

Review

# Natural Products Inhibitors of Monoamine Oxidases—Potential New Drug Leads for Neuroprotection, Neurological Disorders, and Neuroblastoma

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**Abstract:** Monoamine oxidase inhibitors (MAOIs) are an important class of drugs prescribed for treatment of depression and other neurological disorders. Evidence has suggested that patients with atypical depression preferentially respond to natural product MAOIs. This review presents a comprehensive survey of the natural products, predominantly from plant sources, as potential new MAOI drug leads. The psychoactive properties of several traditionally used plants and herbal formulations were attributed to their MAOI constituents. MAO inhibitory constituents may also be responsible for neuroprotective effects of natural products. Different classes of MAOIs were identified from the natural product sources with non-selective as well as selective inhibition of MAO-A and -B. Selective reversible natural product MAOIs may be safer alternatives to the conventional MAOI drugs. Characterization of MAO inhibitory constituents of natural products traditionally used as psychoactive preparations or for treatment of neurological disorders may help in understanding the mechanism of action, optimization of these preparations for desired bioactive properties, and improvement of the therapeutic potential. Potential therapeutic application of natural product MAOIs for treatment of neuroblastoma is also discussed.

**Keywords:** natural products; monoamine oxidases (MAO-A and -B); antidepressant; monoamine oxidase inhibitors; neurological disorders; neuroprotection; Parkinson's disease; neuroblastoma



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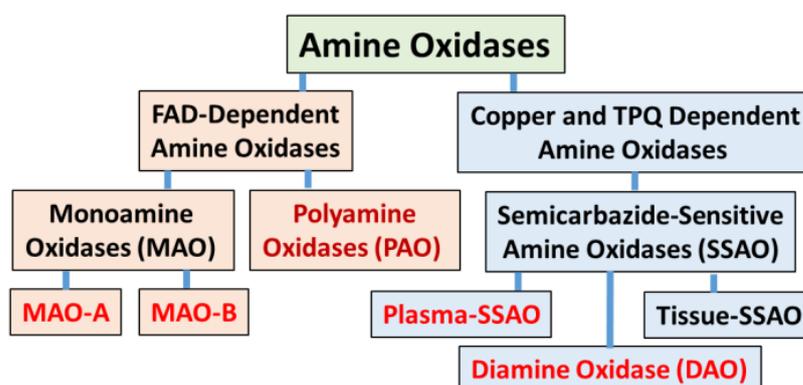
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## 1. Introduction

Amine oxidases are a heterogeneous group of enzymes that metabolize various monoamines, diamines, and polyamines produced endogenously for physiological functions or exogenous xenobiotic substances absorbed through dietary intake [1]. The amine oxidases are distinguished by their co-factor requirements and substrate specificities [2]. The flavin adenine dinucleotide (FAD)-dependent amine oxidases include mitochondrial monoamine oxidase A (MAO-A), monoamine oxidase B (MAO-B), and cytosolic polyamine oxidases (PAOs). Copper and topoquinone (TPQ)-dependent amine oxidases include plasma and tissue enzymes, also referred to as semicarbazide-sensitive amine oxidases (SSAOs) [3] (Figure 1). This review primarily focused on MAO-A and MAO-B due to their predominant role in oxidative deamination of endogenous neurotransmitter biogenic monoamines such as dopamine, epinephrine (EPI), and norepinephrine (NE) [4]. Changes in the physiological levels of these monoamines have been implicated in the pathophysiology of several neurological disorders.

Differential localization of MAO-A and -B in tissues determines their physiological functions. MAO-A and -B play an important role in deamination of biogenic amines in

neural and peripheral tissues [1,5]. MAO-A is more predominant in peripheral tissues such as the intestine, liver, lungs, and placenta and protects the body by oxidation of biogenic monoamines amines in the blood or by preventing the entry of dietary monoamines into circulation [6,7]. MAO-B plays a similar protective role in the micro vessels, acting as a metabolic barrier. In the peripheral and central nervous systems, intra-neuronal MAO-A and -B protect neurons from exogenous amines, regulate the contents of intracellular amine stores, and control pharmacological actions of amine neurotransmitters.

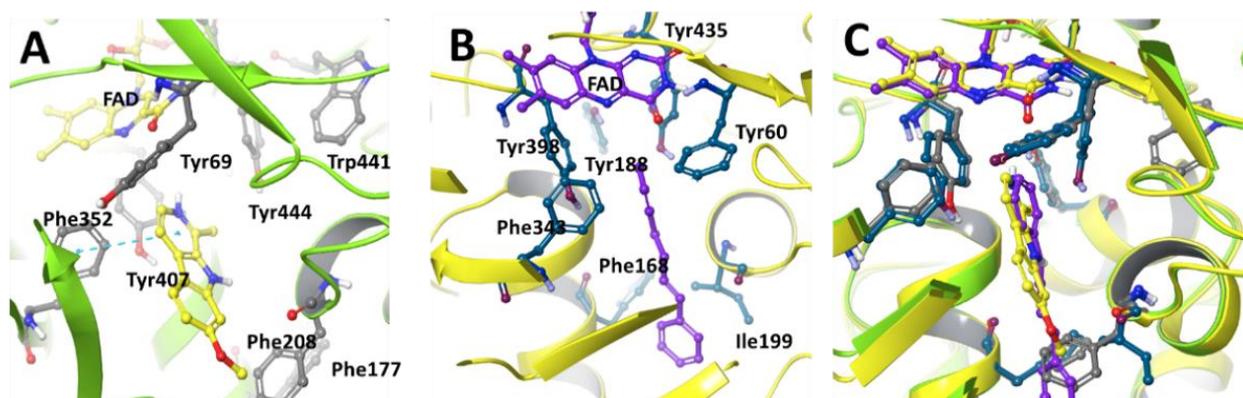


**Figure 1.** General classification of mammalian amine oxidases.

## 2. Structural Difference of MAO-A and MAO-B

MAOs, mitochondrial FAD-containing enzymes, exist as one of two functionally and structurally distinguishable isoforms, either MAO-A or MAO-B. Both isoforms are expressed in all tissues in different ratios. MAO-A is predominantly expressed in the heart, adipose tissue, and skin fibroblast and MAO-B is mainly found in platelets and lymphocytes, but the kidney, liver, and brain express both type of isozymes [8,9]. MAO-A and -B are distinguished by different substrate specificities and inhibitor sensitivities. MAO-A, selectively inhibited by clorgylin, preferentially deaminates norepinephrine and serotonin (5-HT). MAO-B, selectively inhibited by deprenyl and pargyline, metabolizes phenylethylamine and benzylamine. The three-dimensional crystal structures of human MAO-A and MAO-B with their specific inhibitors show significant similarities in their crystalline structure forms (Figure 2). However, important differences can be described in the oligomeric states and structures of substrate binding sites of MAO-A and -B [10,11]. In purified protein form, human MAO-B is dimeric, whereas human MAO-A exists in a monomeric form [12]. The active binding site of human MAO-B is characterized by a hydrophobic dipartite cavity with a substrate entrance cavity ( $\sim 290 \text{ \AA}^3$ ) connected with a larger substrate binding cavity ( $\sim 420 \text{ \AA}^3$ ). The Ile-199 was identified as an important structural component of the enzyme active site because it plays an important role in the accessibility of the second substrate cavity. In the closed conformation, Ile-199 is physically responsible for the separation of two cavities, whereas bulky ligands are present to adapt an open conformation. The binding of clinical inhibitors (R)-deprenyl and rasagiline induces a midspan type of cavity pushing Ile-199 into the open conformation. The active site of human MAO-A differs from human MAO-B in the loop conformation region with residues 108–118 and 210–216. Both these regions are critical components of the enzyme active site [12]. The structural differences incorporating Ile-335 in MAO-A vs. Tyr-326 in MAO-B active sites are responsible for the differential susceptibility of MAO-A and MAO-B toward selective MAO inhibitors [11]. Both human and rat MAO-A contain 16 conserved residues surrounding the substrate and inhibitor cavity. Only 6 of the 16 residues differ between human MAO-A and MAO-B. Substrate and inhibitor selectivity of human MAO-A and MAO-B are determined by Ile-335 and Tyr-326, respectively [13,14]. The selectivity of reversible inhibitors is determined by the size and shape of the substrate and inhibitor cavity, which are related to Ile-335 and Phe-208 in MAO-A and Tyr-326 and Ile-199 in

MAO-B. The differential inhibitor susceptibility of human MAO-A and -B is also attributed to their accommodation by the induced fit of Ile-335, which is similar to that observed for Ile-199 in MAO-B [15]. Neither human MAO-A nor MAO-B contain any disulfide bridges as determined by mass spectrometric analysis [8].



**Figure 2.** Active site configuration of human (A) MAO-A (PDB structure 2Z5X) with harmine and (B) MAO-B (PDB structure 1OJ9) with 1,4,-diphenyl butene, depicting key amino acids lining the enzyme active site. (C) Overlaid view of MAO-A and MAO-B substrate binding site. Harmine and 1,4,-diphenyl butene are selective reversible inhibitors of MAO-A and MAO-B, respectively.

### 2.1. MAO Inhibitors (MAOIs)

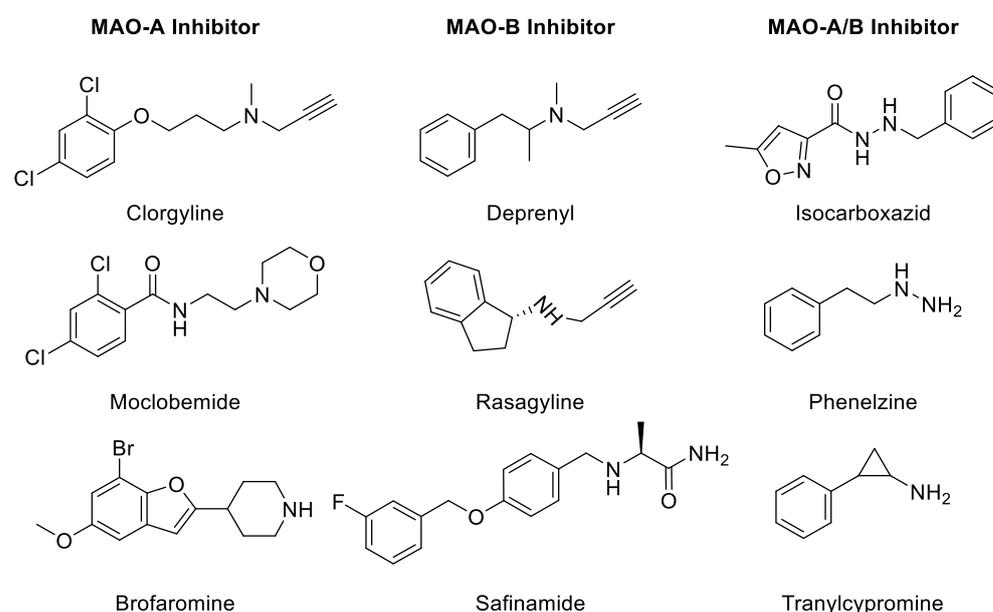
MAO inhibition prevents the degradation of monoamine neurotransmitters in the target cells and reinstates their physiological levels required for normal physiological functions [16]. Derangements in physiological levels and homeostasis of biogenic monoamines lead to pathophysiological consequences. The most important effect of MAO inhibition is a rapid increase in the intracellular concentration of monoamines. A rise in the concentrations of monoamines leads to secondary adaptive consequences, including a reduction in biosynthesis of monoamines via an apparent feedback mechanism, which is most clearly demonstrated for the noradrenergic system [17]. Both reversible as well as irreversible MAOIs have been developed for treatment of disorders caused due to depletion of biogenic amines in the target cells [18–20]. The MAOIs are classified into three types (Table 1): (1) irreversible non-selective inhibitors, such as phenelzine and tranylcypromine, (2) irreversible selective MAOI drugs, such as selegiline and rasagiline, and (3) reversible selective MAO-A inhibitors (RIMAs), such as moclobemide [21,22].

### 2.2. Therapeutic Applications of MAO Inhibitors

The MAOIs are included in a group of drugs, commonly referred as thymoleptic drugs, that favorably modifies mood in serious mood disorders and is primarily prescribed for the treatment of clinical depression or mania [23]. The MAO-A inhibitors show efficacy for treating anxiety and depression while the inhibition of MAO-B appears to be effective for prevention and treatment of Parkinson’s disease [24,25]. MAOIs are also used as a medicine for controlling hypertension or treating depression and other neurological disorders. Psychiatric disorders, such as obsessive-compulsive disorder, somatoform pain, panic disorder, and schizophrenia, have been reported to occasionally respond to treatment with MAOIs.

The progressive death of dopaminergic neurons results in a deficiency of dopamine during the development of Parkinson’s disease (PD), a neurodegenerative disorder of the brain. PD is characterized by a combination of rigidity movements, lack of movements, tremors, and postural instability. Selegiline, a propargylamine, is an irreversible inhibitor of MAO-B which inhibits dopamine metabolism and has been used in the treatment of PD effectively [26,27]. However, the therapeutic utility of selegiline is compromised due to the

generation of potential neurotoxic metabolites [28]. However, the neuroprotective effects of propargylamines in different neuronal models seem to be independent of inhibition of MAO-B. Glyceraldehyde-3-phosphate dehydrogenase (GAPDH), MAO-B, and/or other unknown proteins may be crucial targets in the survival of the injured neurons and may be crucial for the mechanism of neuroprotection by propargylamines. Further analysis of the mechanism(s) involved in the neuroprotective efficacy of MAO-B inhibitors may lead to the development of novel modalities for therapeutic applications. Selected MAO-A and -B inhibitors are listed in Figure 3 and Table 1. Historically, non-selective MAOIs have had in their structure a hydrazine group. The irreversibility of several MAOIs is caused by the triple bond scaffold which interacts directly with the active site and forms a covalent bond, irreversibly inactivating the enzyme (see Table 1) [2,5].



**Figure 3.** Chemical structure of selected MAO inhibitors.

**Table 1.** Monoamine oxidase inhibitors (MAOIs) classified on the basis of their selectivity [28,29].

Inhibitor Type	MAO-A	MAO-B	Non-Selective
Irreversible inhibitors	Clorgyline; Lilly 51641	(-) Deprenyl Lilly 54781 MDL 72145 AGN 1133 AFN 1135 Rasagiline Pargyline Selegiline	Phenelzine Tranylcypromine Isocarboxazid Nialamide Iproniazid Safrazine Metfendrazine
		Harmaline Amiflamine Cimoxatone Moclobemide Brofaromine Ro 11-1163 Toloxatone MD 780515 FLA 336(+)	Safinamide

### 3. MAO Inhibitors in Clinical Development

Several new MAOIs are currently under development and clinical evaluation for the treatment of neurological diseases [26]. Impaired monoamine-mediated neurotransmission is directly connected to neurological disorders such as depression and anxiety as well as susceptibility to stress [27]. The majority of the antidepressants presently used are designed to control one or both of the most important neurotransmitters of the brain (DA and serotonin (5HT)). Inhibition of their degradation to raise the concentrations in synaptic regions using MAOIs restores this function [30,31].

CX157 (3-fluoro-7-(2,2,2-trifluoroethoxy)-phenoxanthin 10, 10-dioxide) is currently under development for the treatment of major neurological disorders. CX157 causes potent and specific MAO-A inhibition in mammalian brain tissue [32]. MAO-A specifically deaminates serotonin, NE, and tyramine and is inhibited selectively at nanomolar concentrations of clorgyline, while MAO-B is insensitive to inhibition by clorgyline [33]. The first selective inhibitor of MAO-B was L-deprenyl (selegiline) [34] patented as an antidepressant and as a psychomotor energizer [35,36]. Advanced basic studies have suggested the therapeutic utility of selegiline for treatment of both Parkinson's disease and depression [37]. The evidence of selegiline having neuroprotective action is still followed and proven by the follow-up pro-pargylamino derivative, rasagiline, as shown recently in a study [37–39]. Researchers discovered isoxazole derivatives that have good MAO-B inhibition in the micromolar range. They also describe the synthesis and formulation of the molecule for MAO inhibition [28].

#### *Limitations of Currently Approved MAOIs and Current Approach for Development of New MAOIs*

MAOIs are powerful antidepressant agents and have been proven to reduce depression symptoms. The most commonly used drugs for neurological disorders are phenelzine, tricyclic antidepressants (TCAs), and benzodiazepines [40,41]. MAOIs are not recommended for use in combination with TCAs. MAOIs in clinical use have different side effects which can interfere with the treatment of depression. The most common adverse effects of MAOIs are nausea, diarrhea, insomnia, drowsiness, dizziness, headaches, and constipation, among others. MAOIs can potentially produce interactions including drug–drug, drug–food, and drug–herbal interactions. Use of antidepressant drugs such as bupropion, paroxetine, fluoxetine, nortriptyline, and amitriptyline should be avoided with MAOIs. MAOIs also show interactions with pain medications such as cyclobenzaprine, mirtazapine, meperidine, tramadol, and St. John's wort. Interactions between MAOIs and foods containing tyramine lead to hypertensive crisis. Hypertension is characterized by severe headache, high blood pressure, sweating, and nausea. To minimize the risk of hypertension effects, patients on MAOIs should take a low-tyramine diet and avoid foods which contain degraded protein such as aged cheeses, sauerkraut, tyramine-rich foods, and smoked meats [42,43]. Other side effects of MAOIs reported are weight gain, impaired sexual functioning, insomnia, and anticholinergic effects (dry mouth and constipation), which occur after long-term treatment with antidepressants [31,44–48]. First-generation MAOIs (non-selective and irreversible) may also cause serious side effects such as hepatotoxicity, orthostatic hypotension, and most importantly, hypertensive crisis that occurs following the consumption of tyramine-rich foods such as aged cheeses [42,49]. Patients using selegiline for Parkinson's disease in combination with levodopa can suffer with the side effects such as anorexia or nausea, dry mouth, dyskinesia, and orthostatic hypotension [49]. Hydrazine derivatives such as phenelzine used as an antidepressant drug have shown serious adverse effects, for example, liver toxicity, hypertensive crises, hemorrhage, and in some cases, death. Liver toxicity has been reported specifically with hydrazine-derived inhibitors. This led to the development of non-hydrazine MAOIs such as tranylcypromine and pargyline. However, hypertensive crises due to treatments with these MAOIs have continued to cause problems [50]. The greatest disadvantages of TCAs and benzodiazepines are overstimulation, heart palpitations, and sweating. Therefore, these drugs have been discontinued for clinical use [50,51].

Antidepressants acting as reversible inhibitors of MAO-A (also referred as RIMAs) have much less impact on the clinical psychopharmacology than other modern classes of medications, such as selective serotonin reuptake inhibitors (SSRIs). RIMAs are distinguished from the previously used MAOIs by their selectivity and reversibility [32]. As suggested with the use of irreversible MAOIs, dietary restrictions are not required during RIMA therapy and hypertensive calamities are relatively less common. Little evidence has emerged to suggest that RIMAs distribute older MAOIs' efficacy for treatment of depression characterized by prominent reverse neurodegenerative features [52]. Based on the available evidence, RIMAs appear to have a limited but important role in the differential therapeutics of depressive disorders [21].

#### 4. Traditional and Psychoactive Medicinal Plants and Herbal Formulations for Treatment of Neurological Disorders

The ancestral traditional medical systems worldwide have widely used medicinal plants for the treatment of different ailments, including neurological disorders. Recent review reports have described the use of psychoactive medicinal plants and herbal formulations for treatment of different neurological disorders [53–58]. Among the medicinal plants with neurological effects, a prominent example is *Banisteriopsis caapi*, a woody vine plant that generally grows in the Amazonian basin. *B. caapi* is an ingredient of the hallucinogenic and sacred drink popularly known as ayahuasca “aya”, a Quechua (South American language) word which means “vine of the dead”; “aya” is also known locally as “hoasca, caapi, oasca” (Brazil) and “yage” (Colombia) [59]. For the preparation of the sacred drink in healing purposes or divine exploration, *B. caapi* is utilized as an auxiliary plant along with primarily *Psychotria viridis* (chacruna) or *Diplopterys cabrerana* (oco yage). *B. caapi* contains beta-carbolines, which have therapeutic properties for neurological disorders [60–62]. Other notable examples are *Peganum harmala* (wild rue), *Rhodiola rosea* (roseroot), and *Crocus sativus* (saffron) for depression; *Passiflora incarnata* (passionflower), *Scutellaria lateriflora* (scullcap), *Ginkgo biloba* (gingko), and *Zizyphus jujuba* (jujube) for early dementia and anxiety disorders; and *Piper methysticum* (kava-kava) for phobic, panic, and obsessive-compulsive disorders [53,54]. Moreover, many species of mind-altering (psychodysleptic) plants have been used by humans throughout the planet to reach mind distortion states; among those, a few examples have been utilized for therapeutic aims, including *Cannabis sativa* (cannabis), *Tabernanthe iboga* (iboga), *psychotria viridis* (chacruna), and *Papaver somniferum* (opium poppy) [63].

MAO-A inhibitors have been demonstrated to be effective antidepressants, and MAO-B inhibitors are used in the treatment of neurodegenerative diseases including PD, as mentioned in Section 3 [35]. The mechanistic aspects of the potential therapeutic applications of MAO-B inhibitors in Alzheimer's disease were reviewed recently [35]. In this context, several medicinal plants have shown pharmacological effects similar to MAOIs. Recent review reports have summarized the medicinal plants as effective as MAOIs [63,64]. This review presents an extensive survey of the chemical properties of MAO inhibitory constituents identified from natural product sources presented as different chemical classes. This may be useful for further follow-up studies with these MAO inhibitory constituents as new drugs leads for structure activity analysis and further optimization of these leads. Utility of the natural product MAO inhibitory constituents as neuroprotective agents and the therapeutic application of natural products for neuroblastoma are also discussed.

Electronic searches were specifically conducted on the literature using major databases including PubMed, Google Scholar, SciFinder®, and Web of Science. The keywords used in the searches were a combination of the words “natural products”, “alkaloids”, “flavonoids”, “phenols”, “terpenes”, “monoamine oxidase inhibitors”, “MAO”, and “MAOI”. We mostly selected published reports that reported natural products or natural product derivatives with inhibition of MAO-A or-B with  $IC_{50} < 100 \mu M$ . These reports were further grouped into different chemical classes of natural product MAOIs. The natural product constituents tested against MAO-A and/or MAO-B are included in the data tables.

## 5. Different Classes of Natural Product MAOIs

Natural products are an important class of nature-based MAOIs. In the present scenario, several research labs are working to identify and characterize the class of natural products with medicinal value of identified lead compounds. In this review, we summarized different classes of natural product MAOIs. The natural products are generally classified based on chemical classes of secondary metabolites, such as alkaloids, flavonoids, coumarins, xanthenes, terpenoids, sterols, and phenolic compounds. Scientists believe that many more compounds of these classes still can be discovered in nature. These natural products and compounds are responsible for a plethora of therapeutic uses, including for treatment of neurological disorders. Various classes of natural product MAOIs are described below. Tables 2–7 include selected natural products, the natural source, their MAO-A or MAO-B  $IC_{50}$  values,  $K_i$  (when available), selective index (SI; MAO-A or -B), and the source of the enzyme used for MAO inhibition analysis.

### 5.1. Alkaloids

Alkaloids are “cyclic compounds containing nitrogen in a negative oxidative state” that are of limited distribution in living organisms [65]. Alkaloids are natural products of non-peptidic origin containing a nitrogen atom, and their basic characteristic is revealed in the name derived from alkaline that means basic. Being one of the most diverse kind of secondary metabolites discovered in living organisms, alkaloids present an array of chemical structures, biosynthetic pathways, and pharmacological activities. While alkaloids have been conventionally isolated from plants, an increasing amount remains to be found in animals, insects, microorganisms (bacteria and fungi), and marine sources (invertebrates and microorganisms) [66]. Several alkaloids have been reported for MAO-A and -B inhibitory activity. Herein, we describe some notable examples of alkaloid types that inhibit MAO enzymes. The chemical structures for the selected alkaloid MAOIs are shown in Figures 4 and 5. Generally, short alkaloids from *Evodia* are moderately selective toward MAO-B. Monoterpene alkaloids, tetra and pentacyclic berberine types, have shown to be selective to MAO-A inhibition, and quinolone and quinoline alkaloids are low- and high-selective MAO-B inhibitors, respectively. Crinine-type alkaloids have been proposed as MAO-A inhibitors; however, no MAO-B inhibition has been reported (Table 2, Figure 4). Harmine-type (beta-carbolines) alkaloids are highly selective toward MAO-A isoenzymes, and tetra and pentacyclic indole alkaloids are also highly potent MAO-A inhibitors; on the contrary, azepine-indole alkaloids are relatively selective toward MAO-B (Table 2, Figure 5).

