

Review

Ocimum Species: A Review on Chemical Constituents and Antibacterial Activity

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Abstract: Infection by bacteria is one of the main problems in health. The use of commercial antibiotics is still one of the treatments to overcome these problems. However, high levels of consumption lead to antibiotic resistance. Several types of antibiotics have been reported to experience resistance. One solution that can be given is the use of natural antibacterial products. There have been many studies reporting the potential antibacterial activity of the *Ocimum* plant. *Ocimum* is known to be one of the medicinal plants that have been used traditionally by local people. This plant contains components of secondary metabolites such as phenolics, flavonoids, steroids, terpenoids, and alkaloids. Therefore, in this paper, we will discuss five types of *Ocimum* species, namely *O. americanum*, *O. basilicum*, *O. gratissimum*, *O. campechianum*, and *O. sanctum*. The five species are known to contain many chemical constituents and have good antibacterial activity against several pathogenic bacteria.

Keywords: *Ocimum* species; chemical constituents; antibacterial



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1. Introduction

Infection by microbes is one of the main problems that causes several diseases. One of the causes of infection is bacteria that can have an impact on public health. Based on the collection of data from 52 sentinel hospitals in North America for 7 years (1998–2004) on contemporary strains of 12,737 strains of pediatric patients under 18 years of age, *E. coli* as a pediatric pathogen ranks in the top six [1]. Mapanguy et al. [2] also reported that oral antibiotics such as cefixime, amoxicillin, and ciprofloxacin were resistant to *E. coli* infection about 50–60%. In addition, microorganisms can also cause wound infections and inhibit healing. Some of the bacteria associated with wound infections are *Staphylococcus aureus*, *Escherichia coli*, *Pseudomonas aeruginosa*, *Klebsiella pneumoniae*, *Streptococcus pyogenes*, *Proteus* spp., *Streptococcus* spp., and *Enterococcus* spp. [3,4]. Bacteria is also can be pathogenic in the skin, such as *Staphylacoccus*, *Micrococcus*, and *Corynebacterium* sp. [5]. In relation to dental and oral health, *Streptococcus sanguinis* and *Streptococcus mutans* can cause dental caries [6].

Some of the infections mentioned can be treated with antibiotics. However, several studies have proven that antibiotics can cause resistance [7]. Antibiotic resistance occurs due to the use of drugs in large quantities, causing selection pressures on human and natural microbial systems. Microbes can undergo mutations to survive, thereby reducing antibiotic sensitivity [8,9]. Infections due to drug resistance have caused the death of up to 700,000 people every year worldwide [10]. There are several antibiotic resistances, namely vancomycin and teicolanin against *S. aureus* [11], gentamicin aminoglycoside

against *E. faecalis* and *E. faecium* with percent resistance of 30% to 50% [12], penicillin against *S. pneumonia* [13], and *E. coli* caused resistance of fluorokuinolon more than 80% and gentamicin more than 40% [12].

Therefore, the use of natural antibacterial ingredients is one solution that has great potential. *Ocimum* species are herbal plants that are available in Indonesia. *Ocimum* species are native to tropical areas such as southern Asia, Africa, and India [14]. *Ocimum* comes from the *Lamiaceae* family, which has about 50 to 150 species [15]. Due to its pharmacological effects, this plant has been widely used traditionally for the treatment of headaches, coughs, diarrhea, constipation, warts, and kidney damage [16]. These properties come from the secondary metabolite components that are abundant in *Ocimum* plants such as steroids, tannins, alkaloids, flavonoids, and phenolics [17]. In addition, the abundant components of essential oils make *Ocimum* a plant that can fight the growth of organisms [18,19]. Therefore, in this paper, we will discuss five species of *Ocimum* that have been tested in vitro to have antibacterial activity against several Gram-positive and Gram-negative bacteria.

2. Chemical Constituents and Antibacterial Activity of *Ocimum* Species

2.1. *Ocimum americanum*

O. americanum is native to Africa and is 15 to 60 cm tall with sub-rectangular striated branches [20,21]. The leaf shape is intact or faintly serrated, lanceolate ellipse, glandular spots, and glabrous. The color of flower is pink, white, or purplish with an elongated circle shape. The fruits are small, pitted notelets, and mucilaginous [21]. It is commonly known as hoary basil or mosquito plant and has three chemotypes, namely spicy, camphoraceous, and floral-lemony [22]. Traditionally, this plant is used for the treatment of digestive, respiratory, and sedative disorders. It also has benefits as a cough medicine, treating bronchitis, immune disorders, relieving toothache, and dysentery, which is commonly used orally [20,23]. The extract of *O. americanum* was also used for tobacco flavoring, tea, and body fragrance. The leaves and branches were used for insecticides against mosquitoes, bees, flies, and other insects. In Africa, the Swahili tribe utilizes the aerial parts of the plant for the treatment of high blood pressure and stomach aches [24]. In addition, local people in the Tamil Nadu area use a decoction of the leaves as a medicine for diabetes, constipation, diarrhea, hemorrhoids, and dysentery [25].

