Supplementary information

PEGylated purpurin 18 with improved solubility: Potent compounds for photodynamic therapy of cancer

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1. CHEMICAL ANALYSIS

1.1. NMR spectra



Figure S1: ¹H-NMR spectra of compound 2



Figure S2: ¹H-NMR spectra of compound 3



Figure S3-1: ¹H-NMR spectra of compound 4



Figure S3-2: ¹³C-NMR spectra of compound 3

¹³C NMR (101 MHz, CD₃OD) δ ppm: 8.82, 10.77, 10.90, 16.03, 17.94, 23.22, 27.34, 31.98, 33.92, 33.96, 38.99, 39.11, 39.75, 46.42, 54.74, 66.69, 69.00, 69.50, 69.64, 69.68, 70.01, 70.04, 78.60, 91.05, 95.47, 104.00, 108.75, 112.83, 120.01, 129.21, 132.98, 134.12, 140.22, 141.42, 142.95, 143.86, 145.01, 147.24, 149.32, 150.04, 157.34, 160.36, 165.82, 170.41, 174.08, 174.37, 174.47.

¹³C-NMR of compound 4

¹³C NMR (101 MHz, CD₃OD) δ ppm: *Major signals were determined from HSQC:* 8.10, 9.76, 9.78, 15.28, 17.21, 22.11, 28.24, 30.91, 30.92, 32.88, 37.94, 46.96, 45.56, 53.74, 57.89, 65.66, 68.85, 69.00, 69.51, 69.64, 69.90, 94.65, 103.44, 108.61, 119.14, 128.15. The carbons of methyls from Boc group at 27.34 ppm disappeared.

1.2.HRMS spectra



Figure S4: HRMS spectra of compound 2



Figure S5: HRMS spectra of compound 3



Figure S6-1: HRMS spectra of compound 4





Figure S6-2: Absorption, excitation and emission spectra of compounds **3** and **4**. Absorption spectra were measured in **A**) CHCl₃-MeOH, 8/2, **B**) phosphate buffered saline (PBS); amber color represents compound **3**, blue color represents compound **4**. **C**, **D**) Excitation and emission spectra were measured in CHCl₃-MeOH, 8/2. Each spectrum was the average of two repeated measurements. The emission and excitation wavelengths were following: **C**) 695 nm (green), 427 nm (amber, compound **3**), **D**) 694 nm (green), 426 nm (blue, compound **4**).



Figure S6-3: Depletion of 9,10-anthracenediyl-bis(methylene)dimalonic acid (**AB**, $7 \cdot 10^{-5}$ M) without presence of photosensitizer-generated singlet oxygen in Dulbecco's Modified Eagle Medium with 10% fetal bovine serum. The experiments were triplicated. (A) solution exposed to light, (B) solution kept in dark. $c_{rel,AB}$ – relative concentration of **AB** (actual concentration with respect to concentration at experiment start).



Figure S6-4: Depletion of 9,10-anthracenediyl-bis(methylene)dimalonic acid (**AB**, $7 \cdot 10^{-5}$ M) with **RB**-generated singlet oxygen in (A) PBS (concentrations of **RB** were: $1.3 \cdot 10^{-7}$ M, $2.6 \cdot 10^{-7}$ M, $3.1 \cdot 10^{-7}$ M) and (B) Dulbecco's Modified Eagle Medium with 10% fetal bovine serum (concentrations of **RB** were $2.6 \cdot 10^{-7}$ M, $5.1 \cdot 10^{-7}$ M, $7.5 \cdot 10^{-7}$

⁷ M). The experiments were duplicated. $\blacksquare \bullet \land$ – solution exposed to light, $\Box \circ \land$ – solution kept in dark.

 $c_{rel,AB}$ – relative concentration of AB (actual concentration with respect to concentration at experiment start)

2. BIOLOGICAL ANALYSIS



Figure S7: Fluorescence microscopy images of the intracellular localization of purpurin 18 (compound 1) and its derivatives (compounds 2–4) at 0.5 μ M concentration in human cancer cell lines of LNCaP (prostate carcinoma) and HaCaT (keratinocytes) after 24 h incubation. In the first and third column, there are bright field images; in the second and fourth column, there is compound localization. The scale bars represent 20 μ m.