#### 5.1.1. MAO Inhibitory Activity of Piperine, Quinolone, and Isoquinoline Alkaloids

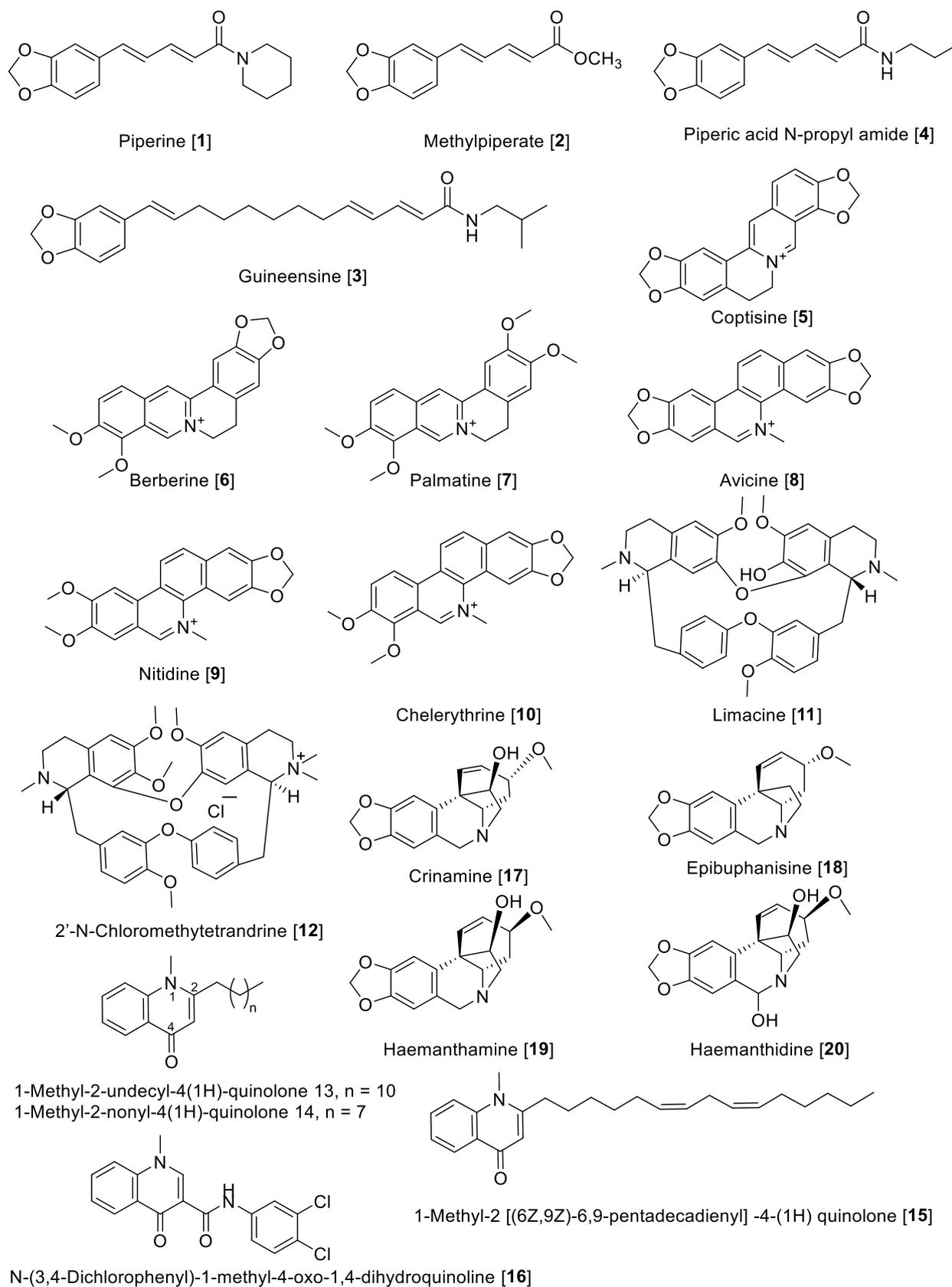
Piperine (**1**) alkaloid is the main component in black pepper (*Piper nigrum*) and in general in the *Piper* genus, which has a wide spectrum of pharmacological activities, including MAO inhibition. For example, from *Piper longum*, piperine **1** and methylpiperate **2** were isolated and their MAO inhibition was evaluated using rat brain mitochondrial fraction [67]. The enzyme inhibition kinetics study for piperine **1** showed that the mode of inhibition for MAO-A was mixed type with a  $K_i$  value of 35.8  $\mu$ M and for MAO-B was competitive with a  $K_i$  value of 79.9  $\mu$ M [68]. In another study, piperine **1** showed an inhibitory effect against MAO-A with an  $IC_{50}$  value of 20.9  $\mu$ M and for MAO-B with an  $IC_{50}$  value of 7.0  $\mu$ M. The MAO inhibition by piperine **1** was reversible as determined by the recovery of enzyme activity after removal of the inhibitor by dialysis of the incubation mixture [69]. The chemical structure of **1** (Figure 4 and Table 2) consists of a free phenolic –OH group, and it was proposed that the inhibition could probably be initiated by the hydrogen bonding of its amide active protons such as –NH–, –OH, and –SH in the active sites of MAO-A and -B [68]. Methylpiperate **2** was isolated from the fruits of *P. longum* and evaluated for inhibitory effects on the MAO-A activity on mouse brain homogenates yielding an  $IC_{50}$  value of 3.6  $\mu$ M; guineesine **3** showed moderate inhibition of total MAO with an  $IC_{50}$  of 139.2  $\mu$ M [69,70]. Several *N*-amide derivatives of piperine **1** were synthesized and tested for MAO inhibition. Piperic acid *N*-propyl amide **4** inhibited MAO isolated from rat brain

mitochondrial homogenate with an  $IC_{50}$  value of 45 nM for MAO-B and an  $IC_{50}$  value of 3.66  $\mu$ M for MAO-A [71].

Coptisine **5**, as well as other isoquinoline alkaloids, were isolated from *Coptis japonica*. The isolated alkaloids from *C. japonica* were evaluated for their inhibitory effect on MAO from mouse brain fraction. The result showed an inhibitory effect of **5** on MAO-A with an  $IC_{50}$  value of 1.8  $\mu$ M but none on the MAO-B enzyme [72]. Earlier reports have mentioned that various alkaloids including berberine **6** and palmatine **7** and **5** inhibited MAO enzymes [73,74]. Berberine **6** and palmatine **7** exhibit a non-competitive inhibition of MAO with  $K_i$  values of 44.2  $\mu$ M and 58.9  $\mu$ M, respectively [75].

Isoquinoline alkaloids including avicine **8**, nitidine **9**, and chelerythrine **10** (Figure 4) isolated from *Zanthoxylum rigidum* showed potent inhibition of human MAO-A with  $IC_{50}$  values of 0.41, 1.89, and 0.55  $\mu$ M, respectively [76]. Limacine **11** and 2'-*N*-chloromethyltetra-ndrine **12**, two benzylisoquinoline dimers isolated from the roots of *Stephania tetrandra*, were found to have moderate inhibitory effects on total MAO with  $IC_{50}$  values of 37.7 and 29.2  $\mu$ M, respectively [77].

Quinolone-type alkaloids also have been reported to possess MAO inhibition; in fact, 1-methyl-2-undecyl-4-(1H) quinolone **13** isolated from *Evodia rutaecarpa* competitively inhibited MAO-B with an  $IC_{50}$  value of 15.3  $\mu$ M but did not inhibit MAO-A [78]. *Evodia rutaecarpa* (Rutaceae) is a well-known traditional Chinese medicine with several therapeutic properties, including analgesic, antiemetic, astringent, hemostatic, uterotonic, cardiogenic, and antihypertensive activities [79]. Seven quinolone alkaloids were isolated from the fruits of *E. rutaecarpa* and their inhibitory effects against MAO enzymes were evaluated using mouse brain mitochondrial fraction. The results found that all compounds were more active against MAO-B compared to MAO-A, with 1-methyl-2-nonyl-4 (1H)-quinolone **14** and 1-methyl-2 [(6Z,9Z)-6,9-pentadecadienyl] -4-(1H) quinolone **15** being the most potent inhibitors for MAO-B with  $IC_{50}$  values of 2.3  $\mu$ M and 3.6  $\mu$ M, respectively [80]. Based on the quinolone base structure, efforts were made to modulate the activity; thus, the synthesis of a series of quinolone derivatives found *N*-(3,4-Dichlorophenyl)-1-methyl-4-oxo-1,4-dihydroquinoline-3 carboxamide **16** as a selective inhibitor of human MAO-B with a selective index (SI) of ~1887 and an  $IC_{50}$  value of 5.3 nM [81]. The crinine-type alkaloids crinamine **17** and epibuphanisine **18** (isolated from *Crossyne guttata*) and haemanthamine **19** and haemanthidine **20** (isolated from *Scadoxus puniceus*) showed inhibition of human MAO-B with  $IC_{50}$  values of 14.9, 39.2, 112.0, and 17.20 nM, respectively, with no inhibition of human MAO-A [82].



**Figure 4.** Chemical structure of selected alkaloid MAO inhibitors with piperine, quinolone, and isoquinoline skeletons.

### 5.1.2. MAO Inhibitory Activity of Beta-Carbolines and Indole-Type Alkaloids

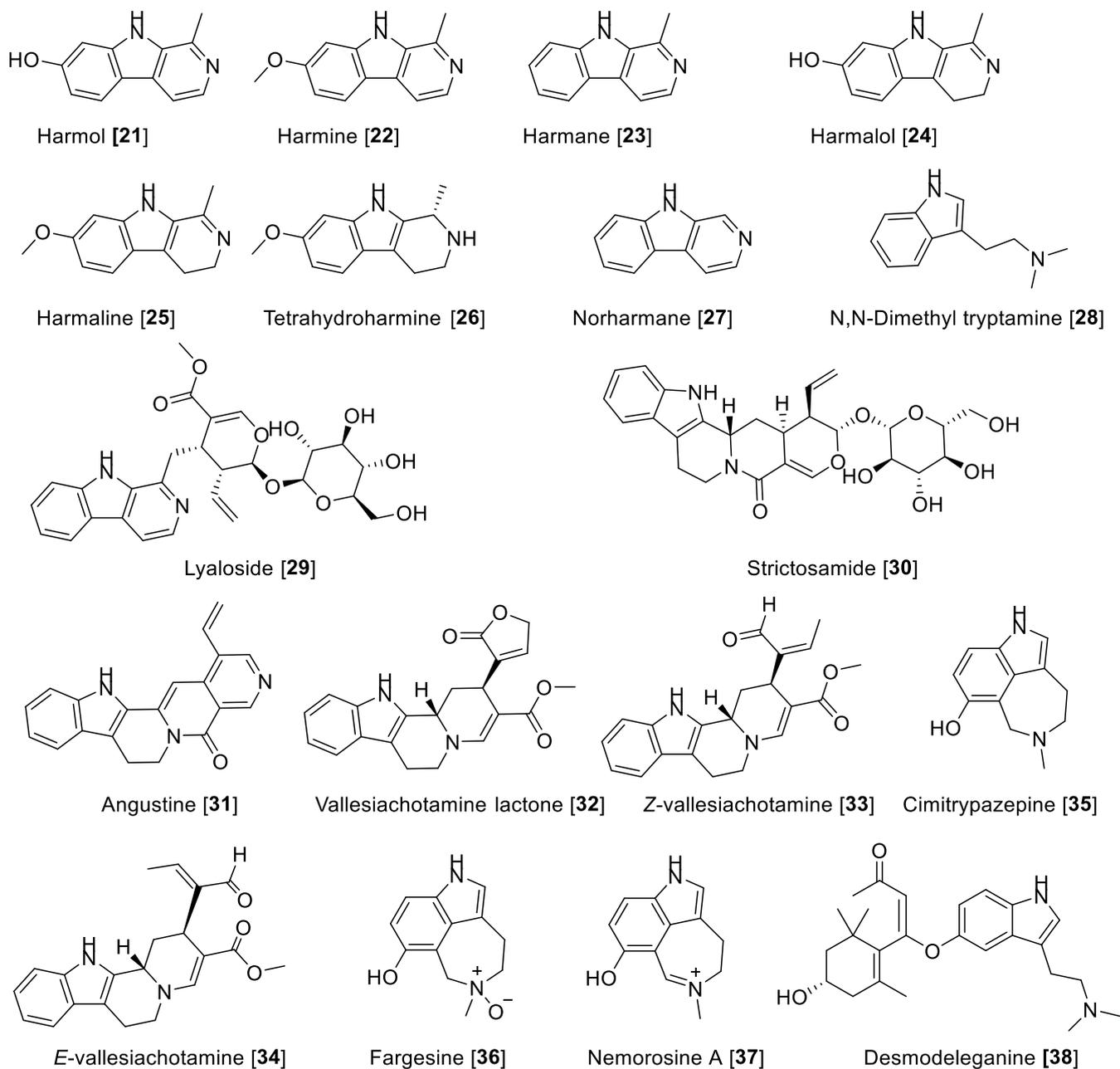
Beta-carbolines derived from indole are endowed with neuropharmacological and neuromodulating properties. Beta-carbolines have been described as potent MAO-A inhibitors [61]. Fernandez de Arriba et al. [83] described the first studies about the kinetic behavior of beta-carboline derivatives as MAO-A inhibitors in the bovine retina. It has also been found that the beta-carbolines harmol **21**, harmine **22**, and harmane **23** are the most potent MAO-A inhibitors, followed by the 3,4-dihydro-beta-carbolines harmalol **24**, harmaline **25**, 1,2,3,4-tetrahydro-beta-carboline (tetrahydro harmine **26**), and norharman **27** [84–86]. The MAO inhibitory activity of beta-carbolines present in the extract of *Banisteriopsis caapi*, a component of “Ayahuasca”, and in mixtures with different compounds or plant extracts was evaluated and also compared with other previously reported MAO inhibition data [87,88]. *B. caapi* extracts and two of its components, **22** and **25**, were tested for MAO inhibition using mouse liver homogenate. *B. caapi* extract and harmaline **25** showed a concentration-dependent inhibition for MAO-A with IC<sub>50</sub> values of 1.24 µg/mL and 4.54 nM, respectively, showing moderate inhibition of MAO-B [88,89]. Phytochemical and biological studies with the specimens of *B. caapi* collected from Hawaii found **22** and **24–26** as the main inhibitors of recombinant human MAO-A with IC<sub>50</sub> values of 2.0, 2.5, 18, and 74 nM, respectively. Only **22** and **25** were found to be moderately active toward recombinant human MAO-B with IC<sub>50</sub> values of 20 and 25 µM, respectively [60].

Harmane **23** was also isolated from the medicinal plant *Uncaria rhynchophylla* (Cat’s claw) and tested for its inhibitory effect on the total MAO activity of mouse brain homogenate. Harmane **23** was found to be a MAO-A inhibitor with an IC<sub>50</sub> value of 11.1 µM [90]. Structural and mechanistic studies have demonstrated that harmine **22** shows reversible inhibition and binds with the active site of the enzyme cavity. Compound **22** interacts with Tyr-69, Asn-181, Phe-208, Val-210, Gln-215, Cys-323, Ile-325, Ile-335, Leu-337, Phe-352, Tyr-407, Tyr-444, and FAD. The seven molecules of water occupy the gap between the inhibitor and these groups. The amide groups of the Gln-215 side chain tightly interact with **22** in human MAO-A or Gln-206 in human MAO-B (Gln-215/206). These results are consistent with previously reported structural analyses of human MAO-A and MAO-B by active site-directed mutagenesis studies [11,12]. The selectivity of the reversible inhibitors is caused by the different size and shape of the substrate and inhibitor cavity regulated by Ile-335 and Phe-208 in MAO-A, which corresponds to Tyr-326 and Ile-199 of MAO-B [91].

*Psychotria* (Rubiaceae) is a complex neotropical genus of remarkable interest due to the high content of alkaloids. *Psychotria viridis* is a component of the hallucinogenic beverage known as ayahuasca. The metabolite responsible for its hallucinogenic effects is the alkaloid *N,N*-dimethyl tryptamine (DMT) **28**, which is biosynthetically based on the indole scaffold. There are several other alkaloid skeletons produced by the *Psychotria* species, some of them with notable bioactivities [92]. The monoterpene indole alkaloids lyaloside **29**, strictosamide **30**, angustine **31**, vallesiachotamine lactone **32**, *E*-vallesiachotamine **33**, and *Z*-vallesiachotamine **34** (Figure 5 and Table 2) were isolated from *P. laciniata* and were found to be MAO inhibitors with a preference toward human MAO-A with IC<sub>50</sub> values of 182, 141, 1.10, 0.87, 2.14, and 0.85 µM, respectively, and negligible or low inhibition toward human MAO-B. These alkaloids were also selective inhibitors of butyrylcholinesterase, suggesting that this scaffold can be a multifunctional agent [93]. Recently, the azepine-indole alkaloids cimitrypazepine **35**, fargesine **36**, and nemorosine A **37** were isolated from *Psychotria nemorosa* among other alkaloids and found to inhibit both MAOs but preferentially human MAO-A with IC<sub>50</sub> values of 1.4, 1.4, and 0.9 µM, respectively [94]. Harmine **22** has been subjected to a clinical trial in combination with DMT **28** to study the network dynamics following the modulation of the serotonin system [95].

Desmodeganine **38**, a derivative of **28**, was found to inhibit both MAOs at micromolar concentrations. Compound **38** was isolated from the traditional Chinese plant “Sha MaHuang” [96]. Several other alkaloids have also been tested for their inhibitory effect on

MAO but have not shown potency of inhibition for MAO-A and -B. Table 2 presents  $IC_{50}$  values of selected alkaloid metabolites for inhibition of MAO-A and -B.



**Figure 5.** Chemical structure of selected alkaloid MAO inhibitors with indole motif.

Table 2. Alkaloid natural product inhibitors of MAO-A and MAO-B.

Compounds	Source	MAO-A		MAO-B		SI	Enzyme Source	References
		IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	MAOA/B		
Piperine [1]	<i>Piper longum</i>	49.3	35.8	91.3	79.9	0.54	A	[68]
	<i>Piper longum</i>	20.9	19.0	7.0	3.19	2.98	A	[69]
Methylpiperate [2]	<i>Piper longum</i>	27.1	23.5	1.6	1.3	16.93	A	[70]
piperic acid <i>N</i> -propyl amide [4]	Piperine derivative	3.66		0.045		81.3	B	[71]
Coptisine [5]	<i>Coptis japonica</i>	1.8	3.3				A	[72]
Avicine [8]	<i>Zanthoxylum rigidum</i>	0.41		>100		>0.0041	F	[76]
Nitidine [9]	<i>Zanthoxylum rigidum</i>	1.89		>300		>0.0063	F	[76]
Chelerythrin [10]	<i>Zanthoxylum rigidum</i>	0.55		>20		>0.0275	F	[76]
1-Methyl-2-undecyl-4-(1H)quinolone [13]	<i>Evodia rutaecarpa</i>	338.2		15.3	9.91	22.10	A	[78]
1-Methyl-2-nonyl-4(1H)-quinolone [14]	<i>Evodia rutaecarpa</i>	240.2		2.3		104.4	A	[80]
1-Methyl-2-[(6Z,9Z)-6,9-pentadecadienyl]-4-(1H)quinolone [15]	<i>Evodia rutaecarpa</i>	>400		3.6	3.8	111.1	A	[80]
Quinoline derivative [16]	Quinolone derivative	>100		0.0053		>18,867	F	[81]
Crinamine [17]	<i>Crossyne guttata</i>	0.014					F	[82]
Epibuphanisine [18]	<i>Crossyne guttata</i>	0.039					F	[82]
Haemanthamine [19]	<i>Scadoxus puniceus</i>	0.112					F	[82]
Haemanthidine [20]	<i>Scadoxus puniceus</i>	0.017					F	[82]
Harmol [21]	<i>Banisteriopsis caapi</i>	0.018					F	[61]
	<i>Banisteriopsis caapi</i>	0.5					F	[84]
	<i>Peganum harmala</i>	0.352					F	[86]
Harmine [22]	<i>Banisteriopsis caapi</i>	0.002		20		0.0001	F	[61]
	Diverse vendors	0.06		NR			F	[84]
	<i>Peganum harmala</i>	0.008		NR			F	[86]
	<i>Banisteriopsis caapi</i>	0.004		>10		>0.0004	A	[89]
Harmane [23]	<i>Banisteriopsis caapi</i>	0.64		NR			F	[84]
Harmalol [24]	<i>Banisteriopsis caapi</i>	0.66		NR			F	[84]
	<i>Peganum harmala</i>	0.48		NR			F	[86]
Harmaline [25]	<i>Banisteriopsis caapi</i>	0.002		25		0.00008	F	[61]
	Diverse vendors	0.09		NR			F	[84]
	<i>Peganum harmala</i>	0.012		NR			F	[86]

Table 2. Cont.

Compounds	Source	MAO-A		MAO-B		SI	Enzyme Source	References
		IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	MAOA/B		
Tetrahydro harmine [26]	Diverse vendors	1.52		NR			F	[84]
Norharmine [27]	Diverse vendors	4.29		NR			F	[84]
Lyaloside [29]	<i>Psychotria. Laciniata</i>	182		>100			F	[93]
Strictosamide [30]	<i>Psychotria laciniata</i>	141		>100			F	[93]
Angustine [31]	<i>Psychotria laciniata</i>	1.10		138		0.0079	F	[93]
Vallesiachotamine lactone [32]	<i>Psychotria laciniata</i>	0.87		34		0.025	F	[93]
<i>E</i> -vallesiachotamine [33]	<i>Psychotria laciniata</i>	2.14		120		0.017	F	[93]
<i>Z</i> -vallesiachotamine [34]	<i>Psychotria laciniata</i>	0.85		126		0.0067	F	[93]
Cimitrypazepine [35]	<i>Psychotria nemorosa</i>	22		1.4		15.71	F	[94]
Fargesine [36]	<i>Psychotria nemorosa</i>	27		1.4		19.28	F	[94]
Nemorosine A [37]	<i>Psychotria nemorosa</i>	31		0.9		34.4	F	[94]
Desmodeganine [38]	<i>Desmonium elegans</i>	9.33		10.16		0.91	F	[95]

Note: natural products tested on total MAO are not listed. Enzyme source: A = mouse brain crude mitochondrial fraction; B = rat brain mitochondrial MAOs; C = rat liver mitochondrial MAOs; D = mouse liver MAOs; E = human MAO-A and -B over-expressed; F = recombinant human MAO-A and -B.

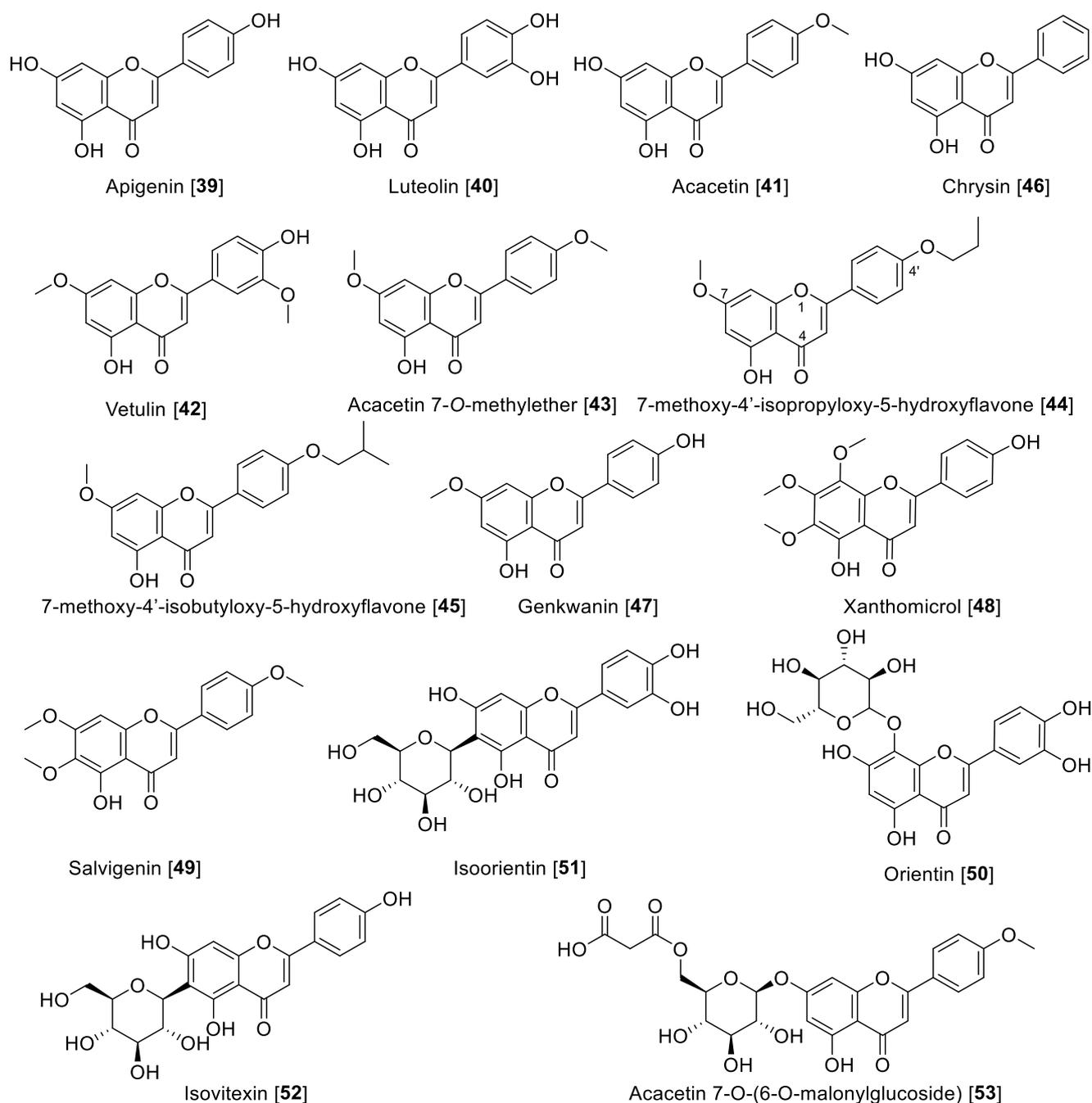
## 5.2. Flavonoids

Flavonoids are included in the polyphenol family, which are made up of two benzene rings and merged with a short three-carbon chain. These types of compounds are water-soluble polyphenolics containing 15 carbon atoms. One of the carbons of the short chain is always connected to a carbon of one of the benzene rings, either directly or through an oxygen bridge, thereby forming a third middle ring, which can be five or six-membered. Flavones, flavanols, flavanones, anthocyanins, isoflavones, and flavans are the subgroups in flavonoids [97], also including the precursors chalcones, neoflavonoids, and dimers and oligomers. Some flavonoids are responsible for the coloring in the plants, including herbs, fruits, vegetables, etc. The most important dietary sources of flavonoids are green tea, fruits, and vegetables. Green and black tea contain about 25% flavonoids. Fruits such as apple (quercetin) and citrus fruits (rutin and hesperidin) are rich sources of flavonoids [27,98]. Flavonoids are also known for their strong antioxidant properties; it is believed that flavonoids are integrated in a network in plants, activating their defenses to fight stress conditions [99,100]. Flavonoids have also been reported as MAO inhibitors. A series of reports has listed several flavonoids to possess MAO inhibition, including a comprehensive structure activity relationship (SAR) study [27,101,102]. Herein, we describe notable examples of several flavonoids that inhibit MAO, separated by class. Notably, flavones depending on the substituents can be MAO-A or -B inhibitors. Those substituted at the C-4' position at ring C are mainly MAO-B inhibitors, and those disubstituted at ring C or per-substituted at ring B-turn are relatively MAO-A selective (Table 3, Figure 6). Flavanols are selective MAO-A inhibitors, while isoflavones are reported to be moderately MAO-B selective (Table 3, Figure 6). The chemical structures for the selected flavonoids are shown in Figures 6–8. The MAO inhibition profiles including IC<sub>50</sub> values against MAO-A and MAO-B of these metabolites are presented in Table 3. The sources of MAO employed in these studies are also specified.