O. americanum has several phytochemical components, such as alkaloids, flavonoids, phenolic, tannins, terpenoids, saponin, steroids and glicosides. Some studies reported that saponin, phenolic, and tannins are found in less polar solvent such as ethyl acetate leave extract [24], while glicosides and steroids are commonly found in methanol extract [25]. Moreover, phenolic, flavonoids, saponin, and tannins were found in the aqueous extract of leaves and flowers [26]. The pharmacological activities found in *O. americanum* are antioxidant, antifungal, antimicrobial, anti-insecticide and larvicide, and gastric cytoprotective antiulcer effect [27,28]. Chemical compounds in *O. americanum* can be seen in Table 1, Figure S1.

Table 1. The Chemical Constituents of *O. americanum*.

Table 1. Cont.

Class of Chemical Constituents	Chemical Constituents	RT (min)	References
Flavonoids	Vicenin-2 (62)	-	[26,35]
	Eriodictyol-7-O-glucoside (63)	-	
	Vitexin (64)	-	
	Rutin (65)	4.55	
	Genkwanin (66)	-	
	Dihydroxy-tetramethoxy(iso)flavone (67)	-	
	Cirsilineol (68)	-	
	Cirsimarinin (69)	-	
	Pilosin (70)	-	
	Riboflavin (71)	-	[26]
Alloxazines and isoalloxazines			

(-) = Test not performed; RT = Retention Time; RI = Retention Index.

2.2. *Ocimum basilicum*

Ocimum basilicum, commonly called sweet basil, is one of the species of genus *Ocimum* from Asia, Africa, and South America regions [36,37]. *O. basilicum* can live in different climates and ecology, grows in cool humid areas to tropical areas with the temperatures between 6 and 24 °C, and also favors warm conditions [38]. This plant is the species of *Ocimum* which is commercially available in the market [39]. It has six different morphologies, namely true basil with green leaves, small-leaf basil, which belongs to green cultivars with short narrow leaves and grows rounded, lettuce-leaf with broad leaves, purple basil A, which has green leaves with purple flowers and stems, purple basil B, where the leaves, flower, and stems are purple, and purple basil C, which has a similarity to purple basil B and has broad listed leaves [40,41]. *O. basilicum* is 20 to 80 cm tall with glabrous and woody stems, large green leaves, and is broadly elliptical, measuring 2.5 to 5 cm × 1 to 2.5 cm. The flowers are red, pink, or white, with a size of 3 mm, and are arranged in terminal spikes [42].

Traditionally, the fruit of *O. basilicum* was used as folk medicine against inflammation, diarrhea, worm infestation, and eye-related disease [43]. Leaves and flowers of *O. basilicum* were used as tonic and vermicifuge, and can also be used as a tea to treat nausea, flatulence, and dysentery. *O. basilicum* contains essential oils that are commonly used to treat colds, seizures, and treatment of wasp stings and snakebites [44]. The polysaccharide component of *O. basilicum* was traditionally used as cancer treatment in China [45,46]. In South Europe, they used *O. basilicum* as Mediterranean food, such as the cuisines of Italian and Greek [47].

O. basilicum contains the main components that are beneficial for health such as calcium, phosphorus, vitamin A, vitamin C, and beta carotene [48]. Phytochemical constituents contained in *O. basilicum* are alkaloids, flavonoids, phenols, saponins, tannins, terpenoids, carbohydrates, cardiac glycosides, cholesterol, glycosides, and phlobatannis [49,50]. Therefore, it has the potential to have anti-inflammatory, antimicrobial, antivirals, anticancer, antifungal, antidiabetic, anti-allergic, analgesic, cardioprotective, and immunomodulatory properties [50–53]. Then, flavonoids and phenolic compounds gave antioxidant activity [54,55]. Chemical compounds in *O. basilicum* can be seen in Table 2, Figure S2.

Table 2. The chemical constituents of *O. basilicum*.

Class of Chemical Constituents	Chemical Constituents	RT (min)	RI	References
Carboxylic acid ester	Cyclohexyl formate (72)	7.30	1304	[56,57]
Aliphatic aldehydes	Citral (16)	6.57	-	[56,58]
Aliphatic alcohol	3-Octanol (73)	11.64	-	[59]
Amines	Phenylethanamine (74)	8.77	-	[56,57]
Phenols	2,3,5-Trimethylphenol (75) Eugenol (33)	7.10 7.19	-	[56,58]
Fatty alcohols	4-Hexen-1-ol acetate (76) 1-Octen-3-ol (77)	8.57 10.78	4.428 979	[56,58,60,61]
Pyrans	2,3-Dehydro-1,8-cineole (78)	11.18	-	[57]
Organoheterocyclic compounds	cis-Linalool oxide (79) 3-Methyl-2-phenylindole (80)	- 12.73	1070 1710	[56–58]
Phenylpropanoids	trans-4-Methoxycinnamaldehyde (81) Methyl cinnamate (82)	8.57 -	-	[56,57,60,62]
Cycloalkenes	Methyl ethyl cyclopentene (83)	5.63	-	[56,58]
Monoterpenoids	α-Terpinol (84)	20.16	-	
	1-Menthone (49)	5.76	-	
	Levomenthol (85)	5.91	-	
	Nerol (86)	-	1229	
	Neral (34)	-	1249	
	Geraniol (38)	-	1256	
	Citral (16)	6.57	1270	
	β-Myrcene (37)	11.38	-	
	p-Menth-3-ene (87)	6.74	977.5	
	Bornyl acetate (88)	-	1286	
	α-Pinene (40)	3.95	-	
	Fenchone (41)	-	1089	
Sesquiterpenoids	Camphor (89)	18.14	-	
	Camphene (90)	8.86	-	
	Sabinene (91)	10.27	-	
	trans-α-Bergamotene (92)	7.73	-	
	β-Pinene (93)	10.36	-	
	α-Phellandrene (46)	11.94	-	
	α-Terpinene (45)	12.52	-	
	γ-Terpinene (94)	14.57	-	
	Terpinolene (95)	16.00	-	
	Estragole (96)	6.06	-	
	1-8-Cineole (97)	13.16	-	
	p-Cymene (44)	12.88	-	
	cis-β-Terpineol (98)	16.30	-	
	β-Copaene (99)	8.10	-	
	α-Humulene (57)	29.26	-	[56–60,62,63,65–70]
	cis-β-Farnesene (100)	29.98	-	
	β-Cubebene (101)	-	1394	
	α-Cadinene (102)	-	1537	
	Aromadendrene (103)	-	1529	
	α-Bisabolene (104)	-	1561	
	β-Bisabolene (56)	8.17	-	
	α-Bisabolol (105)	-	1642	
	Neoisolongifolene (106)	-	-	
	trans-β-Guaiene (107)	-	1499	
	cis-Muurola-3,5-diene (108)	8.31	1502	
	Nerolidol (109)	8.47	-	
	Bicyclogermacrene (110)	30.60	-	
	β-Elemene (111)	-	1387	
	β-Caryophyllene (112)	-	1419	
	δ-Cadinene (113)	31.45	-	
	Spathulenol (114)	32.96	-	
	α-Selinene (115)	30.85	-	
	Bicyclosesquiphellandrene (116)	29.05	-	
	α-Bergamotene (117)	28.79	-	
	Caryopyllene oxide (118)	-	1550	
	1,10-Di-epcubenol (119)	34.06	-	

