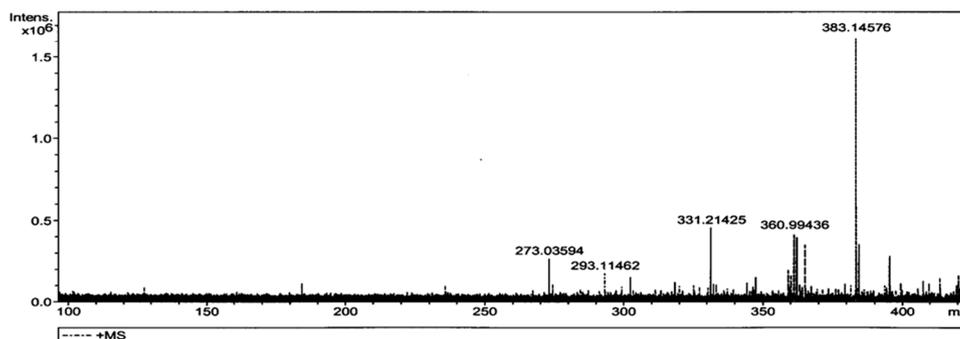
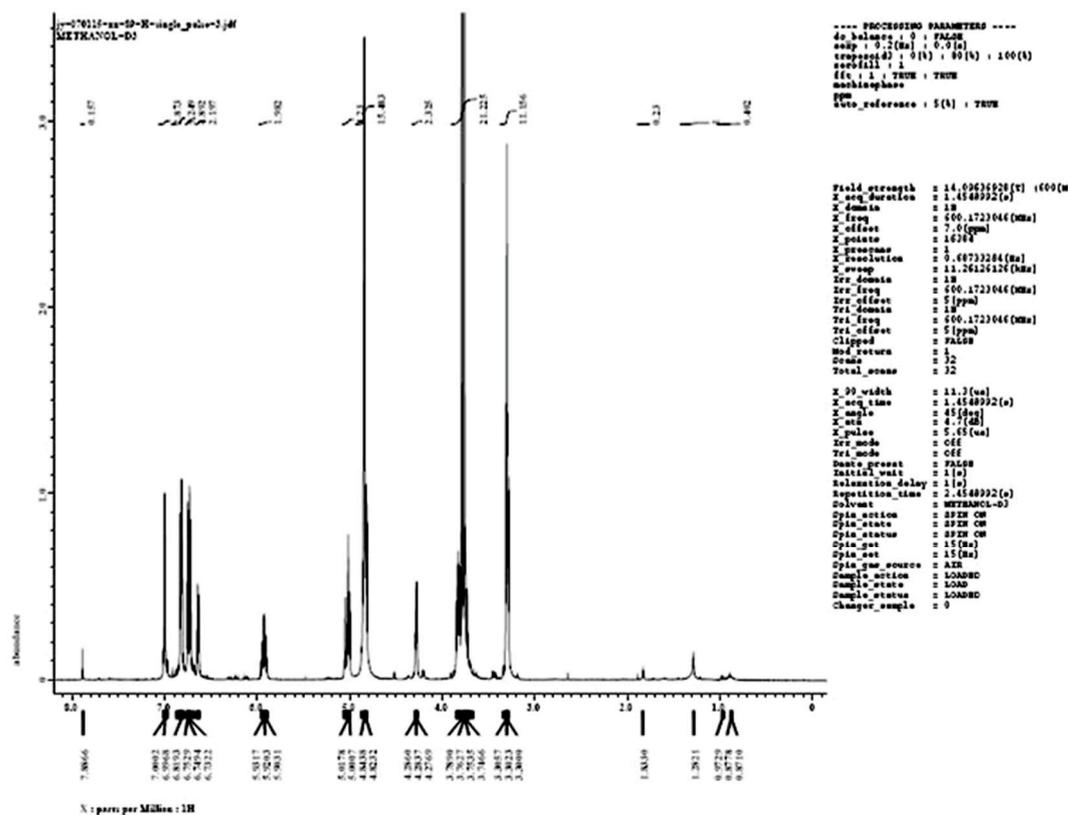
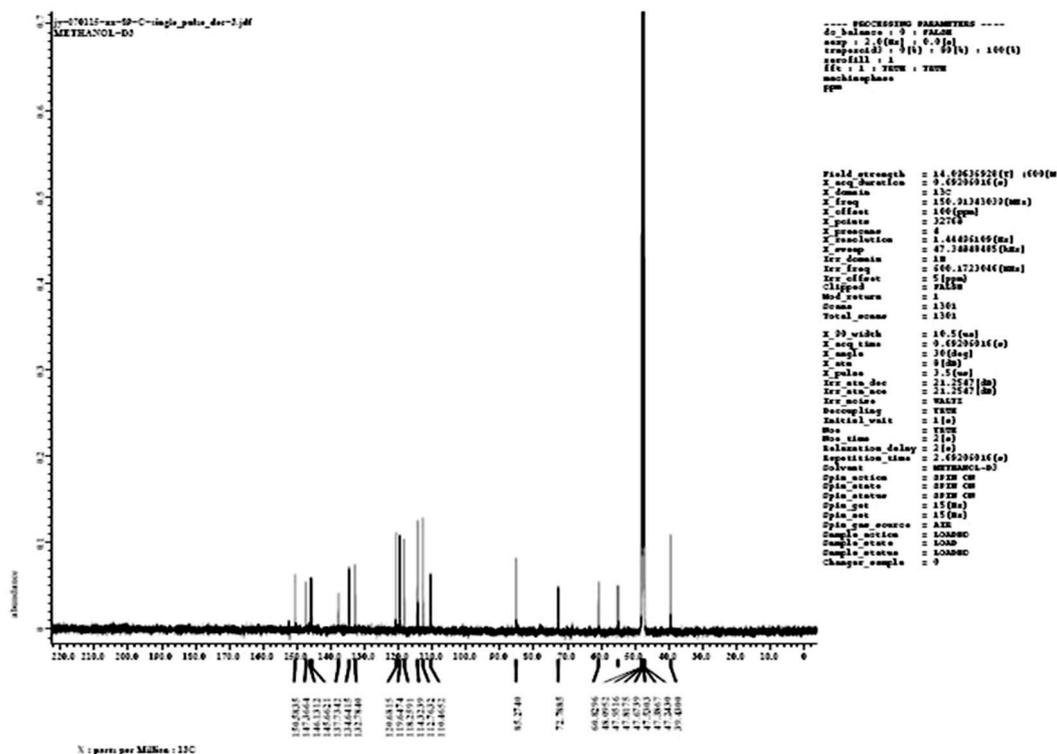
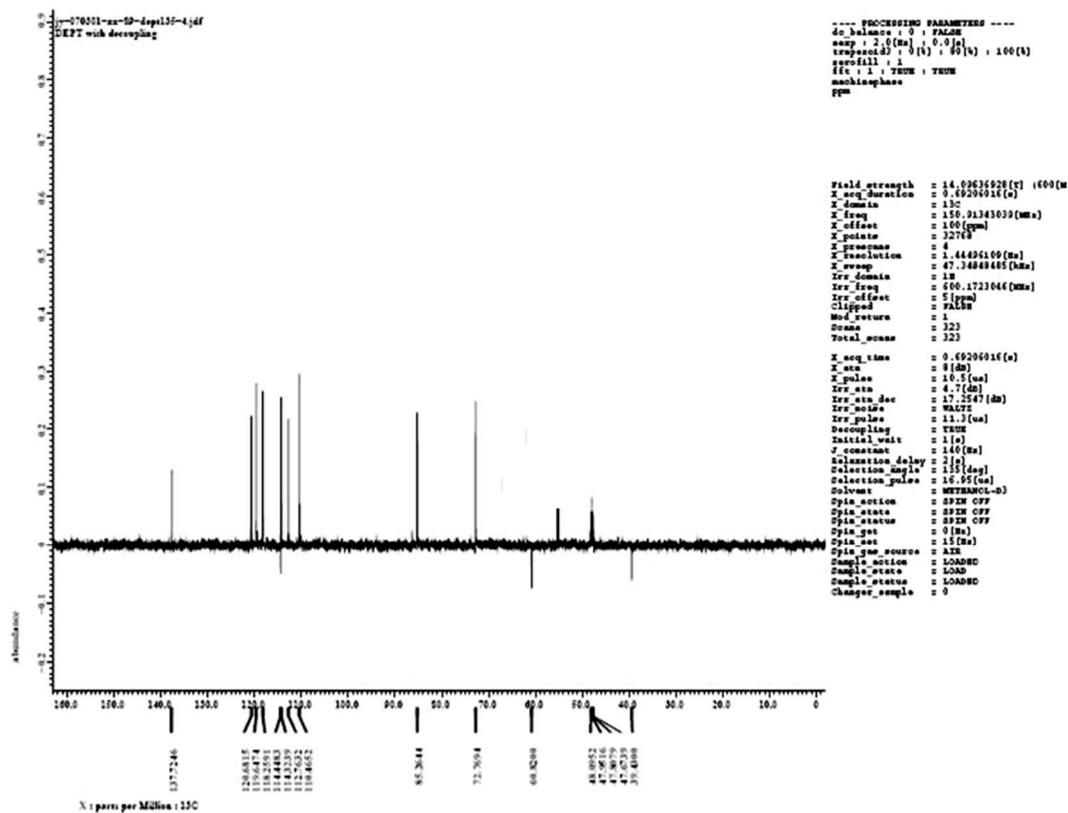
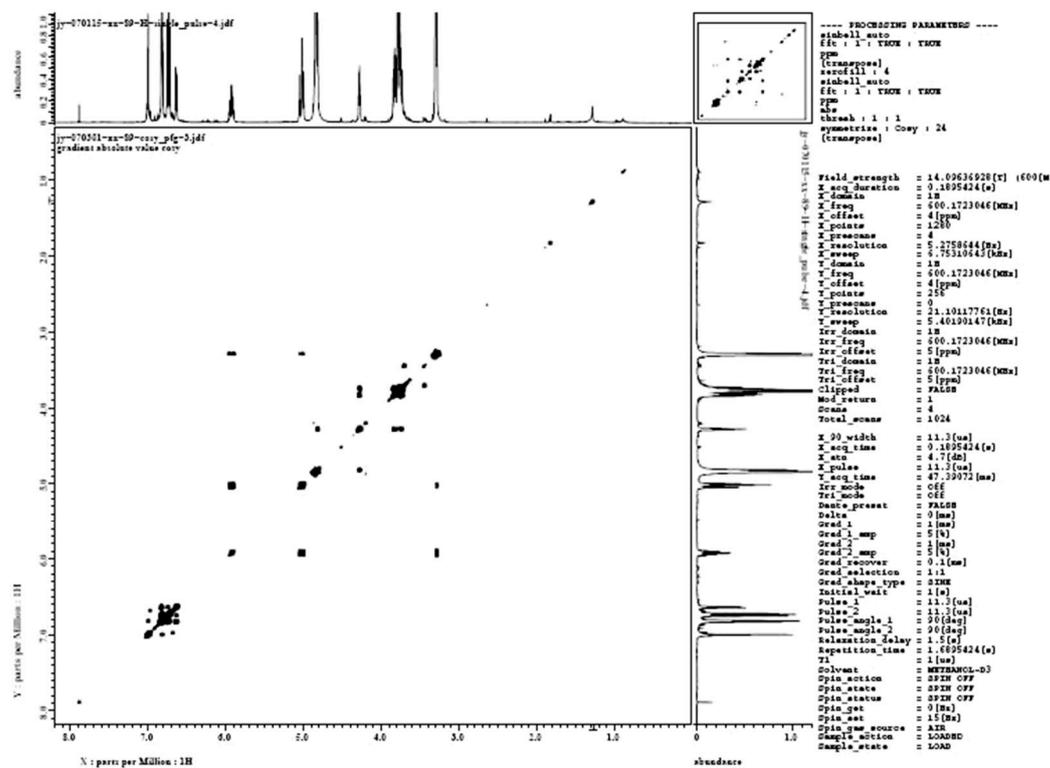
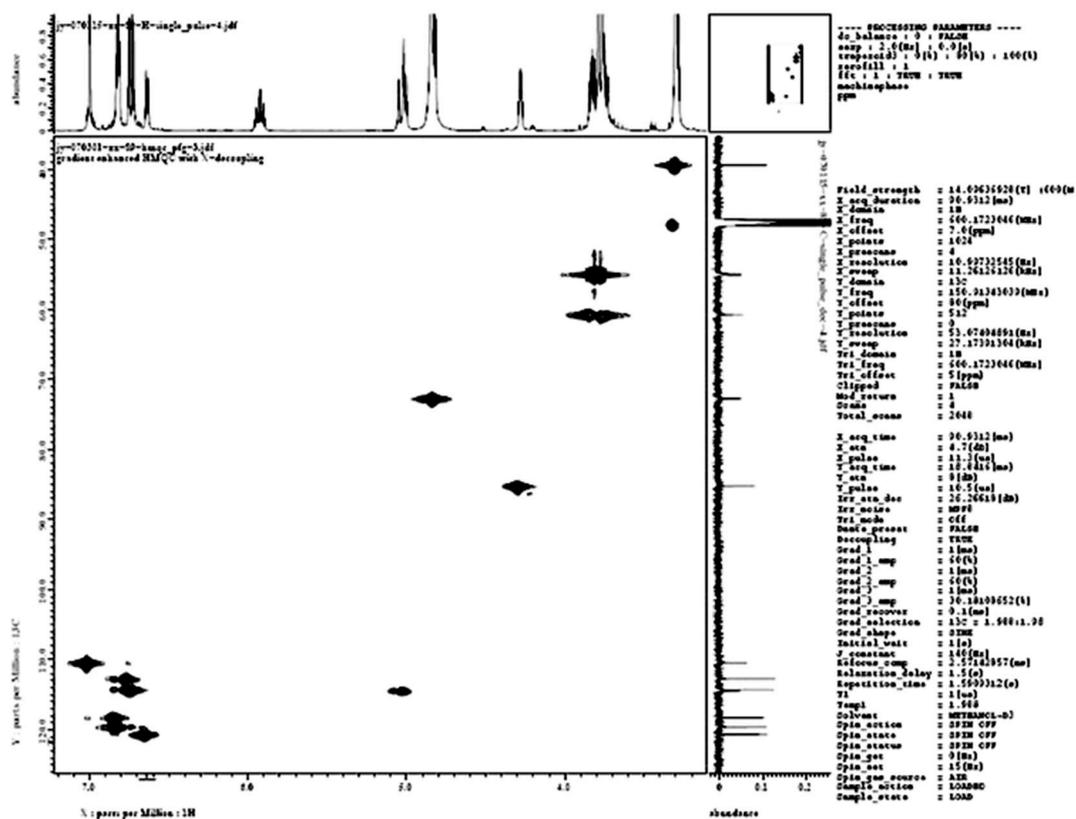
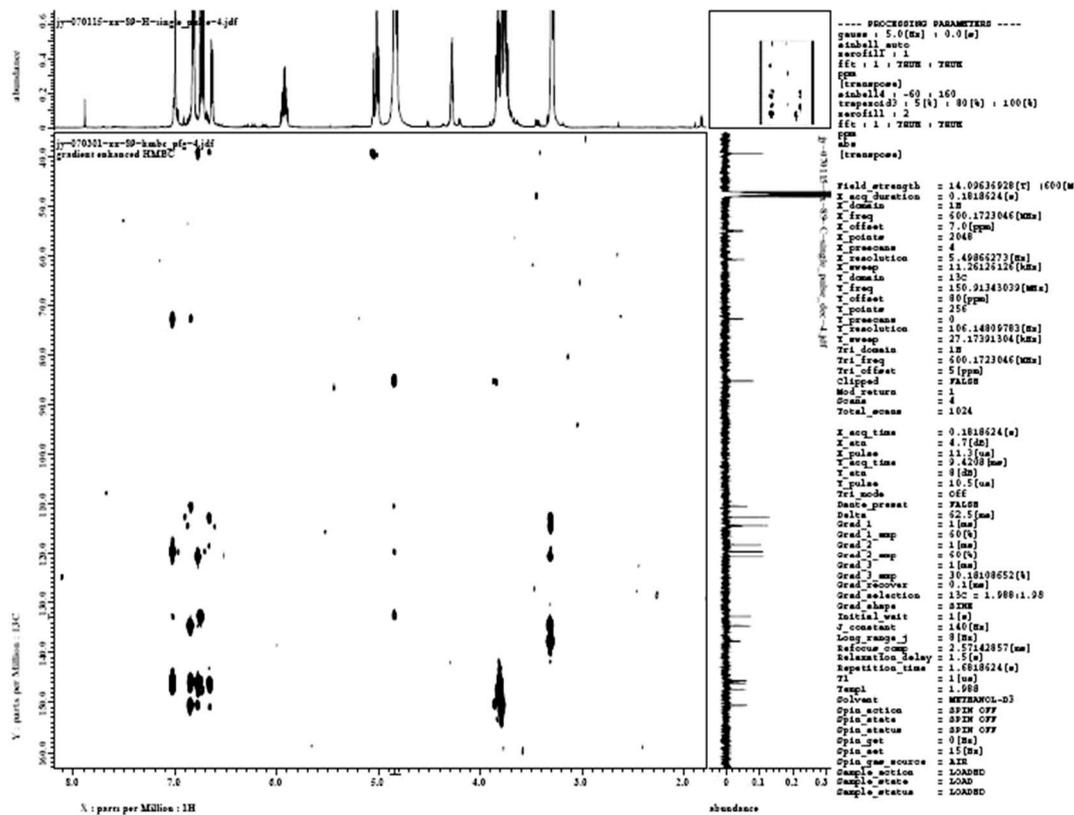
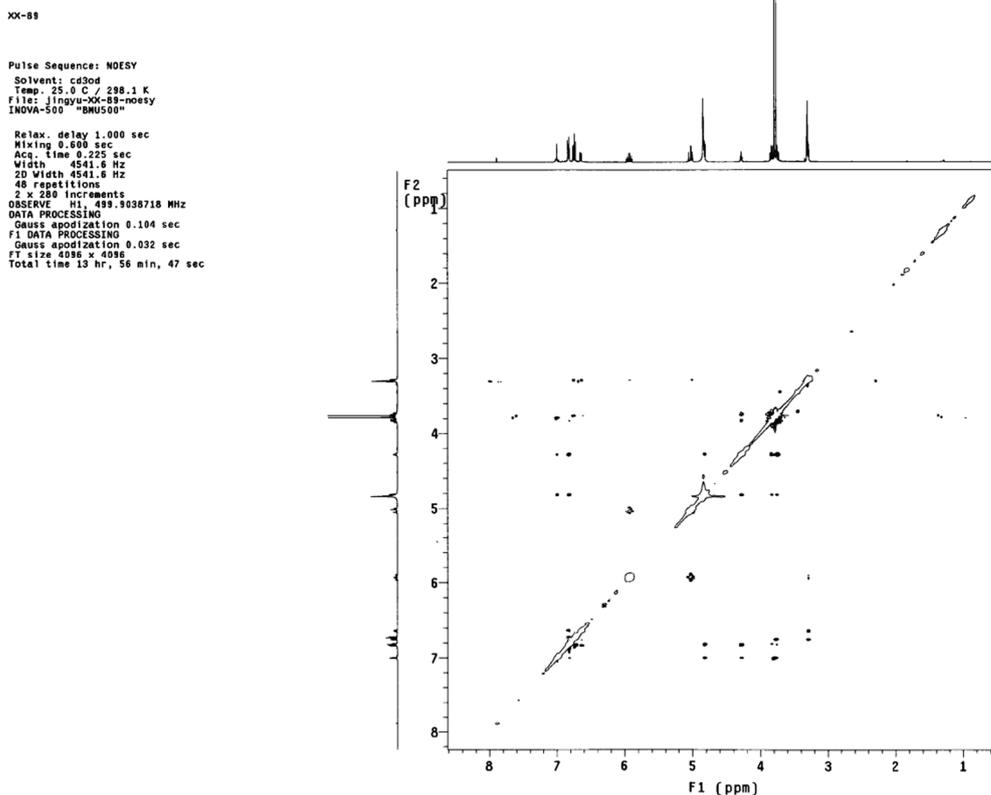


Figure S9. HRMALDIMS spectrum of compound 2.