### 5.2.1. MAO Inhibitory Activity of Flavones

Apigenin **39**, one of the most notable examples of dietary flavone, is heavily consumed by humans. It is found in many fruits, vegetables, and herbs [103]. Apigenin **39** is also one of the most renowned flavones with several nutritional and pharmacological properties [104]. Regarding its MAO inhibitory activity, **39** isolated from propolis samples was reported to have moderate inhibition toward MAO enzymes, being slightly preferential to human MAO-A than MAO-B with IC<sub>50</sub> values of 0.64 and 1.12 μM, respectively [105]. Luteolin **40** is another remarkable dietary flavonoid with great pharmacological potential including its use to treat diabetes, Alzheimer's disease, and depression [106]. Luteolin **40**, isolated from *Cirsium japonicum* var. *maackii* (Compositae) (also known as Korean thistle), was found to be an inhibitor of human MAO-A with an IC<sub>50</sub> value of 8.57 μM [107]. Acacetin **41**, another interesting dietary flavone, was isolated from the Central American medicinal plant *Calea urticifolia* in a bioassay guided fractionation paradigm as the active MAO inhibitor with IC<sub>50</sub> values of 121 and 49 nM for human MAO-A and MAO-B, respectively [108]. Later, the studies with medicinal plant *Turnera diffusa* allowed the isolation of the flavones **41**, vetulin **42**, and acacetin 7-O-methyl ether **43** (Figure 6 and Table 3) as the active bio-components of MAO inhibition. The study found that **42** preferentially inhibited human MAO-B with an IC<sub>50</sub> value of 447 nM and an SI of 42. However, most importantly, **43** was found to be a selective and reversible human MAO-B inhibitor with an IC<sub>50</sub> value of 198 nM and SI of 505 [109]. The search for highly selective reversible MAO-B inhibitors was accomplished based on the computational study of acacetin 7-O-methyl ether-scaffold and the active sites of the MAO enzymes. Several derivatives of **43** were designed and found to have 1000- to 3000-fold selectivity toward human MAO-B; for example, 7-methoxy-4'-isopropoxy-5-hydroxyflavone **44** and 7-methoxy-4'-isobutyloxy-5-hydroxyflavone **45** were found to inhibit human MAO-B with IC<sub>50</sub> values of 33 and 31 nM, respectively, and an SI of 1921 and 3225, respectively [101]. Several other dietary flavones have been isolated and evaluated by their MAO inhibition. For example, the dietary flavone chrysin

**46** isolated from the North African medicinal plant *Cytisus villosus* showed  $IC_{50}$  values of 0.25 and 1.04  $\mu\text{M}$  for human MAO-A and MAO-B, respectively [110]. The phytochemical study of *Prunus padus* yielded the flavone genkwanin **47** as a non-selective MAO inhibitor with  $IC_{50}$  values of 0.14 for MAO-A and 0.35  $\mu\text{M}$  for MAO-B [111]. Xanthomicrol **48** and salvigenin **49** are two well-known flavones present in the *Sideritis* genus. Xanthomicrol **48** and salvigenin **49** were shown to be human MAO-A inhibitors with  $K_i$  values of 0.76 and 0.54  $\mu\text{M}$ , respectively [112].



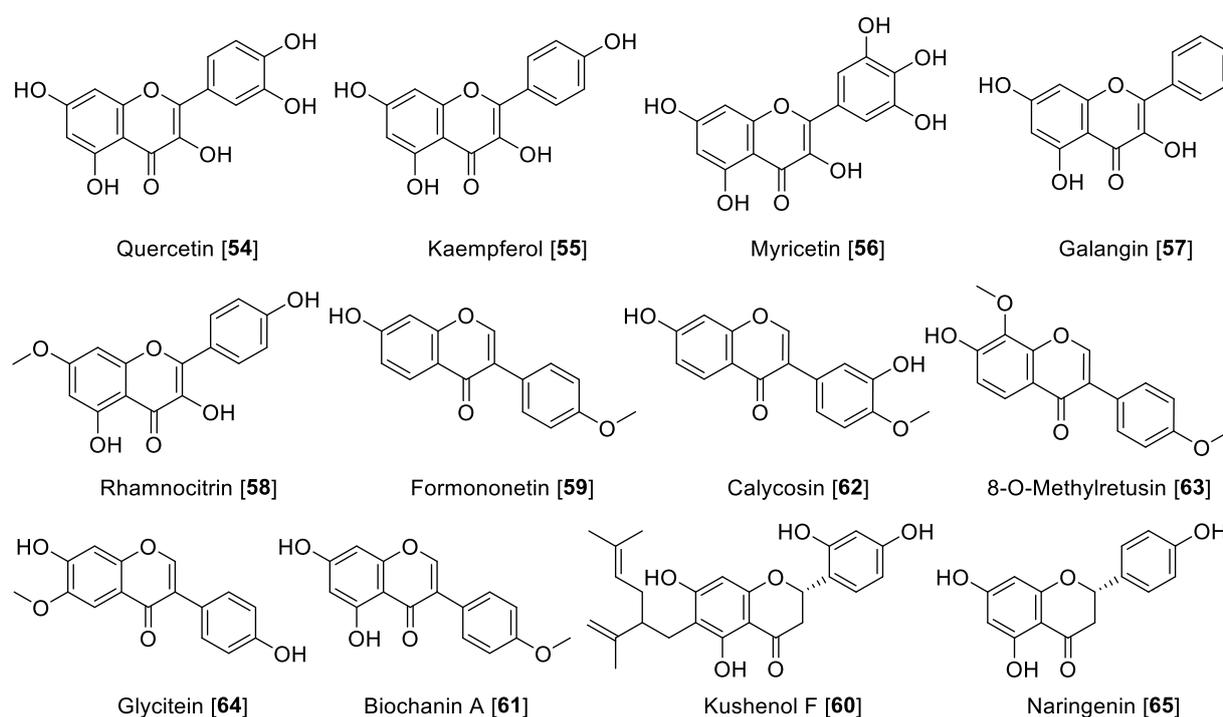
**Figure 6.** Chemical structure of selected flavone MAO inhibitors.

Lately, interest has been growing about the health benefits of glycoside flavones. Flavone O-glycosides or C-glycosides are present in the human diet and are keys to the biosynthesis and functions of plants, several of which have significant reported bioactivities [113]. There are several glycosides reported to display MAO inhibition, for example, orientin **50**, isorientin **51**, and isovitexin **52** are well-known flavone glycosides isolated from *Vitex grandiflora* (grapes). These flavonoid glycosides were reported to be selective human MAO-B inhibitors with IC<sub>50</sub> values of 11.04, 11.08, and 21.3 µM, respectively [114]. From *Agastache rugosa* and using a bioguided assay fractionation, **41** and its glycoside acacetin 7-O-(6-O-malonylglucoside) **53** were isolated and found to be MAO inhibitors. Compound **53** was found to be a non-selective MAO inhibitor with IC<sub>50</sub> values of 2.34 and 1.87 µM toward human MAO-A and MAO-B, respectively [115].

### 5.2.2. MAO Inhibitory Activity of Flavanols, Isoflavones, and Flavanones

Dietary flavanols are mainly referred to quercetin **54**, kaempferol **55**, and myricetin **56** (Figure 7), are specially found in fruits, vegetables, and beverages, and are also the key components in several plants. Similar to flavones, flavanols are also linked with the antioxidant properties of medicinal plants [116]. Regarding their MAO inhibition property, flavanols have been reported widely. Quercetin **54** isolated from *Hypericum hircinum* showed inhibitory activity for MAO-B with an IC<sub>50</sub> value of 20 µM and for MAO-A with an IC<sub>50</sub> value of 10 nM, and the source of the MAO was beef brain mitochondria [117]. Compounds **54** and **55** are common components of wine. A study on the constituents of *Vitis vinifera* and MAO inhibition found that **55** was a potent and selective human MAO-A inhibitor with an IC<sub>50</sub> value of 0.525 µM and no inhibition of MAO-B [118]. This group also found that **54** is a good MAO-A inhibitor with an IC<sub>50</sub> value of 3.98 µM and poor inhibition of MAO-B with an IC<sub>50</sub> of more than 100 µM [118]. The differences in the results of this study [117] and others may be explained by the different sources of the MAO enzymes used for inhibition studies [119]. Myricetin **56** was found to have a moderate inhibitory effect toward human MAO with an IC<sub>50</sub> value of 9.93 µM [110]. Galangin **57**, a flavonol found in propolis, was found to be a preferential MAO-A inhibitor with an IC<sub>50</sub> value of 0.13 µM compared to an IC<sub>50</sub> value of 3.65 µM for human MAO-B [105]. Another interesting flavonol isolated from *Prunus padus* is rhamnocitrin **58**. Rhamnocitrin **58** showed selective reversible inhibition of human MAO-A with an IC<sub>50</sub> value of 51 nM [120].

The isoflavonoid formononetin **59** and the flavanone kushenol F **60** were extracted and isolated from the roots of *Sophora flavescens* and showed significant MAO inhibition in a dose-dependent manner (mouse brain fraction). Formononetin **59** showed IC<sub>50</sub> values of 21.1 and 11.0 µM for MAO-A and MAO-B, respectively. Kushenol F **60** showed a moderate inhibition of MAO-B with an IC<sub>50</sub> value of 63.1 µM [121]. Seed extracts of *Psoralea coryfolia* have been used for centuries as chemoprotective and antioxidant agents. The study carried out to find the MAO-inhibiting bioactive components found biochanin A **61** as notable inhibitor. The isoflavone **61** was found to be a selective reversible MAO inhibitor with IC<sub>50</sub> values of 3.43 and 0.09 µM for human MAO-A and human MAO-B, respectively [122]. The Asian medicinal plant *Maackia amurensis* was reported to be a good source of isoflavonoids. The bioassay guided fractionation of *M. amurensis* bark extract yielded isoflavonoids calycosin **62** and 8-O-methylretusin **63** as reversible and selective inhibitors of human MAO-B with IC<sub>50</sub> values of 0.24 and 0.23 µM, respectively [123]. The isoflavone glycitein **64** is one of the major components in *Glycine max* (soybeans) and showed selective inhibition of human MAO-A with an IC<sub>50</sub> value of 8.30 µM [124]. Naringenin **65**, a common flavanone isolated from *Colvillea racemosae*, was identified as a preferential inhibitor of human MAO-B with an IC<sub>50</sub> value of 0.272 µM [125].



**Figure 7.** Chemical structure of selected flavanol, isoflavone, and flavanone MAO inhibitors.

### 5.2.3. MAO Inhibitory Activity of Miscellaneous Flavonoids and Related Compounds

Miscellaneous flavonoids and precursors (Figure 8) such as chalcones have been recently gaining attention for their effects of inhibiting MAO enzymes; for example, chalcones have been proposed as novel and effective MAO-B inhibitors [126,127]. Catechin 66 and epicatechin 67 have been reported to inhibit MAO enzymes. Compounds 66 and 67 isolated from *Uncaria rhynchophylla* and using rat brain homogenate as a source of MAO enzymes showed inhibition of MAO-B with  $IC_{50}$  values of 58.9 and 88.6  $\mu$ M, respectively [128]. The biflavonoids, morelloflavone 68 and GB-2a 69 isolated from *Garcinia gardneriana*, were reported as preferential human MAO-A inhibitors with  $IC_{50}$  values of 5.05 and 5.47  $\mu$ M, respectively [129]. The bichalcone A (3-3''-linked-(2'-hydroxy-4-O-isoprenylchalcone)-(2'''-hydroxy-4''-O-isoprenyldihydrochalcone) isolated from the dried bark of *Gentiana lutea* was hydrolyzed to form the product (E)-3-(4',6-dihydroxy-5'-(3-(3-(2-hydroxyphenyl)-3-oxopropyl)-[1,1'-biphenyl]-3-yl)-1-(2-hydroxyphenyl)prop-2-en-1-one 70. This hydrolytic product was found to be a MAO inhibitor with an  $IC_{50}$  value of 48.7 for MAO-A and 6.2  $\mu$ M for MAO-B [130]. Isoliquiritigenin 71, a natural chalcone, showed preferential inhibition of human MAO-B with an  $IC_{50}$  value of 0.51  $\mu$ M and an SI of 44.69 compared to human MAO-A [125]. When the chalcone was saturated and hydroxylated as in the case of colveol A 72, the selectivity turned toward MAO-A, as denoted by  $IC_{50}$  values of 0.62 and 29.9  $\mu$ M for MAO-A and MAO-B, respectively [125]. Isoliquiritigenin 71, besides being a potent MAO inhibitor, was also found to have important bioactivity as an antagonist to  $D_1$  dopamine receptors and agonist to dopamine  $D_3$  and vasopressin  $V_{1A}$  receptors. These findings suggest a potential therapeutic utility of Isoliquiritigenin 71 for neurological disorders [131]. The chalcone 4-hydroxyderricin 73, isolated from *Angelica keiskei*, was reported to be a selective human MAO-B inhibitor with an  $IC_{50}$  value of 3.43  $\mu$ M and an SI of 1000-fold difference compared to MAO-A [132]. From *Glycine max* Merrill (soybean), an aurone hispidol 74 was isolated. Compound 74 exhibited a preferential inhibition of human MAO-A with  $IC_{50}$  values of 0.26 and 2.45  $\mu$ M for MAO-A and MAO-B, respectively [133]. Medicarpin 75, a flavonol, was recently reported to have a preferential inhibition of human MAO-B with an  $IC_{50}$  value of 0.3  $\mu$ M [123]. Figure 8 shows the chemical structures and Table 3 presents the  $IC_{50}$  values of selected miscellaneous flavonoids.

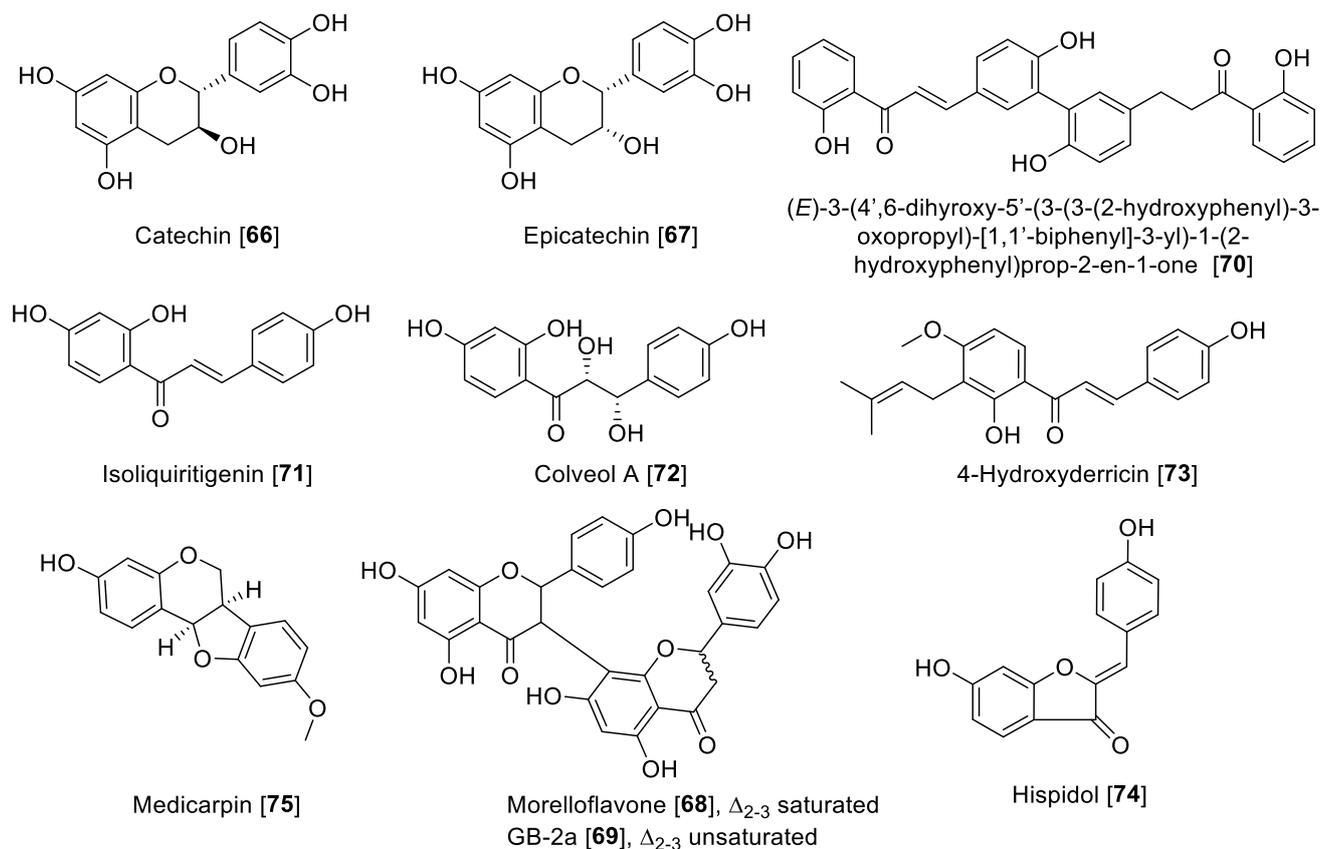
Table 3. Flavonoid natural product inhibitors of MAO-A and MAO-B.

Compounds	Source	MAO-A		MAO-B		SI	Enzyme Source	References
		IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	MAOA/B		
Apigenin [39]	Propolis	0.64	0.125	1.12	0.238	0.57	F	[104]
Luteolin [40]	<i>Cirsium maacki</i>	8.57		>100		>0.0857	F	[106]
Acacetin [41]	<i>Calea urticifolia</i>	0.121	0.0592	0.049	0.049	2.46	F	[107]
Vetulin [42]	<i>Turnera diffusa</i>	18.79		0.447		42.03	F	[108]
Acacetin 7-O-methyl ether [43]	<i>Turnera diffusa</i>	>100		0.198	0.045	<505.05	F	[108]
7-Methoxy-4'-isopropoxy-5-hydroxyflavone [44]	Acacetin derivative	30.74		0.016	0.052	1921.25	F	[101]
7-Methoxy-4'-isobutyloxy-5-hydroxyflavone [45]	Acacetin derivative	>100		0.031	0.037	>3225.8	F	[101]
Chrysin [46]	<i>Cytisus villosus</i>	0.25		1.04	NT	0.24	F	[109]
Genkwanin [47]	<i>Prunus padus</i>	0.14	0.097	0.35	0.12	0.4	F	[109]
Xanthomicrol [48]	<i>Sideritis</i> spp.		0.76		99.54	0.0076	F	[111]
Salvigenin [49]	<i>Sideritis</i> spp.		0.54		6.27	0.086	F	[111]
Orientin [50]	<i>Vitex grandiflora</i>	>100		11.04		>9.05	F	[113]
Isoorientin [51]	<i>Vitex grandiflora</i>	>100		11.08		>9.02	F	[113]
Isovitexin [52]	<i>Vitex grandiflora</i>	>100		21.3		>4.69	F	[113]
Acacetin	<i>Agastache rugosa</i>	2.34	1.06	1.87	0.38	1.25	F	[114]
7-O-(6-O-malonylglucoside) [53]								
Quercetin [54]	<i>Hypericum hircinum</i>	0.010		20		0.0005	I	[116]
	<i>Hypericum afrum</i>	1.52	0.29	28.39		0.053	F	[109]
	<i>Vitis vinifera</i>	3.98		>100		>0.039	F	[117]
Kaempferol [55]	<i>Vitis vinifera</i>	0.525		>100		>0.00525	F	[117]
Myricetin [56]	<i>Hypericum afrum</i>	9.93	2.24	59.34		0.167	F	[109]
Galangin [57]	Propolis	0.13	0.029	3.65	1.998	0.035	F	[104]
Rhamnocitrin [58]	<i>Prunus padus</i>	0.051	0.097	2.97	0.12	0.017	F	[110]
Formononetin [59]	<i>Sophora flavescens</i>	21.2		11.0		1.92	A	[119]
	<i>Maackia amurensis</i>	4.82		0.19		25.36	F	[121]
Kushenol F [60]	<i>Sophora flavescens</i>	103.7		63.1		1.64	A	[119]
Biochanin [61]	<i>Psoralea corylifolia</i>	3.43	0.099	0.09	0.0038	38.11	F	[120]
Calycosin [62]	<i>Maackia amurensis</i>	70.5		0.24		293.75	F	[122]
8-O-Methylretusin [63]	<i>Maackia amurensis</i>	18.7		0.23		81.30	F	[122]

Table 3. Cont.

Compounds	Source	MAO-A		MAO-B		SI	Enzyme Source	References
		IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	MAOA/B		
Glycitein [64]	<i>Pueraria lobata</i>	8.3		24.9		0.33	F	[123]
Naringenin [65]	<i>Colvillea racemosa</i>	8.64		0.272		31.76	F	[124]
Catechin [66]	<i>Uncaria rhynchophylla</i>			88.6	74		B	[127]
Epicatechin [67]	<i>Uncaria rhynchophylla</i>			58.9	21		B	[127]
Morelloflavone [68]	<i>Garcinia gardneriana</i>	5.05		66.2		0.076	F	[127]
GB-2a [69]	<i>Garcinia gardneriana</i>	5.47		56.7		0.20	F	[127]
Bichalcone-derivative [70]	<i>Gentiana lutea</i>	12.5		6.2	1.2	2.01	B	[127]
Isoliquiritigenin [71]	<i>Colvillea racemosa</i>	22.66		0.51		44.43	F	[124]
Colveol A [72]	<i>Colvillea racemosa</i>	0.62		29.90		0.020	F	[124]
4-Hydroxyderricin [73]	<i>Angelica keiskei</i>	>3000		3.43		>874.63	F	[131]
Hispidol [74]	<i>Glycine max</i>	0.26	0.10	2.45	0.51	0.10	F	[132]
Medicarpin [75]	<i>Maackia amurensis</i>	10.2		0.30		34.0	F	[122]

Note: natural products tested on total MAO are not listed. Enzyme source: A = mouse brain crude mitochondrial fraction; B = rat brain mitochondrial MAOs; F = recombinant human MAO-A and -B.



**Figure 8.** Chemical structure of selected miscellaneous flavonoid MAO inhibitors.

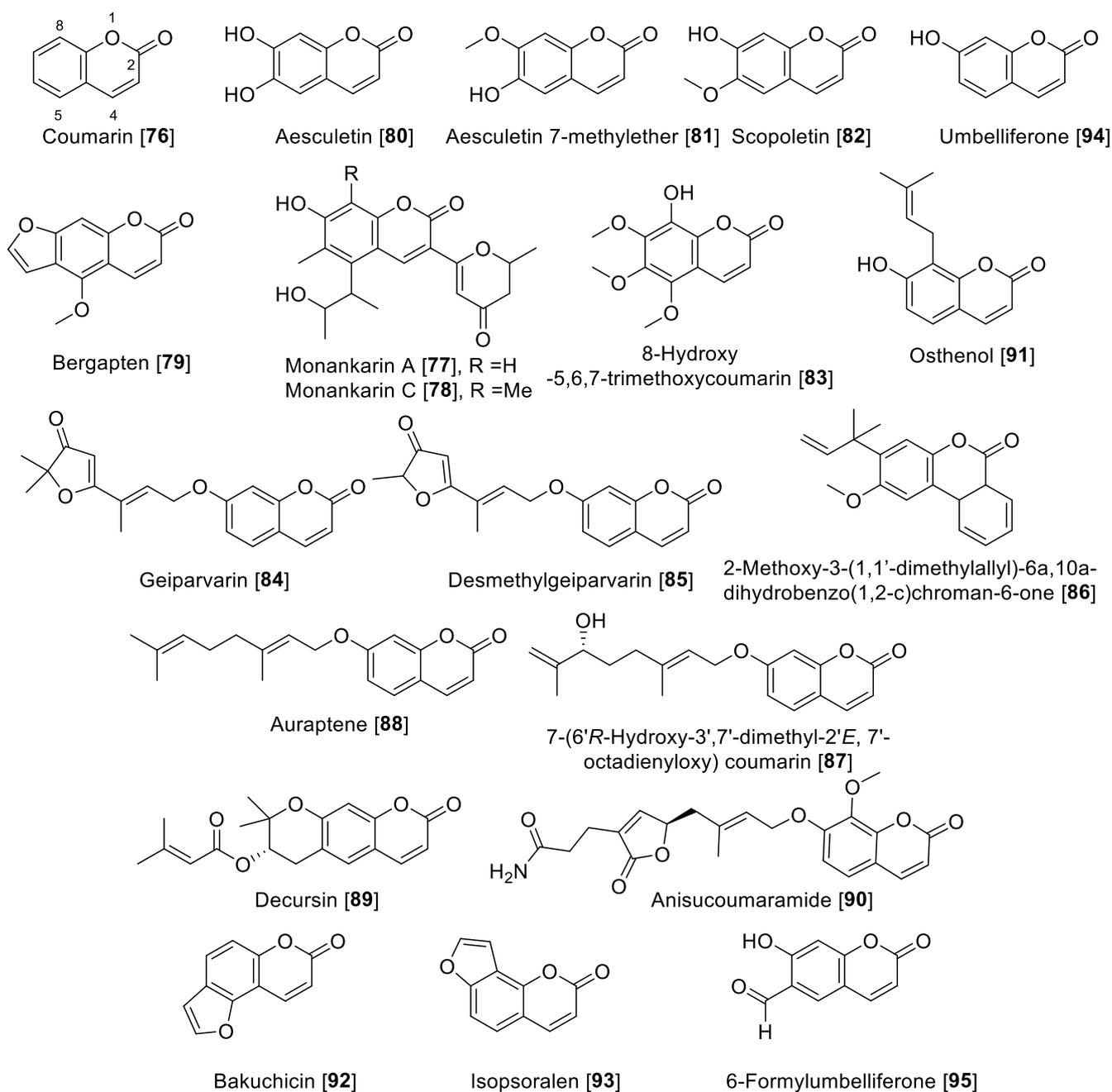
### 5.3. MAO Inhibitory Activity of Coumarins

Coumarins are versatile small lactones, constructed by the combination of a benzene and  $\alpha$  pyrone rings fused with each other, which are biosynthetically phenylpropanoid derivatives [134]. Compound 76 was the first example of natural coumarins reported from the *Dipteryx odorata* (*Coumarona odorata* Syn) beans (tonka beans) way back in 1820. Currently, coumarins are described as important metabolites in natural sources such as medicinal plants and their different parts [135]. The coumarin scaffold shows wide industrial applications and medicinal uses, of which a notable example is warfarin, the most common anticoagulant prescribed worldwide [135]. Coumarins (Figure 9 and Table 4) have been proposed as privilege scaffold and focused on as MAO inhibitors [136,137].