RT = Retention Time; RI = Retention Index.

2.3. *Ocimum gratissimum*

O. gratissimum, with the common name of clove basil, is a species of family *Labiatae* which grows in tropical region, namely India and West Africa [71,72]. It is 1–3 m tall and has leaves that are 3–4 cm × 1–2 cm [73]. The flowers have several colors, such as yellowish white, greenish purple, hairy, calyx greenish purple, brown seeds, and not slimy [74]. In Africa, eastern, central, and western Kenya, *O. gratissimum* is commonly found in scrub and disturbed highland forests at elevations of 600 to 2400 m above sea level [75].

Traditionally, *O. gratissimum* was used for the treatment of cough, fever, snakebites, mosquito repellent, anemia, inflammation, and diarrhea [75,76]. It has several bioactivities, namely antioxidant, anti-inflammation, antimycotoxicogenic, antibacterial, antifungal, antimalaria, and antiseptic activities [77–80]. The phytochemical components of *O. gratissimum* are alkaloids, saponins, tannins, phlobatannins, glycosides, phenols, anthraquinones, flavonoids, and terpenoids [81,82]. Some of the chemical compounds contained in *O. gratissimum* are listed in Table 3, Figure S3.

Table 3. The chemical constituents of *O. gratissimum*.

Class of Chemical Constituents	Chemical Constituents	RT (min)	Percentage (%)	References
Alcohols	Chlorogenic acid (120)	-	-	[83]
Fatty acids	Methyl acetate (121)	30.55	-	[84]
	Palmitic acid (122)	28.35	-	[84]
Flavonoids	Luteolin (123)	9.06	-	[47,84–88]
	Apigenin (124)	12.36	-	
	Quercetin (125)	-	-	
	Epicatechin (126)	-	-	
	Nevadensin (127)	18.45	-	
	Salvigenin (128)	25.13	-	
	Morin (129)	-	-	
	Xanthomicrol (130)	18.23	-	
	Apigenin dimethyl ether (131)	27.46	-	
Phenols	Eugenol (33)	-	74.83	[38]
	Methyl eugenol (132)	6.36	-	
Phenylpropanoids	Sinapic acid (133)	2.72	-	[36,89]
	Rosmarinic acid (134)	3.82	-	
	Nepetoidin A (135)	17.34	-	
Monoterpeneoids	Methyl carvacrol (136)	-	1.19	[89–92]
	Carvacrol (50)	-	0.20–8.40	
	1–8-cineole (97)	-	0.30–23.04	
Sesquiterpenoids	<i>trans</i> - α -Bergamotene (92)	-	0.20–0.70	[83,84,86,90–94]
	δ -Cadinene (113)	-	0.30–3.00	
	β -selinene (148)	-	0.82–7.96	
	β -Caryophyllene (112)	-	0.39–7.23	
	Humulene (57)	-	4.40	
	β -Bergamotene (149)	-	1.03–2.29	
	δ -Cadinene (113)	-	0.30	
	γ -Cadinene (150)	-	0.61	
	Caryopyllene oxide (118)	-	0.50–3.02	
	τ -Cadinol (151)	-	3.5	
	α -Panasinsene (152)	-	-	
	β -Chamigrene (153)	-	1.61–2.84	
	β -Copaene (99)	-	0.27	
	Germacrene-D (154)	-	0.10–29.9	
	β -Bisabolol (155)	-	-	
	β -Cubebene (101)	-	0.08	
	<i>Epi</i> -Cubebol (156)	-	0.21	
	<i>Epi</i> -Cubenol (157)	-	0.23	
	Copaene (158)	-	0.30–7.20	
	β -Bourbonene (159)	-	0.21–0.89	
	β -Selinene (160)	-	0.85–7.96	

Table 3. Cont.