Figure S10. <sup>1</sup>H-NMR spectrum of compound 2 in CD<sub>3</sub>OD.Figure S11. <sup>13</sup>C-NMR spectrum of compound 2 in CD<sub>3</sub>OD.

Figure S12 DEPT NMR spectrum of compound 2 in CD<sub>3</sub>OD.Figure S13. <sup>1</sup>H-<sup>1</sup>H COSY NMR spectrum of compound 2 in CD<sub>3</sub>OD.

Figure S14. HMQC-NMR spectrum of compound 2 in CD<sub>3</sub>OD.Figure S15. HMBC-NMR spectrum of compound 2 in CD<sub>3</sub>OD.

Figure S16. NOESY NMR spectrum of compound 2 in CD<sub>3</sub>OD.

## Display Report

<b>Analysis Info</b>	北京大学医药卫生分析中心	Acquisition Date	7/17/2007 4:37:11 PM
Analysis Name	C:\DATA_Sample\ESI_070711-21\ESI_XX_902	Operator	bpfxsh@bjmu.edu.cn
Method		Instrument	FT_MS_Bruker APEX IV (7.0 T)
Sample Name			
Comment	ESI C20H24O6 MW 360.15727 M/Z ( 509 ) = 186.08732;286.10872::391.28435;441.1669; 509.2547; 611.2613;667.2409;804.48926;826.47121		

## Acquisition Result:

Exact Mass	Measured Mass	Error ( m/e)	Error (ppm)	
383.14651	383.14552	-0.0010	-2.59	M+Na, <sup>-</sup> e
391.28435	391.28363	-0.0007	-1.84	REF ACN

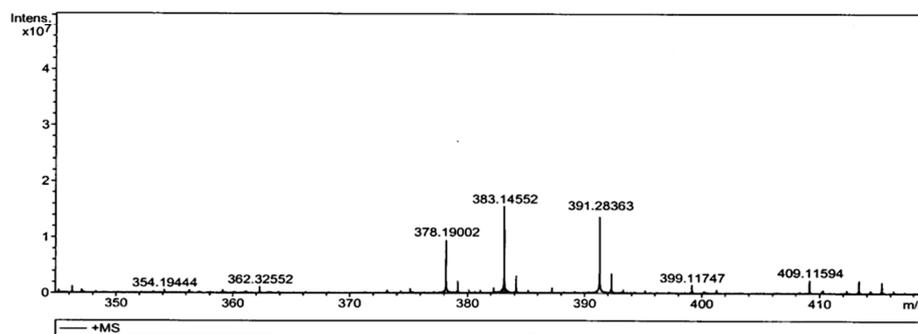


Figure S17. HRESIMS spectrum of compound 3.







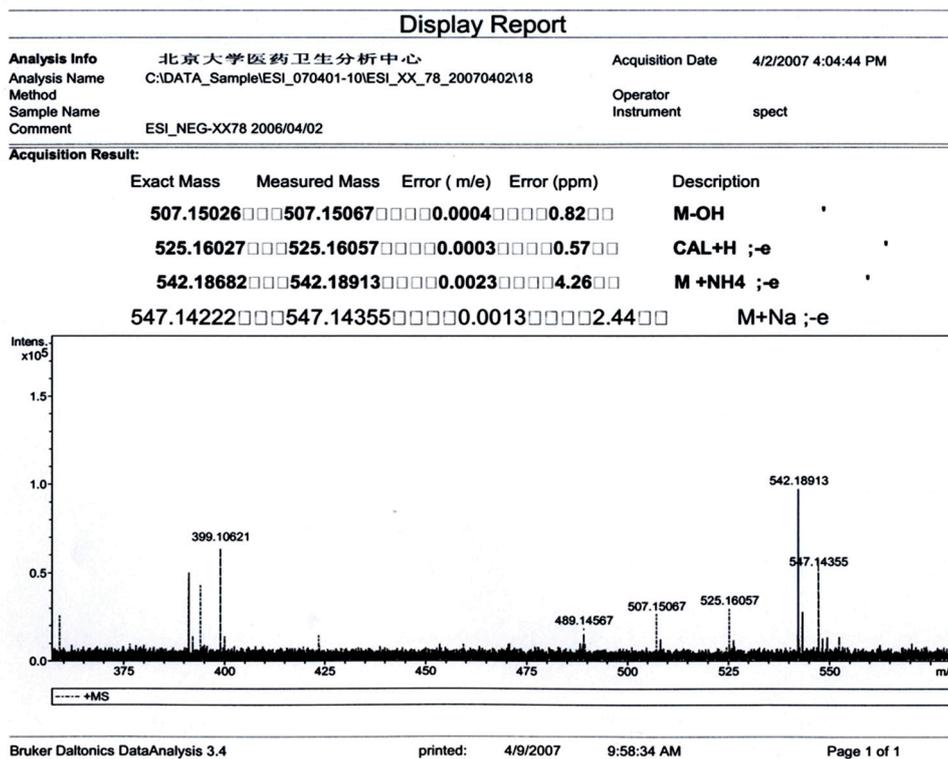
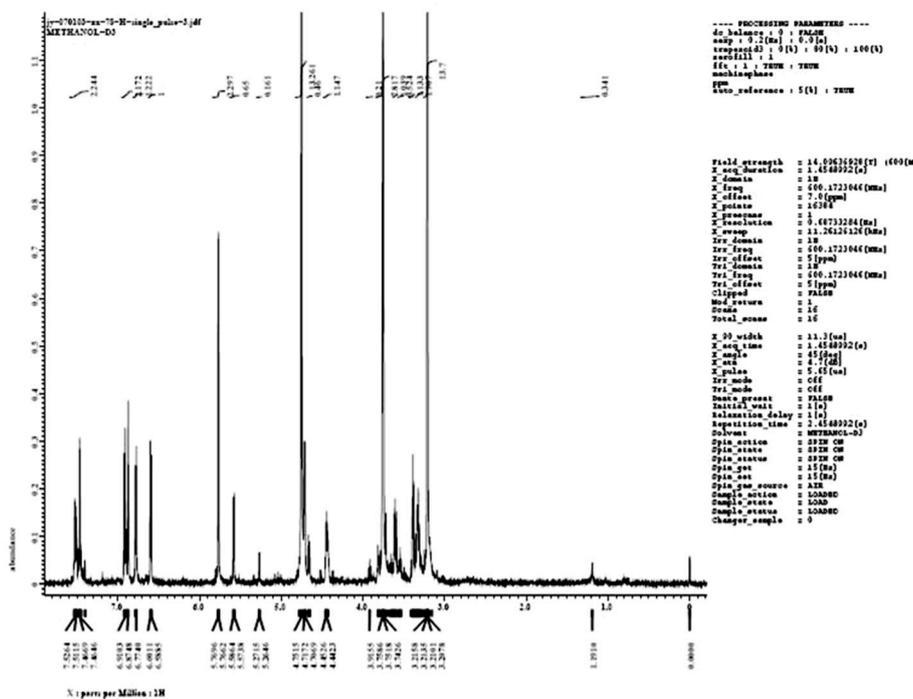


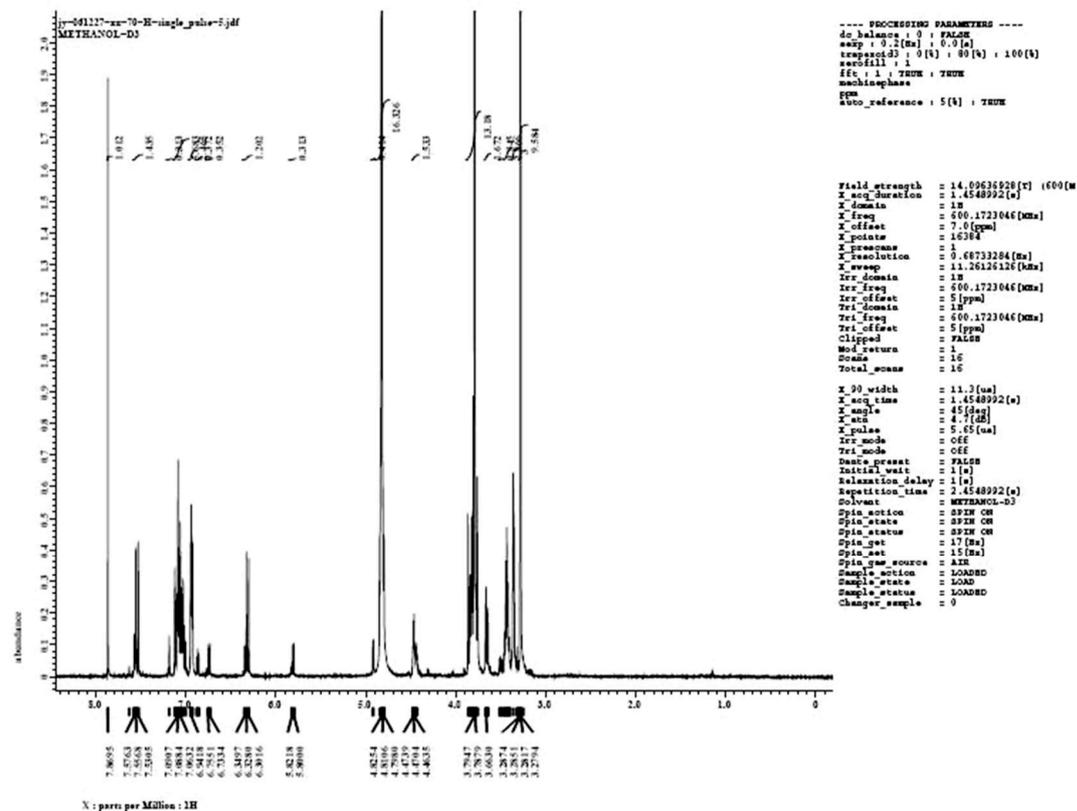
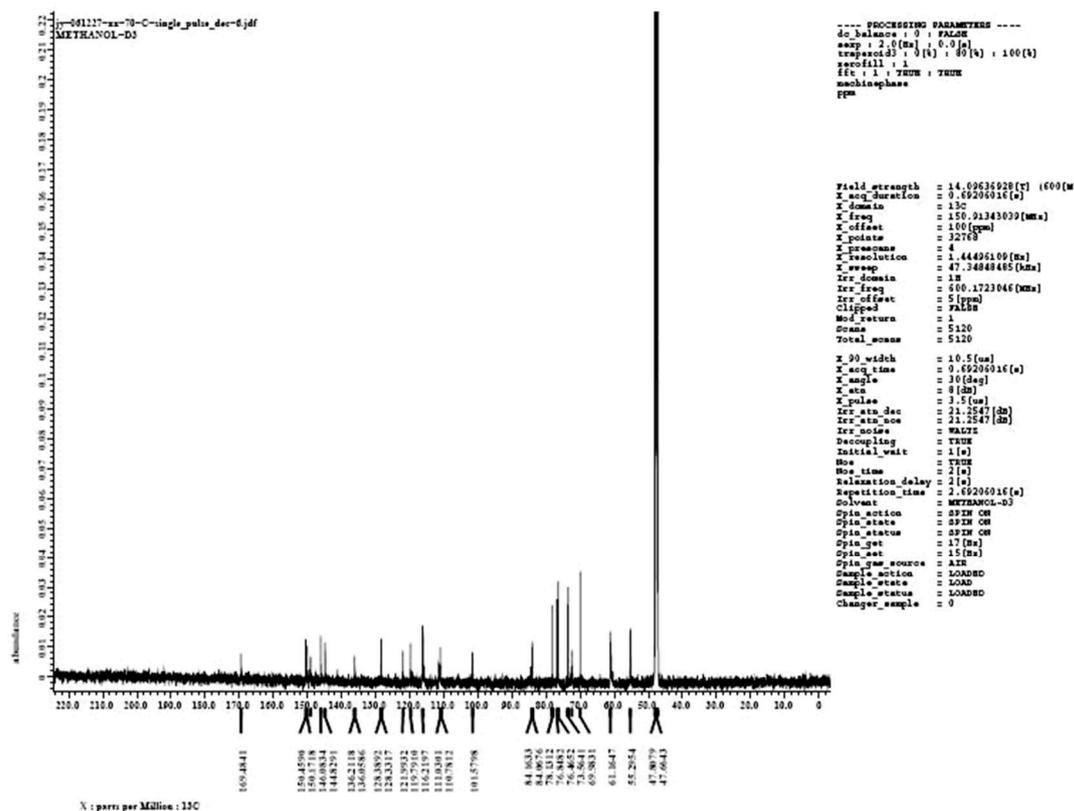
Figure S24. HRESIMS spectrum of compound 4.

Figure S25. <sup>1</sup>H-NMR spectrum of compound 4 in CD<sub>3</sub>OD.

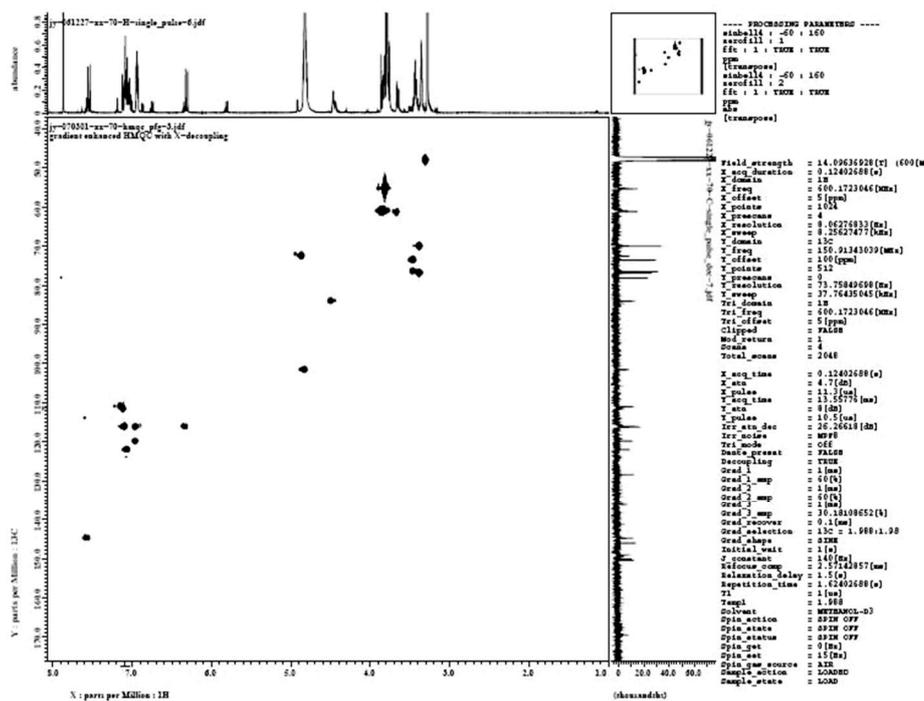
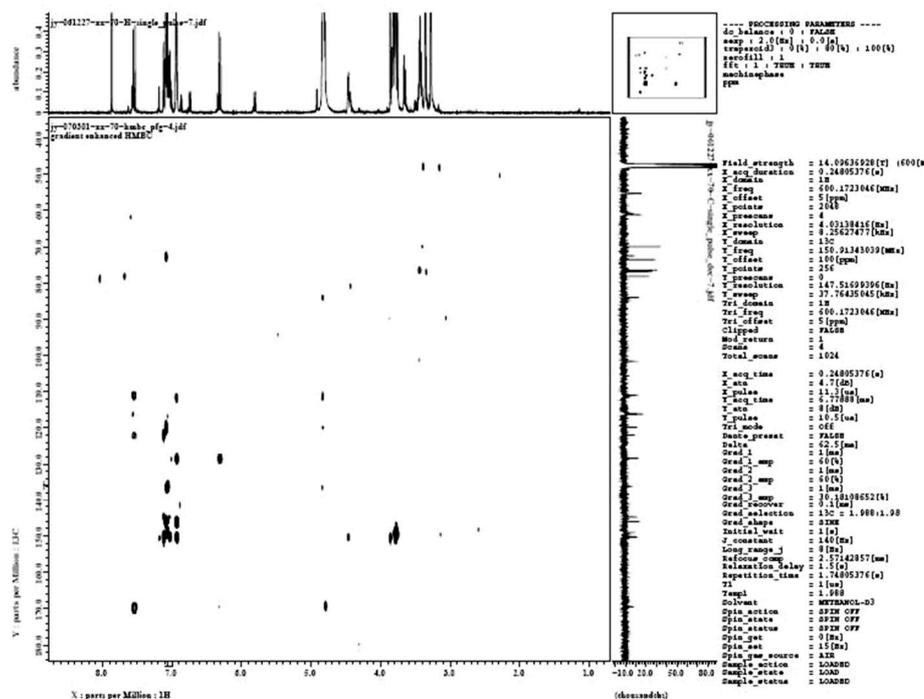






Figure S32. <sup>1</sup>H-NMR spectrum of compound 5 in CD<sub>3</sub>OD.Figure S33. <sup>13</sup>C-NMR spectrum of compound 5 in CD<sub>3</sub>OD.