Several natural coumarins have been reported as MAO inhibitors, including the monankarins, pigments extracted from the fungi *Monascus anka* [138]. In particular, monankarin A 77 and C 78 were found to exhibit inhibitory activities for MAO in mouse brain homogenate with  $IC_{50}$  values of 15.5 and 1.07  $\mu$ M, respectively [138]. From the roots of *Peucedanum japonicum*, a series of coumarins were isolated, and among those, bergapten 79 was found to inhibit total MAO in mouse brain homogenate with an  $IC_{50}$  value of 13.8  $\mu$ M [139]. Additionally, the coumarins aesculetin 80, aesculetin 7-methyl ether 81, and scopoletin 82 showed moderate MAO inhibition with  $IC_{50}$  values of 30.1, 32.2, and 45.0  $\mu$ M, respectively, in mouse brain homogenate. Compounds 80–82 were also isolated from *Artemisia vulgaris*, a well-known European and Asian herb [74]. From the dried flowers of *Hibiscus syriacus*, several coumarins including 8-hydroxy-5,6,7-trimethoxycoumarin 83 and 82 were found to inhibit MAO (brain mouse homogenates) in a dose-dependent manner with  $IC_{50}$  values of 44.5 and 19.4  $\mu$ g/mL, respectively [140]. Geiparvarin 84 and desmethylgeiparvarin 85, two coumarins isolated from *Geijera parviflora*, were found to be selective toward MAO-B. Geiparvarin 84 showed  $IC_{50}$  values of 27  $\mu$ M and 144 nM for

MAO-A and MAO-B, respectively, and desmethylgeiparvarin **85** showed IC<sub>50</sub> values of 24 μM and 28 nM for MAO-A and MAO-B, respectively [141].

From *Gentiana lutea*, the rearranged coumarin 2-methoxy-3-(1,10-dimethylallyl)-6a,10a-dihydrobenzo(1,2-c)chroman-6-one **86** was isolated and found to be a selective inhibitor of MAO-B with an IC<sub>50</sub> value of 3.8 μM [130]. Using bioassay guided fractionation of the Asian medicinal plant *Dictamnus albus*, the coumarins 7-(6'*R*-hydroxy-3',7'-dimethyl-2'*E*,7'-octadienyloxy) coumarin **87** and auraptene **88** were found to have the highest inhibitory MAO activity. Compound **87** inhibited MAO in a non-selective way with IC<sub>50</sub> values of 1.3 and 0.5 μM for MAO-A and MAO-B, respectively, while **88** preferentially inhibited MAO-B with an IC<sub>50</sub> value of 0.6 μM compared to an IC<sub>50</sub> value of 34.6 μM for MAO-A [142]. Decursin **89**, a tetrahydropyrone coumarin isolated from *Angelica gigas*, was found to be selective toward MAO-A with an IC<sub>50</sub> value of 1.89 μM compared to an IC<sub>50</sub> value of 70.5 μM for human MAO-B [143]. From the leaves and twigs of the medicinal plant *Clausena anisum-olens*, anisocoumaramide **90** was isolated. This secondary metabolite **90** showed selective inhibition of human MAO-B with an IC<sub>50</sub> value of 144 nM and SI of more than 696 compared to MAO-A [144]. From another Apiaceae, *Angelica pubescens*, osthonol **91** was isolated and found to be selective toward human MAO-A with an IC<sub>50</sub> value of 0.74 μM and SI of >81 compared to MAO-B [145]. The same group also reported that bakuchicin **92** and isopsoralen **93** coumarins isolated from *Psoralea corylifolia* were non-selective MAO inhibitors and that **92** exhibited IC<sub>50</sub> values of 1.78 and 5.44 μM for human MAO-A and MAO-B, respectively. Compound **93** inhibited human MAO-A and MAO-B with IC<sub>50</sub> values of 0.88 μM and 2.73 μM, respectively [145]. Umbelliferone **94** and 6-formylumbelliferone **95** isolated from *Angelica decursiva* were found to inhibit MAO enzymes with moderate preference toward human MAO-B with IC<sub>50</sub> values of 39.16 compared to 147 μM for human MAO-A. The metabolite **95** showed IC<sub>50</sub> values of 3.23 and 15.31 μM for MAO-A and -B, respectively [145]. The metabolite **95**, besides inhibition of MAO-A, also inhibited lipid peroxidation and Aβ self-aggregation and indicated that 6-formylumbelliferone **95** can be a promising lead for the development of treatments for neurodegenerative diseases [145]. The synthetic studies to modulate the MAO inhibition of coumarins prepared hybrid compounds, making these derivatives more flexible in the case of coumarin-*N*-benzylalkyloxy derivatives [146] and coumarin-chalcones hybrids [147], to mention a few examples. Figure 9 shows the chemical structures for selected coumarins and Table 4 presents the IC<sub>50</sub> values of selected coumarin compounds.



**Figure 9.** Chemical structure of selected coumarin MAO inhibitors.

**Table 4.** Coumarin natural product inhibitors of MAO-A and MAO-B.

Compounds	Source	MAO-A		MAO-B		SI	Enzyme Source	References
		IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	MAOA/B		
Geiparvarin [84]	<i>Geijera parviflora</i>	27		0.144		187.5	B	[142]
Desmethylgeiparvarin [85]	<i>Geijera parviflora</i>	24		0.028		857.1	B	[142]
2-Methoxy-3-(1,10-dimethylallyl)-6a,10a-dihydrobenzo(1,2-c)chroman-6-one [86]	<i>Gentiana lutea</i>	>100	–	2.9	1.1	>34.5	B	[129]
7-(6'R-Hydroxy-3',7'-dimethyl-2'E, 7'-octadienyloxy) coumarin [87]	<i>Dictamnus albus</i>	1.3	–	0.5	0.46	2.6	A	[141]
Auraptene [88]	<i>Dictamnus albus</i>	34.6	–	0.6	0.83	57.6	A	[141]
Decursin [89]	<i>Angelica gigas</i>	0.6		70.5		0.0085	F	[142]
Anisucoumaramide [90]	<i>Clausena anisum-olens</i>	>100		0.143		699.3	F	[143]
Osthenol [91]	<i>Angelica pubescens</i>	0.74	0.26	>60	–	>0.012	F	[144]
Bakuchicin [92]	<i>Angelica pubescens</i>	1.78		5.44		0.32	F	[144]
Isopsoralen [93]	<i>Angelica pubescens</i>	0.88	0.46	2.73		0.32	F	[144]
Umbelliferone [94]	<i>Angelica decursiva</i>	39.16		147.37		0.26	F	[145]
6-Formylumbelliferone [95]	<i>Angelica decursiva</i>	3.23	3.05	15.31	6.81	0.21	F	[145]

Note: natural products tested on total MAO are not listed. Enzyme source: A = mouse brain crude mitochondrial fraction; B = rat brain mitochondrial MAOs; F = recombinant human MAO-A and -B.

#### 5.4. MAO Inhibitory Activity of Xanthenes, Anthraquinones, and Naphthoquinones

Natural xanthenes, anthraquinones, and naphthoquinones are polyphenolic compounds characterized by benzene rings attached to each other by carbonyl and oxygen in the case of xanthenes with a C<sub>6</sub>-C<sub>1</sub>-C<sub>6</sub> skeleton or benzene rings coupled with paraquinones mainly to form naphthoquinone or anthraquinones (C<sub>6</sub>-C<sub>2</sub>-C<sub>6</sub> sequence), configuring a planar ring structure [148]. The biosynthesis for these secondary metabolites is different depending on the natural source but they can be derived through several pathways. In fact, in plants, anthraquinones are derived mainly by polyketide, mevalonic, and methylerythritol 4-phosphate pathways [149]. On the other hand, in plants, shikimate and fatty acid pathways are predominantly found for xanthenes [150]. Recent studies have shown a biosynthetic relationship between xanthenes and anthraquinones, especially in bacteria and fungi [151]. Figures 10 and 11 show the structures and Table 5 presents the IC<sub>50</sub> values of selected xanthenes, anthraquinones, and naphthoquinones against MAOs.

Xanthenes and xanthone glycosides possess numerous biological and pharmacological potentials. Those compounds have been reported to show several therapeutic properties including antiallergic, anti-inflammatory, antituberculous, antitumor, and antiplatelet [152,153]. In earlier studies, a series of natural xanthenes has been evaluated for their activity to inhibit MAO-A and -B [154,155]. Rat brain mitochondrial extract was the source for MAO. 1-Hydroxy-3,8-dimethoxy-xanthone **96**, 1,3-dihydroxy-7,8-dimethoxy xanthone **97**, and bellidifolin **98** were the potent inhibitors of MAO-A. Xanthenes exhibited only moderate inhibition of MAO-B [154,155]. From *Hypericum brasiliense*, a series of compounds was isolated, and among those, 6-deoxyjacareubin **99** and 1,5-dihydroxyxanthone **100** were found to exhibit the best MAO inhibition, using rat brain mitochondria as source of MAO enzymes. Compounds **99** and **100** exhibited slight preferential inhibition of MAO-A with IC<sub>50</sub> values of 12.0 and 0.73 μM, respectively [156].

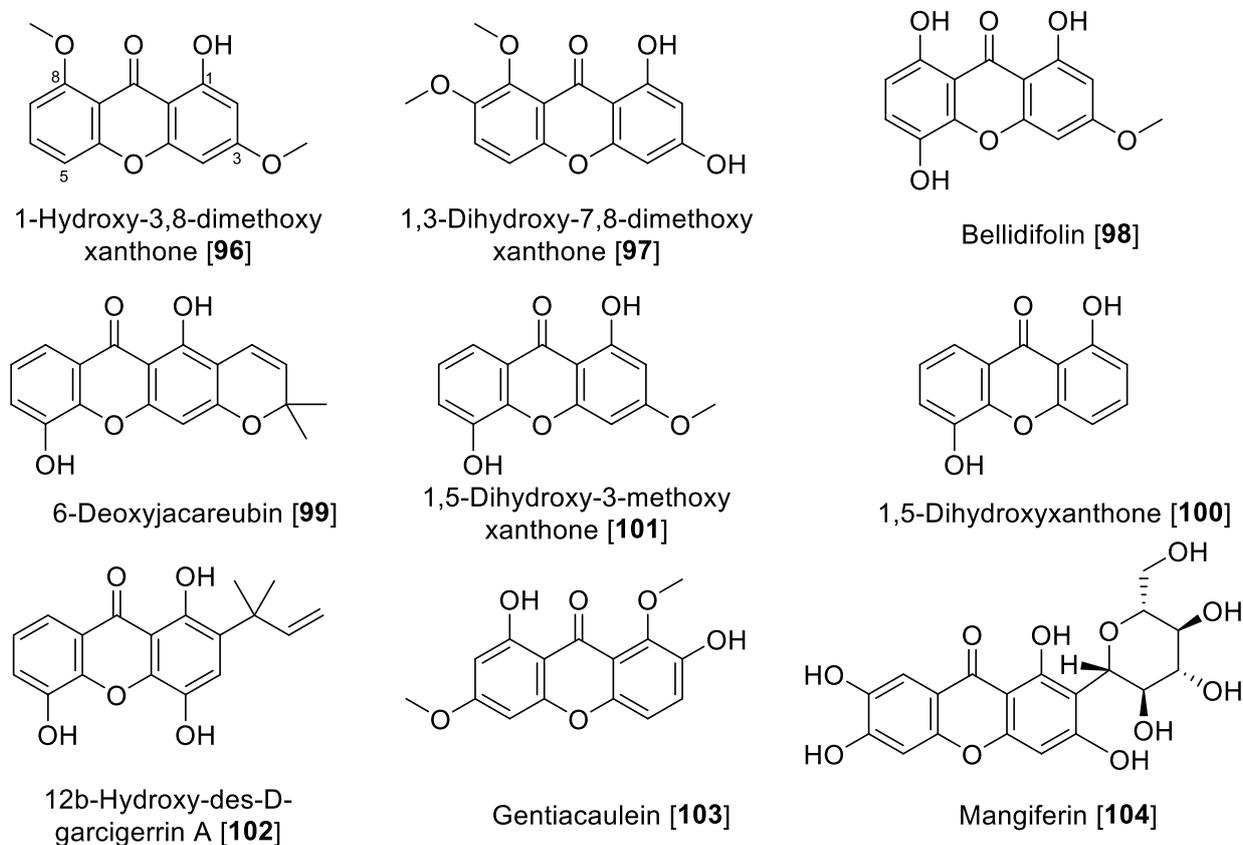
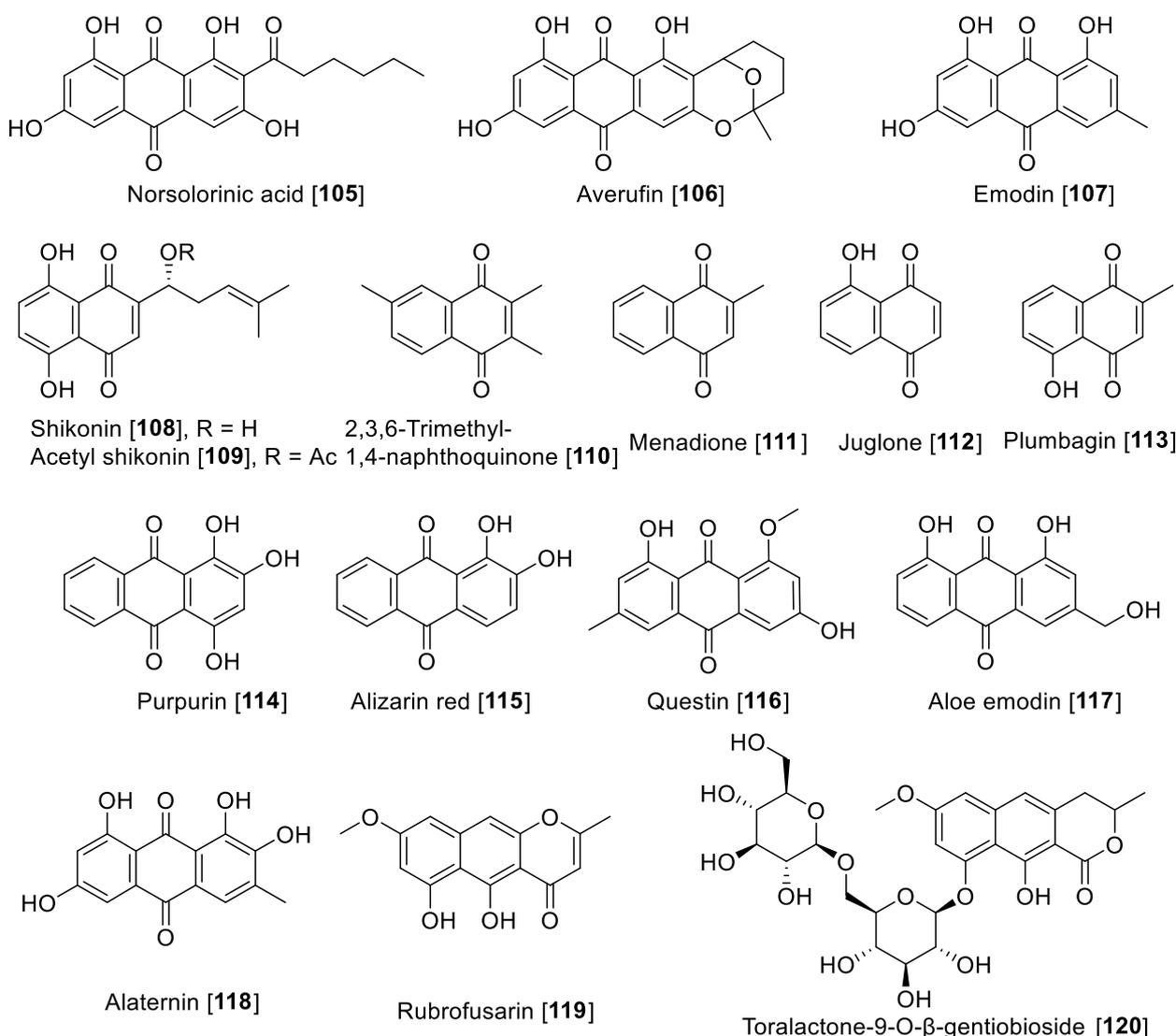


Figure 10. Chemical structure of selected xanthone MAO inhibitors.



**Figure 11.** Chemical structure of selected naphthoquinone and anthraquinone MAO inhibitors.

In a study performed by Gnerre et al., a series of fifty-nine natural and synthetic xanthenes was evaluated for inhibition of MAO-A and -B [157]. It was found that the tested xanthenes in general were preferential toward MAO-A at low micromolar concentrations. It is important to highlight that the natural xanthone 1,5-dihydroxy-3-methoxy xanthone **101** isolated from *Chironia krebssii*, a medicinal plant growing in the tropical East Africa country Malawi [158], was the most active within the series. Compound **101** exhibited inhibition of MAO-A and MAO-B with  $IC_{50}$  values of 40 nM and 33  $\mu$ M, respectively [157]. Another notable example of natural xanthenes is 12b-hydroxy-des-D-garcigerrin A **102**, found in several *Garcinia* species [159], which showed an  $IC_{50}$  value of 3.3  $\mu$ M for MAO-A and no inhibition of MAO-B [157]. Compound **102** also showed antioxidant and antiplasmodial activities [159]. Compound **98** was found to inhibit MAO-A with an  $IC_{50}$  value of 0.66  $\mu$ M [157]. This natural xanthone is frequently isolated from *Gentiana* species [160].

Gentiacaulin **103**, a natural xanthone isolated from *Gentiana kochiana*, was found as one of the two main xanthenes present in the plant. Compound **103** exhibited MAO-A inhibition with an  $IC_{50}$  value of 0.49  $\mu$ M [161]. In addition, several other natural xanthenes were isolated from *Cudrania tricuspidata* and *Gentianella amarella* and evaluated for their ability to inhibit MAO enzymes. The novel xanthenes isolated from *C. tricuspidata* showed poor inhibition of MAO [162], and the only percentage inhibition was reported for the

xanthenes isolated from *G. amarella* [163]. Dimitrov et al. reported that mangiferin **104**, a glucoside xanthone usually found in the *Hypericum* species, showed selective inhibition of MAO-A with an  $IC_{50}$  value of 41  $\mu\text{M}$  and negligible inhibition of MAO-B, using rat liver mitochondria as the MAO source. The authors explained that this MAO inhibition plays a key role in the antidepressant effect of the *Hypericum aucheri* extracts [164].

There are several anthraquinones and naphthoquinones reported to have inhibition toward MAO. Earlier examples are the anthraquinones, norsolorinic acid **105** and averufin **106**, isolated from the fungus *Emericella navahoensis* and which inhibited MAO in mouse liver homogenate. Compound **105** inhibited total MAO with an  $IC_{50}$  value of 0.3  $\mu\text{M}$  [165]. Emodin **107**, a common anthraquinone found in plants, exhibited MAO-B inhibition with an  $IC_{50}$  value of 35.4  $\mu\text{M}$  [68]. *Lithospermum erythrorhizon* roots are a common component in several preparations used in traditional Chinese medicine (TCM) for treatment of wounds and dermatitis [166]. Bioassay guided fractionation studies of *L. erythrorhizon*-root extracts for MAO inhibition using mouse brain homogenate allowed the isolation of naphthoquinones shikonin **108** and acetyl shikonin **109** as the bioactive components and non-selective MAO inhibitors. Shikonin **108** inhibited MAO-A with an  $IC_{50}$  value of 16.4  $\mu\text{M}$  and MAO-B with an  $IC_{50}$  value of 13.6  $\mu\text{M}$ ; **109** had  $IC_{50}$  values of 16.9 and 10.1  $\mu\text{M}$  for MAO-A and MAO-B, respectively [167].

Another notable naphthoquinone, 2,3,6-trimethyl-1,4-naphthoquinone **110**, was extracted and isolated from cured tobacco leaves (*Nicotiana tabacum*) and was found to be a competitive inhibitor of human MAO (human liver mitochondrial preparation) with  $K_i$  values of 3 and 6  $\mu\text{M}$  for MAO-A and MAO-B, respectively [168]. Recent studies found using a human MAO assay that **110** inhibited MAOs with  $IC_{50}$  values of 1.14 and 7.14  $\mu\text{M}$  for MAO-A and MAO-B, respectively [169]. The same authors evaluated the anxiolytic effects of **110** on a zebrafish model, describing that the compound induced anxiolytic and anxiogenic-like effects on the model tested [169]. Menadione **111**, a naphthoquinone formed by metabolism from the dietary vitamin K (menaquinones), was reported as selective MAO-B inhibitor with a  $K_i$  value of 0.4  $\mu\text{M}$  [170]. Mostert et al. in a study with a series of synthetic and natural naphthoquinones reported juglone **112** and plumbagin **113** to inhibit MAO in a non-selective manner. Compound **112** exhibited  $IC_{50}$  values of 1.71 and 4.36  $\mu\text{M}$  for human MAO-A and MAO-B, respectively; plumbagin **113** showed  $IC_{50}$  values of 4.91 and 1.09  $\mu\text{M}$  for human MAO-A and MAO-B, respectively [171]. Two natural dyes anthraquinones purpurin **114** and alizarin red **115** were reported to inhibit MAO, and both anthraquinones showed a selectivity toward MAO-A. Compound **114** exhibited an  $IC_{50}$  value of 2.50  $\mu\text{M}$  and for **115**, an  $IC_{50}$  value of 30.1  $\mu\text{M}$  [172]. In a phytochemical study of the components of *Cassia obtusifolia* seeds, a series of anthraquinones and related compounds were isolated and evaluated for the ability to inhibit human recombinant MAO. The compounds showed selective inhibition of MAO-A, selective and preferential, and notable examples are questin **116** that inhibited human MAO-A with an  $IC_{50}$  value of 0.17  $\mu\text{M}$ , as well as aloe emodin **117**, alaternin **118**, rubrofusarin **119**, and toralactone-9-O- $\beta$ -gentiobioside **120** which were found to be strong inhibitors with  $IC_{50}$  values of 2.47, 5.35, 5.9, and 7.3  $\mu\text{M}$ , respectively [173].

**Table 5.** Xanthone, anthraquinone, and naphthoquinone natural product inhibitors of MAO-A and MAO-B.

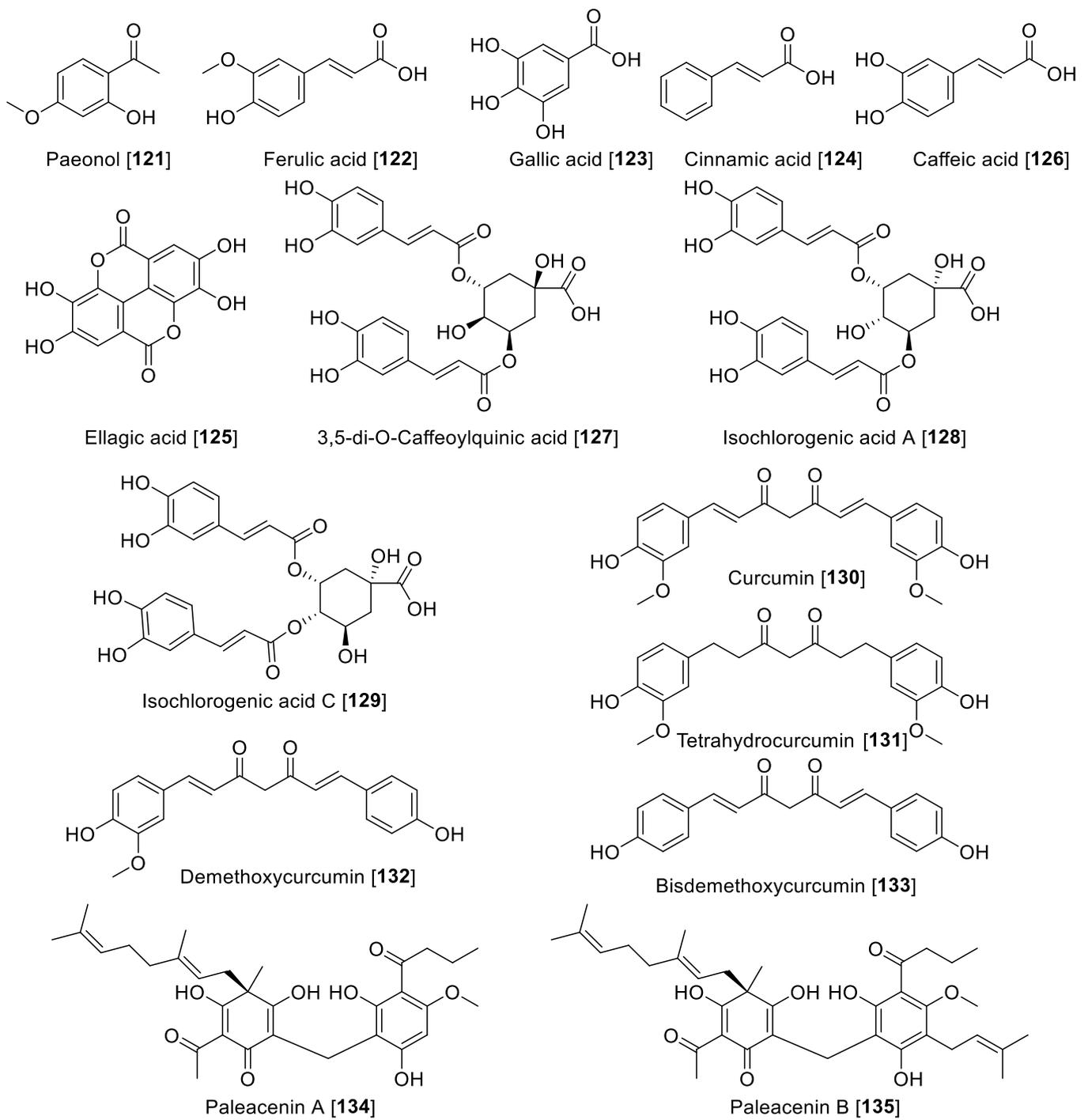
Compounds	Source	MAO-A		MAO-B		SI	Enzyme Source	References
		IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	MAOA/B		
Bellidolin [98]	<i>Gentiana lactea</i> .	0.66		>100		>0.0066	C	[157]
6-Deoxyjacareubin [99]	<i>Hypericum brasiliense</i>	12.0		47.3		0.25	B	[156]
1,5-Dihydroxyxanthone [100]	<i>Hypericum brasiliense</i>	0.73		76.3		0.0095	B	[156]
1,5-dihydroxy-3-methoxy xanthone [101]	<i>Chironia krebsii</i>	0.04		33.0		0.0012	C	[157]
12b-hydroxy-des-D-garcigerrin A [102]	<i>Garcinia gerrardii</i>	3.3		>100		0.033	C	[157]
Genticaulein [103]	<i>Gentiana kochiana</i>	0.22		96.0		0.0022	B	[161]
Mangiferin [104]	<i>Hypericum aucheri</i>	410		>1000		0.41	C	[170]
Emodin [107]	Polygonaceae Fam.			35.4	15.1		B	[68]
Shikonin [108]	<i>Lithospermum erythrorhizon</i>	16.4	12.8	13.6	13.0	1.20	A	[167]
Acetyl shikonin [109]	<i>Lithospermum erythrorhizon</i>	16.9	10.5	10.1	6.3	1.67	A	[167]
2,3,6-Trimethyl-1,4-naphthoquinone [110]	<i>Nicotiana tabacum</i>	3.0		6.0		0.5	E	[168]
		1.14		7.14		1.59	F	[169]
Menadione [111]	Vit K derivative		26.0		0.4	65	F	[169]
		10.2		3.02		3.37	F	[170]
Juglone [112]	<i>Juglans</i> spp.	1.71		4.36		0.39	F	[170]
Plumbagin [113]	<i>Plumbago</i> spp.	4.91		1.09		4.50	F	[170]
Purpurin [114]	<i>Rubia tinctorum</i>	2.50	0.422	>40		>0.062	F	[172]
Alizarin red [115]	<i>Rubia tinctorum</i>	30.1		>60		>0.50	F	[173]
Questin [116]	<i>Cassia obtusifolia</i>	0.17	4.14	10.58		0.016	F	[173]
Aloe emodin [117]	<i>Cassia obtusifolia</i>	2.47	0.50	>400		>0.0061	F	[173]
Alaternin [118]	<i>Cassia obtusifolia</i>	5.35	3.97	4.55		1.17	F	[173]
Rubrofusarin [119]	<i>Cassia obtusifolia</i>	5.90	4.38	91.40		0.064	F	[173]
Toralactone-9-O-β-gentiobioside [120]	<i>Cassia obtusifolia</i>	<b>7.36</b>	<b>4.30</b>	>400		>0.0184	F	[173]

Note: natural products tested on total MAO are not listed. Enzyme source: A = mouse brain crude mitochondrial fraction; B = rat brain mitochondrial MAOs; C = rat liver mitochondrial MAOs; F = recombinant human MAO-A and -B.