Figure S8: Fluorescence microscopy images of purpurin 18 (compound 1) and its derivatives (compounds 2–4) localization in the endoplasmic reticulum of human MCF-7 cells (breast carcinoma). Colocalization of compounds 1–2 (0.5 μ M, 24 h) or compounds 3–4 (0.5 μ M, 24 h) with ER-TrackerTM Blue-White DPX (70 nM, 30 min.). A, E, I, M) Bright-field images; B, F, J, N) localization of the tested compounds; C, G, K, O) ER-TrackerTM Blue-White DPX; D, H, L, P) merge of the fluorescent images. The scale bars represent 20 μ m.



Figure S9: Fluorescence microscopy images of purpurin 18 (compound 1) and its derivatives (compounds 2–4) localization in the endoplasmic reticulum of human immortalized keratinocytes (HaCaT cells). Colocalization of compounds 1–2 (1 μ M, 24 h) or compounds 3–4 (0.5 μ M, 24 h) with ER-TrackerTM Blue-White DPX (70 nM, 30 min.). A, E, I, M) Bright-field images; B, F, J, N) localization of the tested compounds; C, G, K, O) ER-TrackerTM Blue-White DPX; D, H, L, P) merge of the fluorescent images. The scale bars represent 20 μ m.



Figure S10: Fluorescence microscopy images of purpurin 18 (compound 1) and its derivatives (compounds 2–4) localization in the endoplasmic reticulum of human LNCaP cells (prostate carcinoma). Colocalization of compounds 1–2 (0.5 μ M, 24 h) or compounds 3–4 (0.5 μ M, 3 h) and ER-TrackerTM Blue-White DPX (70 nM, 30 min.). A, E, I, M) Bright-field images; B, F, J, N) localization of the tested compounds; C, G, K, O) ER-TrackerTM Blue-White DPX; D, H, L, P) merge of the fluorescent images. The scale bars represent 20 μ m.



Figure S11: Fluorescence microscopy images of purpurin 18 (compound 1) and its derivatives (compounds 2–4) localization in the mitochondria of human MCF-7 cells (breast carcinoma). Colocalization of compounds 1–2 (0.5 μ M, 24 h) or compounds 3–4 (0.5 μ M, 24 h) with a mitosensor (70 nM, 10 min.) based on a dimethinium salt according to Bříza *et al.*¹. A, E, I, M) Bright-field images; B, F, J, N) localization of the tested compounds; C, G, K, O) mitosensor; D, H, L, P) merge of the fluorescent images. The scale bars represent 20 μ m.



Figure S12: Fluorescence microscopy images of purpurin 18 (compound 1) and its derivatives (compounds 2–4) localization in the mitochondria of human LNCaP cells (prostate carcinoma). Colocalization of compounds 1-2 (0.5 μ M, 24 h) or compounds 3–4 (0.5 μ M, 24 h) with MitoTrackerTM Green FM (70 nM, 20 min.). A, E, I, M) Bright-field images; B, F, J, N) localization of the tested compounds; C, G, K, O) MitoTrackerTM Green FM; D, H, L, P) merge of the fluorescent images. The scale bars represent 20 μ m.



Figure S13: Fluorescence microscopy images of purpurin 18 (compound 1) and its derivatives (compounds 2–4) localization in the mitochondria of human keratinocytes HaCaT. Colocalization of compounds 1–2 (0.5 μ M, 3 h) or compounds 3–4 (0.5 μ M, 3 h) and MitoTrackerTM Green FM (70 nM, 20 min.). A, E, I, M) Bright-field images; B, F, J, N) localization of the tested compounds; C, G, K, O) MitoTrackerTM Green FM; D, H, L, P) merge of the fluorescent images. The scale bars represent 20 μ m.



Figure S14: Fluorescence microscopy images of compound **4** localization in the Golgi apparatus in human PC-3 and MCF-7 cells. Colocalization of compound **4** (0.5 μ M, 24 h) and CellLight Golgi-GFP, BacMam 2.0 (2·10⁴ particles per cell.). A, E) Bright-field images; B, F) localization of compound **4**; C, G) CellLight Golgi-GFP, BacMam 2.0; D, H) merge of the fluorescent images. The scale bars represent 20 μ m.



Figure S15: Fluorescence microscopy images of compound **4** localization in lysosomes of human immortalized keratinocytes (HaCaT cells). Colocalization of compound **4** (0.5 μ M, 24 h) and LysoTracker Green DND-26 (70 nM, 20 min.). A) Bright-field images; B) localization of compound **4**; C) LysoTracker Green DND-26; D) merge of the fluorescent images. The scale bars represent 20 μ m.