Figure S36. HMQC-NMR spectrum of compound 5 in CD<sub>3</sub>OD.Figure S37. HMBC-NMR spectrum of compound 5 in CD<sub>3</sub>OD.

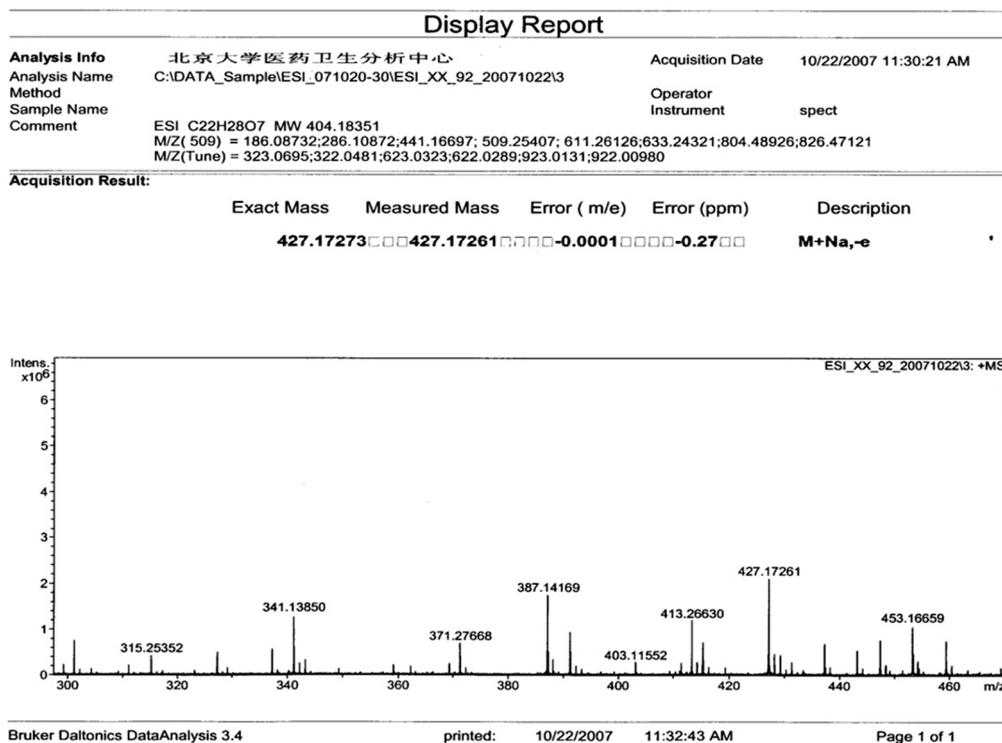
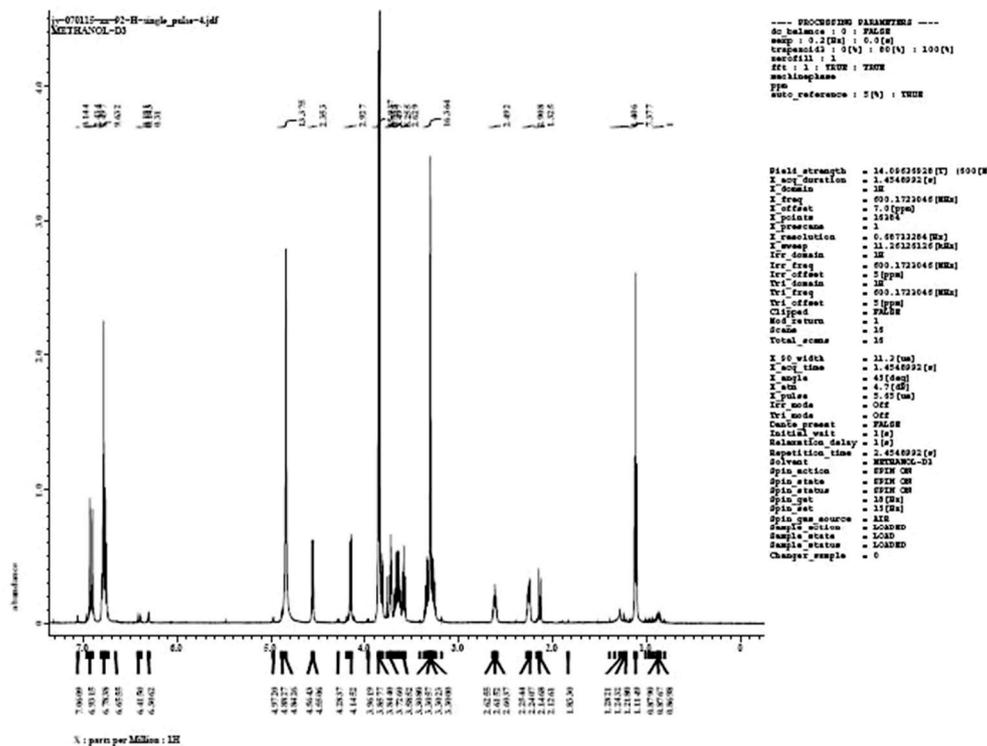
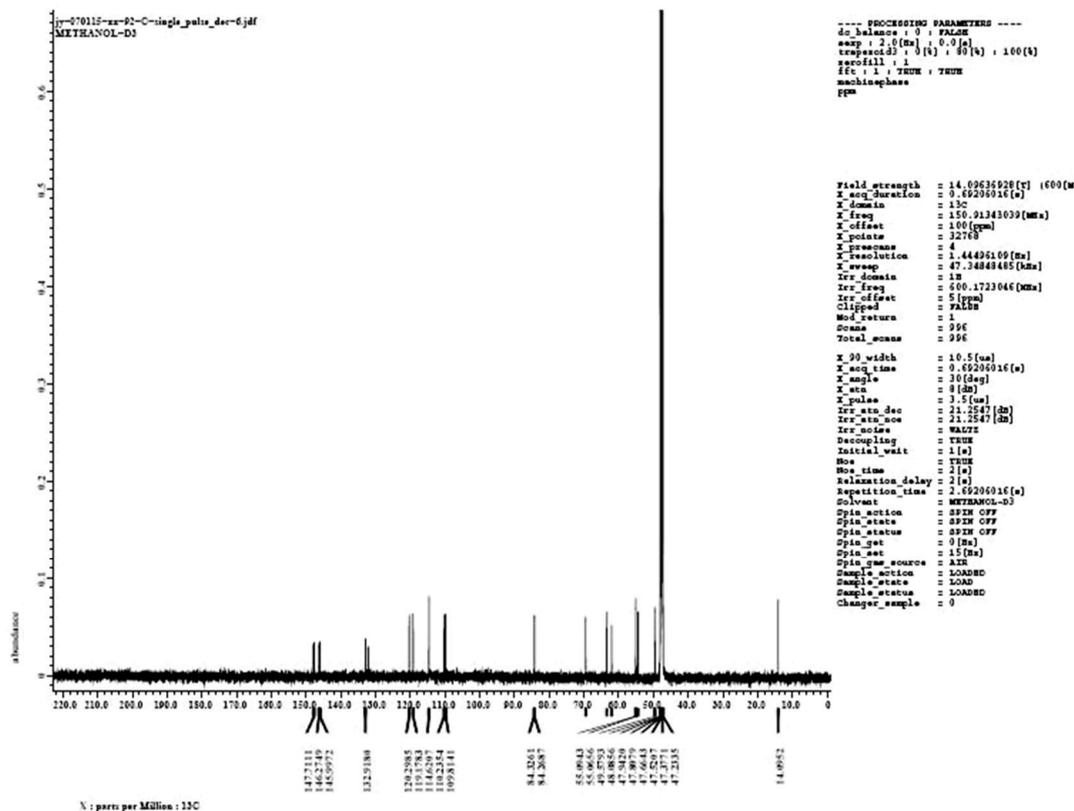
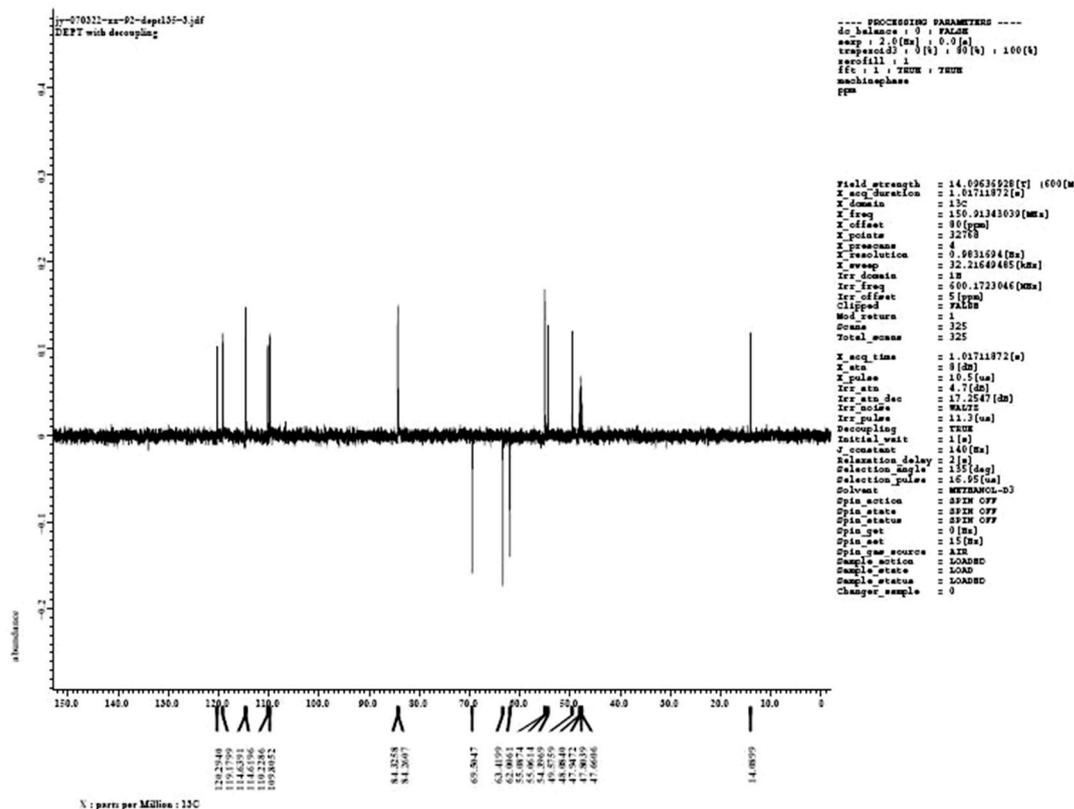
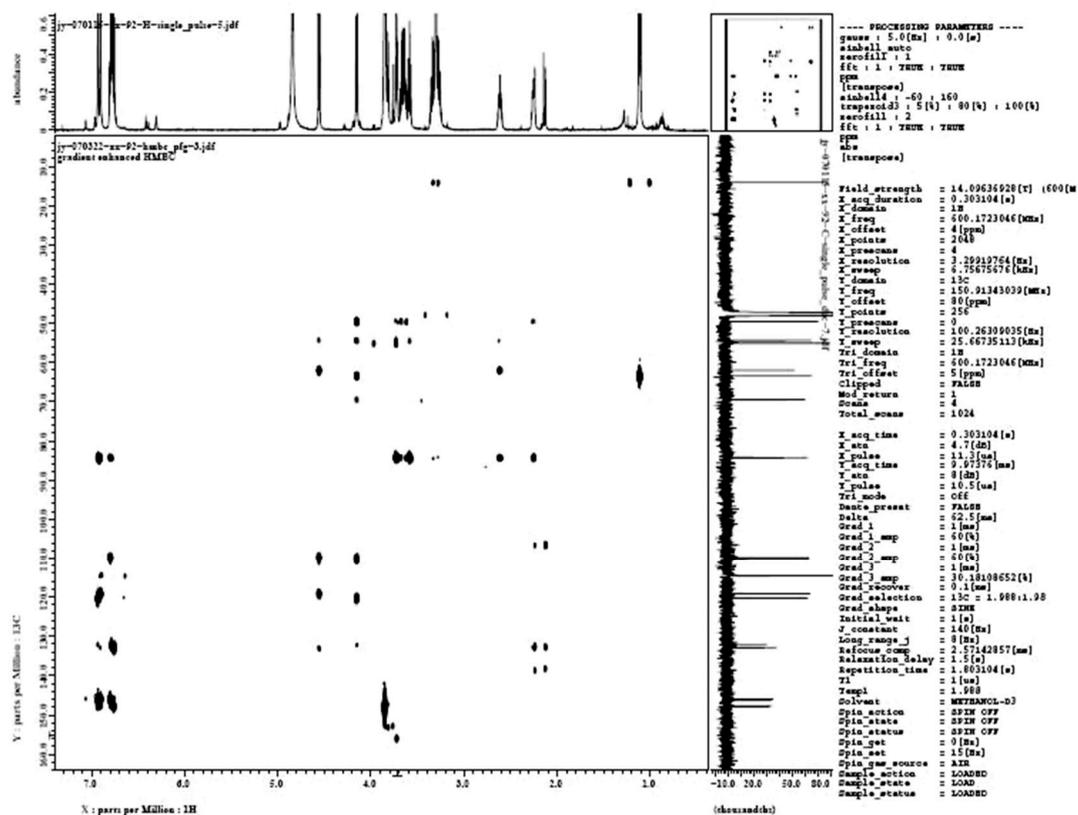
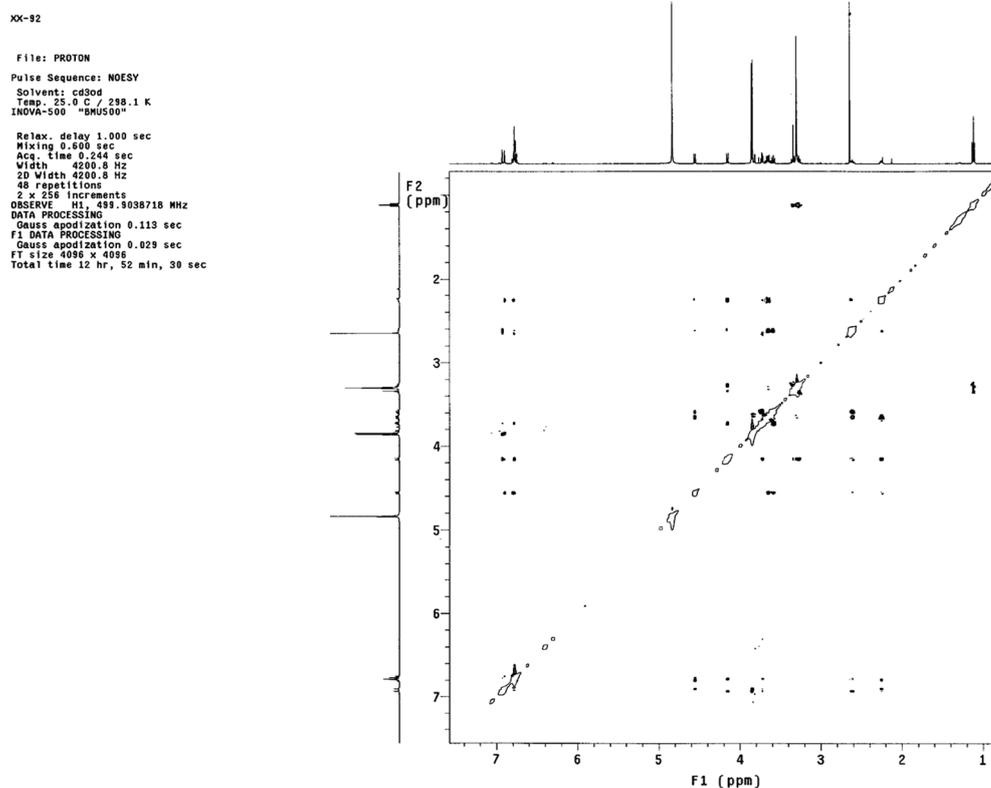


Figure S38. HRESIMS spectrum of compound 6.

Figure S39. <sup>1</sup>H-NMR spectrum of compound 6 in CD<sub>3</sub>OD.

Figure S40.  $^{13}\text{C}$ -NMR spectrum of compound 6 in  $\text{CD}_3\text{OD}$ .Figure S41. DEPT NMR spectrum of compound 6 in  $\text{CD}_3\text{OD}$ .



Figure S44. HMBC-NMR spectrum of compound 6 in CD<sub>3</sub>OD.Figure S45. NOESY NMR spectrum of compound 6 in CD<sub>3</sub>OD.