### 5.5. MAO Inhibitory Phenols and Polyphenolic Compounds

The polyphenolic compounds are simple phenolics and their acids and esters produced predominantly by the shikimate pathway include phenylpropanoids and lignans [174]. Several natural phenols were evaluated on MAO rat brain mitochondrial fraction and only Paeonol **121** showed a moderate inhibition toward MAO-A with a  $K_i$  value of 51.1  $\mu\text{M}$  [68]. Ferulic acid **122** showed to be preferential toward MAO-A with an  $\text{IC}_{50}$  value of 7.55  $\mu\text{M}$  for human MAO-A and 24.0  $\mu\text{M}$  for MAO-B. Gallic acid **123** exhibited  $\text{IC}_{50}$  of 9.49  $\mu\text{M}$  for human MAO-A; *trans*-cinnamic acid **124**, a non-selective MAO inhibitor, exhibited  $\text{IC}_{50}$  values of 6.47 and 1.21  $\mu\text{M}$  for MAO-A and -B, respectively; ellagic acid **125** was a selective MAO-B inhibitor with an  $\text{IC}_{50}$  value of 0.40  $\mu\text{M}$ ; and caffeic acid **126** was shown to be a non-selective MAO inhibitor with  $\text{IC}_{50}$  values of 11.72 and 22.88  $\mu\text{M}$  for human MAO-A and MAO-B, respectively [175].

A series of chlorogenic acids was isolated from the flowers of *Lonicera macranthoides*, and from the isolated compounds, only 3,5-di-*O*-caffeoylquinic acid **127** exhibited moderate inhibition with an  $\text{IC}_{50}$  value of 20.04  $\mu\text{M}$  for MAO-B and no inhibition for MAO-A [176]. From *Lonicera japonica*, using a magnetic nanoparticle MAO-B immobilized affinity solid-phase extraction method, two new chlorogenic acids isochlorogenic A **128** and C **129** were identified along with **122** as the bioactive component in *L. japonica* [177]. Isochlorogenic acid A **128** and C **129** were found to be mixed-type inhibitors for MAO-B with  $\text{IC}_{50}$  values of 29.0 and 29.77  $\mu\text{M}$ , respectively [177]. Turmeric is a commonly used spice prepared from the rhizomes of *Curcuma longa*. Several bioactivities are attributed to turmeric, and those responsible for that bioactivity are the curcuminoid-type compounds, and in particular, curcumin **130** [178]. The inhibitory effects of **130** and tetrahydrocurcumin **131** on MAO-B were evaluated in a Parkinson's disease rodent model induced by 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine. Depletion of dopamine and 3,4-dihydroxy phenyl acetic acid occurs with increased MAO-B activity. Use of **130** and **131** reversed the decrease in dopamine and 3,4-dihydroxy phenyl acetic acid induced by the model [179]. Recently, the ability of **130**, demethoxycurcumin **132**, and bisdemethoxycurcumin **133** to inhibit MAO was reported, where the three compounds inhibited MAO moderately in a non-selective manner. Compound **130** exhibited  $\text{IC}_{50}$  values of 3.64 and 3.36  $\mu\text{M}$  for human MAO-A and MAO-B, respectively; **131** exhibited  $\text{IC}_{50}$  values of 3.09 and 2.59  $\mu\text{M}$ , respectively; and **132** exhibited  $\text{IC}_{50}$  values of 3.24 and 2.45  $\mu\text{M}$  for MAO-A and MAO-B, respectively [180]. Paleacinins A **134** and B **135** are two novel acylphloroglucinols isolated from *Elaphoglossum paleaceum* rhizome, and these two undescribed compounds exhibited high MAO inhibition [181]. Compounds **134** and **135** showed to be a non-selective MAO inhibitor in rat brain mitochondria, and **134** exhibited  $\text{IC}_{50}$  values of 31.0 and 4.7  $\mu\text{M}$  for MAO-A and -B, respectively. Meanwhile, **135** showed  $\text{IC}_{50}$  values of 1.3 and 4.4  $\mu\text{M}$  for MAO-A and -B, respectively. Besides their MAO inhibition, **134** and **135** also showed activity toward several cancer cell lines including the prostate, cervix, breast, and colon, denoting a multitarget effect for paleacinins [181]. Figure 12 shows the selected additional polyphenolic compound MAO inhibitors and Table 6 presents the  $\text{IC}_{50}$  values of selected phenol and polyphenolic compounds.



**Figure 12.** Chemical structure of miscellaneous polyphenol MAO inhibitors.

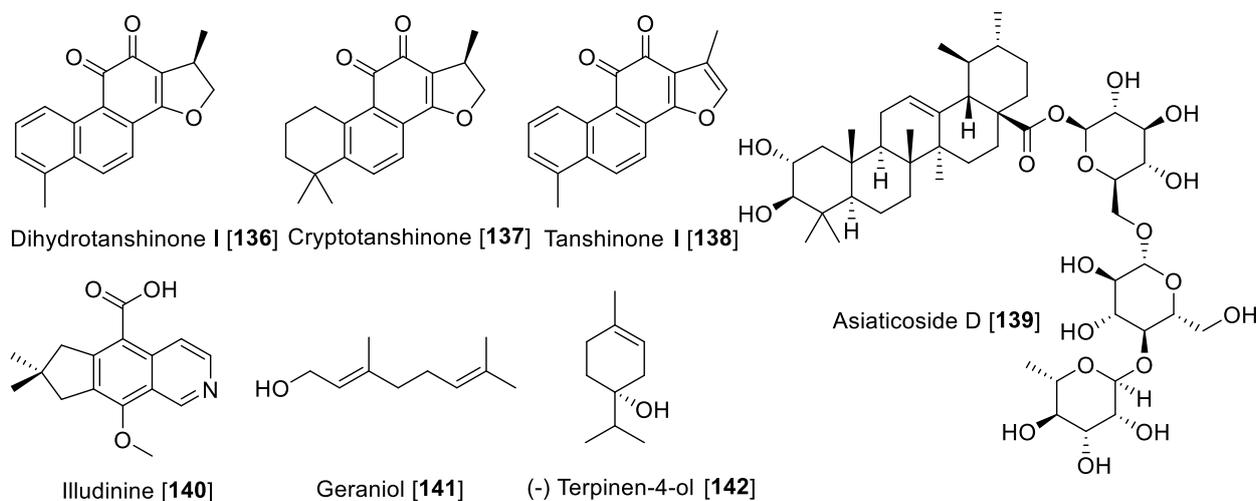
**Table 6.** Phenol and polyphenolic natural product inhibitors of MAO-A and MAO-B.

Compounds	Source	MAO-A		MAO-B		SI	Enzyme Source	References
		IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	MAOA/B		
Paeonol [121]	<i>Paeonia</i> spp.	54.6	51.1	42.5		1.28	B	[68]
Ferulic acid [122]			7.55		24.0	0.31	F	[175]
Gallic acid [123]		9.49		NR			F	[175]
<i>t</i> -Cinnamic acid [124]		6.47		1.21		5.34	A	[175]
Ellagic acid [125]				0.40			B	[175]
Caffeic acid [126]		11.7		22.9		0.51	F	[175]
3,5-di- <i>O</i> -caffeoylquinic acid [127]	<i>Lonicera macranthoides</i>			20.04			B	[176]
Isochlorogenic acid A [128]	<i>Lonicera japonica</i>			29.05	9.55		J	[177]
Isochlorogenic acid C [129]	<i>Lonicera japonica</i>			29.77	9.53		J	[177]
Curcumin [130]	<i>Curcuma longa</i>	3.64		3.36		1.08	F	[180]
Demethoxycurcumin [132]	<i>Curcuma longa</i>	3.09	0.91	2.59	0.86	1.19	F	[180]
bis-Demethoxycurcumin [133]	<i>Curcuma longa</i>	3.24	1.40	2.45	0.80	1.32	F	[180]
Paleacerins A [134]	<i>Elaphoglossum paleaceum</i>	31.0		4.7		6.59	B	[181]
Paleacerins C [135]	<i>Elaphoglossum paleaceum</i>	1.3		4.4		0.29	B	[181]

Note: natural products tested on total MAO are not listed. Enzyme source: A = mouse brain crude mitochondrial fraction; B = rat brain mitochondrial MAOs; F = recombinant human MAO-A and -B; J = liver porcine.

### 5.6. MAO Inhibitory Terpenes and Terpenoids

Terpenes or terpenoids are one of the broad families of natural products with uncountable pharmacological and biological uses. These secondary metabolites are built based on isoprene units (five carbon building blocks) [182]. There are few terpenoids that have been reported to exhibit MAO inhibition. In a study exploring the use of HPLC-based bioguided activity profiling, *Salvia miltiorrhiza* was screened. The extract of *S. miltiorrhiza* showed a high inhibitory effect on rat liver monoamine oxidase fraction with a preference toward MAO A. From the active fractions based on the HPLC-bioguided assay, the diterpenes dihydrotanshinone I **136**, cryptotanshinone **137**, and tanshinone I **138** were identified as the bioactive compounds with  $IC_{50}$  values of 23, 80, and 84  $\mu\text{M}$ , respectively [183]. Recently, a triterpene saponin asiaticoside D **139**, isolated from *Centella asiatica* (a herbal product used in Ayurvedic medicine) following a bioassay guided fractionation, exhibited MAO inhibition [184]. Compound **139** was found to be a non-selective inhibitor with  $IC_{50}$  values of 4.0 and 1.3  $\mu\text{g}/\text{mL}$  for MAO-A and MAO-B, respectively [184]. Illudinine **140**, a sesquiterpene-alkaloid with an illudalane skeleton, was found to be an inhibitor of human MAO-B with an  $IC_{50}$  value of 18.3  $\mu\text{M}$ . Several synthetic derivatives of illudinine have been synthesized; however, none of these compounds were found to be superior to illudinine [185]. In a bioassay guided fractionation study of the *Zingiber officinale* rhizomes, several monoterpenes showed potential MAO-A inhibitory properties [186]. The monoterpenes geraniol **141** and (-) terpinen-4-ol **142** were found to be inhibitors of MAO-A, but only a percentage of inhibition was reported [186]. Figure 13 shows the chemical structures for selected terpenoid MAO inhibitors and Table 7 presents the  $IC_{50}$  values of selected terpenoid compounds.

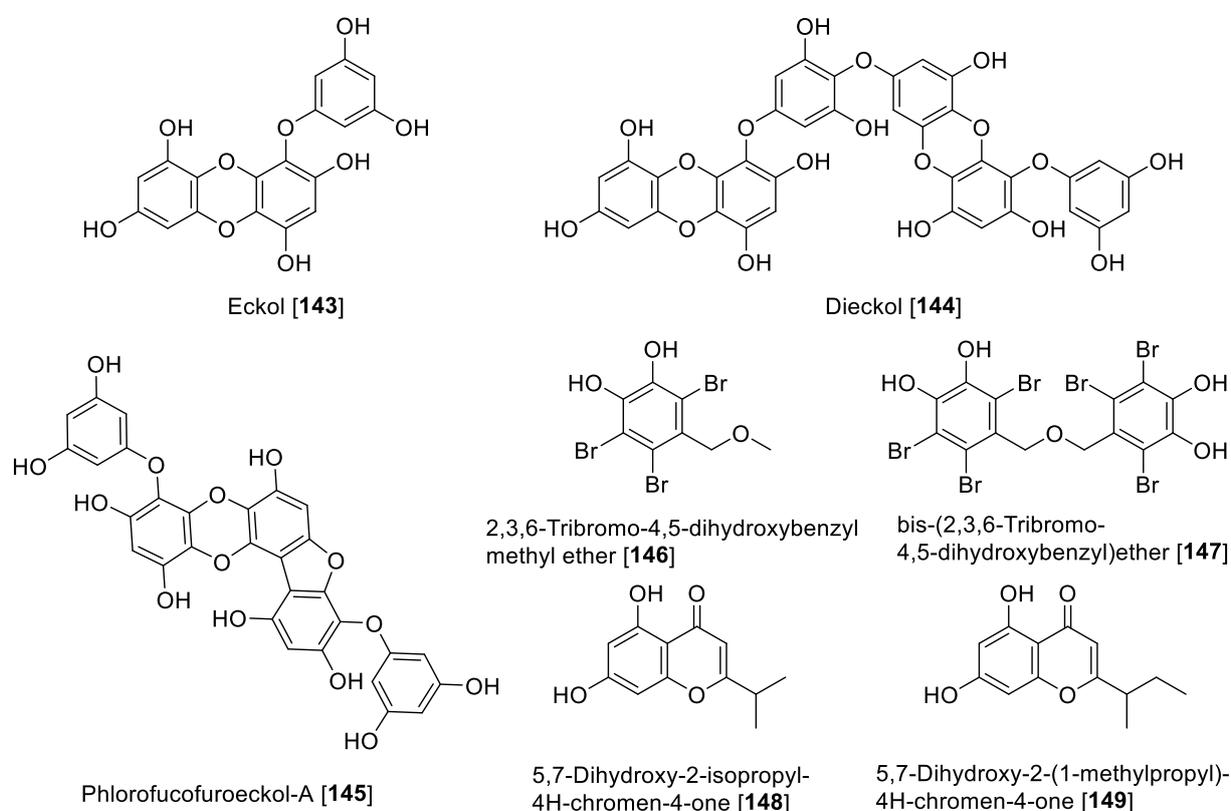


**Figure 13.** Chemical structure of selected terpenoid MAO inhibitors.

### 5.7. MAO Inhibitors from Marine Sources

In the last three decades, the study of marine environments has been the source of great chemical diversity, and at least six isolated compounds from marine organisms have been FDA approved, with several others currently in clinical trials [187,188]. Several marine natural products have been reported with inhibitory activity toward MAO enzymes, including aplysinopsins, piloquinones, anthiaactins, bromopyrroles, caulerpins, and astaxanthins, which have been compiled in a recent review [189]. As an update of the review mentioned before, two well-known phlorotannins isolated from the edible brown alga *Eisenia bicyclis* were found to be MAO inhibitors [190]. The two phlorotannins named eckol **143** and dieckol **144** showed a preferential human MAO-A inhibition with  $IC_{50}$  values of 7.2 and 11.43  $\mu\text{M}$ , respectively [190]. Phlorofucofuroeckol-A **145** (PFF-A), another phlorotannin isolated from *Ecklonia stolonifera* (edible brown alga), showed non-selective MAO inhibition. PFF-A **145**

showed  $IC_{50}$  values of 9.22 and 4.89  $\mu\text{M}$  for human MAO-A and MAO-B, respectively [191]. Compounds **144** and **145** also exhibited dopamine  $D_3R$  and  $D_4R$  agonism as well as  $D_1$ , serotonin  $5\text{-HT}_{1A}$ , and neurokinin  $NK_1$  antagonism [191]. From the red alga *Symphyclocladia latiuscula*, a series of bromophenols was isolated and 2,3,6-tribromo-4,5-dihydroxybenzyl methyl ether **146** and bis-(2,3,6-tribromo-4,5-dihydroxybenzyl) ether **147** exhibited moderate inhibition of human MAO-A with  $IC_{50}$  values of 63.2 and 89.3  $\mu\text{M}$ , respectively [192]. Two chromenones isolated from the marine microorganism *Streptomyces* sp (CNQ-031), 5,7-dihydroxy-2-isopropyl-4H-chromen-4-one **148** and 5,7-dihydroxy-2-(1-methylpropyl)-4H-chromen-4-one **149**, were found to be inhibitors for human MAO. Compound **148** inhibited human MAO-A with a 10-fold difference than for MAO-B with  $IC_{50}$  values of 2.7 and 27.0  $\mu\text{M}$ , respectively, while **149** showed a non-selective inhibition with  $IC_{50}$  values of 6.92 and 3.42  $\mu\text{M}$  for human MAO-A and MAO-B, respectively [123]. Figure 14 shows marine compounds with MAO inhibition and Table 7 presents the  $IC_{50}$  values of marine MAO inhibitors.

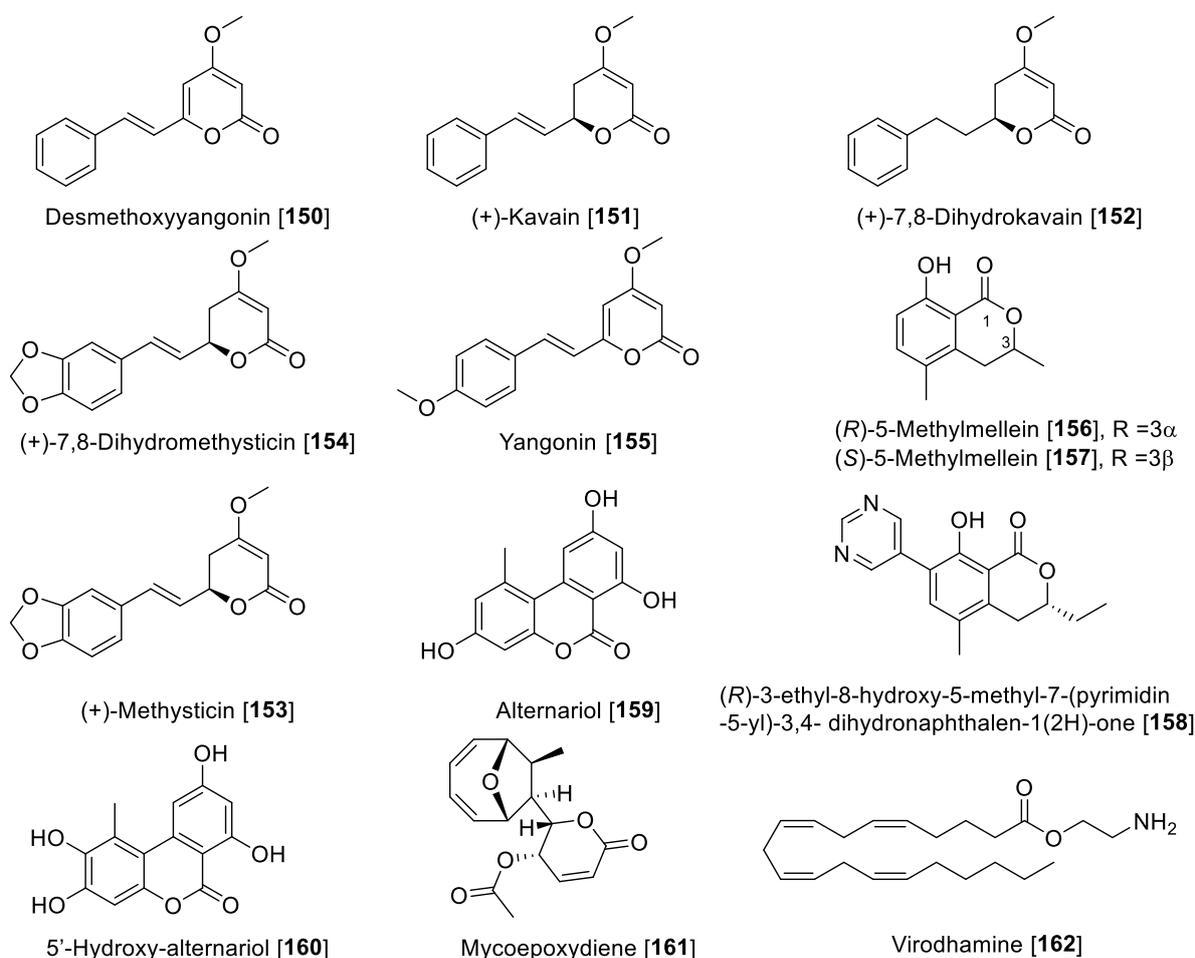


**Figure 14.** Chemical structure of selected marine isolated compound MAO inhibitors.

### 5.8. MAO Inhibitors from Miscellaneous Classes of Natural Products

Small polyketides have been reported as MAOIs; for example, desmethoxyyangonin **150** isolated from the medicinal plant *Renealmia Alpinia* was found to be a MAO inhibitor, being selective toward human MAO-B with an  $IC_{50}$  value of 0.12  $\mu\text{M}$  compared to an  $IC_{50}$  value of 1.85  $\mu\text{M}$  for MAO-A [18]. Desmethoxyyangonin **150** belongs to the well-known kavalactone-type secondary metabolites which are the major constituents isolated predominantly from *Piper methysticum* roots, the principal ingredient of the kava drink [193]. The kava drink has been used for centuries by the natives of the pacific islands of Vanuatu, Fiji, Tonga, Samoa, and Micronesia due to its psychoactive and medicinal properties. The kava drink is also a popular drink in Western countries [194]. Besides **150**, the kavalactones: (+)-kavain **151**, (+)-7,8-dihydrokavain **152**, (+)-methysticin **153**, (+)-7,8-dihydromethysticin **154**, and yangonin **155** were reported to be selective MAO-B inhibitors with  $IC_{50}$  values of 4.34, 8.23, 0.42, 0.85, and 0.085  $\mu\text{M}$ , respectively. Only **153** and **155** exhibited inhibition of human

MAO-A with  $IC_{50}$  values of 8.12 and 1.29  $\mu\text{M}$ , respectively [195]. Another small polyketide, (*S*)-5-methylmellein **156**, a small lactone isolated from the endogenous fungus *Rosellinia corticium* within the lichen *Pseudevernia furfuracea*, exhibited MAO inhibition. Compound **156** was identified through bioassay guided fractionation and exhibited  $IC_{50}$  values of 5.31 and 9.15  $\mu\text{M}$  for human MAO-A and MAO-B, respectively [196]. Its enantiomer, (*R*)-5-methylmellein **157**, was also isolated from the fermented mycelia of *Xylaria nigripes*, and **157** showed moderate selectivity toward MAO-A with  $IC_{50}$  values of 4.6 and 38.5  $\mu\text{M}$  for human MAO-A and MAO-B, respectively [197]. The enantiomers were subjected to a synthetic study aiming to find better inhibitory agents, and the study found a selective human MAO-B inhibitor in a pyrimidin derivative, (*R*)-3-ethyl-8-hydroxy-5-methyl-7-(pyrimidin-5-yl)-3,4-dihydronaphthalen-1(2H)-one **158**, with an  $IC_{50}$  value of 0.06  $\mu\text{M}$  for human MAO-B and SI value of >830 [197]. From another endogenous fungus *Diaporthe mahothocarpus* isolated from the lichen *Cladonia symphyocarpia*, three polyketides were isolated with potent MAO inhibition. The polyketides alternariol **159**, 5'-hydroxy-alternariol **160**, and mycoepoxydiene **161** were isolated following a bioassay guided paradigm and were found to be MAO-A inhibitors. Compound **159** exhibited the best value with an  $IC_{50}$  of 0.020  $\mu\text{M}$ , while **160** and **161** exhibited  $IC_{50}$  values of 0.31 and 8.7  $\mu\text{M}$  for human MAO-A, respectively [198]. The in vitro evaluation of several endogenous endocannabinoids found that virodhamine **162** inhibited both MAO-A and -B, being preferential to MAO-B with  $IC_{50}$  values of 38.70 and 0.71  $\mu\text{M}$  for human MAO-A and MAO-B, respectively [199]. Figure 15 shows the structures and Table 7 presents the  $IC_{50}$  values of selected miscellaneous natural product classes on natural product inhibitors.



**Figure 15.** Chemical structure of selected miscellaneous compound MAO inhibitors.

Table 7. Terpenoids, marine sources, and miscellaneous natural product inhibitors of MAO-A and MAO-B.