Class of Chemical Constituents	Chemical Constituents	RT (min)	Percentage (%)	References
	Bicyclogermacrene (110)	-	0.40–2.90	
	Calamenene (161)	-	0.38	
	α -Thujone (33)	-	0.1	
	β -Myrcene (37)	-	0.14	
	Camphepane (90)	-	0.10–0.60	
	Sabinene (91)	-	0.18–0.90	
	2-Carene (137)	-	-	
	Camphor (89)	-	0.10–0.60	
	β -Pinene (93)	-	0.11–2.22	
	<i>trans</i> -Longipinocarveol (138)	-	-	
	γ -Terpinene (94)	-	8.23–22.90	
	4-Carene (139)	-	-	
	β -(E)-Ocimene (140)	-	0.10–0.43	
	β -(Z)-Ocimene (141)	-	0.21–4.00	
	Allo-Ocimene (142)	-	-	
	4-Methylstyrene (143)	-	-	
	Styrene (144)	-	-	
	Limonene (48)	-	1.25–5.27	
	<i>p</i> -Cymene (44)	-	12.04–25.00	
	Terpinene-4-ol (145)	-	1.90–4.35	
	Borneol (51)	-	0.20–0.55	
	Umbellulone (146)	-	-	
	α -Terpineol (147)	-	0.10–0.92	
	Estragole (96)	-	0.20–1.50	
	Thymol (29)	-	8.50–46.99	
	α -Pinene (40)	-	0.24–2.66	
	α -Curcumene (162)	-	0.26	
	Isolongifolol (163)	-	0.10	
	Isopinocamphone (164)	-	0.40	
	Longifolene (165)	-	3.00	
	γ -Murolidine (166)	-	0.16–3.88	
	α - <i>p</i> -Dimethyl styrene (167)	-	1.19	
	β -Ylangene (168)	-	2.7	
Triterpenoids	Oleanolic acid (169)	33.60	-	
Stigmastanes	Basilimoside (170)	29.12	-	

(-) = Test not performed; RT = Retention Time; RI = Retention Index.

2.4. *Ocimum campechianum*

O. campechianum is a plant of the Lamiaceae family group which originates from the tropics of South and Central America. This plant is commonly known as “Albahaca de campo” or “Albahaca silvestre”, used by local people for traditional medicine or culinary purposes [95,96]. This plant has a height of 1 m and contains essential oil components with two types of aromatic leaves, namely glandular trichomes, peltate, and capitate. In addition, *O. campechianum* contains components of flavonoids, polyphenols, and tannins [97,98]. Traditionally, this plant is used as a decoction of leaves, ointments, and for the treatment of fever, cough, bronchitis, diarrhea, dysentery, and hypertension. In addition, *O. campechianum* can also be used as an emmenagogue that helps childbirth [99]. In terms of pharmacological effects, this plant extract is known to have antifungal, antioxidant, antiradical, antiproliferative, and analgesic activities [96,100,101]. In addition, it also has potential as a natural larvicide and pesticide [102]. The data of chemical constituents in *O. campechianum* can be seen in Table 4, Figure S4.

Table 4. The chemical constituents of *O. campechianum*.

Class of Chemical Constituents	Chemical Constituents	KI	Percentage (%)	References
Aliphatic alcohol	3-Octanol (73)	988	-	[99]
Fatty alcohol	1-Vinylhexanol (77)	974	-	[99]
	(3Z)-Hexenol (171)	850	-	[99]
Monocyclic ketone	Carvone (13)	1239	-	[99]
Benzeneoids	Benzene acetaldehyde (172)	1036	-	[99]
Phenols	Eugenol (33)	-	9.00	[99]
	Methyl eugenol (132)	-	12.00	
Polymethoxyflavones	5-Demethylnobiletin (173)	-	-	[99,103]
	5-Demethylsinensetin (174)	-	-	
Organooxygen compounds	(2E)-Hexenal (175)	846	-	[99]
	3-Octanone (176)	979	-	
Carboxylic acid esters	(3E)-Hexenyl acetate (177)	1001	-	[104]
Monoterpoids	δ-3-Carene (178)	1008	-	[99,104,105]
	α-Terpinene (45)	1014	-	
	p-Cymene (44)	1020	-	
	β-(E)-Ocimene (139)	1044	-	
	β-(Z)-Ocimene (140)	1032	-	
	Neoalloocimene (179)	1140	-	
	Terpinolene (95)	1086	-	
	Limonene (48)	-	0.30	
	α-Terpinol (84)	-	0.30	
	Estragole (96)	-	-	
	1,8-Cineole (97)	-	3.30	
	Isoborneol (180)	1155	-	
	(+)-4-Terpinal (144)	1174	-	
	Linalool (36)	-	2.90	
	Myrcene (37)	-	0.20	
	β-Pinene (93)	-	0.80	
	α-Thujene (181)	924	-	
	Tricyclene (182)	921	-	
	Cis-sabinen hydrate (183)	1065	-	
	α-Pinene (40)	-	0.20	
	Camphene (90)	-	0.40	
	trans-α-Bergamotene (92)	1432	-	
	Sabinene (91)	-	0.10	
	Camphor (89)	1141	-	
	Sesquisabinene (184)	-	0.20	
Sesquiterpenoids	Humulene epoxide II (185)	1608	-	[99,105]
	β-Bisabolene (56)	1505	-	
	Germacrene-D (154)	-	10.10	
	Bicyclogermacrene (110)	1500	-	
	Germacrene-B (186)	1559	-	
	α-Humulene (57)	-	2.80	
	Aromadendrene (103)	1439	-	
	α-Copaene (158)	-	1.90	
	β-Bourbonene (159)	1387	-	
	α-Gurjunene (187)	1409	-	
	Viridiflorol (188)	1592	-	
	β-Eudesmol (189)	1649	-	
	3,7(11)-Eudesmadiene (190)	-	-	
	α-Guaiene (191)	-	5.60	
	γ-Muurolene (192)	-	0.30	
	α-Bulnesene (193)	-	7.10	
	Spathulenol (114)	-	0.40	
	τ-Cadinol (151)	-	-	
	Junenol (194)	1618	-	
	β-elinene (148)	-	-	
	δ-Cadinene (113)	-	2.00	
	Caryophyllene oxide (195)	-	0.40	
	E-Caryophyllene (112)	-	4.05	
	β-Elemene (111)	-	0.53	
	γ-Elemene (196)	-	0.60	
	Bicycloelemene (197)	-	0.20	