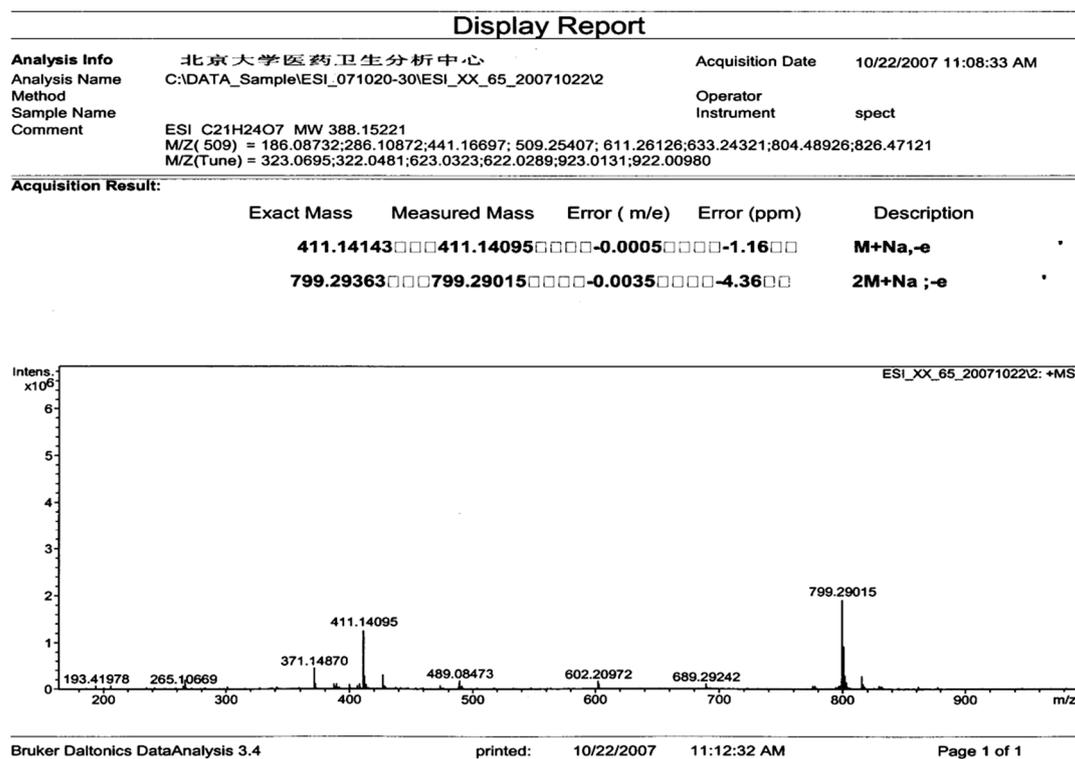
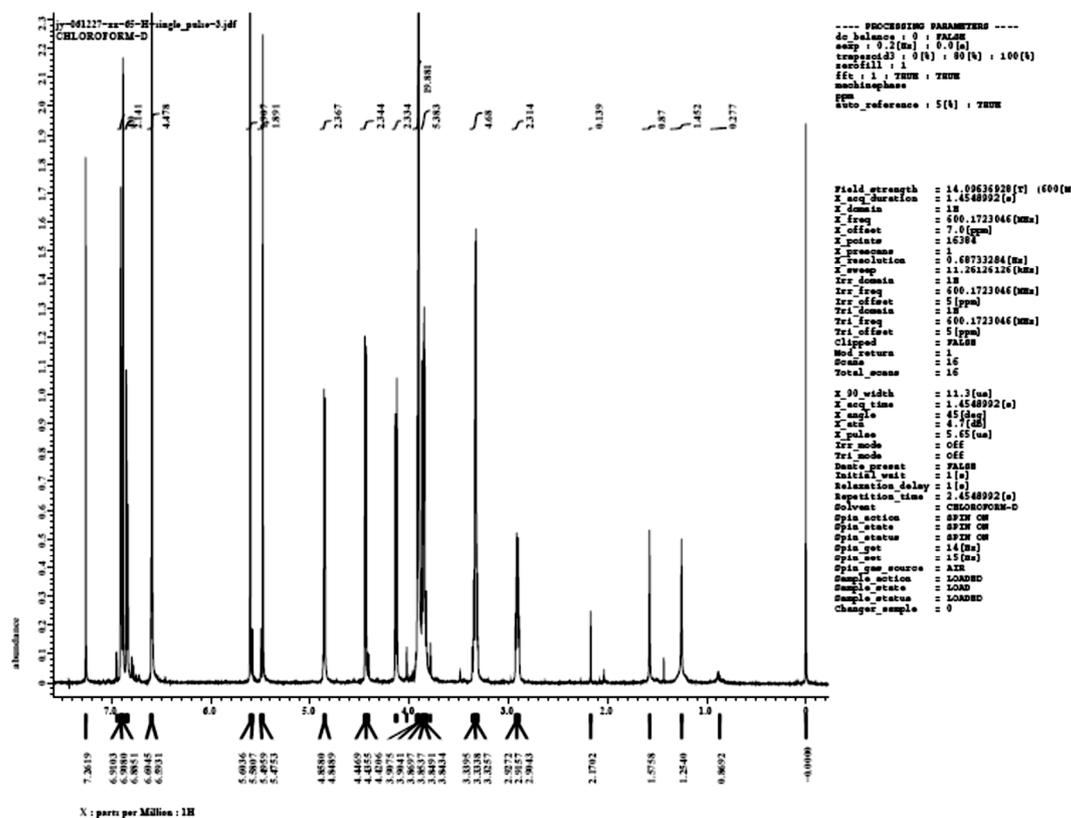


Figure S46. HRESIMS spectrum of compound 7.

Figure S47. <sup>1</sup>H-NMR spectrum of compound 7 in CDCl<sub>3</sub>.

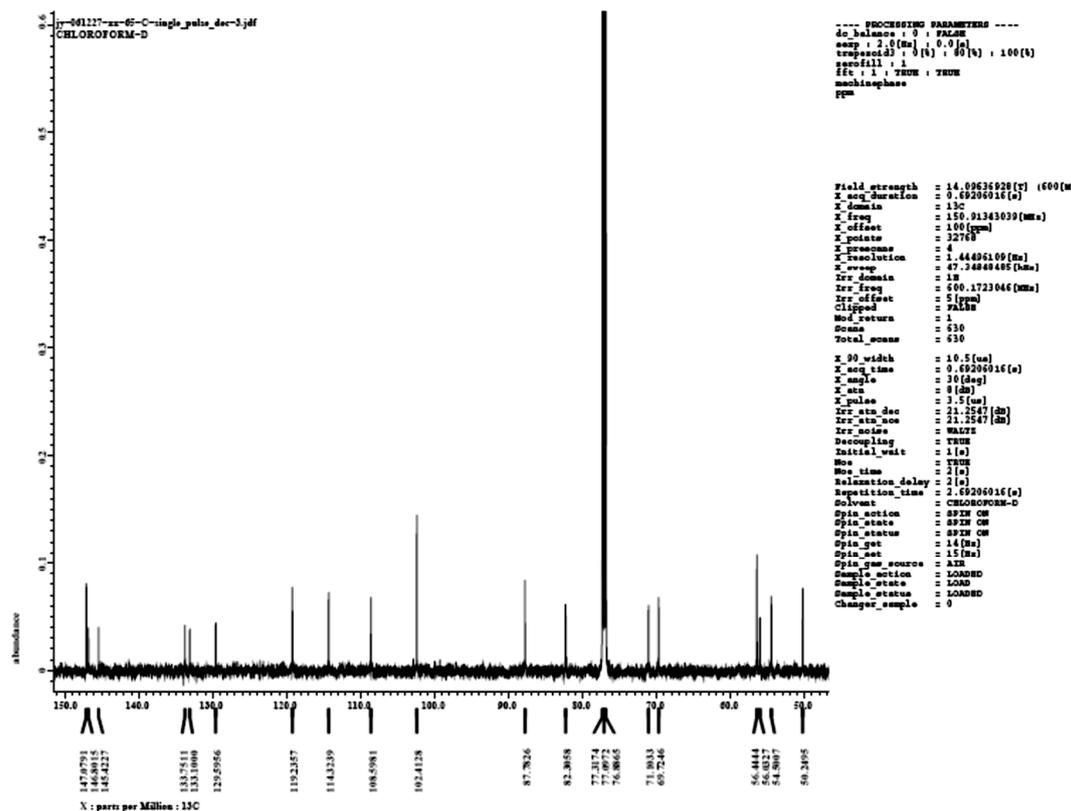
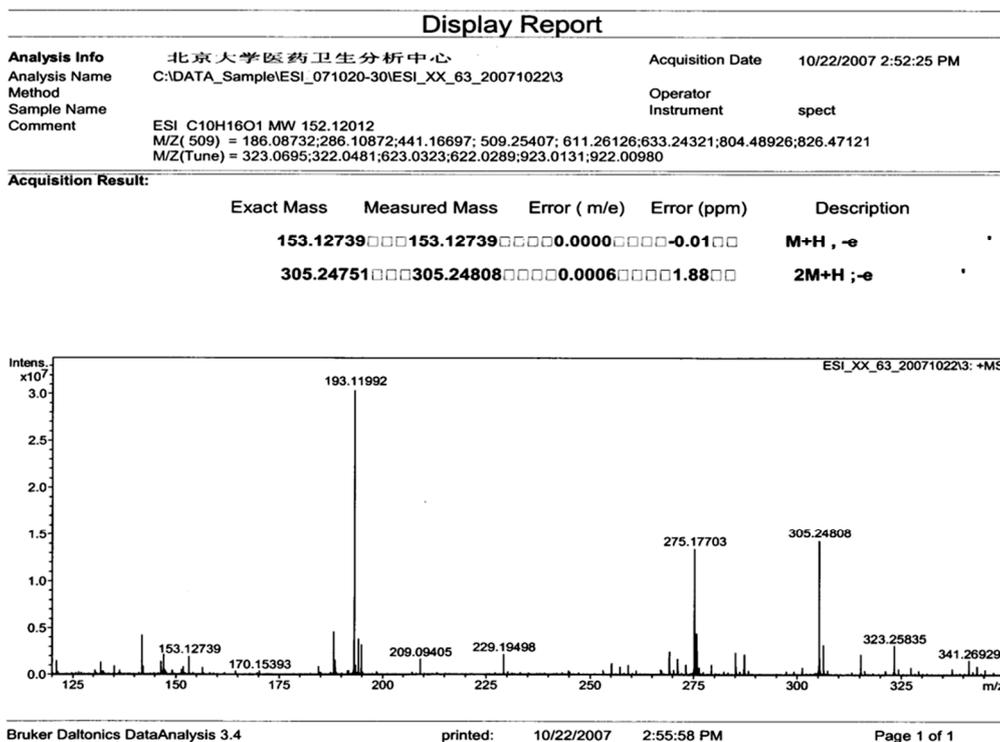
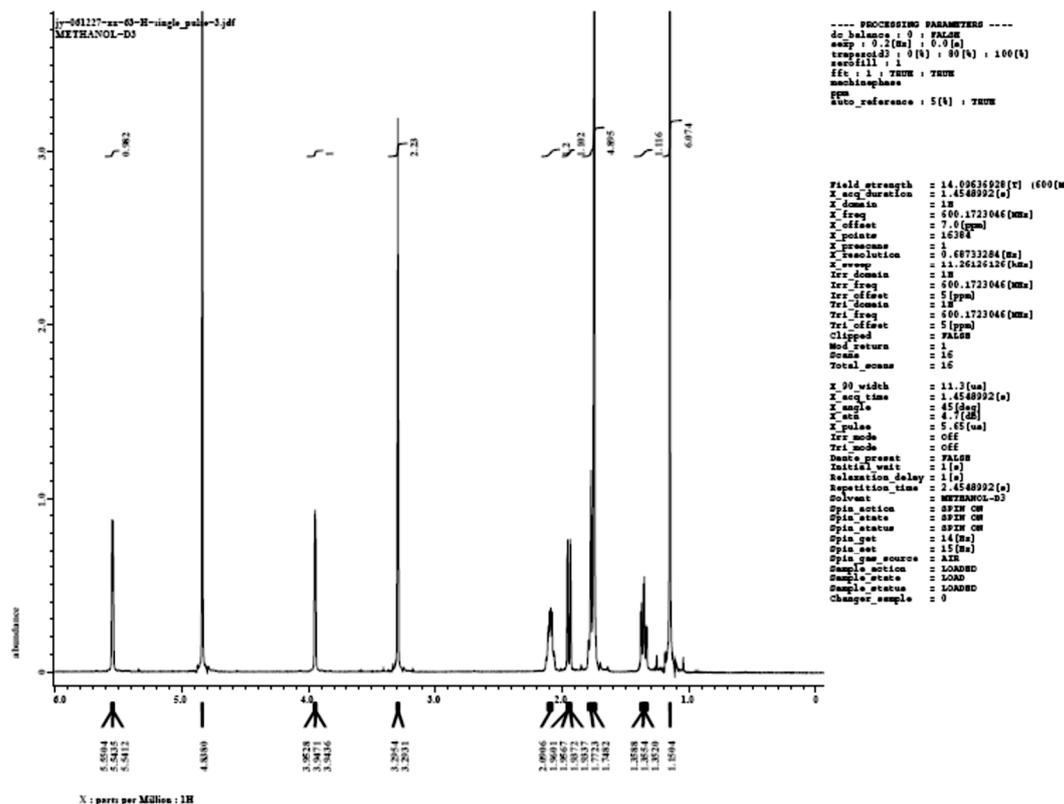
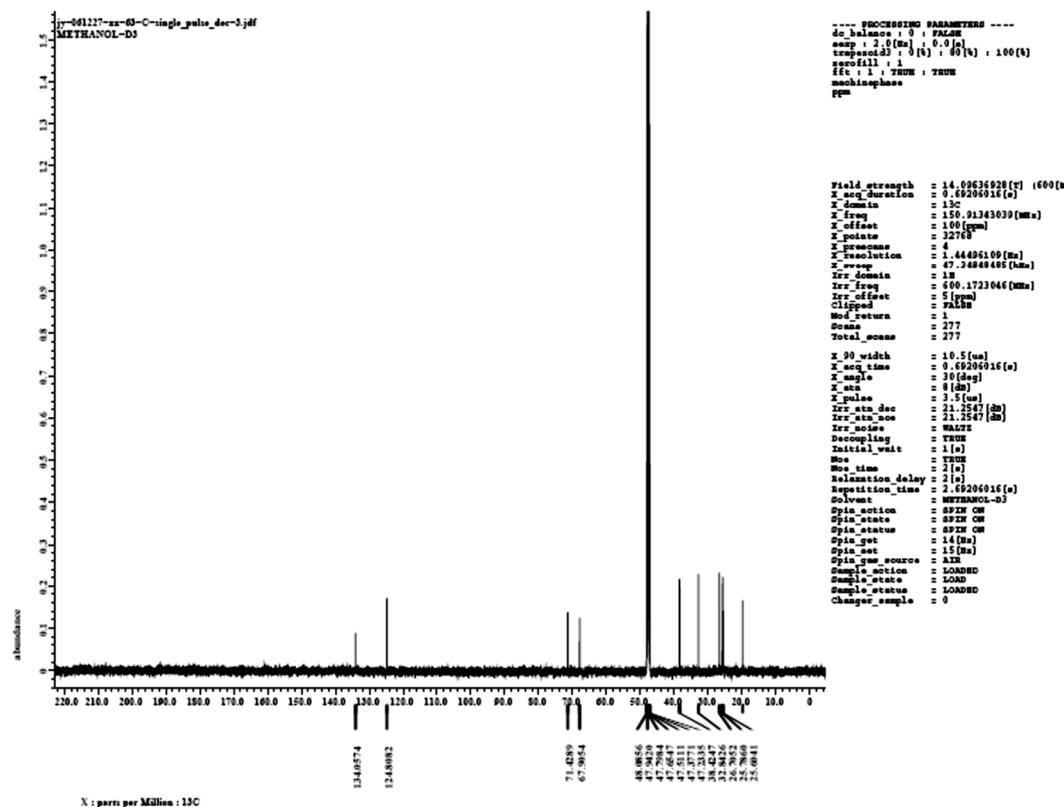
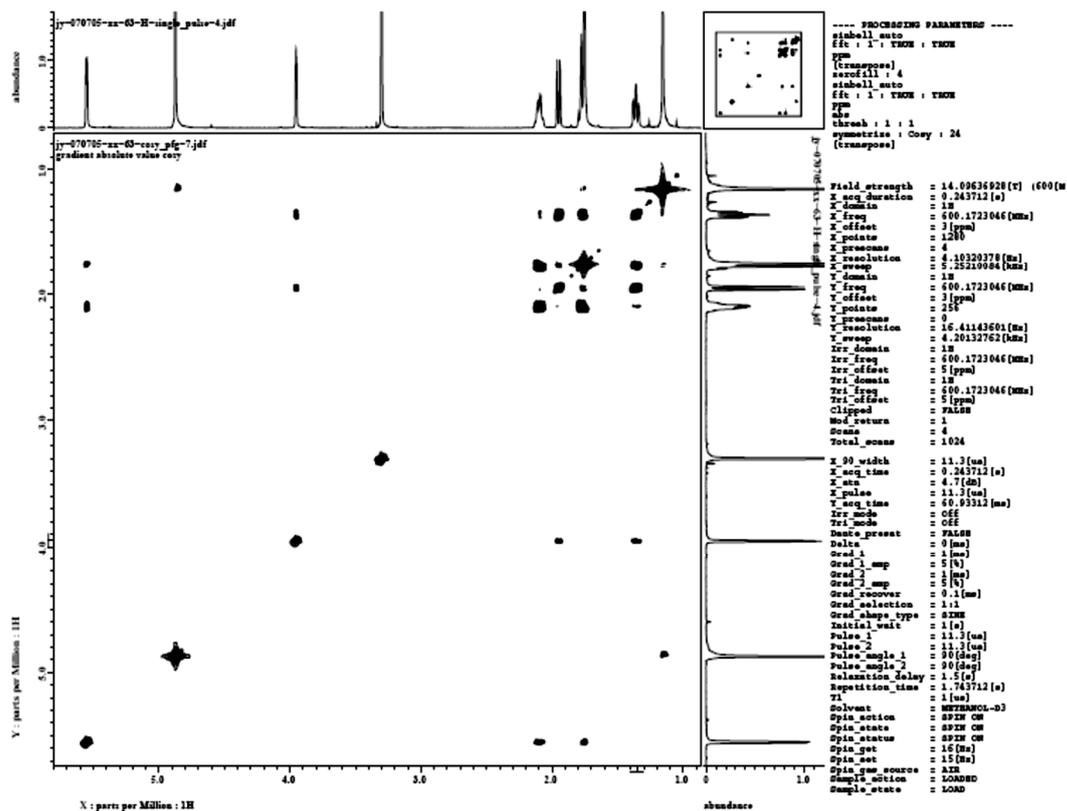
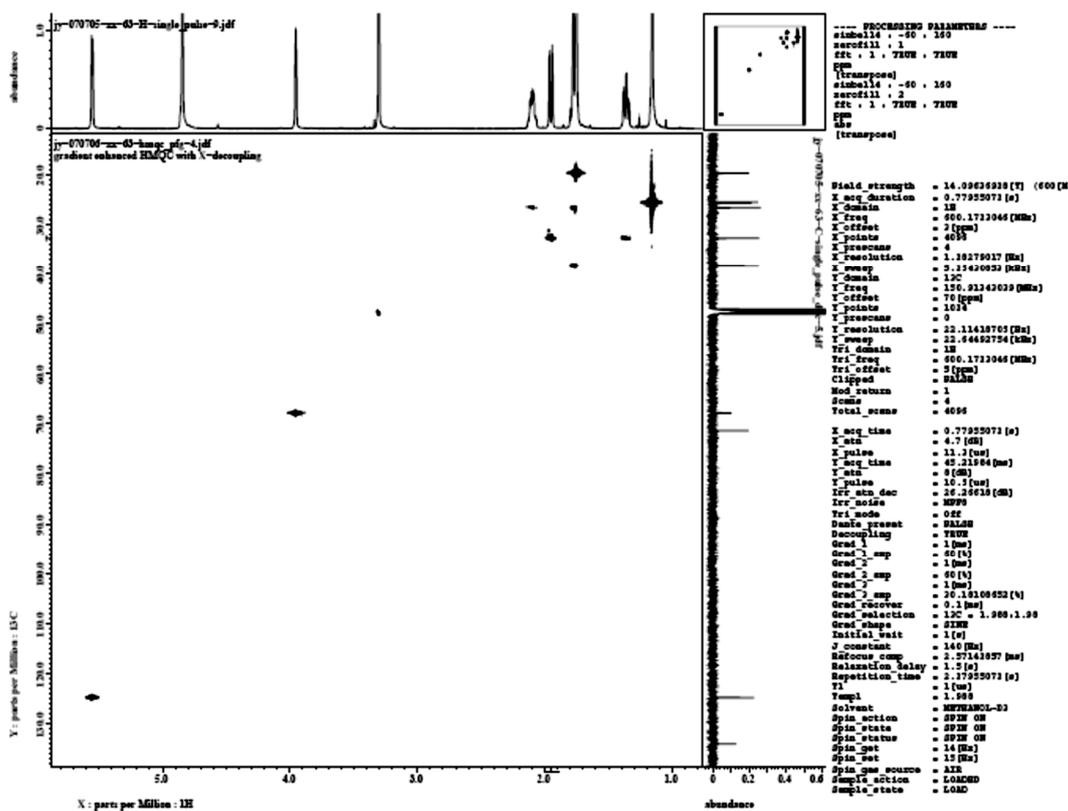
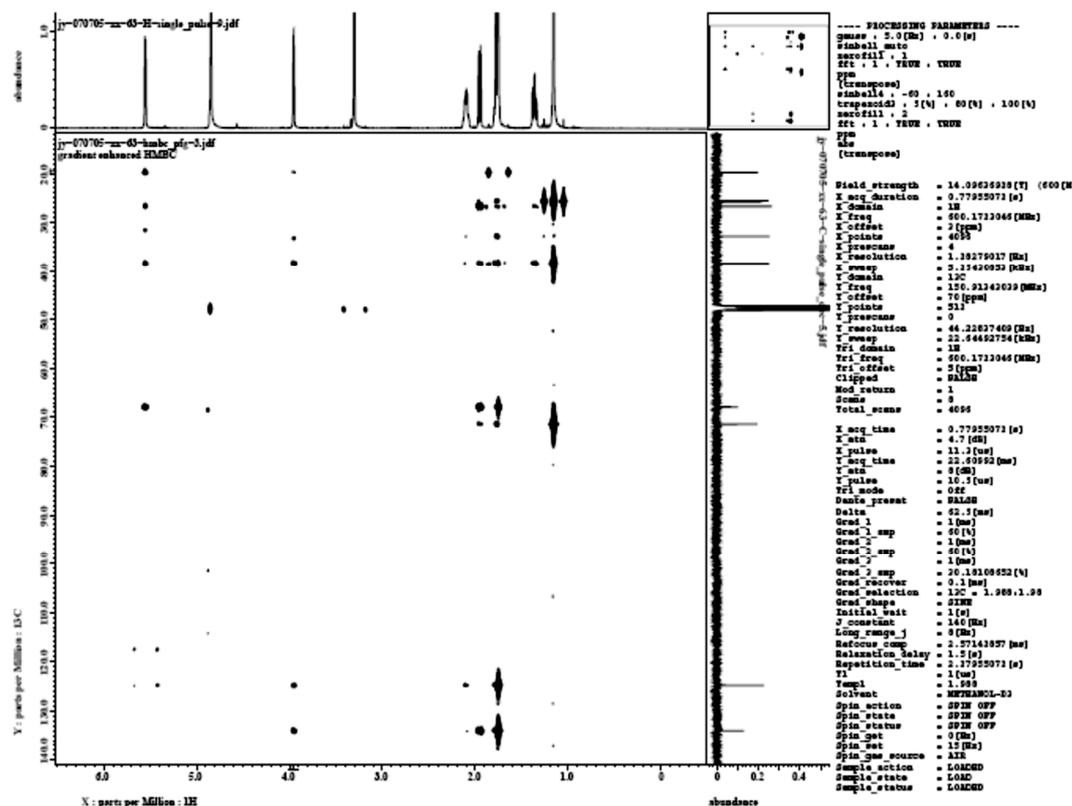
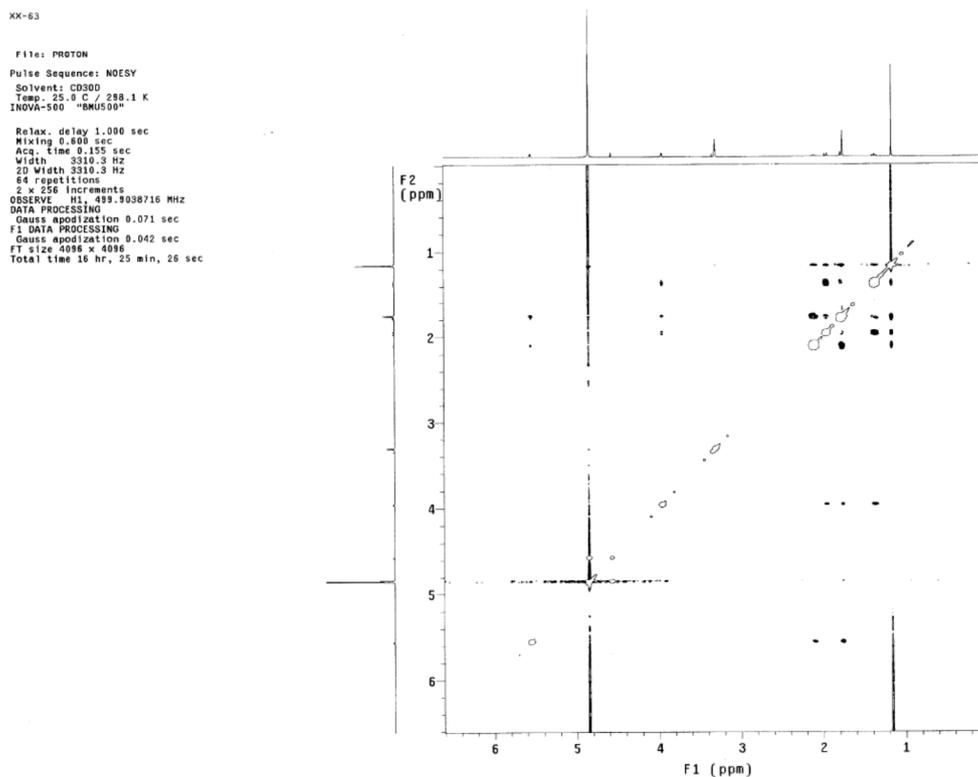
Figure S48. <sup>13</sup>C-NMR spectrum of compound 7 in CDCl<sub>3</sub>.

