



Supplementary Material

## Benzotriazine di-oxide prodrugs for exploiting hypoxia and low extracellular pH in tumors

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			Oxic IC <sub>50</sub>	Anoxic			PCR <sup>c</sup>	
Cmpd	Na	pНe	(µM)	IC50 (µM)	ACR <sup>b</sup>	PCR <sup>c</sup> oxic	anoxic	TME ratio <sup>d</sup>
CHL (1)	4	7.4	$6.34 \pm 0.88$	$5.05 \pm 0.38$	$1.26\pm0.20$			
		6.5	$0.8 \pm 0.17$	$0.85\pm0.06$	0.94 ±0.21	$7.93 \pm 2.01$	$5.94 \pm 0.61$	$7.46 \pm 1.16$
SN30000	3	7.4	$477 \pm 65$	$5.92 \pm 2.42$	$80.6\pm34.7$			
(2)		6.5	$190 \pm 42$	$6.29 \pm 1.79$	$30.2 \pm 10.9$	$2.51 \pm 0.65$	$0.94 \pm 0.47$	$75.8 \pm 23.9$
4a	4	7.4	$863 \pm 66$	$225 \pm 26$	$3.84 \pm 0.53$			
		6.5	$297 \pm 74$	97.6 ± 12.3	$3.04 \pm 0.85$	$2.91 \pm 0.76$	$2.31 \pm 0.39$	$8.84 \pm 1.30$
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Table S1. Cytotoxicity of chlorambucil, SN30000 and BTO acid **4a** against UT-SCC-74B cells by IC<sub>50</sub> assay after 4 h exposure at pHe 7.4 or 6.5 under oxic (20% O<sub>2</sub>) or anoxic (<0.01% O<sub>2</sub>) conditions.

Footnotes: <sup>a</sup> Number of experiments. <sup>b</sup> Anoxic cytotoxicity ratio = oxic IC<sub>50</sub>/anoxic IC<sub>50</sub>. <sup>c</sup> pH cytotoxicity ratio = IC<sub>50</sub> at pHe 7.4/ IC<sub>50</sub> at pHe 6.5. <sup>d</sup> Overall ratio = oxic IC<sub>50</sub> pHe 7.4 / anoxic IC<sub>50</sub> pHe 6.5. Values are means and errors are SEM. For ratios, the errors are root mean square SEM.



**Figure S1.** pKa dependence of selectivity of weak acids for extracellular acidosis. The ordinate is the ratio of intracellular concentration at extracellular pH (pHe) 6.5 to that at pHe 7.4. Squares represent the values under the assumption that pHi is 7.1 and is independent of pHe ( $\Delta$ pHi, the difference in pHi values at pHe 7.4 and 6.4 is zero, i.e. cells fully control pHi under acidosis) or that pHi = 7.3 at pHe 7.4 and pHi = 6.9 at pHe 6.5 ( $\Delta$ pHi = 0.4) as in the present study.



**Figure S2.** Effect of pH on the change in radical absorption at 490 nm upon one-electron reduction of **7a** (150 mM) by the CO<sub>2</sub>- radical anion following pulse radiolysis (3 Gy in 200 ns) of N<sub>2</sub>O-saturated solutions containing sodium formate (0.1 M) and buffers (10 mM). Data points, obtained ca. 20 ms after the pulse, are fitted to a sigmoidal curve yielding a pKa value of 5.69±0.15.



**Figure S3.** The radical anion was generated by pulse radiolysis of N<sub>2</sub>O-saturated solutions containing **7a** (150 mM), sodium formate (0.1 M), phosphate buffer (10 mM), pH 7.0. The plot shows the dependence on increasing radiation dose (radical concentration) of the reciprocal of the first half-life of decay of the radical anion of **7a** measured at 490 nm. The intercept of the plot on the ordinate gives  $k_1 = \ln 0.5 \times 1/(t_{0.5})$ , from which the first order rate constant  $k_1$  is estimated as  $78 \pm 4 \text{ s}^{-1}$ , while the gradient gives the second-order rate constant,  $k_2 = 3.78 \pm 0.73 \times 10^7 \text{ M}^{-1} \text{ s}^{-1}$ .