Compounds	Source	MAO-A		MAO-B		SI	Enzyme Source	References
		IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	IC <sub>50</sub> (μM)	K <sub>i</sub> (μM)	MAOA/B		
Dihydrotanshinone I [136]	<i>Salvia miltiorrhiza</i>	23					F	[183]
Cryptotanshinone [137]	<i>Salvia miltiorrhiza</i>	80					F	[183]
Tanshinone I [138]	<i>Salvia miltiorrhiza</i>	84					F	[183]
Illudinine [140]	<i>Clitocybe illudens</i>			18.3			F	[185]
Eckol [143]	<i>Eisenia bicyclis</i>	7.20	20.26	83.44	162.8	0.086	F	[185]
Dieckol [144]	<i>Eisenia bicyclis</i>	11.43	20.28	43.42	18.50	0.26	F	[190]
Phlorofucofuroeckol-A [145]	<i>Ecklonia stolonifera</i>	9.22	5.18	4.89	2.69	1.88	F	[190]
2,3,6-Tribromo-4,5-dihydroxybenzyl methyl ether [146]	<i>Symphycocladia latiuscula</i>	63.16	25.4	105.13	40.7	0.60	F	[191]
bis-(2,3,6-Tribromo-4,5-dihydroxybenzyl) ether [147]	<i>Symphycocladia latiuscula</i>	89.31	22.8	102.53	35.5	0.87	F	[191]
5,7-Dihydroxy-2-isopropyl-4H-chromen-4-one [148]	<i>Streptomyces</i> spp.	2.70		27.0		0.10	F	[192]
5,7-Dihydroxy-2-(1-methylpropyl)-4H-chromen-4-one [149]	<i>Streptomyces</i> spp.	6.92		3.42		2.02	F	[192]
Desmethoxyyangonin [150]	<i>Renealmia alpinia</i>	1.85	0.922	0.12	0.031	15.41	F	[22]
(+)-Kavain [151]	<i>Piper methysticum</i>	19.0		5.34		3.55	F	[193]
(+)-7,8-Dihydrokavain [152]	<i>Piper methysticum</i>	>100		8.23		12.15	F	[193]
(+)-Methysticin [153]	<i>Piper methysticum</i>	8.12		0.42		19.33	F	[193]
(+)-7,8-Dihydromethysticin [154]	<i>Piper methysticum</i>	23.2		0.855		27.13	F	[193]
Yangonin [155]	<i>Piper methysticum</i>	1.29	1.12	0.085	0.226	15.17	F	[193]
(S)-5-Methymellein [156]	<i>Rosellinia corticium</i>	5.31	2.45	9.15		0.58	F	[194]
(R)-5-Methymellein [157]	<i>Xylaria nigripes</i>	4.6		38.5		0.11	F	[195]
(R)-3-Ethyl-8-hydroxy-5-methyl-7-(pyrimidin-5-yl)-3,4-dihydronaphthalen-1(2H)-one [158]	Methymellein derivative	0.06		>50		>0.012	F	[195]
Alternariol [159]	<i>Diaporthe mahothoncarpus</i>	0.020	0.0075	20.7		0.00096	F	[196]
5'-Hydroxy-alternariol [160]	<i>Diaporthe mahothoncarpus</i>	0.31	0.116	>40		0.00775	F	[196]
Mycopoxydiene [161]	<i>Diaporthe mahothoncarpus</i>	8.7	3.76	>40		>0.21	F	[196]
Virodhamine [162]		38.7		0.71	0.258	54.50	F	[196]

Note: natural products tested on total MAO are not listed. Enzyme source: F = recombinant human MAO-A and -B.

## 6. Natural MAO Inhibitors in Neuroblastoma

Therapeutic applications of MAOIs have been established primarily for the treatment of depression and other neurological disorders. Oxidative deamination of monoamines by MAO produces oxidative stress [200]. Recent studies have suggested the role of MAO-A in antitumor immunity [201]. An enhanced T cell immunity and suppression of tumor growth were reported in MAO-A knockout mice. Treatment with clinically approved MAO-A inhibitors such as phenelzine, moclobemide, and clorgyline produced tumor suppression in preclinical animal models [201]. The role of MAOs in tumor progression and metastasis suggests these enzymes as potential anticancer drug targets [202]. Neuroblastoma, known to originate from the undifferentiated neural crest cells which differentiate into malignant neuroblastoma, is the most common extracranial solid tumor in children. Excess production of catecholamines in neuroblastoma tumor cells has been reported [203]. This may be the cause for severe hypertension associated with neuroblastoma [204–206]. Elevations in the levels of urinary metabolites of catecholamines vanillylmandelic acid and homovanillic acid are routinely assessed as prognostic biomarkers of neuroblastoma [207,208]. A report from the children oncology group suggested a correlation between the elevated expression of the vesicular monoamine transporter with clinical features, tumor biology, and metaiodobenzylguanidine (MIBG) avidity in neuroblastoma [84]. MAO-A has been shown to promote protective autophagy in human SH-SY5Y neuroblastoma cells through Bcl-2 phosphorylation [209]. These studies suggest the potential utility of MAO-A inhibitors for treatment of neuroblastoma. Isatin, an endogenous MAO inhibitor, caused a dose-dependent switch from apoptosis to necrosis in human neuroblastoma cells [210]. The natural product MAO inhibitors may also have potential utility for treatment of neuroblastoma tumors. Harmine **22**, the potent natural product alkaloid inhibitor of MAO-A, has been shown to induce apoptosis and inhibit cell proliferation of several human cancer cell lines [211,212]. Harmine **22** also induced apoptosis in different neuroblastoma cell lines such as SKNBE and KELLY (MYCN amplified) and SKNAS and SKNFI (MYCN non-amplified) [213]. Induction of progressive apoptosis of human neuroblastoma cells by harmine was also attributed to the activation of caspase-3/7 and caspase-9 [214]. Selective inhibition of dual-specificity tyrosine phosphorylation-regulated kinase (DYRK) family proteins and mitogen-activated protein kinase by harmine [214] led to the investigation of the expression of DYRK family kinases in neuroblastoma tumors [213]. A clinical study suggested the role of DYRK2 in tumorigenesis of neuroblastoma. Inhibition of DYRK2 by **22** [212,214] and activation of caspase-mediated apoptosis in neuroblastoma cells [213] suggest the treatment of neuroblastoma by **22**. Norharman **27**, a potential MAO-A inhibitor, also induced apoptotic cell death in human neuroblastoma cells [215]. Berberine treatment produced the induction of neuronal cell differentiation, attenuated cancer stemness markers, and potentiated G<sub>0</sub>/G<sub>1</sub> cell cycle arrest in neuro2a (N2a) neuroblastoma cells [216]. Concurrent inhibition of  $\beta$ -adrenergic signaling pathways with a  $\beta$ -blocker and inhibition of MAO-A with berberine **6** produced attenuation of tumor growth and an increase in differentiation of cells [217].

## 7. Concluding Remarks

This review presents a comprehensive survey of natural products, predominantly isolated or purified from plant sources, which have shown promising MAO inhibitory activity. The natural product MAOIs may be utilized as potential leads for new drug discovery and safer alternatives for treatment of neurological disorders. The neuroactive effects of several traditionally used herbal formulations have been attributed to the presence of the constituents with selective inhibition of MAOs; for example, the potent inhibition of MAO-A by harmala alkaloids is responsible for the hallucinogenic effects of ayahuasca. The survey was further extended to analyze different classes of natural product MAOIs and selectivity of the inhibition by these constituents. Natural products with selective and reversible inhibition of MAOs may be advanced to clinical evaluation for treatment

of disorders linked to pathophysiological consequences of the metabolism of biogenic monoamines.

Selective MAO-A and -B inhibitors isolated from natural sources may be safer alternatives for the treatment of neurological disorders. A greater understanding of the natural product inhibitors for MAOs may improve the drug discovery and treatment of neurological disorders. Evaluation of MAOIs as constituents of natural product ingredients used in dietary supplements and traditional herbal preparations may also be important for preventing adverse drug–dietary supplement interactions.

Naturally occurring MAOIs are good alternatives for the treatment of depression, anxiety, Parkinson’s disease, and neurodegenerative diseases. Non-selective and irreversible classical MAOIs are characterized by the risk of inducing hypertensive crisis when foods rich in tyramine are ingested. However, selective MAO-B inhibitors do not show such interactions. Several natural products with selective reversible inhibition of MAO-B have been identified. The selective MAO-B inhibitors and RIMAs are potential and safer alternatives to irreversible MAOIs. A greater understanding of the pharmacodynamic and pharmacokinetic properties of the natural product MAOIs may improve treatment of patients in the future. Another challenge for future basic and clinical research is to examine the potential preventive medications that slow the physiological and other age-related activities. The natural products are also an important source of MAOIs that can slow the age-related behavioral problems and decrease susceptibility to senile depression, Parkinson’s disease, and Alzheimer’s disease. Characterization of MAO inhibitory constituents of natural products traditionally used as psychoactive preparations or for treatment of neurological disorders may help in understanding the mechanisms of action and in optimization of these preparations for desired bioactive properties. Potential therapeutic applications of natural product MAOIs for the treatment of neuroblastoma are also discussed.

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## References

1. Edmondson, D.E.; Binda, C. Monoamine Oxidases. *Subcell Biochem.* **2018**, *87*, 117–139. [[CrossRef](#)] [[PubMed](#)]
2. Ramsay, R.R. Monoamine oxidases: The biochemistry of the proteins as targets in medicinal chemistry and drug discovery. *Curr. Top. Med. Chem.* **2012**, *12*, 2189–2209. [[CrossRef](#)] [[PubMed](#)]
3. Klinman, J.P. The multi-functional topa-quinone copper amine oxidases. *Biochim. Biophys. Acta* **2003**, *1647*, 131–137. [[CrossRef](#)]
4. Abell, C.W.; Kwan, S.W. Molecular characterization of monoamine oxidases A and B. *Prog. Nucleic Acid. Res. Mol. Biol.* **2001**, *65*, 129–156. [[CrossRef](#)]

5. Goldstein, D.S.; Castillo, G.; Sullivan, P.; Sharabi, Y. Differential Susceptibilities of Catecholamines to Metabolism by Monoamine Oxidases. *J. Pharmacol. Exp. Ther.* **2021**, *379*, 253–259. [[CrossRef](#)]
6. Ramonet, D.; Rodriguez, M.; Saura, J.; Lizcano, J.M.; Romera, M.; Unzeta, M.; Finch, C.; Billett, E.; Mahy, N. Localization of monoamine oxidase A and B and semicarbazide-sensitive amine oxidase in human peripheral tissues. *Inflammopharmacology* **2003**, *11*, 111–117. [[CrossRef](#)]
7. Rodriguez, M.J.; Saura, J.; Billett, E.E.; Finch, C.C.; Mahy, N. Cellular localization of monoamine oxidase A and B in human tissues outside of the central nervous system. *Cell Tissue Res.* **2001**, *304*, 215–220. [[CrossRef](#)]
8. Hubalek, F.; Pohl, J.; Edmondson, D.E. Structural comparison of human monoamine oxidases A and B: Mass spectrometry monitoring of cysteine reactivities. *J. Biol. Chem.* **2003**, *278*, 28612–28618. [[CrossRef](#)]
9. Bach, A.W.; Lan, N.C.; Johnson, D.L.; Abell, C.W.; Bembenek, M.E.; Kwan, S.W.; Seeburg, P.H.; Shih, J.C. cDNA cloning of human liver monoamine oxidase A and B: Molecular basis of differences in enzymatic properties. *Proc. Natl. Acad. Sci. USA* **1988**, *85*, 4934–4938. [[CrossRef](#)]
10. Hubalek, F.; Binda, C.; Khalil, A.; Li, M.; Mattevi, A.; Castagnoli, N.; Edmondson, D.E. Demonstration of isoleucine 199 as a structural determinant for the selective inhibition of human monoamine oxidase B by specific reversible inhibitors. *J. Biol. Chem.* **2005**, *280*, 15761–15766. [[CrossRef](#)]
11. Son, S.Y.; Ma, J.; Kondou, Y.; Yoshimura, M.; Yamashita, E.; Tsukahara, T. Structure of human monoamine oxidase A at 2.2-Å resolution: The control of opening the entry for substrates/inhibitors. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 5739–5744. [[CrossRef](#)] [[PubMed](#)]
12. De Colibus, L.; Li, M.; Binda, C.; Lustig, A.; Edmondson, D.E.; Mattevi, A. Three-dimensional structure of human monoamine oxidase A (MAO A): Relation to the structures of rat MAO A and human MAO B. *Proc. Natl. Acad. Sci. USA* **2005**, *102*, 12684–12689. [[CrossRef](#)] [[PubMed](#)]
13. Ma, J.; Yoshimura, M.; Yamashita, E.; Nakagawa, A.; Ito, A.; Tsukahara, T. Structure of rat monoamine oxidase A and its specific recognitions for substrates and inhibitors. *J. Mol. Biol.* **2004**, *338*, 103–114. [[CrossRef](#)] [[PubMed](#)]
14. Ma, J.; Kubota, F.; Yoshimura, M.; Yamashita, E.; Nakagawa, A.; Ito, A.; Tsukahara, T. Crystallization and preliminary crystallographic analysis of rat monoamine oxidase A complexed with clorgyline. *Acta Crystallogr. D Biol. Crystallogr.* **2004**, *60*, 317–319. [[CrossRef](#)] [[PubMed](#)]
15. Binda, C.; Li, M.; Hubalek, F.; Restelli, N.; Edmondson, D.E.; Mattevi, A. Insights into the mode of inhibition of human mitochondrial monoamine oxidase B from high-resolution crystal structures. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 9750–9755. [[CrossRef](#)]
16. Jones, D.N.; Raghanti, M.A. The role of monoamine oxidase enzymes in the pathophysiology of neurological disorders. *J. Chem. Neuroanat* **2021**, *114*, 101957. [[CrossRef](#)]
17. Fowler, J.S.; Volkow, N.D.; Logan, J.; Franceschi, D.; Wang, G.J.; MacGregor, R.; Shea, C.; Garza, V.; Pappas, N.; Carter, P.; et al. Evidence that L-deprenyl treatment for one week does not inhibit MAO A or the dopamine transporter in the human brain. *Life Sci.* **2001**, *68*, 2759–2768. [[CrossRef](#)]
18. Chaurasiya, N.D.; Leon, F.; Ding, Y.; Gomez-Betancur, I.; Benjumea, D.; Walker, L.A.; Cutler, S.J.; Tekwani, B.L. Interactions of Desmethoxyyangonin, a Secondary Metabolite from *Renalemia alpinia*, with Human Monoamine Oxidase-A and Oxidase-B. *Evid. Based Compl. Alternat Med.* **2017**, *2017*, 1–10. [[CrossRef](#)]
19. Liebowitz, M.R.; Hollander, E.; Schneier, F.; Campeas, R.; Welkowitz, L.; Hatterer, J.; Fallon, B. Reversible and irreversible monoamine oxidase inhibitors in other psychiatric disorders. *Acta Psychiatr. Scand. Suppl.* **1990**, *360*, 29–34. [[CrossRef](#)]
20. Priest, R.G.; Gimbrett, R.; Roberts, M.; Steinert, J. Reversible and selective inhibitors of monoamine oxidase A in mental and other disorders. *Acta Psychiatr. Scand. Suppl.* **1995**, *386*, 40–43. [[CrossRef](#)]
21. Amrein, R.; Hetzel, W.; Stabl, M.; Schmid-Burgk, W. RIMA: A safe concept in the treatment of depression with moclobemide. *Can. J. Psychiatry* **1992**, *37* (Suppl. S1), 7–11. [[PubMed](#)]
22. Mann, J.J.; Aarons, S.F.; Frances, A.J.; Brown, R.D. Studies of selective and reversible monoamine oxidase inhibitors. *J. Clin. Psychiatry* **1984**, *45*, 62–66. [[PubMed](#)]
23. Suchting, R.; Tirumalajaru, V.; Gareeb, R.; Bockmann, T.; de Dios, C.; Aickareth, J.; Pinjari, O.; Soares, J.C.; Cowen, P.J.; Selvaraj, S. Revisiting monoamine oxidase inhibitors for the treatment of depressive disorders: A systematic review and network meta-analysis. *J. Affect Disord.* **2021**, *282*, 1153–1160. [[CrossRef](#)] [[PubMed](#)]
24. Silverman, R.B.; Ding, C.Z.; Gates, K.S. Design and mechanism of monoamine oxidase inactivators from an organic chemical perspective. In *Perspectives in Medicinal Chemistry*; Testa, B., Kyburz, E., Fuhrer, W., Giger, R., Eds.; Verlag Helvetica Chimica: Basel, Switzerland, 1993; pp. 73–86.
25. Kanazawa, I. Short review on monoamine oxidase and its inhibitors. *Eur. Neurol.* **1994**, *34*, 36–39. [[CrossRef](#)] [[PubMed](#)]
26. Binda, C.; Newton-Vinson, P.; Hubalek, F.; Edmondson, D.E.; Mattevi, A. Structure of human monoamine oxidase B, a drug target for the treatment of neurological disorders. *Nat. Struct. Biol.* **2002**, *9*, 22–26. [[CrossRef](#)]
27. Dhiman, P.; Malik, N.; Sobarzo-Sanchez, E.; Uriarte, E.; Khatkar, A. Quercetin and Related Chromenone Derivatives as Monoamine Oxidase Inhibitors: Targeting Neurological and Mental Disorders. *Molecules* **2019**, *24*, 418. [[CrossRef](#)]
28. Bolasco, A.; Carradori, S.; Fioravanti, R. Focusing on new monoamine oxidase inhibitors. *Exp. Opin. Ther. Pat.* **2010**, *20*, 909–939. [[CrossRef](#)]
29. Riederer, P.L.G. MAO-inhibitors in Parkinson's Disease. *Exp. Neurobiol.* **2011**, *20*, 1–17. [[CrossRef](#)]

30. Kirwin, J.L.; Gören, J.L. Duloxetine: A dual serotonin-norepinephrineuptake inhibitor for treatment of major depressive disorder. *Pharmacotherapy* **2005**, *25*, 396–410. [[CrossRef](#)]
31. Moret, C.; Isaac, M.; Briley, M. Problems associated with long-term treatment with selective serotonin reuptake inhibitors. *J. Psychopharmacol.* **2009**, *23*, 967–974. [[CrossRef](#)]
32. Fowler, J.S.; Logan, J.; Azzaro, A.J.; Fielding, R.M.; Zhu, W.; Poshusta, A.K.; Burch, D.; Brand, B.; Free, J.; Asgharnejad, M.; et al. Reversible inhibitors of monoamine oxidase-A (RIMAs): Robust, reversible inhibition of human brain MAO-A by CX157. *Neuropsychopharmacology* **2010**, *35*, 623–631. [[CrossRef](#)] [[PubMed](#)]
33. Gleiter, C.H.; Muhlbauer, B.; Schulz, R.M.; Nilsson, E.; Antonin, K.H.; Bieck, P.R. Monoamine oxidase inhibition by the MAO-A inhibitors brofaromine and clorgyline in healthy volunteers. *J. Neural Transm. Gen. Sect.* **1994**, *95*, 241–245. [[CrossRef](#)] [[PubMed](#)]
34. Ebadi, M.; Brown-Borg, H.; Ren, J.; Sharma, S.; Shavali, S.; El ReFaey, H.; Carlson, E.C. Therapeutic efficacy of selegiline in neurodegenerative disorders and neurological diseases. *Curr. Drug Targets* **2006**, *7*, 1513–1529. [[CrossRef](#)] [[PubMed](#)]
35. Riederer, P.; Lachenmayer, L.; Laux, G. Clinical applications of MAO-inhibitors. *Curr. Med. Chem.* **2004**, *11*, 2033–2043. [[CrossRef](#)]
36. Youdim, M.B.; Bakhle, Y.S. Monoamine oxidase: Isoforms and inhibitors in Parkinson's disease and depressive illness. *Br. J. Pharmacol.* **2006**, *147* (Suppl. S1), S287–S296. [[CrossRef](#)]
37. Stocchi, F. Rasagiline: Defining the role of a novel therapy in the treatment of Parkinson's disease. *Int. J. Clin. Pract.* **2006**, *60*, 215–221. [[CrossRef](#)]
38. Lecht, S.; Haroutiunian, S.; Hoffman, A.; Lazarovici, P. Rasagiline—a novel MAO B inhibitor in Parkinson's disease therapy. *Ther. Clin. Risk Manag.* **2007**, *3*, 467–474.
39. Chen, J.J.; Ly, A.V. Rasagiline: A second-generation monoamine oxidase type-B inhibitor for the treatment of Parkinson's disease. *Am. J. Health Syst. Pharm.* **2006**, *63*, 915–928. [[CrossRef](#)]
40. McGrath, P.J.; Blood, D.K.; Stewart, J.W.; Harrison, W.; Quitkin, F.M.; Tricamo, E.; Markowitz, J. A comparative study of the electrocardiographic effects of phenelzine, tricyclic antidepressants, mianserin, and placebo. *J. Clin. Psychopharmacol.* **1987**, *7*, 335–339. [[CrossRef](#)]
41. Tibrewal, P.; Kanigere, M.K.; Looi, J.C.; Allison, S.; Bastiampillai, T. The evidence for the use of long-term benzodiazepines in the setting of treatment-refractory anxiety disorders. *Aust. N. Z. J. Psychiatry* **2022**, *56*, 723–724. [[CrossRef](#)]
42. Sathyanarayana Rao, T.S.; Yeragani, V.K. Hypertensive crisis and cheese. *Indian J Psychiatry* **2009**, *51*, 65–66. [[CrossRef](#)] [[PubMed](#)]
43. Flockhart, D.A. Dietary restrictions and drug interactions with monoamine oxidase inhibitors: An update. *J. Clin. Psychiatry* **2012**, *73* (Suppl. S1), 17–24. [[CrossRef](#)] [[PubMed](#)]
44. Baldwin, D.S. Sexual dysfunction associated with antidepressant drugs. *Exp. Opin. Drug Saf.* **2004**, *3*, 457–470. [[CrossRef](#)] [[PubMed](#)]
45. Cohen, S.; Kuhn, K.U.; Strater, B.; Scherbaum, N.; Weig, W. Adverse side-effect on sexual function caused by psychotropic drugs and psychotropic substances. *Nervenarzt* **2010**, *81*, 1129–1137. [[CrossRef](#)]
46. Mago, R.T.N.; Andrade, C. Cardiovascular adverse effects of newer antidepressants. *Exp. Rev. Neurother.* **2014**, *14*, 539–551. [[CrossRef](#)]
47. Reichenpfader, U.G.G.; Morgan, L.C.; Greenblatt, A.; Nussbaumer, B.; Hansen, R.A. Sexual dysfunction associated with second-generation antidepressants in patients with major depressive disorder: Results from a systematic review with network meta-analysis. *Drug Saf.* **2014**, *37*, 19–31. [[CrossRef](#)]
48. Cappetta, K.B.C.; Johnson, J.A.; Bloch, M.H. Meta-analysis: Risk of dry mouth with second generation antidepressants. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **2018**, *84*, 282–293. [[CrossRef](#)]
49. Rapaport, M.H. Dietary restrictions and drug interactions with monoamine oxidase inhibitors: The state of the art. *J. Clin. Psychiatry* **2007**, *68* (Suppl. S8), 42–46.
50. Remick, R.A. Anticholinergic side effects of tricyclic antidepressants and their management. *Prog. Neuropsychopharmacol. Biol. Psychiatry* **1988**, *12*, 225–231. [[CrossRef](#)]
51. Brambilla, P.C.A.; Hotopf, M.; Barbui, C. Side-effect profile of fluoxetine in comparison with other SSRIs, tricyclic and newer antidepressants: A meta-analysis of clinical trial data. *Pharmacopsychiatry* **2005**, *38*, 69–77. [[CrossRef](#)]
52. Nair, N.P.; Ahmed, S.K.; Kin, N.M.; West, T.E. Reversible and selective inhibitors of monoamine oxidase A in the treatment of depressed elderly patients. *Acta Psychiatrica Scand. Suppl.* **1995**, *386*, 28–35. [[CrossRef](#)] [[PubMed](#)]
53. Sarris, J. Herbal medicines in the treatment of psychiatric disorders: 10-year updated review. *Phytother. Res.* **2018**, *32*, 1147–1162. [[CrossRef](#)] [[PubMed](#)]
54. Moragrega, I.; Rios, J.L. Medicinal Plants in the Treatment of Depression: Evidence from Preclinical Studies. *Planta Med.* **2021**, *87*, 656–685. [[CrossRef](#)] [[PubMed](#)]
55. Martins, J.S.B. Phytochemistry and pharmacology of anti-depressant medicinal plants: A review. *Biomed. Pharmacother.* **2018**, *104*, 343–365. [[CrossRef](#)]
56. Geck, M.S.; Lecca, D.; Marchese, G.; Casu, L.; Leonti, M. Ethnomedicine and neuropsychopharmacology in Mesoamerica. *J. Ethnopharmacol.* **2021**, *278*, 114243. [[CrossRef](#)]
57. Rehman, M.U.; Wali, A.F.; Ahmad, A.; Shakeel, S.; Rasool, S.; Ali, R.; Rashid, S.M.; Madkhali, H.; Ganaie, M.A.; Khan, R. Neuroprotective Strategies for Neurological Disorders by Natural Products: An update. *Curr. Neuropharmacol.* **2019**, *17*, 247–267. [[CrossRef](#)]