Figure S49. HRESIMS spectrum of compound 8.

Figure S50. <sup>1</sup>H-NMR spectrum of compound 8 in CD<sub>3</sub>OD.Figure S51. <sup>13</sup>C-NMR spectrum of compound 8 in CD<sub>3</sub>OD.

Figure S52.  $^1\text{H}$ - $^1\text{H}$  COSY NMR spectrum of compound 8 in  $\text{CD}_3\text{OD}$ .Figure S53. HMQC-NMR spectrum of compound 8 in  $\text{CD}_3\text{OD}$ .

Figure S54. HMBC-NMR spectrum of compound 8 in CD<sub>3</sub>OD.Figure S55. NOESY NMR spectrum of compound 8 in CD<sub>3</sub>OD.

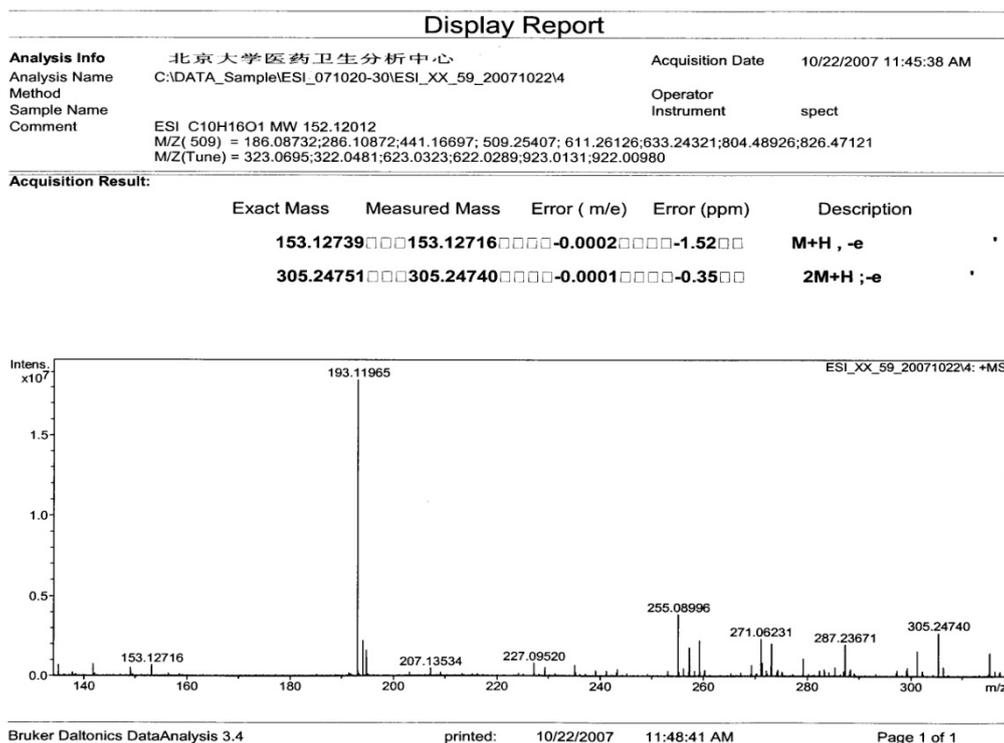
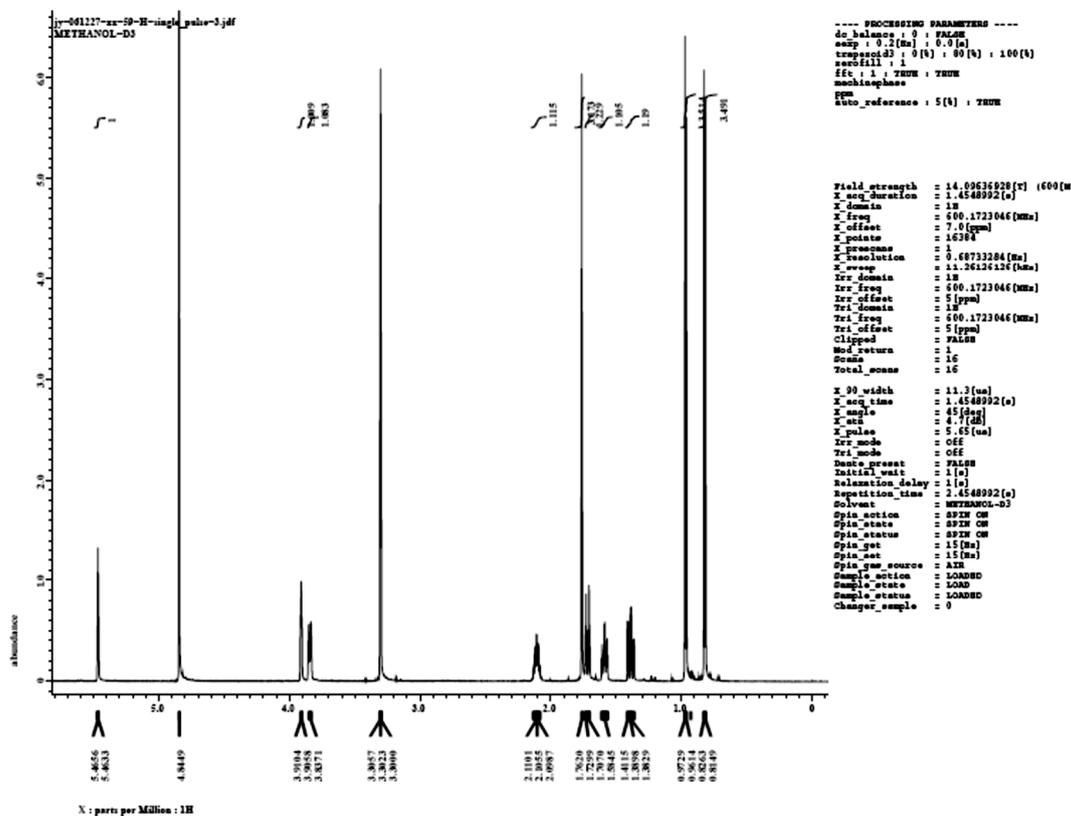


Figure S56. HRESIMS spectrum of compound 9.

Figure S57. <sup>1</sup>H-NMR spectrum of compound 9 in CD<sub>3</sub>OD.





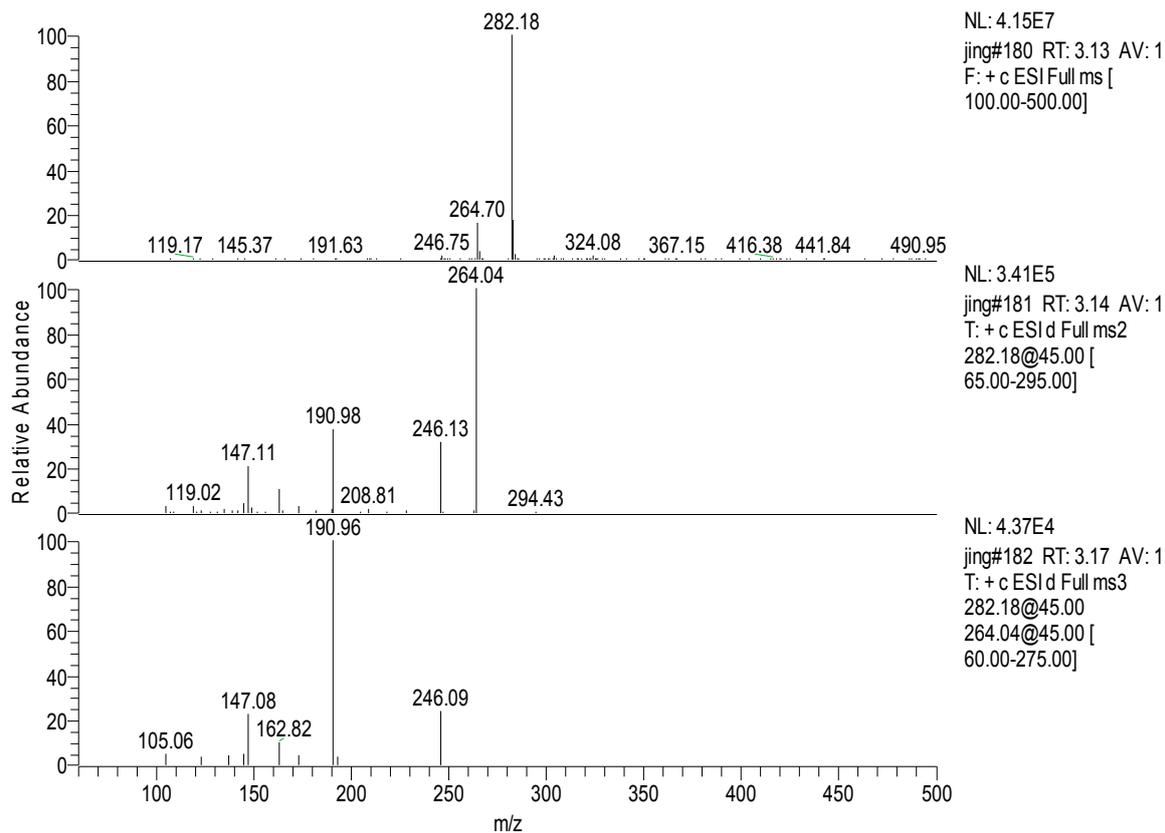


Figure S62. HPLC-MS/MS spectrum of compound 10.