(-) = Test not performed; KI = Kovat's Index.

2.5. *Ocimum sanctum*

O. sanctum is a plant from the Lamiaceae family, commonly known as basil or tulsi, which is native to India and is widely distributed as a cultivated plant throughout Southeast Asia [106]. It is known as a sacred plant in India and a symbol of purity. It got the name of "Tulasi" from Tulasi Devi, one of the Lord Krishna's eternal consorts. Tulasi was a Gopi who was said to have fallen in love with Krishna and was cursed by his wife Radha. He is very similar to Vishnu [107]. In India, it is used for religious plants and important events such as weddings [108]. *O. sanctum* is 30–75 cm tall with an herbaceous shape, and is erect, more-branched, and hairy-soft [107]. It has pointed or blunt leaves, the leaves are oval, and the flowers are tightly coiled with a pale red or dark red color [109].

Traditionally, *O. sanctum* was used for the treatment of diarrhea, chronic fever, malaria, skin disease, bronchitis, dysentery, insect bite, arthritis, and bronchial asthma [110,111]. *O. sanctum* has several bioactivities such as anticancer, antispasmodic, antifertility, antimicrobial, anti-inflammatory, antioxidant, antifungal, analgesic, antidiabetic, cardio protective, adaptogenic, antiemetic, and hepatoprotective [112–115]. It has several phytochemicals, namely terpenoids, phenolic, flavonoids, glycosides, and propenyl phenols [116]. Moreover, it also contains vitamin C, A, and minerals such as zinc, iron, and calcium [117]. The protein content in *O. sanctum* is 4.2 g, then 0.5 g fat, 25 mg carbohydrates, 287 mg phosphorus, 25 mg calcium, vitamin C per 100 g, and 15.1 mg iron [118]. The chemical constituents included in *O. sanctum* are listed in Table 5, Figure S5.

Table 5. The chemical constituents of *O. sanctum*.

Class of Chemical Constituents	Chemical Constituents	RT (min)	RI	References
Fatty acids	Ocimumnaphthanoic acid (198)	-	-	[119–121]
	Methyl 9-methyltetradecanoate (199)	-	-	
	Ethyl 13-methyl-tetradecanoate (200)	-	-	
	Methyl 7-Z-hexadecanoate (201)	-	-	
	Ethyl palmitate (202)	-	-	
	Ethyl isovalerate (203)	-	-	
Carboxylic acids	(Z)-3-Hexenil acetate (204)	-	1005	[122]
Aliphatic aldehyde	Citronellal (17)	-	-	[113]
Benzeneoids	Protocatechuic acid (205)	-	-	[113,120]
Unsaturated hydrocarbons	Di- <i>n</i> -2-propylpentylphthalate (206)	-	-	
	1,2,4- Trimethylcyclohexane (207)	13.36	-	[113]
Organooxygen compounds	1,2-Cyclopentanedione (208)	-	-	[119,120]
	Citrusin C (209)	-	-	
Phenolic	Vanillin (31)	-	-	[113,115,123]
	Methylisoeugenol (210)	7.50	-	
	Vanilllic acid (211)	-	-	
	Sinapic acid (132)	7.20	-	
	<i>p</i> -coumeric acid (212)	-	-	
	<i>p</i> -hydroxybenzoic acid (213)	-	-	
	Ferulic acid (214)	-	-	
	Eugenol (33)	-	1382	
	Bieugenol (215)	-	-	
Flavonoids	Galuteolin (216)	-	-	[113,119]
	Isovitolin (217)	-	-	
	Chrysoeriol (218)	-	-	
	Cirsilineol (68)	-	-	
	Isothymunin (219)	-	-	
	Isothymusin (220)	-	-	
	Cirsimarinin (221)	-	-	
	Molludistin (222)	-	-	
	Vitexin (64)	-	-	
	Orientin (223)	-	-	

Table 5. Cont.