Figure S16: Corrected total cell fluorescence (CTCF) of compounds 1-4 (1 µM, 24 h) localized in PC-3 cells

Table S1: Dose-dependent mechanisms of cell death in MCF-7 cells induced by compounds **1–4** after 24 h treatment and light induction (Light) measured by flow cytometry. Control represents untreated cells and cells incubated with the same compounds without illumination (Dark). The total light dose was of 4 J·cm⁻²

	Dark		rk	Light	
Compound	Concentration [µM]	Apoptosis	Necrosis	Apoptosis	Necrosis
Control	0	9.5 ± 1.0	0	10.6 ± 0.6	0
1	0.10	7.6 ± 1.4	0	11.0 ± 2.4	0.1±0.1
	0.25	8.5 ± 2.1	0	14.2 ± 2.7	0
	0.50	8.2 ± 1.1	0	32.6 ± 6.1	0.1±0
	1.00	7.3 ± 0.2	0	51.7±4.7	0.1 ± 0.1
2	0.10	10.5 ± 2.3	0	14.2 ± 2.1	0.1 ± 0.1
	0.25	12.1 ± 1.9	0.1 ± 0.1	18.2 ± 2.6	0
	0.50	12.8 ± 3.8	0	25.8 ± 4.0	0.4 ± 0.2
	1.00	14.3 ± 5.1	0	31.9 ± 4.1	0.3 ± 0.3
3	0.10	16.3 ± 3.5	1.7 ± 1.4	36.7 ± 12.1	0
	0.25	10.5 ± 0.8	0	56.1 ± 14.9	0
	0.50	13.7 ± 1.5	0	65.0 ± 9.5	0.1 ± 0
	1.00	21.2 ± 7.6	0	61.1 ± 7.6	0.3 ± 0.2
4	0.10	14.5 ± 3.7	0.1 ± 0.1	32.0 ± 2.6	0
	0.25	19.8 ± 7.7	0	49.8 ± 2.9	0
	0.50	19.0 ± 7.6	0.1 ± 0.1	57.8 ± 9.8	0
	1.00	22.4± 8.0	0.1 ± 0.1	68.0 ± 8.4	0



Figure S17: Photo- and dark toxicity of compounds **1–4** *in vitro*. Cell viability determined by WST-1 assay after 48 h of incubation with the tested compounds (24 h prior illumination + 24 h after illumination). Left panel dark toxicity (cells kept in the dark after the compound treatment), right panel phototoxicity (cell viability after compound photoactivation). A, B) HeLa, C, D) LNCaP, E, F) MCF-7 cells.



Figure S18: Photo- and dark toxicity of compounds **1–4** *in vitro*. Cell viability determined by WST-1 assay after 48 h of incubation with the tested compounds (24 h prior illumination + 24 h after illumination). Left panel dark toxicity (cells kept in the dark after the compound treatment), right panel phototoxicity (cell viability after compound photoactivation). A, B) MIA PaCa-2, C, D) PC-3, E, F) U-2 OS cells.

Flow cytometry – raw data
































Tube: Dark 1 100 nM			
Population	#Events	%Parent	%Total
All Events	10,000	####	100.0
	8,582	85.8	85.8
P2	7,147	83.3	71.5
- 🖂 Q1	0	0.0	0.0
🖂 necrosis	1	0.0	0.0
	6,557	91.7	65.6
🏧 🖂 apoptosis	589	8.2	5.9





Tube: Dark 1 500nM			
Population	#Events	%Parent	%Total
All Events	10,000	####	100.0
	8,585	85.9	85.9
	7,063	82.3	70.6
	0	0.0	0.0
🖂 necrosis	2	0.0	0.0
	6,654	94.2	66.5
🏧 🛛 apoptosis	407	5.8	4.1





Tube: Dark 2 T00nM			
Population	#Events	%Parent	%Total
All Events	10,000	####	100.0
	8,412	84.1	84.1
P2	6,881	81.8	68.8
	0	0.0	0.0
🖂 necrosis	5	0.1	0.0
	6,491	94.3	64.9
apoptosis	385	5.6	3.8





Tube: Dark 2 500nM			
Population	#Events	%Parent	%Total
All Events	10,000	####	100.0
	8,206	82.1	82.1
P2	6,687	81.5	66.9
- 🖂 Q1	0	0.0	0.0
🗆 🖂 necrosis	3	0.0	0.0
	6,325	94.6	63.2
🛄 🛛 apoptosis	359	5.4	3.6