## Display Report

<b>Analysis Info</b>	北京大学医药卫生分析中心	Acquisition Date	10/22/2007 4:17:33 PM
Analysis Name	C:\DATA_Sample\ESI_071020-30\ESI_XX35_20071022\2	Operator	
Method		Instrument	spect
Sample Name			
Comment	ESI C16H27N1O3 MW 281.19909 M/Z ( 509 ) = 186.08732;286.10872;441.16697; 509.25407; 611.26126;633.24321;804.48926;826.47121 M/Z(Tune) = 323.0695;322.0481;623.0323;622.0289;923.0131;922.00980		

## Acquisition Result:

Exact Mass	Measured Mass	Error (m/e)	Error (ppm)	Description
282.20637	282.20694	0.0006	2.02	M+H, -e
304.18832	304.18839	0.0001	0.25	M+Na, -e

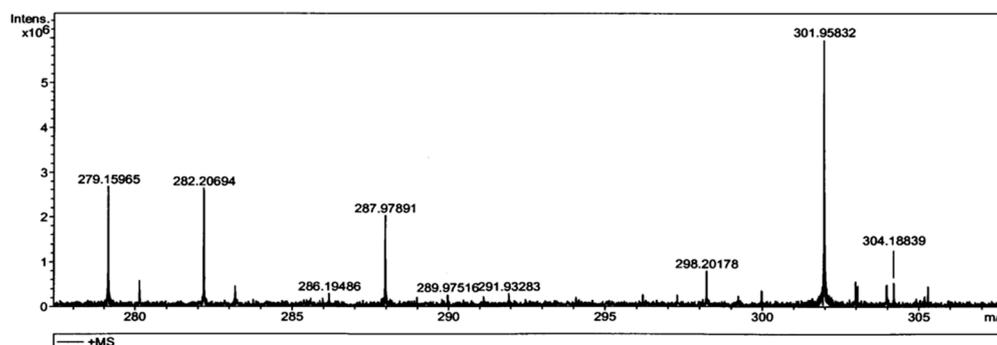
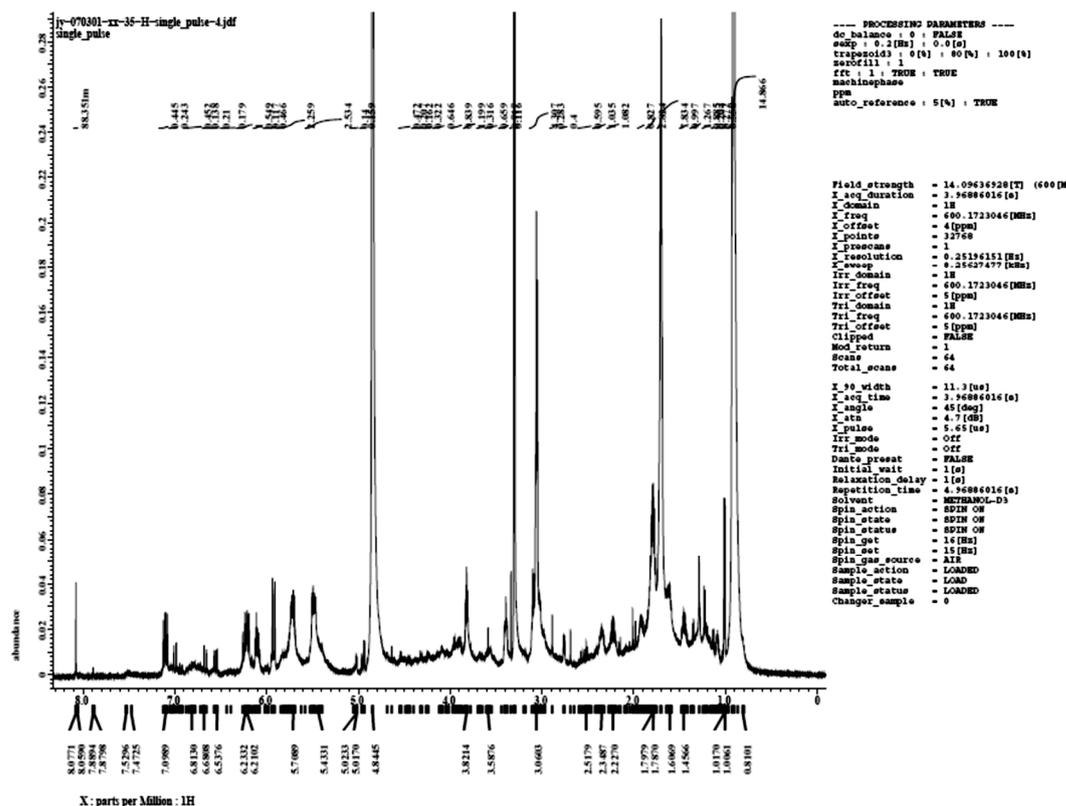
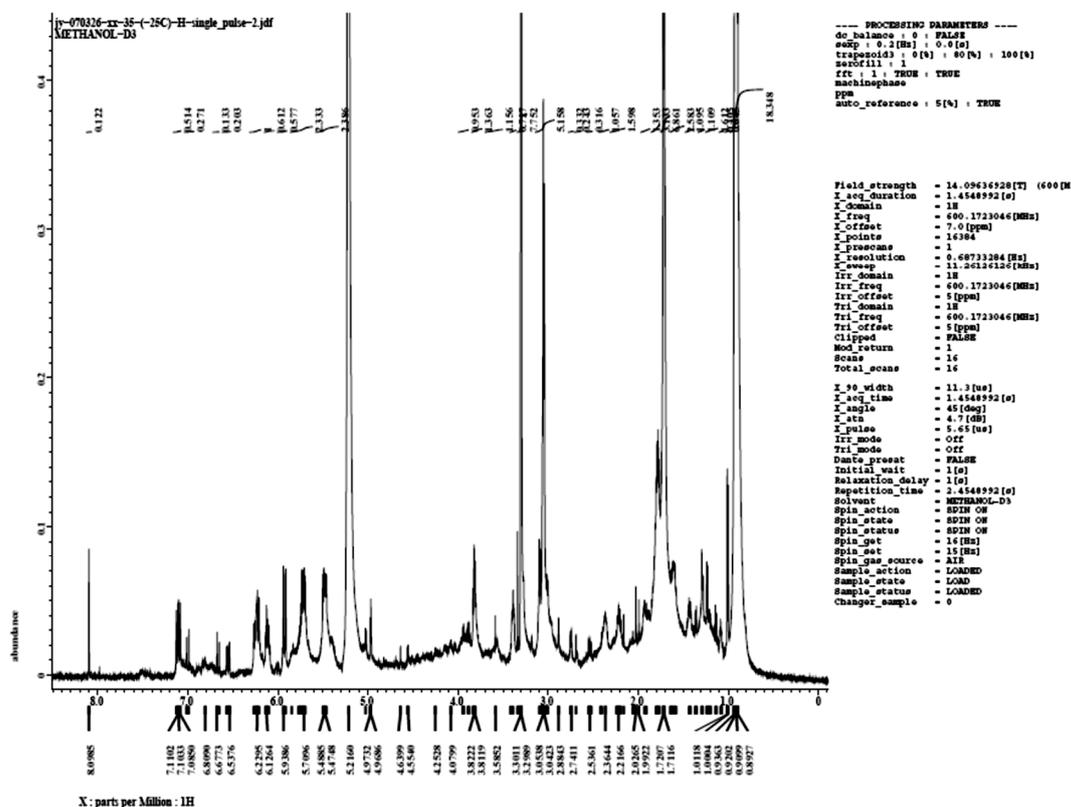
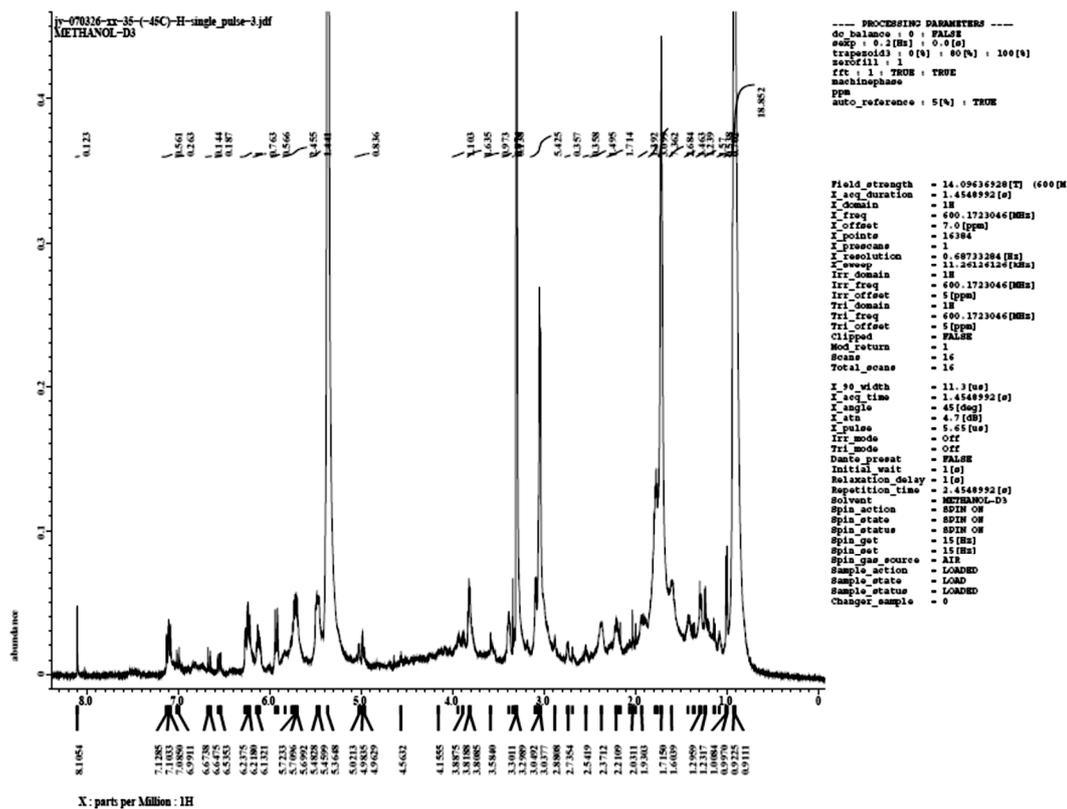
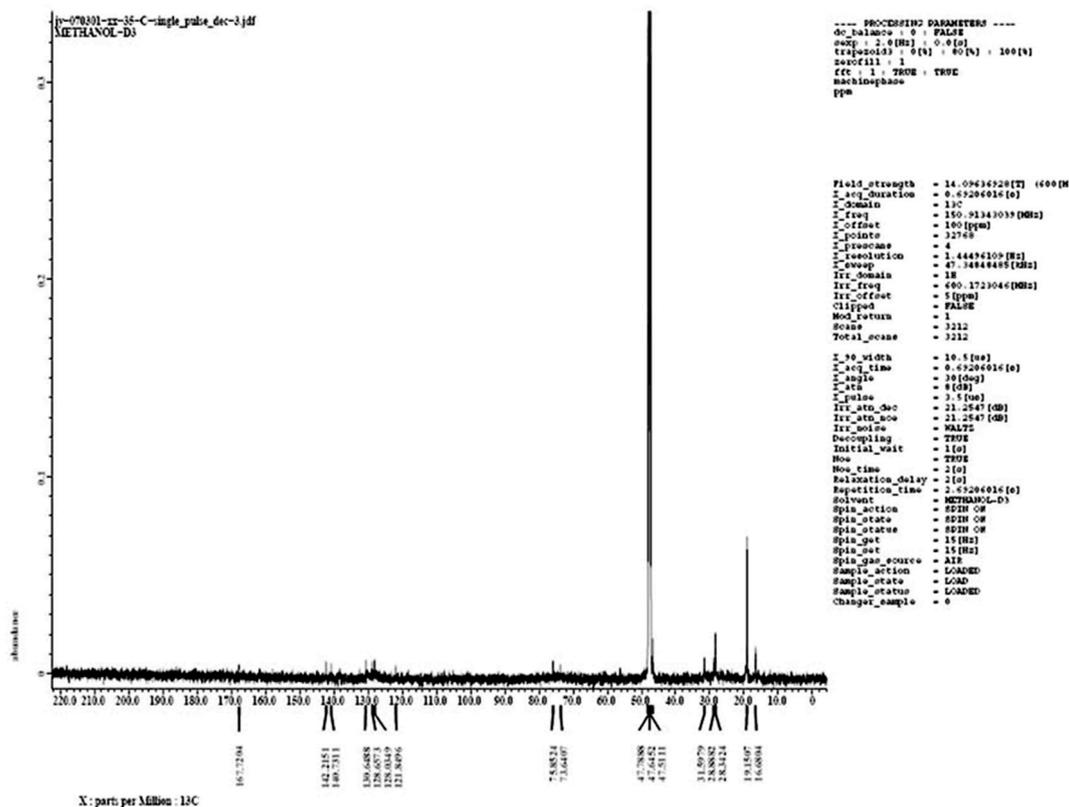
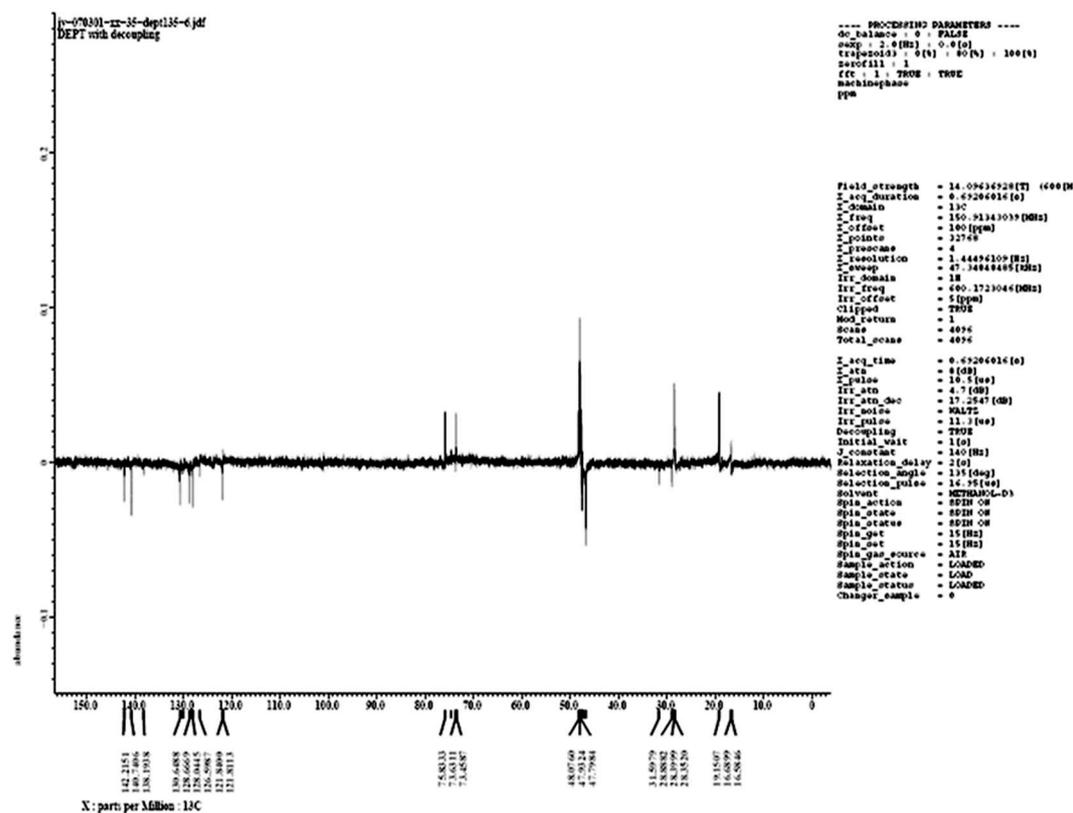
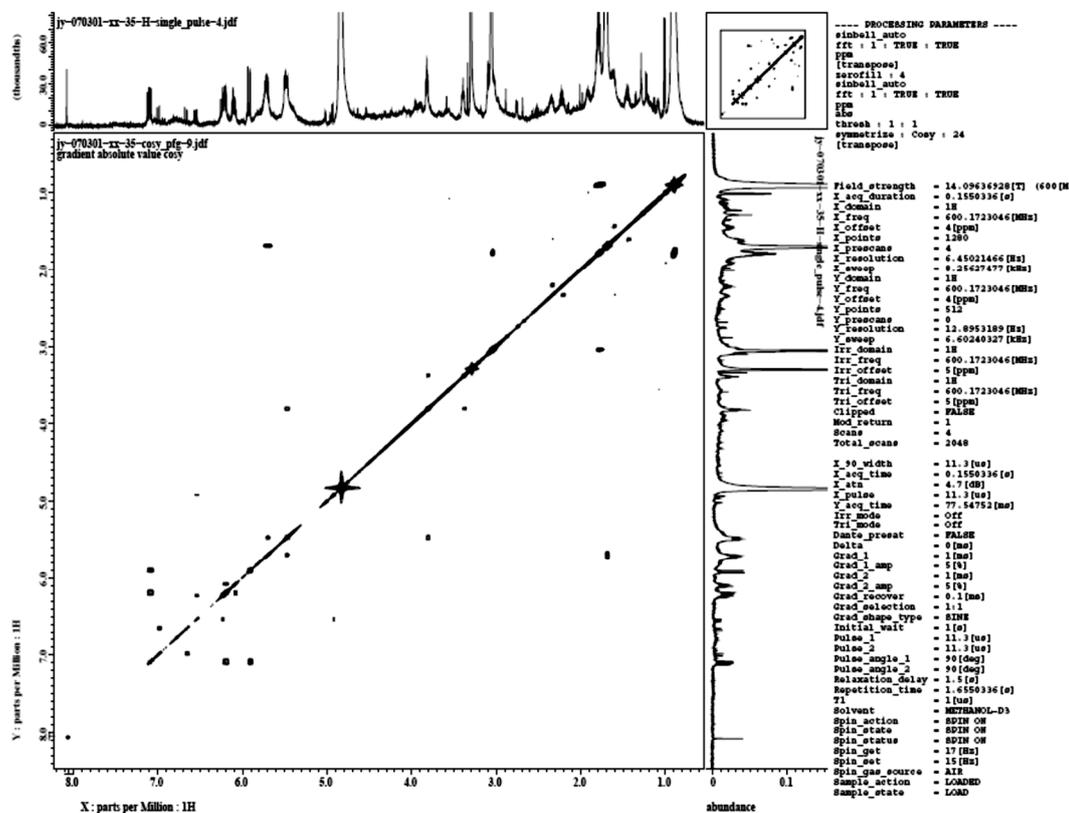
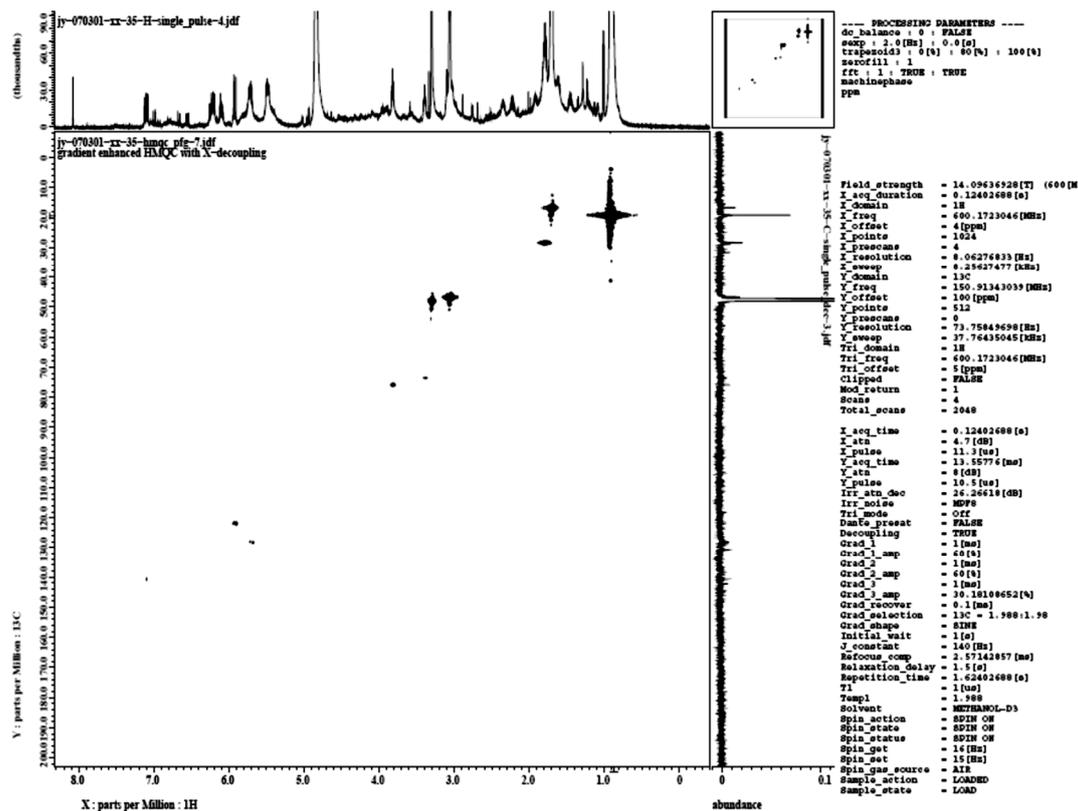