Class of Chemical Constituents	Chemical Constituents	RT (min)	RI	References
Phenylpropanoids	Isoorientin (224)	-	-	
	Vicenin (62)	-	-	
	Luteolin-5-glucoside (225)	-	-	
	Esculin (226)	-	-	
	Esculetin (227)	-	-	
	Rosmarinic acid (133)	6.20	-	[115,124]
	Estragole (96)	-	1229	
	Carvacrol (50)	-	-	
	Linalool (36)	-	1109	
	α -Terpinyl acetate (228)	-	-	
Sesquiterpenoids	Terpiniolene (95)	-	-	
	Bornyl acetate (88)	-	1289	
	α -Bergamotene (117)	-	1469	[113,115,119–121,124,125]
	Spathulenol (114)	-	1578	
	α -Humulene (57)	-	1482	
	α -Guaiol (229)	-	-	
	2-Methylene-4,8,8-trimethyl-4-vinylbicyclo[5.2.0]nonane (230)	-	-	
Triterpenoids	Oleanolic acid (169)	-	-	
	Ursolic acid (231)	8.50	-	

(-) = Test not performed; RT = Retention Time; RI = Retention Index.

3. Antibacterial Activity of *Ocimum* Species

Tests of antibacterial activity against *Ocimum* species have been carried out on both Gram-positive and Gram-negative bacteria. Based on the data in Tables 6–8, the *Ocimum* extracts that were widely tested and gave relatively high activity were essential oil extracts. In general, the five *Ocimum* species have good antibacterial activity. However, based on the inhibition zone data in Table 6, it can be seen that *O. americanum*, *O. basilicum*, and *O. sanctum* had higher activity than the other two species, where the diameter of the inhibition zone reached 20 to 40 mm. In Gram-positive bacteria, *O. americanum*, *O. basilicum*, and *O. sanctum* gave the highest inhibition zone against *S. aureus*, while *O. gratissimum* gave the highest inhibition zone against *E. faecalis*, and *O. campechianum* gave the highest inhibition against *L. ivanovii*. Furthermore, in Gram-negative bacteria, *O. americanum* gave the highest inhibition zone against *P. gingivalis* and *P. intermedia*, *O. basilicum* against *P. aeruginosa*, *O. gratissimum* against *P. mirabilis*, *O. campechianum* against *E. coli*, and *O. sanctum* against *Yersinia enterocolitica*.

Table 6. Data of inhibition zone of *Ocimum* species.

Microorganism	Extract	Inhibition Zone (mm)					Positive Control (mm)	Reference
		1	2	3	4	5		
Gram-Positive Bacteria								
<i>S. mutans</i>	Essential oil	28	-	14	-	-	^{1a} 21	
	Methanol	-	8.30–9.40	-	-	-	^{2a} 10.90	[126–128]
	Ethyl acetate	-	-	-	-	24	^{5b} 29.40	
<i>L. casei</i>	Essential oil	19	-	-	-	-	-	[126]
<i>E. faecalis</i>	Essential oil	15.67	-	-	10	-	-	
	Methanol	-	6.90–10.40	20–26	-	-	^{2a} 15.50, ^{3c} 20	[41,129–131]
<i>E. faecium</i>	Essential oil	15.67	-	-	9	-	-	[41,131]
<i>S. aureus</i>	Essential oil	33.33	29.20–30.56	-	11.67	41.50	^{2e} 23.93	
	70% hydroethanol	9.33–11.17	-	-	-	-	-	[41,128,131–136]
	Hexane	-	-	-	-	2.36	-	
	Ethyl acetate	-	-	-	-	30	^{5b} 29.30	
	Ethanol	-	-	-	-	12	-	

Table 6. Cont.