**Figure S4.** Dependence of the first-order rate constant measured at 490 nm for the decay of the radical anion of **7a** at pH 7.0, *k*, on the concentration of O<sub>2</sub> in solution following pulse radiolysis (10 Gy in 200 ns). Solutions contained **7a** (0.5 mM), sodium formate (0.1 M), phosphate buffer (5 mM), pH 7.0 and saturated with mixtures of O<sub>2</sub>/N<sub>2</sub>O. The second order rate constant, *k*O<sub>2</sub>, calculated from the gradient of the plot,  $kO_2 = 5.39 \pm 0.40 \times 10^6$  M<sup>-1</sup> s<sup>-1</sup>.



**Figure S5. (a)** EPR spectrum (average of 100, 2 min scans) obtained upon reduction of **7a** (17 mM) by sPOR protein (7 ng mL<sup>-1</sup>) in the presence of DMSO (2 M). The anaerobic solution at 310 K contained DETAPAC (100 mM), SOD (300 units mL<sup>-1</sup>), catalase (1500 units mL<sup>-1</sup>), glucose-6-phosphate dehydrogenase (13 units mL<sup>-1</sup>), glucose-6-phosphate (10 mM) and NADPH (1 mM). **(b)** Simulation of the spectrum fitted to the combination of two species, (i) a C-centred radical and (ii) an aryl-type radical, in the ratio 0.65:0.35. The hyperfine coupling constants (HFC) of (i) and (ii) are aH 3.7 G, aN 16.5 G and aH 4.3 G , aN 15.9 G respectively,. r = 0.980.



**Figure S6.** Growth of control (non-drug-treated) SiHa cells following seeding 1500 cells/well in 96well plates, exposure to hypoxia ( $0.2\% O_2$ ) for 24 h or anoxia ( $<0.01\% O_2$ ) for 6 h or continuous growth under oxia ( $20\% O_2$ ), followed by growth in fresh medium at pHe 7.4 for 5 days under oxia before staining with SRB. Values are mean and errors are SEM for 6 experiments. Differences between pHe values and gas phase O<sub>2</sub> concentrations were not significant (P < 0.05) by two-way ANOVA.



**Figure S7.** Protein binding of **7a** in plasma from NIHIII mice and fetal bovine serum (FBS) assessed by equilibrium dialysis using a 12-14KDa cut-off dialysis membrane (HTDialysis, Gales Ferry, CT) as described by Banker et al., *J. Pharm. Sci.* 92, 967-974. Briefly, 100  $\mu$ L samples of undiluted plasma, plasma diluted in PBS, and  $\alpha$ MEM with or without FBS containing **7a** at 100  $\mu$ M were dialysed against 100  $\mu$ L PBS or  $\alpha$ MEM (pH 7.4) in an incubator (Innova-42, New Brunswick Scientific, US ) at 37°C with shaking at 80 rpm for 6 h. Proteins were precipitated with 3 vol of ice-cold MeCN, the centrifuged supernatants were analysed by HPLC as described in Section 4.12.



**Figure S8.** IC<sub>50</sub> values of SN30000 and two BTO acids (**4a** and **7a**) in MDCK-II cells with forced expression of BCRP or Pgp. The cell lines were gifts from A/Professor James Paxton, University of Auckland. Cells were seeded at 300 cells/well in DMEM with 10% FBS in 96 well plates in an anaerobic chamber, allowed to attach for 2 h then exposed to drugs for 4 h under anoxia. Cells were then washed with fresh medium and grown for 5 days under aerobic conditions before staining with SRB. Values are mean ± SEM from 2 separate experiments.



**Figure S9**. Effect of phenylpyruvate (PPV, 100 or 500  $\mu$ M) on proliferation of SiHa cells following 4 h exposure under anoxia to PPV alone (A) or with cmpd **4a** at pHe 7.4 (B) or pHe 6.5 (C). 3000 cells/well were seeded in 96 well plates, and IC<sub>50</sub> values determined as in Figure S6. Each data point is mean ± SEM for three replicate cultures in the same experiment. There is some evidence in panel B for an effect of 500  $\mu$ M PPV on the cytotoxicity of **4a** at pHe 6.5, but PPV alone showed selective cytotoxicity at pHe 6.5 relative to pHe 7.4 (panel A) which makes this difficult to interpret. No effect of PPV was observed under non-toxic conditions (pHe 7.4, or 100  $\mu$ M PPV at pHe 6.5).