Figure S63. HRESIMS spectrum of compound 10.

Figure S64. <sup>1</sup>H-NMR spectrum of compound 10 in CD<sub>3</sub>OD (25 °C).Figure S65. <sup>1</sup>H-NMR spectrum of compound 10 in CD<sub>3</sub>OD (-25 °C).

Figure S66. <sup>1</sup>H-NMR spectrum of compound 10 in CD<sub>3</sub>OD (−45 °C).Figure S67. <sup>13</sup>C-NMR spectrum of compound 10 in CD<sub>3</sub>OD.

Figure S68. DEPT NMR spectrum of compound 10 in CD<sub>3</sub>OD.Figure S69. <sup>1</sup>H-<sup>1</sup>H COSY NMR spectrum of compound 10 in CD<sub>3</sub>OD.

Figure S70. HMQC-NMR spectrum of compound 10 in CD<sub>3</sub>OD.

## Display Report

Analysis Info		Acquisition Date	10/22/2007 4:08:35 PM
Analysis Name	北京大学医药卫生分析中心	Operator	spect
Method	C:\DATA_Sample\ESI_071020-30\ESI_XX32_20071022\5	Instrument	spect
Sample Name	ESI C16H27N1O3 MW 281.19909		
Comment	M/Z (509) = 186.08732;286.10872;441.16697; 509.25407; 611.26126;633.24321;804.48926;826.47121 M/Z(Tune) = 323.0695;322.0481;623.0323;622.0289;923.0131;922.00980		

## Acquisition Result:

Exact Mass	Measured Mass	Error (m/e)	Error (ppm)	Description
282.20637	282.20632	-0.0001	-0.18	M+H, -e
304.18832	304.18857	0.0003	0.84	M+Na, -e

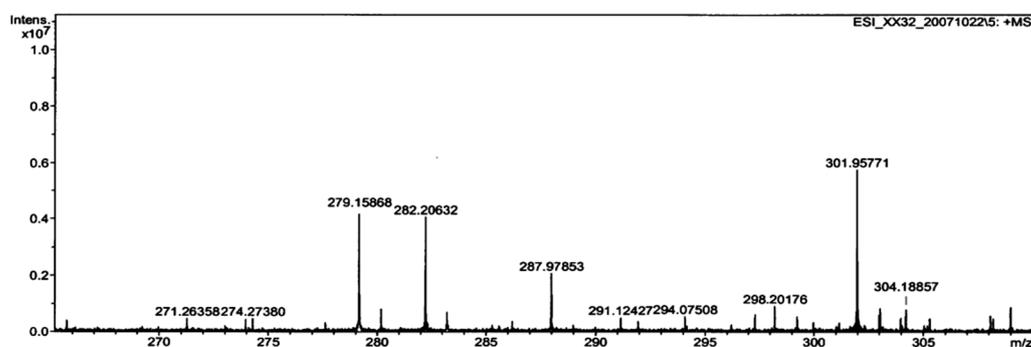
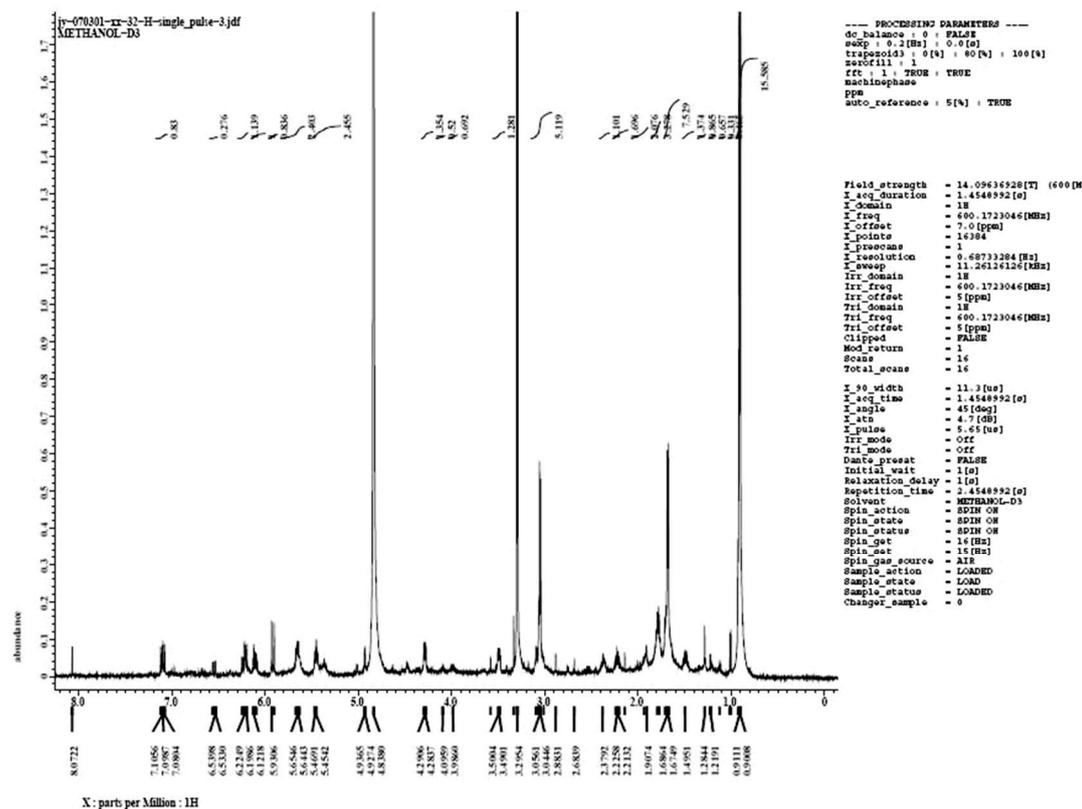
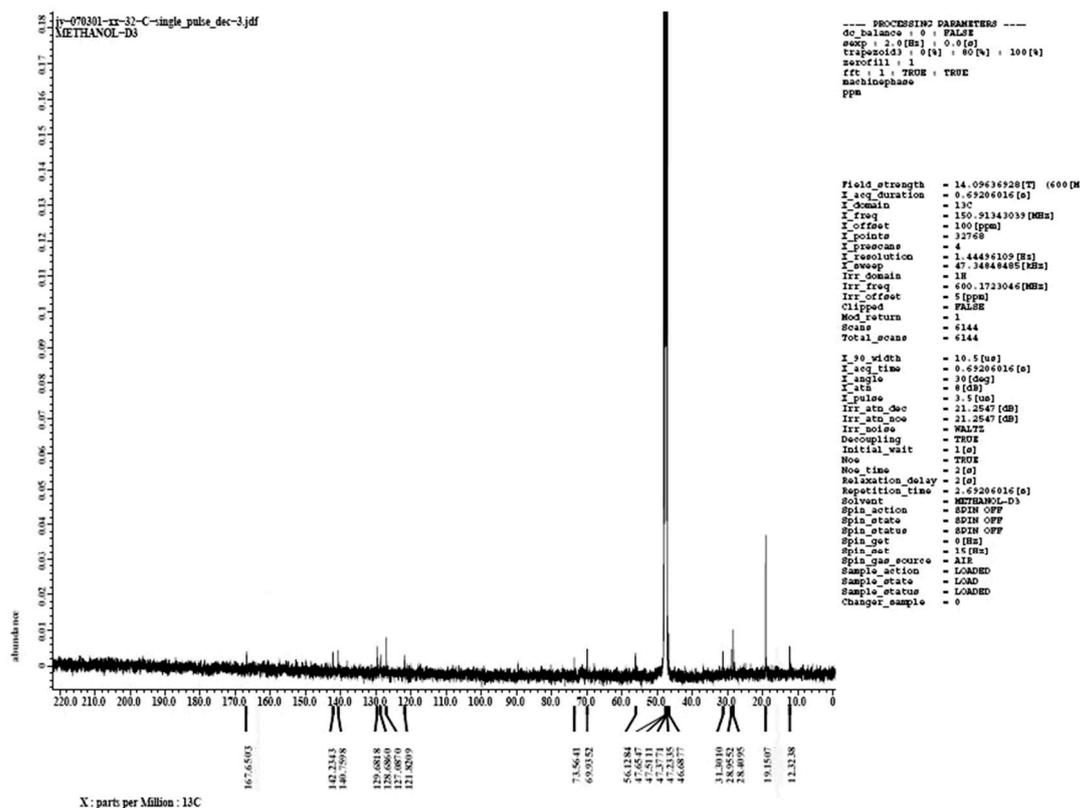
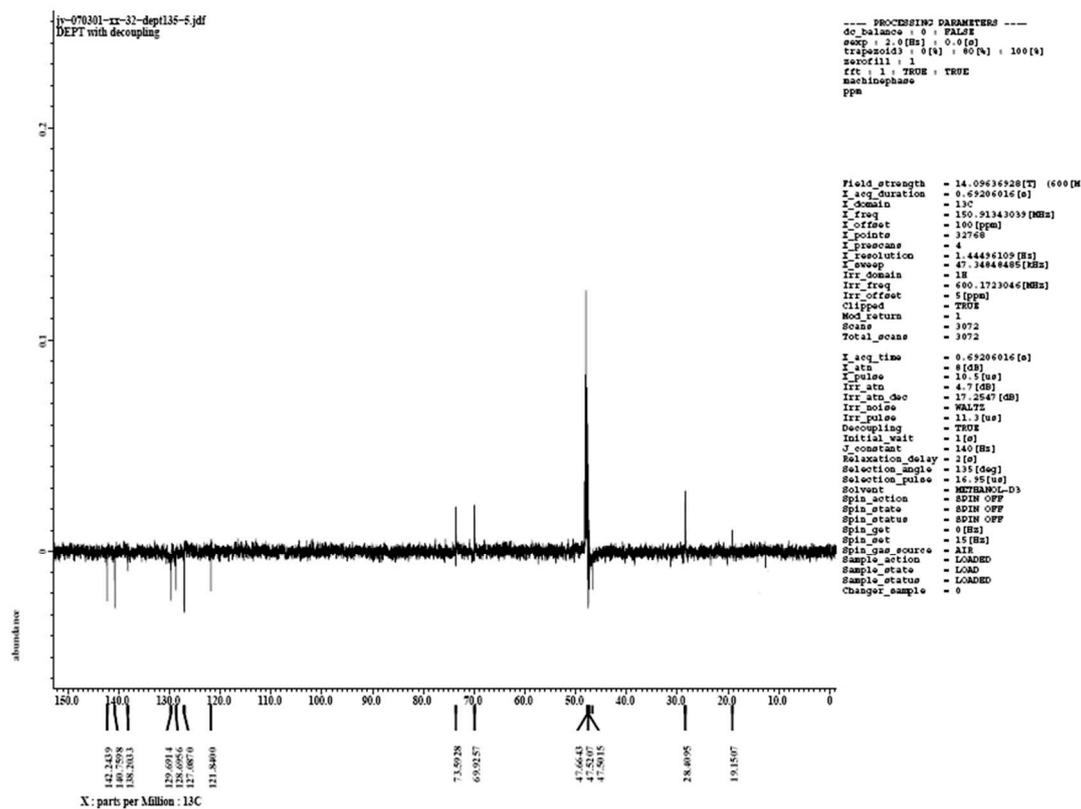
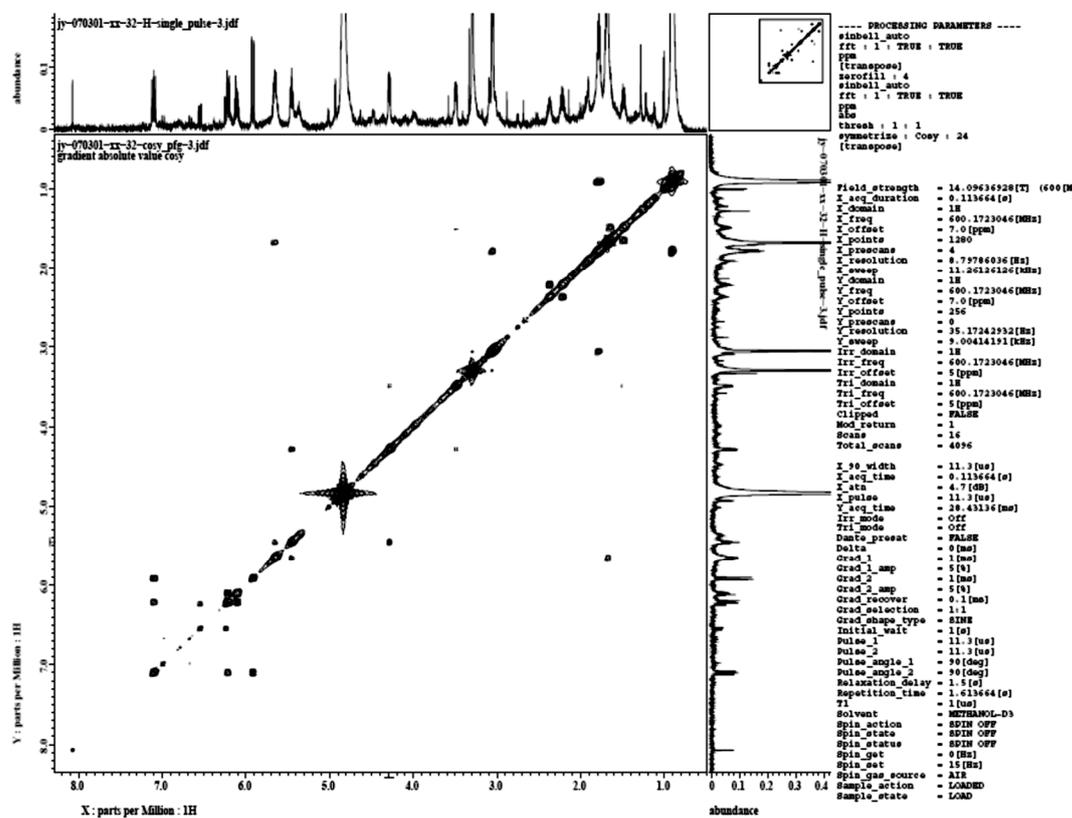


Figure S71. HRESIMS spectrum of compound 11.