Microorganism	Extract	Inhibition Zone (mm)					Positive Control (mm)	Reference
		1	2	3	4	5		
<i>B. cereus</i>	Ethyl acetate	10	-	-	-	21	^{5b} 29.30	
	70% hydroethanol	9.50–16.33	-	-	-	-	-	[25,128,132,136,137]
	Essential oil	-	10.66–16.11	-	-	-	^{2e} 20.53	
	Chloroform	-	-	12	-	-	^{3f} 15	
<i>Clostridium perfringens</i>	Ethyl acetate	10	-	-	-	-	-	[138]
<i>S. phyogenes</i>	Essential oil	-	19.00	-	-	-	^{2g} 11.00	[139,140]
<i>Ethanol</i>	-	-	6–10	-	-	-	-	[139,140]
<i>Cutibacterium acnes</i>	Essential oil	-	16.78–18.13	-	-	-	^{2f} 13.13	[141]
<i>Lactococcus garvieae</i>	Ethanol	-	1.90	-	-	-	²ⁱ 20.00	[142]
<i>B. subtilis</i>	Methanol	-	13.58	-	-	-	^{2d} 22.01	
	Ethyl acetate	-	-	-	-	-	^{5b} 28.70	[128,143]
<i>S. sanguinis</i>	Methanol	-	7.80–11.40	-	-	-	^{2a} 17.90	[129]
<i>S. faecalis</i>	Ethanol	-	-	10	-	-	-	[144]
<i>L. monocytogenes</i>	Essential oil	12	10.60–11.70	-	8	-	^{1j} 33, ^{2j} 33.70	[92,131,138]
	Ethanol	-	-	8–26	-	-	-	
<i>L. ivanovii</i>	Essential oil	13.67	-	-	12.67	-	-	[41,131]
<i>S. epidermidis</i>	Essential oil	22.67	-	-	11	-	-	[41,131,135,145]
	Ethanol	-	-	-	-	-	^{2k} 22	
<i>M. luteus</i>	Essential oil	14	-	-	-	-	^{1c} 26	[30]
<i>C. perfringens</i>	Essential oil	-	-	-	-	-	^{5c} 41.39	[25]
<i>Lb. plantarum</i>	Essential oil	-	-	-	-	-	^{5c} 15.38	[138]
<i>S. agalactiae</i>	Ethyl acetate	-	-	-	-	-	^{5b} 30.40	[128]
Gram-Negative Bacteria								
<i>P. gingivalis</i>	Essential oil	30	-	-	-	-	-	
<i>P. intermedia</i>	Essential oil	30	-	-	-	-	-	[146]
<i>F. nucleatum</i>	Essential oil	24	-	-	-	-	-	
<i>P. vulgaris</i>	Essential oil	-	-	-	11.33	-	-	[41,131]
<i>E. coli</i>	Essential oil	10.67	17.48–23.58	12	11.67	15.40	-	[41,128,131–134,147]
	Ethyl acetate	9	-	-	-	15.70	^{5b} 31.30	
	Hexane	-	-	-	-	2.26	-	[25]
<i>Klebsiella pneumonia</i>	Ethyl acetate	10	-	-	-	15.30	^{5b} 29.30	[128,148]
	Methanol	-	-	16–23	-	-	-	
<i>Salmonella paratyphi</i>	Ethyl acetate	9	-	-	-	-	-	[25]
<i>V. chlorea</i>	Essential oil	18	-	-	-	-	-	[30]
<i>S. typhimurium</i>	Essential oil	13.00–20.10	20.00	-	-	-	^{1j} 28.80; ^{2h} 11.00	[92,128,139]
	Ethyl acetate	-	-	-	-	16.70	^{5b} 29.70	
<i>S. dysenteria</i>	Essential oil	18	-	-	-	-	^{1c} 17	
	Bark	-	16	-	-	-	^{2k} 25	[30,145]
<i>P. aeruginosa</i>	Essential oil	-	Max	-	-	12	^{2e} 12.64	
	Leaves	-	-	16	-	-	^{3k} 39	[128,132,135,149,150]
	Ethyl acetate	-	-	-	-	10.30	^{5b} 22.30	
	Ethanol	-	-	-	-	11	-	
<i>A. baumannii</i>	Essential oil	-	15.30	-	-	-	-	[151]
<i>Salmonella enteritidis</i>	Essential oil	-	18.00	-	-	-	-	[152]
<i>S. typhi</i>	Essential oil	-	24.80	-	-	-	-	[136,153]
<i>P. multocida</i>	Chloroform	-	-	14	-	-	^{3c} 16	
<i>Listonella anguillarum</i>	Essential oil	-	13.60–18.40	-	-	-	^{2h} 31.10	[154]
<i>Yersinia ruckeri</i>	Ethanol	-	3.90	-	-	-	²ⁱ 20.00	[142]
	Ethanol	-	1.50	-	-	-	²ⁱ 20.80	[142]

Table 6. Cont.

Microorganism	Extract	Inhibition Zone (mm)					Positive Control (mm)	Reference
		1	2	3	4	5		
<i>P. mirabilis</i>	Leaves	-	-	11–22	-	-	^{3d} 15 mm; ^{3l} 29 mm	[128,148,155]
	Methanol	-	-	25–32	-	-	-	
	Ethyl acetate	-	-	-	-	15.10	^{5b} 30.30	
<i>Shigella</i> <i>gemineri</i>	Essential oil	-	-	16	-	-	^{3m} 29	[156]
<i>Proteus</i> <i>mirabilis</i>	Flavonoids	-	-	12–28	-	-	^{3e} 24	[148]
<i>Shigalla</i> sp.	Ethanol	-	-	8	-	-	-	
<i>Salmonella</i> sp.	Ethanol	-	-	6	-	-	-	
<i>Shigella</i> <i>dysenteriae</i>	Chloroform	-	-	2	-	-	³ⁿ 14	[136]
<i>P. fluorescens</i>	Essential oil	-	-	-	-	20	^{5m} 12.33	[135]
<i>Shigella</i> <i>boydii</i>	Ethyl acetate	-	-	-	-	14.30	^{5b} 30.70	[128]
<i>Yersinia</i> <i>enterocolitica</i>	Ethyl acetate	-	-	-	-	20.10	^{5b} 28.30	
<i>Aeromonas</i> <i>hydrophila</i>	Ethanol	-	-	-	-	15	-	[135]
<i>Vibrio harveyi</i>	Ethanol	-	-	-	-	12	-	

(-) = Test not performed; 1 = *O. americanum*, 2 = *O. basilicum*, 3 = *O. gratissimum*, 4 = *O. campechianum*, 5 = *O. sanctum*. Positive control: ^a(Chlorhexidine), ^b(Ciprofloxacin), ^c(Gentamicin), ^d(Ciprofloxacin), ^e(Tetracycline), ^f(Clindamycin), ^g(Rifampicin), ^h(Amoxicillin), ⁱ(Oxytetracycline), ^j(Ampicillin), ^k(Streptomycin), ^l(Ketoconazole), ^m(Penicillin), ⁿ(Cloxacilllin).

