Supplementary Material

Synthesis of bisphenol neolignans inspired by honokiol as antiproliferative agents

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Preliminary experiments for bromination

2-Methoxy-4-propylphenol (10a) has been subjected to preliminary reactions as reported:

1) compound **10a** (21 mg; 126 μ mol) was solubilized in CH₃CN (700 μ L) and treated with I₂ (7 mg; 20%) and *N*-bromosuccinimide (NBS; 31.5 mg; 175 μ mol) at rt;

2) compound **10a** (21 mg; 126 μ mol) was solubilized in CHCl₃ (700 μ L) and treated with I₂ (7 mg; 20%) and NBS (31.5 mg; 175 μ mol) at rt;

3) compound **10a** (21 mg; 126 μ mol) was solubilized in CH₃CN (700 μ L) and treated with AlCl₃ (7 mg; 40%) NBS (31 mg; 170 μ mol) at rt;

4) compound **10a** (21 mg; 126 μ mol) was solubilized in CHCl₃ (700 μ L) and treated with AlCl₃ (7 mg; 40%) and NBS (31.5 mg; 175 μ mol) at rt;

5) the phenol **10a** (21 mg; 126 μ mol) solubilized in CH₃CN (700 μ L) was mixed with AlCl₃ (7 mg; 40%) and Br₂ (10 μ L; 210 μ mol) at rt;

6) the phenol 10a (21 mg; 126 μmol) was solubilized in CHCl₃ (700 μL) and treated with Br₂ (10 μL ; 210 μmol) at rt;

7) the phenol **10a** (21 mg; 126 μ mol) was solubilized in CHCl₃ (700 μ L) and treated with Br₂ (10 μ L; 210 μ mol) at 0 °C;

8) the phenol **10a** (21 mg; 126 μ mol) was solubilized in acetone (500 μ L) and treated with NaBr (26.1 mg; 252 μ mol) and a solution of oxone (100.2 mg) in water (500 μ L) at -10 °C.

The mixtures were stirred at room temperature and monitored by TLC (85:15 *n*-hexane/acetone) for 6 h. Then each mixture was diluted with CH₂Cl₂ (1 mL) and partitioned with a saturated Na₂S₂O₃ solution. The organic layer of experiment 7 was purified on silica gel column chromatography (cyclohexane:EtOAc 98:2 \rightarrow cyclohexane:EtOAc 96:4) to give **11a**. The pure product was used to create a calibration curve via HPLC-UV, to determine the yield of other reactions (see Table 1).

Preliminary experiments for Suzuki-Miyaura cross-coupling

Preliminary experiments for S-M reaction were performed employing compound **11a** as starting material (8.3 mg, 50 μ mol) in presence of 4-hydroxyphenylboronic acid (10.3 mg, 75 μ mol), dppf (8.3 mg, 15 μ mol), Pd(OAc)₂ (1.1 mg, 5 μ mol) as catalyst and K₂CO₃ (34.6 mg, 250 μ mol). The solvent and temperature were varied as reported:

- 1) the reaction was carried out in THF (500 μ L) at 25 °C;
- 2) the reaction was carried out in THF (500 μ L) at 70 °C;
- 3) the mixture was stirred in THF:H₂O 10:1 (500 μ L and 50 μ L) at 70 °C;
- 4) the mixture was stirred in THF:H₂O 10:1 (910 μ L and 90 μ L) at 70 °C;
- 5) the reaction was carried out in 1,4-dioxane (500 μ L) at 70 °C;
- 6) the reaction was carried out in 1,4-dioxane (500 μ L) at 180 °C.

The course of the reactions was followed by TLC for 24h, then they were partitioned between H₂O:EtOAc ($3 \times 1 \text{ mL}$). The organic layer obtained from experiment 4) was purified on silica gel column chromatography (petroleum ether \rightarrow petroleum ether:acetone 92:8) to furnish the product **12a**. The pure bisphenol was used to create a calibration curve via HPLC-UV, to determine the yield of other reactions (see Table 2).



Figure S1. HRESIMS (+) spectrum of **6**.