Figure S72. <sup>1</sup>H-NMR spectrum of compound 11 in CD<sub>3</sub>OD.Figure S73. <sup>13</sup>C-NMR spectrum of compound 11 in CD<sub>3</sub>OD.

Figure S74. DEPT NMR spectrum of compound 11 in CD<sub>3</sub>OD.Figure S75. <sup>1</sup>H-<sup>1</sup>H COSY NMR spectrum of compound 11 in CD<sub>3</sub>OD.

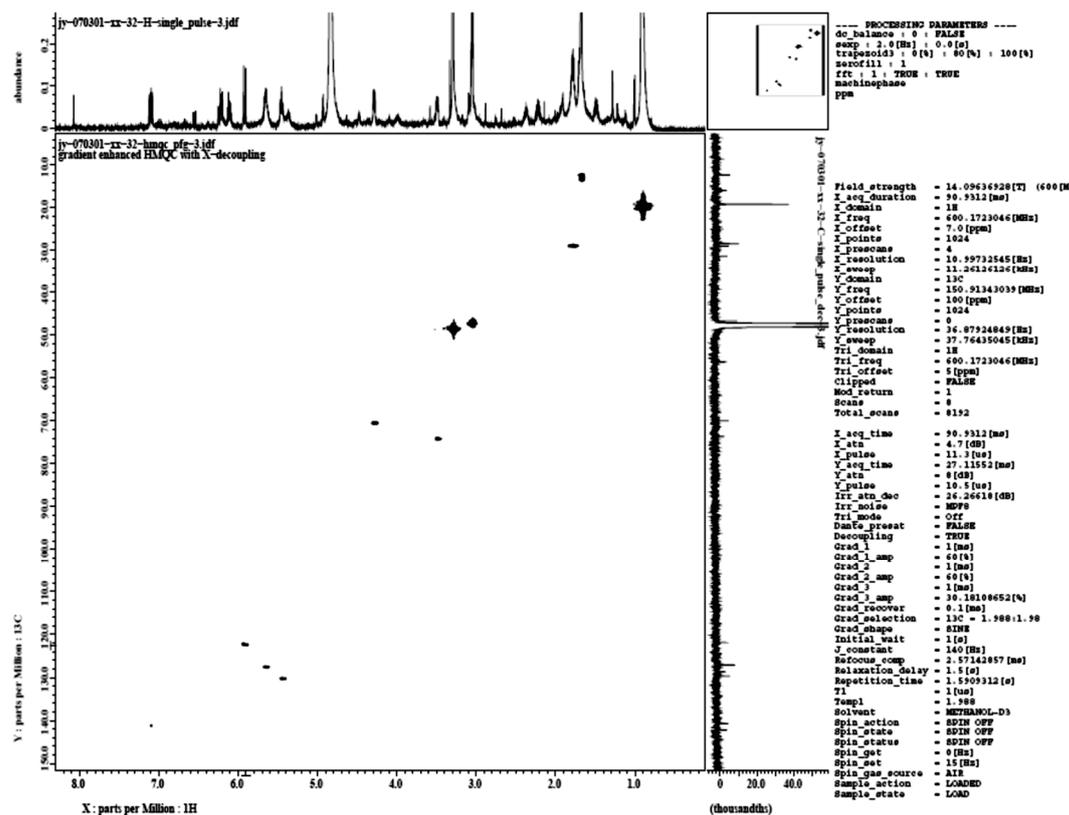


Figure S76. HMQC-NMR spectrum of compound **11** in CD<sub>3</sub>OD.

## 2. Extraction and Isolation of Known Compounds 13–47

The air-dried and powdered roots and rhizomes of *A. heterotropoides* var. *mandshuricum* (36 kg) were extracted three times (2 h, 1.5 h, 1.5 h for each) under reflux with 95% ethanol and then three times (2 h, 1.5 h, 1.5 h for each) with 50% ethanol successively. The combined extracts were concentrated under reduced pressure to give a dark brown residue (9.2 kg), then 7.8 kg of it was suspended in H<sub>2</sub>O (16.5 L) and partitioned sequentially with petroleum ether (Pet.) (60–90 °C °C) (4 × 5 L), CHCl<sub>3</sub> (4 × 5 L), EtOAc (4 × 5 L), and *n*-BuOH (4 × 5 L), respectively. The CHCl<sub>3</sub> layer (231 g) was fractionated on silica gel CC eluting with a gradient of petroleum ether–EtOAc (10:1 to 0:1) to obtain 16 fractions C1–C16. Fraction C5 and C6 were left to stand overnight and colorless columnar crystal as precipitate were collected, the colorless crystal was purified by recrystallized from EtOAc to afford compound **14** (10 g). Fraction C7 produced two different colorless needle crystals when standing overnight, and then the two needle crystals were further recrystallized from EtOAc to obtain compounds **15** (1 g) and **16** (0.8 g). The residue of fraction C7 was subjected to silica gel column chromatography eluting with a gradient of petroleum ether–EtOAc (100:15, 100:30, 0:100) to yield seven subfractions, C7-1–C7-7. C7-7 was further separated by silica gel CC and purified on Sephadex LH-20 eluting with CHCl<sub>3</sub>–MeOH (6:4) to afford compound **45** (17 mg). Fraction C10 was then subjected to silica gel CC eluting with a gradient of CHCl<sub>3</sub>–MeOH (1:40 to 1:30) to provide three subfractions, C10-1–C10-3. C10-1 was further separated by silica gel CC and purified on Sephadex LH-20 eluting with CHCl<sub>3</sub>–MeOH (1:1) to afford compound **13** (12 mg). C10-3 was chromatographed over silica gel with petroleum ether–MeOH (1:30) and further separated by Sephadex LH-20 (CHCl<sub>3</sub>–MeOH, 1:1) to obtain compound **41** (31 mg). Fraction C11 was subjected to silica gel CC eluting with a gradient of CHCl<sub>3</sub>–MeOH (1:30 to 1:35) to yield four subfractions, C11-1–C11-4. C11-1 was subjected to chromatography on silica gel CC eluting with petroleum ether–Me<sub>2</sub>CO (4:1) to produce seven subfractions, C11-1-1–C11-1-7. C11-1-3 was further separated by silica gel column chromatography using CHCl<sub>3</sub>–EtOAc (9:1) to yield four subfractions, C11-1-3-1–C11-1-3-4. C11-1-3-3

was then separated by Sephadex LH-20 (CHCl<sub>3</sub>–MeOH, 1:1) and semi-preparative HPLC eluted with a gradient of MeOH–H<sub>2</sub>O to yield compound **21** (21 mg). C11-1-4 was separated on silica gel CC eluting with CHCl<sub>3</sub>–EtOAc (9:1) and further purified by Sephadex LH-20 (CHCl<sub>3</sub>–MeOH, 6:4) to afford compound **18** (33 mg). C11-1-5 was chromatographed over silica gel with petroleum ether–Me<sub>2</sub>CO (2.5:1) as eluent, and subfraction was then separated by Sephadex LH-20 (CHCl<sub>3</sub>–MeOH, 6:4) and semi-preparative HPLC eluted with a gradient of MeOH–H<sub>2</sub>O to yield compound **17** (6.3 mg). C11-3 yielded a colorless needle crystal, collected by filtering, and further purified by washing with CHCl<sub>3</sub> to afford compound **39** (33 mg). The residue of C11-3 was subjected to silica gel CC eluting with a gradient of CHCl<sub>3</sub>–MeOH to yield three subfractions, C11-3-1–C11-3-3. C11-3-2 was subjected to silica gel CC eluting with petroleum ether–Me<sub>2</sub>CO (4.5:1) to yield compound **46** (6 mg). C11-3-3 was also chromatographed on silica gel with petroleum ether–Me<sub>2</sub>CO (4.5:1) as eluent to obtain compounds **34** (7.2 mg) and **35** (11mg). Fraction C12 was chromatographed on silica gel with a gradient of CHCl<sub>3</sub>–MeOH to give four fractions, C12-1–C12-4. C12-3 was further divided into five subfractions, C12-3-1–C12-3-5, with repeated silica gel CC eluting with CHCl<sub>3</sub>–MeOH (80:1 to 15:1). Chromatography of C12-3-1 on silica gel with CHCl<sub>3</sub>–EtOAc (5:1, 4:1) and petroleum ether–Me<sub>2</sub>CO (2:1) as eluent successively and purified by semi-preparative HPLC eluting with a gradient of MeOH–H<sub>2</sub>O to yield compound **25** (7.8 mg). C12-3-3 was chromatographed over silica gel with CHCl<sub>3</sub>–EtOAc (4:1, 1:1) as eluent, and the subfraction was then separated by Sephadex LH-20 (CHCl<sub>3</sub>–MeOH, 6:4) and semi-preparative HPLC eluted with a gradient of MeOH–H<sub>2</sub>O to yield compound **44** (21 mg). C12-4 was separated on silica gel CC eluting with CHCl<sub>3</sub>–MeOH (70:1 to 15:1) and further purified by Sephadex LH-20 eluting with (CHCl<sub>3</sub>–MeOH, 1:1) to afford compound **47** (28 mg). Fraction C13 was chromatographed with a gradient of CHCl<sub>3</sub>–MeOH (35:1, 20:1, 5:1) as eluent to give six subfractions, C13-1–C13-6. C13-6 was then subjected to silica gel CC with petroleum ether–Me<sub>2</sub>CO (30:1) to obtain five subfractions, C13-6-1–C13-6-5, and subfraction C13-6-4 was further separated by Sephadex LH-20 (petroleum ether–CHCl<sub>3</sub>–MeOH, 2:1:1) and semi-preparative HPLC with a gradient of MeOH–H<sub>2</sub>O as the mobile phase to yield compounds **20** (2.7 mg) and **23** (34 mg). Fraction C14 was subjected to silica gel CC eluting with a gradient of CHCl<sub>3</sub>–Me<sub>2</sub>CO (25:1 to 10:1) to yield five subfractions, C14-1–C14-5. C14-4 was chromatographed over silica gel with CHCl<sub>3</sub>–Me<sub>2</sub>CO (10:1 to 5:1) and further separated by Sephadex LH-20 (petroleum ether–CHCl<sub>3</sub>–MeOH, 2:1:1) to obtain compound **22** (18 mg). C14-5 was subjected to silica gel CC eluting with CHCl<sub>3</sub>–Me<sub>2</sub>CO (10:1 to 5:1) to give seven subfractions, C14-5-1–C14-5-7. C14-5-5 was purified by Sephadex LH-20 eluting with petroleum ether–CHCl<sub>3</sub>–MeOH (2:1:1) and semi-preparative HPLC with a gradient of MeOH–H<sub>2</sub>O as eluent to afford compounds **38** (6.8 mg) and **19** (9.8 mg). C14-5-7 was further purified by Sephadex LH-20 (petroleum ether–CHCl<sub>3</sub>–MeOH, 2:1:1) to obtain compound **24** (21 mg). Fraction C15 was subjected to silica gel CC eluting with a gradient of CHCl<sub>3</sub>–MeOH (25:1 to 10:1) and further separated by chromatographed over silica gel with CHCl<sub>3</sub>–Me<sub>2</sub>CO (5:1) and Sephadex LH-20 (CHCl<sub>3</sub>–MeOH, 6:4) to obtain compound **37** (69 mg).

The EtOAc layer (50 g) was fractionated on silica gel CC eluting with a gradient of CHCl<sub>3</sub>–MeOH (10:1 to 0:1) to yield 10 fractions, E1–E10. Fraction E3 was separated by silica gel CC with CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O (16:1:0.1) to give compound **36** (2.5 mg). Fraction E5 was subjected to silica gel CC eluting with CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O (15:1:0.1, 10:1:0.1) to yield E5-1–E5-4, and E5-4 was separated on Sephadex LH-20 eluting with MeOH to yield compound **33** (8 mg). E5-5 was further purified by semi-preparative HPLC with MeOH–H<sub>2</sub>O as eluent to obtain compound **40** (8.2 mg). Fraction E7 yielded a yellow powder when standing overnight, and then the yellow powder was further purified to obtain compound **26** (62 mg). The residue of Fraction E7 was chromatographed on silica gel CC eluting with CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O (9:1:0.1, 8:1:0.1, 5:1:0.1), and separated by semi-preparative HPLC with a gradient of MeOH–H<sub>2</sub>O as eluent to afford compounds **27** (6.7 mg), **28** (7 mg), and **29** (41 mg). Fraction E8 was separated on silica gel CC eluting with CHCl<sub>3</sub>–MeOH–H<sub>2</sub>O (8:1:0.1, 5:1:0.1) to give nine subfractions, E8-1–E8-9, and then E8-7 was further fractionated on silica gel CC (CH<sub>2</sub>Cl<sub>2</sub>–isopropanol–H<sub>2</sub>O, 4:1:0.05 to 2:1:0.05) and purified by Sephadex LH-20 with MeOH as eluent to yield compound **32** (13 mg).

The *n*-BuOH layer (400 g) was fractionated on silica gel CC eluting with a gradient of CHCl<sub>3</sub>–MeOH (7:1 to 0:1) to yield 14 fractions, B1–B14. Fraction B5 yielded a white powder, collected by

filtering, which was then purified by washing with MeOH to obtain compound 43 (120mg). Fraction B6 was further separated on silica gel CC eluting with CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O (5:1:0.1 to 2:1:0.1) to give nine subfractions, B6-1-B6-9. B6-7 was further divided by silica gel CC using CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O (5:1:0.1), and purified by Sephadex LH-20 eluting with MeOH to afford compound 42 (14 mg). Fraction B12 was separated on repeated silica gel CC eluting with CHCl<sub>3</sub>-MeOH-H<sub>2</sub>O (2:1:0.1, 1:1:0.1) and further purified by silica gel CC (CH<sub>2</sub>Cl<sub>2</sub>-isopropanol-H<sub>2</sub>O, 4:1:0.1) and semi-preparative HPLC with MeOH-H<sub>2</sub>O as eluent to yield compounds 30 (7 mg) and 31 (6 mg), respectively.

### 3. Anti-Inflammatory Activity Assay In Vitro

**Table S1.** Anti-inflammatory activity assay in vitro <sup>a</sup>.

Plate	Compound	OD Value <sup>b</sup>	I.R. (%)	Plate	Compound	OD Value <sup>b</sup>	I.R. (%)
1 <sup>#</sup>	control	0.643 ± 0.064		3 <sup>#</sup>	control	0.559 ± 0.044	
	model	0.923 ± 0.032			model	0.930 ± 0.079	
	Ginkgolide B <sup>c</sup>	0.712 ± 0.015 ***	75.4		Ginkgolide B <sup>c</sup>	0.649 ± 0.028 **	75.7
	Pet. extract <sup>d</sup>	0.861 ± 0.085	22.1		15	0.850 ± 0.020	21.6
	CHCl <sub>3</sub> extract <sup>d</sup>	0.708 ± 0.139 **	76.8		19	0.427 ± 0.024 ***	135.6 <sup>e</sup>
	EtOAc extract <sup>d</sup>	0.847 ± 0.099	27.1		27	0.832 ± 0.042	26.4
	<i>n</i> -BuOH extract <sup>d</sup>	0.867 ± 0.076	20.0		29	0.788 ± 0.044 *	38.3
	14	0.808 ± 0.069 ***	41.1		41	0.806 ± 0.013 *	33.4
	17	0.845 ± 0.029 **	27.9		control	0.710 ± 0.033	
	18	0.819 ± 0.023 **	37.1		model	1.046 ± 0.115	
2 <sup>#</sup>	control	0.619 ± 0.034		Ginkgolide B <sup>c</sup>	0.774 ± 0.025 **	80.9	
	Model	0.861 ± 0.053		2	0.915 ± 0.051	39.0	
	Ginkgolide B <sup>c</sup>	0.706 ± 0.015 **	64.1	4	0.811 ± 0.014 **	69.9	
	1	0.763 ± 0.048 *	40.5	4 <sup>#</sup>	22	0.879 ± 0.052 *	49.7
	5	0.799 ± 0.055	25.6	32	0.903 ± 0.041 *	42.6	
	7	0.721 ± 0.075 **	57.9	33	0.897 ± 0.010 *	44.4	
	8	0.726 ± 0.038 **	55.8	40	0.802 ± 0.055 **	72.6	
	23	0.795 ± 0.024	27.3	43	0.821 ± 0.096 **	66.9	
	24	0.777 ± 0.053 *	34.7	45	0.855 ± 0.042 **	56.9	
	25	0.737 ± 0.067 *	51.2				
	30	0.773 ± 0.064 *	36.4				
	42	0.786 ± 0.014 *	31.0				
	44	0.796 ± 0.142	26.9				
	46	0.693 ± 0.039 **	69.4				

<sup>a</sup> All samples were assigned to five different 96-well plates, 1<sup>#</sup>-4<sup>#</sup>; The Student's *t*-test for unpaired observations between model (stimulated by PAF alone) and control (cultured in medium alone) or tested samples was carried out to identify statistical differences; \* *p* < 0.05, \*\* *p* < 0.01, \*\*\* *p* < 0.001; *p* < 0.05 were considered as significantly different; All the compounds were tested at 10<sup>-5</sup> mol/L. <sup>b</sup> OD values were expressed as mean ± SD (for control and sample, *n* = 3; for model, *n* = 4). <sup>c</sup> Ginkgolide B, positive control. <sup>d</sup> Pet. extract, CHCl<sub>3</sub> extract, EtOAc extract, and *n*-BuOH extract represent the extract described in "Extraction and Isolation" (tested at 10 µg/mL). <sup>e</sup> The tests were repeated several times, and the results were reproducible.