Tube: Light control			
Population	#Events	%Parent	%Total
All Events	10,000	####	100.0
	6,626	66.3	66.3
	5,485	82.8	54.8
	0	0.0	0.0
🖂 necrosis	18	0.3	0.2
	4,874	88.9	48.7
🏧 🛛 apoptosis	593	10.8	5.9





Tube: Light I 250nM			
Population	#Events	%Parent	%Total
All Events	10,000	####	100.0
	7,595	75.9	75.9
P2	6,345	83.5	63.4
- 🖂 Q1	0	0.0	0.0
🗆 🖂 necrosis	9	0.1	0.1
	5,789	91.2	57.9
····· 🔀 apoptosis	547	8.6	5.5





Tube: Light I 1000nivi			
Population	#Events	%Parent	%Total
All Events	10,000	####	100.0
	8,207	82.1	82.1
P2	6,950	84.7	69.5
	0	0.0	0.0
	31	0.4	0.3
	5,804	83.5	58.0
apoptosis	1,115	16.0	11.2





Tube: Light 2 250nM			
Population	#Events	%Parent	%Total
All Events	10,000	####	100.0
	8,129	81.3	81.3
P2	6,733	82.8	67.3
🔀 Q1	0	0.0	0.0
- 🖂 necrosis	17	0.3	0.2
🖂 Q3	5,800	86.1	58.0
🔤 🖂 apoptosis	916	13.6	9.2





Tube: Light 2 1000nM			
Population	#Events	%Parent	%Total
All Events	10,000	####	100.0
	7,764	77.6	77.6
P2	6,751	87.0	67.5
- 🖂 Q1	0	0.0	0.0
🖂 necrosis	70	1.0	0.7
	5,143	76.2	51.4
apoptosis	1,538	22.8	15.4



















🔀 apoptosis

####

78.1

84.3

0

6,240

346

0.0

0.0

5.3

94.7

100.0

78.1

65.9

0.0

0.0

62.4





🔀 apoptosis

100.0

75.3

63.5

0.0

0.0

2.8

60.7

4.5





🔀 apoptosis

100.0

73.1

62.6

0.0

0.0

2.8

59.8

4.5





🔀 apoptosis

####

76.1

83.9

0.0

0.0

9.5

90.5

5,778

608

100.0

76.1

63.9

0.0

0.0

57.8





🔀 apoptosis

####

75.9

91.7

0

5,608

1,354

0.0

0.0

80.6

19.4

100.0

75.9

69.6

0.0

0.0

56.1





Tube: Light3500nM			
Population	#Events	%Parent	%Tota
All Events	10,000	####	100.0
	2,940	29.4	29.4
	2,832	96.3	28.3
- 🔀 Q1	0	0.0	0.0
🖂 necrosis	0	0.0	0.0
	1,273	45.0	12.7
🔤 🖂 apoptosis	1,559	55.0	15.6





ube. Light 4 roomvi			
opulation	#Events	%Parent	%Total
All Events	10,000	####	100.0
	7,366	73.7	73.7
	6,728	91.3	67.3
	0	0.0	0.0
🖂 necrosis	0	0.0	0.0
	5,216	77.5	52.2
🔤 🖂 apoptosis	1,512	22.5	15.1





🛛 apoptosis

10 ² 10 ³ 10 ⁴ 10 ⁵ 488-530/30-A			
:: Light 4 500nM			
ation	#Events	%Parent	%Total
Events	10,000	####	100.0
P1	4,521	45.2	45.2
- P2	4,112	91.0	41.1
- 🖂 Q1	0	0.0	0.0
- 🖂 necrosis	0	0.0	0.0
5 7			

1,535

2,577

37.3

62.7

15.4

22-8-2019-Light 4 1000nM				
Q1	necrosis			
29 29 29 29 29 20 20 20 20 20 20 20 20 20 20 20 20 20	apoptosis			
222				
$\begin{bmatrix} 4 3 6 6 6 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7 7$				
488	-530/30-A			
Tube: Light 4 1000nM				
Population		#Events	%Parent	%Total
All Events		10,000	####	100.0
En P1		5,101	51.0	51.0
P2		4,465	87.5	44.6
- 🖸 Q1		0	0.0	0.0
		0	0.0	0.0
		3,633	81.4	36.3
i 🛛 apoptosis		832	18.6	8.3










































References

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