Figure S2. ¹H NMR spectrum (500 MHz, CDCl₃) of 6.



Figure S3. ¹³C NMR spectrum (125 MHz, CDCl₃) of 6.



Figure S4. HRESIMS (-) spectrum of 12a.



Figure S5. ¹H NMR spectrum (500 MHz, CDCl₃) of **12a**.



Figure S6. ¹³C NMR spectrum (125 MHz, CDCl₃) of **12a**.



Figure S7. gHSQC spectrum of 12a.



Figure S8. gHMBC spectrum of 12a.



Figure S9. HRESIMS (-) spectrum of 12b.



Figure S10. ¹H NMR spectrum (500 MHz, (CD₃)₂CO) of 12b.



Figure S11. ¹³C NMR spectrum (125 MHz, (CD₃)₂CO) of 12b.



FigureS12. gCOSY spectrum of 12b.



FigureS13. gHMBC spectrum of 12b.



FigureS14. HRESIMS (-) spectrum of 12c.



Figure S15. ¹H NMR spectrum (500 MHz, (CD₃)₂CO) of 12c.



Figure S16. ¹³C NMR spectrum (125 MHz, (CD₃)₂CO) of 12c.



Figure S17. gCOSY spectrum of 12c.



Figure S18. gHSQC spectrum of 12c.



Figure S19. gHMBC spectrum of 12c.



Figure S20. HRESIMS (+) spectrum of 13a.



Figure S21. ¹H NMR spectrum (500 MHz, CDCl₃) of 13a.



Figure S22. ¹³C NMR spectrum (125 MHz, CDCl₃) of 13a.



Figure S23. gCOSY spectrum of 13a.



Figure S24 gHMBC spectrum of 13a.



Figure S25 HRESIMS (+) spectrum of 13b.



Figure S26. ¹H NMR spectrum (500 MHz, CDCl₃) of 13b.



Figure S27. ¹³C NMR spectrum (125MHz, CDCl₃) of 13b.



Figure S28. gCOSY spectrum of 13b.



Figure S29. gHSQC spectrum of 13b.



Figure S30. gHMBC spectrum of 13b.



Figure S31. HRESIMS (-) spectrum of 13c.



Figure S32. ¹H NMR spectrum (500 MHz, (CD₃)₂CO) of 13c.



Figure S33. ¹³C NMR spectrum (125 MHz, (CD₃)₂CO) of **13c**.



Figure S34. gCOSY spectrum of 13c.



Figure S35. gHSQC spectrum of 13c.



Figure S36. gHMBC spectrum of 13c.



Figure S37. HRESIMS (-) spectrum of 14a.



Figure S38. ¹H NMR spectrum (500 MHz, CDCl₃) of 14a.



Figure S39. ¹³C NMR spectrum (125 MHz, CDCl₃) of 14a.



Figure S40. gCOSY spectrum of 14a.



Figure S41. gHMBC spectrum of 14a.



Figure S42. HRESIMS (-) spectrum of 14b.



Figure S43. ¹H NMR spectrum (500 MHz, CDCl₃) of 14b.



Figure S44. ¹³C NMR spectrum (125 MHz, CDCl₃) of 14b.



Figure S45. gCOSY spectrum of 14b.



Figure S46. gHSQC spectrum of 14b.



Figure S47. gHMBC spectrum of 14b.



Figure S48. HRESIMS (-) spectrum of 14c.



Figure S49. ¹H NMR spectrum (500 MHz, (CD₃)₂CO) of 14c.



Figure S50. ¹³C NMR spectrum (125 MHz, (CD₃)₂CO) of 14c.



Figure S51. gCOSY spectrum of 14c.



Figure S52. gHSQC spectrum of 14c.



Figure S53. gHMBC spectrum of 14c.



Figure S54. HRESIMS (-) spectrum of 15a.



Figure S55. ¹H NMR spectrum (500 MHz, CDCl₃) of **15a**.



Figure S56. ¹³C NMR spectrum (125 MHz, CDCl₃) of 15a.



Figure S57. gCOSY spectrum of 15a.



Figure S58. gHMBC spectrum of 15a.