58. Ji, H.F.; Zhang, H.Y. Multipotent natural agents to combat Alzheimer's disease. Functional spectrum and structural features. *Acta Pharmacol. Sin.* **2008**, *29*, 143–151. [[CrossRef](#)]
59. Schultes, R.E.; Raffauf, R.F. *The Healing Forest: Medicinal and Toxic Plants of the Northwest Amazonia*; Dioscorides Press: Portland, OR, USA, 1990.
60. Samoylenko, V.; Rahman, M.M.; Tekwani, B.L.; Tripathi, L.M.; Wang, Y.H.; Khan, S.I.; Khan, I.A.; Miller, L.S.; Joshi, V.C.; Muhammad, I. Banisteriopsis caapi, a unique combination of MAO inhibitory and antioxidative constituents for the activities relevant to neurodegenerative disorders and Parkinson's disease. *J. Ethnopharmacol.* **2010**, *127*, 357–367. [[CrossRef](#)]
61. Glover, V.; Liebowitz, J.; Armando, I.; Sandler, M. beta-Carbolines as selective monoamine oxidase inhibitors: In vivo implications. *J. Neural Transm.* **1982**, *54*, 209–218. [[CrossRef](#)]
62. Morales-Garcia, J.A.; de la Fuente Revenga, M.; Alonso-Gil, S.; Rodriguez-Franco, M.I.; Feilding, A.; Perez-Castillo, A.; Riba, J. The alkaloids of Banisteriopsis caapi, the plant source of the Amazonian hallucinogen Ayahuasca, stimulate adult neurogenesis in vitro. *Sci. Rep.* **2017**, *7*, 5309. [[CrossRef](#)]
63. Das, T.; Saha, S.C.; Sunita, K.; Majumder, M.; Ghorai, M.; Mane, A.B.; Prasanth, D.A.; Kumar, P.; Pandey, D.K.; Al-Tawaha, A.R.; et al. Promising botanical-derived monoamine oxidase (MAO) inhibitors: Pharmacological aspects and structure-activity studies. *S. Afr. J. Bot.* **2022**, *146*, 127–145. [[CrossRef](#)]
64. Ege, T.S.H.D. Monoamine oxidase inhibitory effects of medicinal plants in management of Alzheimer's dis-ease. *J. Turk. Chem. Soc.* **2021**, *8*, 239–248.
65. Verpoorte, R. *Alkaloids in Encyclopedia of Analytical Sciences*, 2nd ed.; Worsfold, P., Townshend, A., Poole, C., Eds.; Elsevier Sciences: London, UK, 2005.
66. Fattorusso, E.; Tagliatela-Scafati, O. *Modern Alkaloids: Structure, Isolation, Synthesis, and Biology*; Wiley-VCH Verlag GmbH & Co. KGaA: Weinheim, Germany, 2008; p. 689.
67. Schurr, A.; Livne, A. Differential inhibition of mitochondrial monoamine oxidase from brain by hashish components. *Biochem. Pharmacol.* **1976**, *25*, 1201–1203. [[CrossRef](#)]
68. Kong, L.D.; Cheng, C.H.; Tan, R.X. Inhibition of MAO A and B by some plant-derived alkaloids, phenols and anthraquinones. *J. Ethnopharmacol.* **2004**, *91*, 351–355. [[CrossRef](#)] [[PubMed](#)]
69. Lee, S.A.; Hong, S.S.; Han, X.H.; Hwang, J.S.; Oh, G.J.; Lee, K.S.; Lee, M.K.; Hwang, B.Y.; Ro, J.S. Piperine from the fruits of Piper longum with inhibitory effect on monoamine oxidase and antidepressant-like activity. *Chem. Pharm. Bull.* **2005**, *53*, 832–835. [[CrossRef](#)]
70. Lee, S.A.; Hwang, J.S.; Han, X.H.; Lee, C.; Lee, M.H.; Choe, S.G.; Hong, S.S.; Lee, D.; Lee, M.K.; Hwang, B.Y. Methylpiperate derivatives from Piper longum and their inhibition of monoamine oxidase. *Arch. Pharm. Res.* **2008**, *31*, 679–683. [[CrossRef](#)]
71. Mu, L.H.; Wang, B.; Ren, H.Y.; Liu, P.; Guo, D.H.; Wang, F.M.; Bai, L.; Guo, Y.S. Synthesis and inhibitory effect of piperine derivatives on monoamine oxidase. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 3343–3348. [[CrossRef](#)]
72. Ro, J.S.; Lee, S.S.; Lee, K.S.; Lee, M.K. Inhibition of type A monoamine oxidase by coptisine in mouse brain. *Life Sci.* **2001**, *70*, 639–645. [[CrossRef](#)]
73. Lee, S.S.; Lee, M.K. Effects of natural isoquinoline alkaloids on monoamine oxidase activity in mouse brain: Inhibition by berberine and palmatine. *Med. Sci. Res.* **1999**, *27*, 749–751.
74. Lee, S.J.; Chung, H.Y.; Lee, I.K.; Oh, S.U.; Yoo, I.D. Phenolics with Inhibitory Activity on Mouse Brain Monoamine Oxidase (MAO) from Whole Parts of Artemisia vulgaris L (Mugwort). *Food Sci. Biotechnol.* **2000**, *9*, 179–182.
75. Lee, S.S.; Yun-Choi, H.S.; Kim, E.I.; Lee, M.K. Inhibition of monoamine oxidase by higenamine. *Med. Sci. Res.* **1999**, *27*, 71–72.
76. Plazas, E.; Hagenow, S.; Avila Murillo, M.; Stark, H.; Cuca, L.E. Isoquinoline alkaloids from the roots of Zanthoxylum rigidum as multi-target inhibitors of cholinesterase, monoamine oxidase A and Abeta1-42 aggregation. *Bioorg. Chem.* **2020**, *98*, 103722. [[CrossRef](#)] [[PubMed](#)]
77. Yang, L.J.; Yang, Z.D.; Li, Z.J.; Yang, S.H.; Shu, Z.M. Stephtetrandrine A-D, bisbenzylisoquinoline alkaloids from Stephania tetrandra. *Nat. Prod. Res.* **2021**, *5*, 1–12. [[CrossRef](#)] [[PubMed](#)]
78. Lee, M.K.; Hwang, B.Y.; Lee, S.A.; Oh, G.J.; Choi, W.H.; Hong, S.S.; Lee, K.S.; Ro, J.S. 1-methyl-2-undecyl-4(1H)-quinolone as an irreversible and selective inhibitor of type B monoamine oxidase. *Chem. Pharm. Bull.* **2003**, *51*, 409–411. [[CrossRef](#)] [[PubMed](#)]
79. Chen, C.F.; Chiou, W.F.; Chou, C.J.; Liao, J.F.; Lin, L.C.; Wang, G.J.; Ueng, Y.F. Pharmacological effects of Evodia rutaecarpa and its bioactive components. *Chin. Pharm. J.* **2002**, *54*, 419–435.
80. Han, X.H.; Hong, S.S.; Lee, D.; Lee, J.J.; Lee, M.S.; Moon, D.C.; Han, K.; Oh, K.W.; Lee, M.K.; Ro, J.S.; et al. Quinolone alkaloids from evodiae fructus and their inhibitory effects on monoamine oxidase. *Arch. Pharm. Res.* **2007**, *30*, 397–401. [[CrossRef](#)]
81. Mesiti, F.; Maruca, A.; Silva, V.; Rocca, R.; Fernandes, C.; Remiao, F.; Uriarte, E.; Alcaro, S.; Gaspar, A.; Borges, F. 4-Oxoquinolines and monoamine oxidase: When tautomerism matters. *Eur. J. Med. Chem.* **2021**, *213*, 113183. [[CrossRef](#)]
82. Naidoo, D.; Roy, A.; Slavětinská, L.P.; Chukwujekwu, J.C.; Gupta, S.; Van Staden, J. New role for crinamine as a potent, safe and selective inhibitor of human monoamine oxidase B: In vitro and in silico pharmacology and modeling. *J. Ethnopharmacol.* **2020**, *248*, 112305. [[CrossRef](#)]
83. Fernandez de Arriba, A.; Lizcano, J.M.; Balsa, M.D.; Unzeta, M. Inhibition of monoamine oxidase from bovine retina by beta-carbolines. *J. Pharm. Pharmacol.* **1994**, *46*, 809–813. [[CrossRef](#)]

84. Temple, W.; Mendelsohn, L.; Kim, G.E.; Nekritz, E.; Gustafson, W.C.; Lin, L.; Giacomini, K.; Naranjo, A.; Van Ryn, C.; Yanik, G.A.; et al. Vesicular monoamine transporter protein expression correlates with clinical features, tumor biology, and MIBG avidity in neuroblastoma: A report from the Children's Oncology Group. *Eur. J. Nucl. Med. Mol. Imaging* **2016**, *43*, 474–481. [CrossRef]
85. Herraiz, T.; Chaparro, C. Analysis of monoamine oxidase enzymatic activity by reversed-phase high performance liquid chromatography and inhibition by beta-carboline alkaloids occurring in foods and plants. *J. Chromatogr.* **2006**, *1120*, 237–243. [CrossRef]
86. Herraiz, T.; González, D.; Ancín-Azpilicueta, C.; Arán, V.J.; Guillén, H. Beta-Carboline alkaloids in *Peganum harmala* and inhibition of human monoamine oxidase (MAO). *Arch. Pharm. Res.* **2010**, *48*, 839–845.
87. McKenna, D.J.; Towers, G.N.; Abbott, F. Monoamine oxidase inhibitors in south American Hallucinogenic Plants Part 2: Constituents of orally active myristicaceous hallucinogens. *J. Ethnopharmacol.* **1984**, *12*, 179–211. [CrossRef]
88. Denis, J.M.; Towers, G.H.N.; Abbott, F.S. Monoamine oxidase inhibitors in south American Hallucinogenic Plants: Tryptamine and beta-carboline constituents of *Ayahuasca*. *J. Ethnopharmacol.* **1984**, *10*, 195–223.
89. Schwarz, M.J.; Houghton, P.J.; Rose, S.; Jenner, P.; Lees, A.D. Activities of extract and constituents of *Banisteriopsis caapi* relevant to parkinsonism. *Pharmacol. Biochem. Behav.* **2003**, *75*, 627–633. [CrossRef]
90. Hong, S.S.; Han, X.H.; Park, S.Y.; Choi, W.H.; Lee, M.K.; Hur, J.D.; Ro, J.S. Monoamine Oxidase inhibitors from *Uncaria rhynchophylla*. *Nat. Prod. Sci.* **2005**, *11*, 145–149.
91. Reniers, J.; Robert, S.; Frederick, R.; Masereel, B.; Vincent, S.; Wouters, J. Synthesis and evaluation of beta-carboline derivatives as potential monoamine oxidase inhibitors. *Bioorg. Med. Chem.* **2011**, *19*, 134–144. [CrossRef]
92. Calixto, N.O.; Pinto, M.E.F.; Ramalho, S.D.; Burger, M.; Bobey, A.F.; Young, M.C.M.; Pinto, A.C. The genus *Psychotria*: Phytochemistry, chemotaxonomy, ethnopharmacology and biological properties. *J. Braz. Chem. Soc.* **2016**, *27*, 1355–1378.
93. Passos, C.S.; Simoes-Pires, C.A.; Nurisso, A.; Soldi, T.C.; Kato, L.; de Oliveira, C.M.; de Faria, E.O.; Marcourt, L.; Gottfried, C.; Carrupt, P.A.; et al. Indole alkaloids of *Psychotria* as multifunctional cholinesterases and monoamine oxidases inhibitors. *Phytochemistry* **2013**, *86*, 8–20. [CrossRef]
94. Klein-Junior, L.C.; Cretton, S.; Vander Heyden, Y.; Gasper, A.L.; Nejad-Ebrahimi, S.; Christen, P.; Henriques, A.T. Bioactive Azepine-Indole Alkaloids from *Psychotria nemorosa*. *J. Nat. Prod.* **2020**, *83*, 852–863. [CrossRef]
95. Milan, S. Neurodynamics of Prosocial Emotional Processing Following Serotonergic Stimulation with N,N-Dimethyltryptamine (DMT) and Harmine in Healthy Subjects. Available online: <https://clinicaltrials.gov/ct2/show/NCT04716335> (accessed on 29 April 2022).
96. Zhi, K.K.; Yang, Z.D.; Shi, D.F.; Yao, X.J.; Wang, M.G. Desmodeganine, a new alkaloid from the leaves of *Desmodium elegans* as a potential monoamine oxidase inhibitor. *Fitoterapia* **2014**, *98*, 160–165. [CrossRef]
97. Kumar, S.; Pandey, A.K. Chemistry and biological activities of flavonoids: An overview. *Sci. World J.* **2013**, *2013*, 162750. [CrossRef]
98. Nijveldt, R.J.; van Nood, E.; van Hoorn, D.E.; Boelens, P.G.; van Norren, K.; van Leeuwen, P.A. Flavonoids: A review of probable mechanisms of action and potential applications. *Am. J. Clin. Nutr.* **2001**, *74*, 418–425. [CrossRef]
99. Dias, M.C.; Pinto, D.; Silva, A.M.S. Plant Flavonoids: Chemical Characteristics and Biological Activity. *Molecules* **2021**, *26*, 5377. [CrossRef]
100. Agati, G.; Brunetti, C.; Fini, A.; Gori, A.; Guidi, L.; Landi, M.; Sebastiani, F.; Tattini, M. Are Flavonoids Effective Antioxidants in Plants? Twenty Years of Our Investigation. *Antioxidants* **2020**, *9*, 1098. [CrossRef]
101. Gogineni, V.; Nael, M.A.; Chaurasiya, N.D.; Elokely, K.M.; McCurdy, C.R.; Rimoldi, J.M.; Cutler, S.J.; Tekwani, B.L.; Leon, F. Computationally Assisted Lead Optimization of Novel Potent and Selective MAO-B Inhibitors. *Biomedicines* **2021**, *9*, 1304. [CrossRef]
102. Carradori, S.; Gidaro, M.C.; Petzer, A.; Costa, G.; Guglielmi, P.; Chimenti, P.; Alcaro, S.; Petzer, J.P. Inhibition of Human Monoamine Oxidase: Biological and Molecular Modeling Studies on Selected Natural Flavonoids. *J. Agric. Food Chem.* **2016**, *64*, 9004–9011. [CrossRef]
103. Wang, M.; Firrman, J.; Liu, L.; Yam, K. A Review on Flavonoid Apigenin: Dietary Intake, ADME, Antimicrobial Effects, and Interactions with Human Gut Microbiota. *Biomed. Res. Int.* **2019**, *2019*, 7010467. [CrossRef]
104. Salehi, B.; Venditti, A.; Sharifi-Rad, M.; Kregiel, D.; Sharifi-Rad, J.; Durazzo, A.; Lucarini, M.; Santini, A.; Souto, E.B.; Novellino, E.; et al. The Therapeutic Potential of Apigenin. *Int. J. Mol. Sci.* **2019**, *20*, 1305. [CrossRef]
105. Chaurasiya, N.D.; Ibrahim, M.A.; Muhammad, I.; Walker, L.A.; Tekwani, B.L. Monoamine oxidase inhibitory constituents of propolis: Kinetics and mechanism of inhibition of recombinant human MAO-A and MAO-B. *Molecules* **2014**, *19*, 18936–18952. [CrossRef]
106. Lopez-Lazaro, M. Distribution and biological activities of the flavonoid luteolin. *Mini Rev. Med. Chem.* **2009**, *9*, 31–59. [CrossRef]
107. Park, S.E.; Paudel, P.; Wagle, A.; Seong, S.H.; Kim, H.R.; Fauzi, F.M.; Jung, H.A.; Choi, J.S. Luteolin, a Potent Human Monoamine Oxidase-A Inhibitor and Dopamine D4 and Vasopressin V1A Receptor Antagonist. *J. Agric. Food Chem.* **2020**, *68*, 10719–10729. [CrossRef]
108. Chaurasiya, N.D.; Gogineni, V.; Elokely, K.M.; Leon, F.; Nunez, M.J.; Klein, M.L.; Walker, L.A.; Cutler, S.J.; Tekwani, B.L. Isolation of Acacetin from *Calea urticifolia* with Inhibitory Properties against Human Monoamine Oxidase-A and -B. *J. Nat. Prod.* **2016**, *79*, 2538–2544. [CrossRef]
109. Chaurasiya, N.D.; Zhao, J.; Pandey, P.; Doerksen, R.J.; Muhammad, I.; Tekwani, B.L. Selective Inhibition of Human Monoamine Oxidase B by Acacetin 7-Methyl Ether Isolated from *Turnera diffusa* (Damiana). *Molecules* **2019**, *24*, 810. [CrossRef]

110. Larit, F.; Elokely, K.M.; Chaurasiya, N.D.; Benyahia, S.; Nael, M.A.; Leon, F.; Abu-Darwish, M.S.; Efferth, T.; Wang, Y.H.; Belouahem-Abed, D.; et al. Inhibition of human monoamine oxidase A and B by flavonoids isolated from two Algerian medicinal plants. *Phytomedicine* **2018**, *40*, 27–36. [[CrossRef](#)]
111. Baek, S.C.; Kang, M.G.; Park, J.E.; Lee, J.P.; Lee, H.; Ryu, H.W.; Kim, H. Osthenol, a prenylated coumarin, as a monoamine oxidase A inhibitor with high selectivity. *Bioorg. Med. Chem. Lett.* **2019**, *29*, 839–843. [[CrossRef](#)]
112. Turkmenoglu, F.P.; Baysal, I.; Ciftci-Yabanoglu, S.; Yelekci, K.; Temel, H.; Pasa, S.; Ezer, N.; Calis, I.; Ucar, G. Flavonoids from *Sideritis* Species: Human Monoamine Oxidase (hMAO) Inhibitory Activities, Molecular Docking Studies and Crystal Structure of Xanthomicrol. *Molecules* **2015**, *20*, 7454–7473. [[CrossRef](#)]
113. Jiang, N.; Doseff, A.I.; Grotewold, E. Flavones: From Biosynthesis to Health Benefits. *Plants* **2016**, *5*, 27. [[CrossRef](#)]
114. Bello, O.M.; Ogbesejana, A.B.; Adetunji, C.O.; Oguntoye, S.O. Flavonoids Isolated from *Vitex grandifolia*, an Underutilized Vegetable, Exert Monoamine A & B Inhibitory and Anti-inflammatory Effects and Their Structure-activity Relationship. *Turk J. Pharm. Sci.* **2019**, *16*, 437–443. [[CrossRef](#)]
115. Lee, H.W.; Ryu, H.W.; Baek, S.C.; Kang, M.G.; Park, D.; Han, H.Y.; An, J.H.; Oh, S.R.; Kim, H. Potent inhibitions of monoamine oxidase A and B by acacetin and its 7-O-(6-O-malonylglucoside) derivative from *Agastache rugosa*. *Int. J. Biol. Macromol.* **2017**, *104*, 547–553. [[CrossRef](#)]
116. Aherne, S.A.; O'Brien, N.M. Dietary flavonols: Chemistry, food content, and metabolism. *Nutrition* **2002**, *18*, 75–81. [[CrossRef](#)]
117. Chimenti, F.; Cottiglia, F.; Bonsignore, L.; Casu, L.; Casu, M.; Floris, C.; Secci, D.; Bolasco, A.; Chimenti, P.; Granese, A.; et al. Quercetin as the active principle of *Hypericum hircinum* exerts a selective inhibitory activity against MAO-A: Extraction, biological analysis, and computational study. *J. Nat. Prod.* **2006**, *69*, 945–949. [[CrossRef](#)] [[PubMed](#)]
118. Gidaro, M.C.; Astorino, C.; Petzer, A.; Carradori, S.; Alcaro, F.; Costa, G.; Artese, A.; Rafele, G.; Russo, F.M.; Petzer, J.P.; et al. Kaempferol as Selective Human MAO-A Inhibitor: Analytical Detection in Calabrian Red Wines, Biological and Molecular Modeling Studies. *J. Agric. Food Chem.* **2016**, *64*, 1394–1400. [[CrossRef](#)] [[PubMed](#)]
119. Novaroli, L.; Reist, M.; Favre, E.; Carotti, A.; Catto, M.; Carrupt, P.A. Human recombinant monoamine oxidase B as reliable and efficient enzyme source for inhibitor screening. *Bioorg. Med. Chem.* **2005**, *13*, 6212–6217. [[CrossRef](#)] [[PubMed](#)]
120. Baek, S.C.; Park, M.H.; Ryu, H.W.; Lee, J.P.; Kang, M.G.; Park, D.; Park, C.M.; Oh, S.R.; Kim, H. Rhamnocitrin isolated from *Prunus padus* var. *seoulensis*: A potent and selective reversible inhibitor of human monoamine oxidase A. *Bioorg. Chem.* **2019**, *83*, 317–325. [[CrossRef](#)]
121. Hwang, J.S.; Lee, S.A.; Hong, S.S.; Lee, K.S.; Lee, M.K.; Hwang, B.Y.; Ro, J.S. Monoamine oxidase inhibitory components from the roots of *Sophora flavescens*. *Arch. Pharm. Res.* **2005**, *28*, 190–194. [[CrossRef](#)]
122. Zarmouh, N.O.; Eyunni, S.K.; Soliman, K.F. The Benzopyrone Biochanin-A as a reversible, competitive, and selective monoamine oxidase B inhibitor. *BMC Compl. Altern Med.* **2017**, *17*, 34. [[CrossRef](#)]
123. Oh, J.M.; Lee, C.; Nam, S.J.; Kim, H. Chromenone Derivatives as Monoamine Oxidase Inhibitors from Marine-Derived MAR4 Clade *Streptomyces* sp. CNQ-031. *J. Microbiol. Biotechnol.* **2021**, *31*, 1022–1027. [[CrossRef](#)]
124. Prajapati, R.; Park, S.E.; Park, H.J.; Jung, H.A.; Choi, J.S. Identification of a potent and selective human monoamine oxidase A inhibitor, glycitein, an isoflavone isolated from *Pueraria lobata* flowers. *ACS Food Sci. Technol.* **2021**, *1*, 538–550. [[CrossRef](#)]
125. Mohamed, E.I.; Zaki, M.A.; Chaurasiya, N.D.; Owis, A.I.; AbouZid, S.; Wang, Y.H.; Avula, B.; Seida, A.A.; Tekwani, B.L.; Ross, S.A. Monoamine oxidases inhibitors from *Colvillea racemosa*: Isolation, biological evaluation, and computational study. *Fitoterapia* **2018**, *124*, 217–223. [[CrossRef](#)]
126. Minders, C.; Petzer, J.P.; Petzer, A.; Lourens, A.C. Monoamine oxidase inhibitory activities of heterocyclic chalcones. *Bioorg. Med. Chem. Lett.* **2015**, *25*, 5270–5276. [[CrossRef](#)] [[PubMed](#)]
127. Guglielmi, P.; Mathew, B.; Secci, D.; Carradori, S. Chalcones: Unearthing their therapeutic possibility as monoamine oxidase B inhibitors. *Eur. J. Med. Chem.* **2020**, *205*, 112650. [[CrossRef](#)] [[PubMed](#)]
128. Hou, W.C.; Lin, R.D.; Chen, C.T.; Lee, M.H. Monoamine oxidase B (MAO-B) inhibition by active principles from *Uncaria rhynchophylla*. *J. Ethnopharmacol.* **2005**, *100*, 216–220. [[CrossRef](#)] [[PubMed](#)]
129. Recalde-Gil, M.A.; Klein-Junior, L.C.; Passos, C.D.S.; Salton, J.; Bordignon, S.A.L.; Monace, F.D.; Cechinel, V.; Teresinha Henriquesa, A. Monoamine Oxidase Inhibitory Activity of Biflavonoids from Branches of *Garcinia gardneriana* (Clusiaceae). *Nat. Prod. Commun.* **2017**, *12*, 505–508. [[CrossRef](#)] [[PubMed](#)]
130. Haraguchi, H.; Tanaka, Y.; Kabbash, A.; Fujioka, T.; Ishizu, T.; Yagi, A. Monoamine oxidase inhibitors from *Gentiana lutea*. *Phytochemistry* **2004**, *65*, 2255–2260. [[CrossRef](#)]
131. Prajapati, R.; Seong, S.H.; Park, S.E.; Paudel, P.; Jung, H.A.; Choi, J.S. Isoliquiritigenin, a potent human monoamine oxidase inhibitor, modulates dopamine D1, D3, and vasopressin V1A receptors. *Sci. Rep.* **2021**, *11*, 23528. [[CrossRef](#)]
132. Kim, J.H.; Son, Y.K.; Kim, G.H.; Hwang, K.H. Xanthoangelol and 4-Hydroxyderricin Are the Major Active Principles of the Inhibitory Activities against Monoamine Oxidases on *Angelica keiskei* K. *Biomol. Ther.* **2013**, *21*, 234–240. [[CrossRef](#)]
133. Baek, S.C.; Lee, H.W.; Ryu, H.W.; Kang, M.G.; Park, D.; Kim, S.H.; Cho, M.L.; Oh, S.R.; Kim, H. Selective inhibition of monoamine oxidase A by hispidol. *Bioorg. Med. Chem. Lett.* **2018**, *28*, 584–588. [[CrossRef](#)]
134. Stefanachi, A.; Leonetti, F.; Pisani, L.; Catto, M.; Carotti, A. Coumarin: A Natural, Privileged and Versatile Scaffold for Bioactive Compounds. *Molecules* **2018**, *23*, 250. [[CrossRef](#)]

135. Sharifi-Rad, J.; Cruz-Martins, N.; Lopez-Jornet, P.; Lopez, E.P.; Harun, N.; Yeskaliyeva, B.; Beyatli, A.; Sytar, O.; Shaheen, S.; Sharopov, F.; et al. Natural Coumarins: Exploring the Pharmacological Complexity and Underlying Molecular Mechanisms. *Oxid. Med. Cell Longev.* **2021**, *2021*, 6492346. [[CrossRef](#)]
136. Rauhamaki, S.; Postila, P.A.; Niinivehmas, S.; Kortet, S.; Schildt, E.; Pasanen, M.; Manivannan, E.; Ahinko, M.; Koskimies, P.; Nyberg, N.; et al. Structure-Activity Relationship Analysis of 3-Phenylcoumarin-Based Monoamine Oxidase B Inhibitors. *Front. Chem.* **2018**, *6*, 41. [[CrossRef](#)] [[PubMed](#)]
137. Koyiparambath, V.P.; Prayaga Rajappan, K.; Rangarajan, T.M.; Al-Sehemi, A.G.; Pannipara, M.; Bhaskar, V.; Nair, A.S.; Sudevan, S.T.; Kumar, S.; Mathew, B. Deciphering the detailed structure-activity relationship of coumarins as Monoamine oxidase enzyme inhibitors—An updated review. *Chem. Biol. Drug. Des.* **2021**, *98*, 655–673. [[CrossRef](#)] [[PubMed](#)]
138. Hossain, C.F.; Okuyama, E.; Yamazaki, M. A new series of coumarin derivatives having monoamine oxidase inhibitory activity from *Monascus anka*. *Chem. Pharm. Bull.* **1996**, *44*, 1535–1539. [[CrossRef](#)]
139. Huong, D.T.; Choi, H.C.; Rho, T.C.; Lee, H.S.; Lee, M.K.; Kim, Y.H. Inhibitory activity of monoamine oxidase by coumarins from *Peucedanum japonicum*. *Arch. Pharm. Res.* **1999**, *22*, 324–326. [[CrossRef](#)] [[PubMed](#)]
140. Yun, B.S.; Lee, I.K.; Ryoo, I.J.; Yoo, I.D. Coumarins with monoamine oxidase inhibitory activity and antioxidative coumarinolignans from *Hibiscus syriacus*. *J. Nat. Prod.* **2001**, *64*, 1238–1240. [[CrossRef](#)]
141. Carotti, A.; Carrieri, A.; Chimichi, S.; Boccalini, M.; Cosimelli, B.; Gnerre, C.; Carotti, A.; Carrupt, P.A.; Testa, B. Natural and synthetic geiparvarins are strong and selective MAO-B inhibitors. Synthesis and SAR studies. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 3551–3555. [[CrossRef](#)]
142. Jeong, S.H.; Han, X.H.; Hong, S.S.; Hwang, J.S.; Hwang, J.H.; Lee, D.; Lee, M.K.; Ro, J.S.; Hwang, B.Y. Monoamine oxidase inhibitory coumarins from the aerial parts of *Dictamnus albus*. *Arch. Pharm. Res.* **2006**, *29*, 1119–1124. [[CrossRef](#)]
143. Lee, H.W.; Ryu, H.W.; Kang, M.G.; Park, D.; Lee, H.; Shin, H.M.; Oh, S.R.; Kim, H. Potent inhibition of monoamine oxidase A by decursin from *Angelica gigas* Nakai and by wogonin from *Scutellaria baicalensis* Georgi. *Int. J. Biol. Macromol.* **2017**, *97*, 598–605. [[CrossRef](#)]
144. Wang, Y.S.; Li, B.T.; Liu, S.X.; Wen, Z.Q.; Yang, J.H.; Zhang, H.B.; Hao, X.J. Anisucoumaramide, a Bioactive Coumarin from *Clausena anisum-olens*. *J. Nat. Prod.* **2017**, *80*, 798–804. [[CrossRef](#)]
145. Seong, S.H.; Ali, M.Y.; Jung, H.A.; Choi, J.S. Umbelliferone derivatives exert neuroprotective effects by inhibiting monoamine oxidase A, self-amyloidbeta aggregation, and lipid peroxidation. *Bioorg. Chem.* **2019**, *92*, 103293. [[CrossRef](#)]
146. Farina, R.; Pisani, L.; Catto, M.; Nicolotti, O.; Gadaleta, D.; Denora, N.; Soto-Otero, R.; Mendez-Alvarez, E.; Passos, C.S.; Muncipinto, G.; et al. Structure-Based Design and Optimization of Multitarget-Directed 2H-Chromen-2-one Derivatives as Potent Inhibitors of Monoamine Oxidase B and Cholinesterases. *J. Med. Chem.* **2015**, *58*, 5561–5578. [[CrossRef](#)] [[PubMed](#)]
147. Rehuman, N.A.; Oh, J.M.; Nath, L.R.; Khames, A.; Abdelgawad, M.A.; Gambacorta, N.; Nicolotti, O.; Jat, R.K.; Kim, H.; Mathew, B. Halogenated Coumarin-Chalcones as Multifunctional Monoamine Oxidase-B and Butyrylcholinesterase Inhibitors. *ACS Omega* **2021**, *6*, 28182–28193. [[CrossRef](#)] [[PubMed](#)]
148. Perveen, S.; Al-Taweel, A.M. Phenolic Compounds from the Natural Sources and Their Cytotoxicity. In *Phenolic Compounds—Natural Sources, Importance and Applications*; Soto-Hernandez, M., Palma-Tenango, M., Garcia-Mateos, M.d.R., Eds.; IntechOpen: London, UK, 2017.
149. Han, Y.S.; Van der Heijden, R.; Verpoorte, R. Biosynthesis of anthraquinones in cell cultures of the Rubiaceae. *Plant Cell Tissue Organ. Cult.* **2001**, *67*, 201–220. [[CrossRef](#)]
150. El-Awaad, I.; Bocola, M.; Beuerle, T.; Liu, B.; Beerhues, L. Bifunctional CYP81AA proteins catalyse identical hydroxylations but alternative regioselective phenol couplings in plant xanthone biosynthesis. *Nat. Commun.* **2016**, *7*, 11472. [[CrossRef](#)] [[PubMed](#)]
151. Matsuda, Y.; Gotfredsen, C.H.; Larsen, T.O. Genetic Characterization of Neosartorin Biosynthesis Provides Insight into Heterodimeric Natural Product Generation. *Org. Lett.* **2018**, *20*, 7197–7200. [[CrossRef](#)]
152. Huang, Q.; Wang, Y.; Wu, H.; Yuan, M.; Zheng, C.; Xu, H. Xanthone Glucosides: Isolation, Bioactivity and Synthesis. *Molecules* **2021**, *26*, 5575. [[CrossRef](#)]
153. Ruan, J.; Zheng, C.; Liu, Y.; Qu, L.; Yu, H.; Han, L.; Zhang, Y.; Wang, T. Chemical and Biological Research on Herbal Medicines Rich in Xanthenes. *Molecules* **2017**, *22*, 1698. [[CrossRef](#)]
154. Suzuki, O.; Katsumata, Y.; Oya, M.; Chari, V.M.; Vermes, B.; Wagner, H.; Hostettmann, K. Inhibition of type A and type B monoamine oxidases by naturally occurring xanthenes. *Planta Med.* **1981**, *42*, 17–21. [[CrossRef](#)]
155. Schaufelberger, D.; Hostettmann, K. Chemistry and pharmacology of *Gentiana lactea*. *Planta Med.* **1988**, *54*, 219–221. [[CrossRef](#)]
156. Rocha, L.; Marston, A.; Kaplan, M.A.; Stoeckli-Evans, H.; Thull, U.; Testa, B.; Hostettmann, K. An antifungal gamma-pyrone and xanthenes with monoamine oxidase inhibitory activity from *Hypericum brasiliense*. *Phytochemistry* **1994**, *36*, 1381–1385. [[CrossRef](#)]
157. Gnerre, C.; Thull, U.; Gaillard, P.; Carrupt, P.A.; Testa, B.; Fernandes, E.; Cruciani, G. Natural and synthetic xanthenes as monoamine oxidase inhibitors: Biological assay and 3D-QSAR. *Helv. Chim. Acta* **2001**, *84*, 552–570. [[CrossRef](#)]
158. Wolfender, J.L.; Hamburger, M.; Msonthi, J.D.; Hostettmann, K. Xanthenes from *Chironia krebisii*. *Phytochemistry* **1991**, *30*, 3625–3629. [[CrossRef](#)]
159. Wairata, J.; Sukandar, E.R.; Fadlan, A.; Purnomo, A.S.; Taher, M.; Ersam, T. Evaluation of the Antioxidant, Antidiabetic, and Antiplasmodial Activities of Xanthenes Isolated from *Garcinia forbesii* and Their In Silico Studies. *Biomedicines* **2021**, *9*, 1380. [[CrossRef](#)]

160. Yang, J.L.; Liu, L.L.; Shi, Y.P. Phytochemicals and biological activities of *Gentiana* species. *Nat. Prod. Commun.* **2010**, *5*, 649–664. [[CrossRef](#)] [[PubMed](#)]
161. Tomic, M.; Tovilovic, G.; Butorovic, B.; Krstic, D.; Jankovic, T.; Aljancic, I.; Menkovic, N. Neuropharmacological evaluation of diethylether extract and xanthenes of *Gentiana kochiana*. *Pharmacol. Biochem. Behav.* **2005**, *81*, 535–542. [[CrossRef](#)] [[PubMed](#)]
162. Hwang, J.H.; Hong, S.S.; Han, X.H.; Hwang, J.S.; Lee, D.; Lee, H.; Yun, Y.P.; Kim, Y.; Ro, J.S.; Hwang, B.Y. Prenylated xanthenes from the root bark of *Cudrania tricuspidata*. *J. Nat. Prod.* **2007**, *70*, 1207–1209. [[CrossRef](#)]
163. Urbain, A.; Marston, A.; Grilo, L.S.; Bravo, J.; Purev, O.; Purevsuren, B.; Batsuren, D.; Reist, M.; Carrupt, P.A.; Hostettmann, K. Xanthenes from *Gentianella amarella* ssp. *acuta* with acetylcholinesterase and monoamine oxidase inhibitory activities. *J. Nat. Prod.* **2008**, *71*, 895–897. [[CrossRef](#)]
164. Dimitrov, M.; Nikolova, I.; Benbasat, N.; Kitanov, G.; Danchev, N. Acute toxicity, antidepressive and MAO inhibitory activity of mangiferin isolated from *Hypericum aucheri*. *Biothechnol. Biotechnol. Equip.* **2011**, *25*, 2668–2671. [[CrossRef](#)]
165. Yamazaki, M.; Satoh, Y.; Maebayashi, Y.; Horie, Y. Monoamine oxidase inhibitors from a fungus, *Emericella navahoensis*. *Chem. Pharm. Bull.* **1988**, *36*, 670–675. [[CrossRef](#)]
166. Andujar, I.; Recio, M.C.; Giner, R.M.; Rios, J.L. Traditional chinese medicine remedy to jury: The pharmacological basis for the use of shikonin as an anticancer therapy. *Curr. Med. Chem.* **2013**, *20*, 2892–2898. [[CrossRef](#)]
167. Choi, W.H.; Hong, S.S.; Lee, S.A.; Han, X.H.; Lee, K.S.; Lee, M.K.; Hwang, B.Y.; Ro, J.S. Monoamine oxidase inhibitory naphthoquinones from the roots of *Lithospermum erythrorhizon*. *Arch. Pharm. Res.* **2005**, *28*, 400–404. [[CrossRef](#)] [[PubMed](#)]
168. Khalil, A.A.; Steyn, S.; Castagnoli, N., Jr. Isolation and characterization of a monoamine oxidase inhibitor from tobacco leaves. *Chem. Res. Toxicol.* **2000**, *13*, 31–35. [[CrossRef](#)] [[PubMed](#)]
169. Jaka, O.; Iturria, I.; van der Toorn, M.; Hurtado de Mendoza, J.; Latino, D.; Alzualde, A.; Peitsch, M.C.; Hoeng, J.; Koshibu, K. Effects of Natural Monoamine Oxidase Inhibitors on Anxiety-Like Behavior in Zebrafish. *Front. Pharmacol.* **2021**, *12*, 669370. [[CrossRef](#)] [[PubMed](#)]
170. Coelho Cerqueira, E.; Netz, P.A.; Diniz, C.; Petry do Canto, V.; Follmer, C. Molecular insights into human monoamine oxidase (MAO) inhibition by 1,4-naphthoquinone: Evidences for menadione (vitamin K3) acting as a competitive and reversible inhibitor of MAO. *Bioorg. Med. Chem.* **2011**, *19*, 7416–7424. [[CrossRef](#)] [[PubMed](#)]
171. Mostert, S.; Petzer, A.; Petzer, J.P. Evaluation of Natural and Synthetic 1,4-naphthoquinones as Inhibitors of Monoamine Oxidase. *Chem. Biol. Drug Des.* **2016**, *87*, 737–746. [[CrossRef](#)]
172. Lee, H.W.; Ryu, H.W.; Kang, M.G.; Park, D.; Oh, S.R.; Kim, H. Selective inhibition of monoamine oxidase A by purpurin, an anthraquinone. *Bioorg. Med. Chem. Lett.* **2017**, *27*, 1136–1140. [[CrossRef](#)]
173. Paudel, P.; Seong, S.H.; Shrestha, S.; Jung, H.A.; Choi, J.S. In Vitro and in Silico Human Monoamine Oxidase Inhibitory Potential of Anthraquinones, Naphthopyrones, and Naphthalenic Lactones from *Cassia obtusifolia* Linn Seeds. *ACS Omega* **2019**, *4*, 16139–16152. [[CrossRef](#)]
174. Tohge, T.; Watanabe, M.; Hoefgen, R.; Fernie, A.R. Shikimate and phenylalanine biosynthesis in the green lineage. *Front. Plant Sci.* **2013**, *4*, 62. [[CrossRef](#)]
175. Dhiman, P.; Malik, N.; Khatkar, A. Hybrid caffeic acid derivatives as monoamine oxidases inhibitors: Synthesis, radical scavenging activity, molecular docking studies and in silico ADMET analysis. *Chem. Cent. J.* **2018**, *12*, 112. [[CrossRef](#)]
176. Mei, Y.; Pan, D.; Jiang, Y.; Zhang, W.; Yao, X.; Dai, Y.; Yu, Y.; Yao, X. Target discovery of chlorogenic acid derivatives from the flower buds of *Lonicera macranthoides* and their MAO B inhibitory mechanism. *Fitoterapia* **2019**, *134*, 297–304. [[CrossRef](#)]
177. Wu, G.F.; Jiang, X.L.; Gong, Y.Z.; Hu, Y.D.; Bai, X.L.; Liao, X. Ligand fishing of anti-neurodegenerative components from *Lonicera japonica* using magnetic nanoparticles immobilised with monoamine oxidase B. *J. Sep. Sci.* **2019**, *42*, 1289–1298. [[CrossRef](#)] [[PubMed](#)]
178. Xu, X.Y.; Meng, X.; Li, S.; Gan, R.Y.; Li, Y.; Li, H.B. Bioactivity, Health Benefits, and Related Molecular Mechanisms of Curcumin: Current Progress, Challenges, and Perspectives. *Nutrients* **2018**, *10*, 1553. [[CrossRef](#)] [[PubMed](#)]
179. Rajeswari, A.; Sabesan, M. Inhibition of monoamine oxidase-B by the polyphenolic compound, curcumin and its metabolite tetrahydrocurcumin, in a model of Parkinson's disease induced by MPTP neurodegeneration in mice. *Inflammopharmacology* **2008**, *16*, 96–99. [[CrossRef](#)] [[PubMed](#)]
180. Baek, S.C.; Choi, B.; Nam, S.J.; Kim, H. Inhibition of monoamine oxidase A and B by demethoxycurcumin and bisdemethoxycurcumin. *J. Appl. Biol. Chem.* **2018**, *61*, 187–190. [[CrossRef](#)]
181. Arvizu-Espinosa, M.G.; von Poser, G.L.; Henriques, A.T.; Mendoza-Ruiz, A.; Cardador-Martinez, A.; Gesto-Borroto, R.; Nunez-Aragon, P.N.; Villarreal-Ortega, M.L.; Sharma, A.; Cardoso-Taketa, A. Bioactive Dimeric Acylphloroglucinols from the Mexican Fern *Elaphoglossum paleaceum*. *J. Nat. Prod.* **2019**, *82*, 785–791. [[CrossRef](#)]
182. Perveen, S. *Introductory Chapter: Terpenes and Terpenoids in Terpenes and Terpenoids*; Perveen, S., Al-Taweel, A., Eds.; IntechOpen: London, UK, 2018; p. 79683.
183. Dittmann, K.; Gerhauser, C.; Klimo, K.; Hamburger, M. HPLC-based activity profiling of *Salvia miltiorrhiza* for MAO A and iNOS inhibitory activities. *Planta Med.* **2004**, *70*, 909–913. [[CrossRef](#)]
184. Subaraja, M.; Vanisree, A.J. The novel phytochemical asiaticoside-D isolated from *Centella asiatica* exhibits monoamine oxidase-B inhibiting potential in the rotenone degenerated cerebral ganglions of *Lumbricus terrestris*. *Phytomedicine* **2019**, *58*, 152833. [[CrossRef](#)]

185. Gaston, R., Jr.; Geldenhuys, W.J.; Dudley, G.B. Synthesis of Illudinine from Dimedone and Identification of Activity as a Monoamine Oxidase Inhibitor. *J. Org. Chem.* **2020**, *85*, 13429–13437. [[CrossRef](#)]
186. Kukula-Koch, W.; Koch, W.; Czernicka, L.; Glowniak, K.; Asakawa, Y.; Umeyama, A.; Marzec, Z.; Kuzuhara, T. MAO-A Inhibitory Potential of Terpene Constituents from Ginger Rhizomes—A Bioactivity Guided Fractionation. *Molecules* **2018**, *23*, 1301. [[CrossRef](#)]
187. Jimenez, C. Marine Natural Products in Medicinal Chemistry. *ACS Med. Chem. Lett.* **2018**, *9*, 959–961. [[CrossRef](#)]
188. Lu, W.Y.; Li, H.J.; Li, Q.Y.; Wu, Y.C. Application of marine natural products in drug research. *Bioorg. Med. Chem.* **2021**, *35*, 116058. [[CrossRef](#)] [[PubMed](#)]
189. Hong, A.; Tu, L.C.; Yang, I.; Lim, K.M.; Nam, S.J. Marine natural products with monoamine oxidase (MAO) inhibitory activity. *Pharm. Biol.* **2020**, *58*, 716–720. [[CrossRef](#)] [[PubMed](#)]
190. Jung, H.A.; Roy, A.; Jung, J.H.; Choi, J.S. Evaluation of the inhibitory effects of eckol and dieckol isolated from edible brown alga *Eisenia bicyclis* on human monoamine oxidases A and B. *Arch. Pharm. Res.* **2017**, *40*, 480–491. [[CrossRef](#)] [[PubMed](#)]
191. Seong, S.H.; Paudel, P.; Choi, J.W.; Ahn, D.H.; Nam, T.J.; Jung, H.A.; Choi, J.S. Probing Multi-Target Action of Phlorotannins as New Monoamine Oxidase Inhibitors and Dopaminergic Receptor Modulators with the Potential for Treatment of Neuronal Disorders. *Mar. Drugs* **2019**, *17*, 377. [[CrossRef](#)]
192. Paudel, P.; Park, S.E.; Seong, S.H.; Jung, H.A.; Choi, J.S. Bromophenols from *Symphocladia latiuscula* Target Human Monoamine Oxidase and Dopaminergic Receptors for the Management of Neurodegenerative Diseases. *J. Agric. Food Chem.* **2020**, *68*, 2426–2436. [[CrossRef](#)]
193. Mikell, J.R.; Schaneberg, B.T.; Khan, I.A. Isolation and purification of Kava lactones by high performance centrifugal partition chromatography. *J. Liq. Chromatogr. Relat. Technol.* **2003**, *26*, 3069–3074. [[CrossRef](#)]
194. Aporosa, S.A. De-mythologizing and re-branding of kava as the new world drug of choice. *Drug Sci. Pol. Law* **2019**, *5*, 1–13. [[CrossRef](#)]
195. Prinsloo, D.; van Dyk, S.; Petzer, A.; Petzer, J.P. Monoamine Oxidase Inhibition by Kavalactones from Kava (*Piper Methysticum*). *Planta Med.* **2019**, *85*, 1136–1142. [[CrossRef](#)]
196. Jeong, G.S.; Lee, E.Y.; Kang, M.G.; Nam, S.J.; Park, D.; Kim, H. (S)-5-Methymellein isolated from an endogenous lichen fungus *Rosellinia corticum* as a potent inhibitor of human monoamine oxidase A. *Processes* **2022**, *10*, 166. [[CrossRef](#)]
197. Huang, C.; Xiong, J.; Guan, H.D.; Wang, C.H.; Lei, X.; Hu, J.F. Discovery, synthesis, biological evaluation and molecular docking study of (R)-5-methylmellein and its analogs as selective monoamine oxidase A inhibitors. *Bioorg. Med. Chem.* **2019**, *27*, 2027–2040. [[CrossRef](#)]
198. Jeong, G.S.; Hillman, P.F.; Kang, M.G.; Hwang, S.; Park, J.E.; Nam, S.J.; Park, D.; Kim, H. Potent and Selective Inhibitors of Human Monoamine Oxidase A from an Endogenous Lichen Fungus *Diaporthe mahothocarpus*. *J. Fungi* **2021**, *7*, 876. [[CrossRef](#)] [[PubMed](#)]
199. Pandey, P.; Chaurasiya, N.D.; Tekwani, B.L.; Doerksen, R.J. Interactions of endocannabinoid virodhamine and related analogs with human monoamine oxidase-A and -B. *Biochem. Pharmacol.* **2018**, *155*, 82–91. [[CrossRef](#)] [[PubMed](#)]
200. Fitzgerald, J.C.; Ugun-Klusek, A.; Allen, G.; De Girolamo, L.A.; Hargreaves, I.; Ufer, C.; Abramov, A.Y.; Billett, E.E. Monoamine oxidase-A knockdown in human neuroblastoma cells reveals protection against mitochondrial toxins. *FASEB J.* **2014**, *28*, 218–229. [[CrossRef](#)]
201. Wang, X.; Li, B.; Kim, Y.J.; Wang, Y.C.; Li, Z.; Yu, J.; Zeng, S.; Ma, X.; Choi, I.Y.; Di Biase, S.; et al. Targeting monoamine oxidase A for T cell-based cancer immunotherapy. *Sci. Immunol.* **2021**, *6*, 2383. [[CrossRef](#)]
202. Aljanabi, R.; Alsous, L.; Sabbah, D.A.; Gul, H.I.; Gul, M.; Bardaweel, S.K. Monoamine Oxidase (MAO) as a Potential Target for Anticancer Drug Design and Development. *Molecules* **2021**, *26*, 6019. [[CrossRef](#)] [[PubMed](#)]
203. Romero, M.; Kapur, G.; Baracco, R.; Valentini, R.P.; Mattoo, T.K.; Jain, A. Treatment of Hypertension in Children With Catecholamine-Secreting Tumors: A Systematic Approach. *J. Clin. Hypertens.* **2015**, *17*, 720–725. [[CrossRef](#)] [[PubMed](#)]
204. Shinohara, M.; Shitara, T.; Hatakeyama, S.I.; Suzuki, N.; Maruyama, K.; Kobayashi, T.; Tsuchida, Y. An infant with systemic hypertension, renal artery stenosis, and neuroblastoma. *J. Pediatr. Surg.* **2004**, *39*, 103–106. [[CrossRef](#)] [[PubMed](#)]
205. Moon, S.B. Childhood neuroblastoma masquerading as pheochromocytoma: Case report. *Int. Med. Case Rep. J.* **2016**, *9*, 65–67. [[CrossRef](#)]
206. Harding, M.; Deyell, R.J.; Blydt-Hansen, T. Catecholamines in neuroblastoma: Driver of hypertension, or solely a marker of disease? *Cancer Rep.* **2021**, e1569. [[CrossRef](#)]
207. Laug, W.E.; Siegel, S.E.; Shaw, K.N.; Landing, B.; Baptista, J.; Gutenstein, M. Initial urinary catecholamine metabolite concentrations and prognosis in neuroblastoma. *Pediatrics* **1978**, *62*, 77–83. [[CrossRef](#)]
208. Liu, Y.L.; Cheng, A.T.; Chen, H.R.; Hsu, Y.P. Simultaneous HPLC of twelve monoamines and metabolites shows neuroblastoma cell line releases HVA and HIAA. *Biomed. Chromatogr.* **2000**, *14*, 544–548. [[CrossRef](#)]
209. Ugun-Klusek, A.; Theodosi, T.S.; Fitzgerald, J.C.; Burte, F.; Ufer, C.; Boocock, D.J.; Yu-Wai-Man, P.; Bedford, L.; Billett, E.E. Monoamine oxidase-A promotes protective autophagy in human SH-SY5Y neuroblastoma cells through Bcl-2 phosphorylation. *Redox. Biol.* **2019**, *20*, 167–181. [[CrossRef](#)] [[PubMed](#)]
210. Igosheva, N.; Lorz, C.; O’Conner, E.; Glover, V.; Mehmet, H. Isatin, an endogenous monoamine oxidase inhibitor, triggers a dose- and time-dependent switch from apoptosis to necrosis in human neuroblastoma cells. *Neurochem. Int.* **2005**, *47*, 216–224. [[CrossRef](#)] [[PubMed](#)]

211. Matthay, K.K.; Maris, J.M.; Schleiermacher, G.; Nakagawara, A.; Mackall, C.L.; Diller, L.; Weiss, W.A. Neuroblastoma. *Nat. Rev. Dis. Primers* **2016**, *2*, 16078. [[CrossRef](#)] [[PubMed](#)]
212. Himpel, S.; Panzer, P.; Eirnbter, K.; Czajkowska, H.; Sayed, M.; Packman, L.C.; Blundell, T.; Kentrup, H.; Grotzinger, J.; Joost, H.G.; et al. Identification of the autophosphorylation sites and characterization of their effects in the protein kinase DYRK1A. *Biochem. J.* **2001**, *359*, 497–505. [[CrossRef](#)]
213. Uhl, K.L.; Schultz, C.R.; Geerts, D.; Bachmann, A.S. Harmine, a dual-specificity tyrosine phosphorylation-regulated kinase (DYRK) inhibitor induces caspase-mediated apoptosis in neuroblastoma. *Cancer Cell Int.* **2018**, *18*, 82. [[CrossRef](#)]
214. Gockler, N.; Jofre, G.; Papadopoulos, C.; Soppa, U.; Tejedor, F.J.; Becker, W. Harmine specifically inhibits protein kinase DYRK1A and interferes with neurite formation. *FEBS J.* **2009**, *276*, 6324–6337. [[CrossRef](#)]
215. Uezono, T.; Maruyama, W.; Matsubara, K.; Naoi, M.; Shimizu, K.; Saito, O.; Ogawa, K.; Mizukami, H.; Hayase, N.; Shiono, H. Norharman, an indoleamine-derived beta-carboline, but not Trp-P-2, a gamma-carboline, induces apoptotic cell death in human neuroblastoma SH-SY5Y cells. *J. Neural Transm.* **2001**, *108*, 943–953. [[CrossRef](#)]
216. Naveen, C.R.; Gaikwad, S.; Agrawal-Rajput, R. Berberine induces neuronal differentiation through inhibition of cancer stemness and epithelial-mesenchymal transition in neuroblastoma cells. *Phytomedicine* **2016**, *23*, 736–744. [[CrossRef](#)]
217. Calvani, M.; Subbiani, A.; Bruno, G.; Favre, C. Beta-Blockers and Berberine: A Possible Dual Approach to Contrast Neuroblastoma Growth and Progression. *Oxid. Med. Cell Longev.* **2020**, *7*, 534–693. [[CrossRef](#)]