Furthermore, MIC is an antibacterial test used to determine its inhibitory activity. Based on de Aguiar et al. [157], MIC concentrations in the range 101–500 µg/mL have strong activity and in the range 500–1000 µg/mL have moderate activity. As for Table 7, testing of *O. basilicum* against several bacteria, such as *S. mutans*, *S. aureus*, *B. cereus*, *L. monocytogenes*, *E. coli*, *P. aeruginosa*, and *S. typhi*, showed MIC values below 100 µg/mL, which means that they provide very strong activity, as well as in *O. sanctum*. The other three *Ocimum* species only showed strong to weak activity.

Table 7. Data of minimum inhibitory concentration (MIC) of *Ocimum* species.

Microorganism	Extract	MIC (µg/mL)					Positive Control (µg/mL)	Reference
		1	2	3	4	5		
Gram-Positive Bacteria								
<i>S. mutans</i>	Essential oil	0.04% v/v	18	-	-	-	-	[126,129,158]
	Lauric acid	-	156	-	-	-	-	
	β-Sitosterol	-	25,000	-	-	-	-	
<i>L. casei</i>	Essential oil	0.04% v/v	-	-	-	-	-	[126]
<i>E. faecalis</i>	Essential oil	500	-	-	-	-	-	[41,129]
	β-Sitosterol	-	25,000	-	-	-	-	
<i>E. faecium</i>	Essential oil	500	-	-	-	-	-	[41]
<i>S. aureus</i>	Essential oil	200	-	1000	-	2.50	-	
	70% hydroethanolic	1840	-	-	-	-	-	
	Ethanol	-	-	-	-	4280	^{5e} 6	[41,55,132– 135,137,147, 159]
	Hexane	-	-	-	-	2.30	-	
	Linalool	-	32	-	-	-	-	
	Methanol	-	-	-	>2000	-	-	
	Rosmarinic acid	-	-	-	>2000	-	-	
	Eugenol	-	-	-	1000	-	-	
	Caryophyllene	-	-	-	-	50	-	

Table 7. Cont.

Microorganism	Extract	MIC ($\mu\text{g/mL}$)					Positive Control ($\mu\text{g/mL}$)	Reference
		1	2	3	4	5		
<i>B. cereus</i>	70% hydroethanolic	1540	-	-	-	-	-	[132,137,159]
	Essential oil	-	18–36	-	-	-	-	[132,137,159]
	Eugenol	-	-	-	-	25	-	[132,137,159]
<i>S. epidermidis</i>	Essential oil	300	-	-	-	-	-	[41,145]
	Bark	-	500	-	-	-	-	[41,145]
<i>S. phyogenes</i>	Essential oil	-	50	-	-	-	-	[139,140]
	Ethanol	-	-	7	-	-	-	[139,140]
<i>Listeria monocytogenes</i>	Essential oil	-	36	-	-	-	-	[41,153,160]
	Ethanol	-	-	2150	-	-	-	[41,153,160]
	Hydroethanolic	-	10,000	-	-	-	^{2b} > 150 ^{2b} < 150	[41,153,160]
<i>B. subtilis</i>	Methanol	-	625	-	-	-	-	[143]
<i>S. sanguinis</i>	Lauric acid	-	78	-	-	-	-	[158,161]
	Nevadensin	-	3750	-	-	-	-	[158,161]
<i>S. faecalis</i>	Ethanol	-	-	125	-	-	-	[144]
Gram-Negative Bacteria								
<i>P. gingivalis</i>	Essential oil	350	-	-	-	-	-	[146]
<i>P. intermedia</i>	Essential oil	350	-	-	-	-	-	[146]
<i>F. nucleatum</i>	Essential oil	700	-	-	-	-	-	[146]
<i>P. vulgaris</i>	Essential oil	400	-	-	-	-	-	[41]
<i>A. baumannii</i>	Essential oil	1000	-	-	-	-	-	[151]
<i>E. coli</i>	Essential oil	-	9–18	1000	-	2.25	-	[132–134,159,162]
	Hexane	-	-	-	-	2.50	-	[132–134,159,162]
	(–)- β -elememe	-	-	-	-	>200	-	[132–134,159,162]
<i>S. typhimurium</i>	Methanol:DMSO	>200	-	-	-	-	-	[26,139,159,163]
	Essential oil	-	1600	-	-	-	-	[26,139,159,163]
	Leave	-	-	60–250	-	-	-	[26,139,159,163]
	(–)- β -elememe	-	-	-	-	100	-	[26,139,159,163]
<i>S. dysenteriae</i>	Bark	-	10,000	-	-	-	-	[145]
<i>P. aeruginosa</i>	Essential oil	-	9–18	-	-	>4.50	-	[55,132,162]
	Linalool	-	1024	-	-	-	-	[55,132,162]
<i>P. multocida</i>	Essential oil	-	2300	-	-	-	-	[154]
<i>K. pneumoniae</i>	Hydroethanolic	-	20,000	-	-	-	^{2b} 10,000 ^{2o} < 7.80	[164]
<i>M. morganii</i>	Hydroethanolic	-	10,000	-	-	-	^{2o} 20,000 ^{2o} < 7.80	[164]
<i>P. mirabilis</i>	Hydroethanolic	-	>20,000	-	-	-	^{2b} < 150 ^{2o} < 7.80	[164]
<i>Coliform bacilli</i>	Seed	-	-	2500–7000	-	-	-	[165]
<i>Shigella</i> sp.	Ethanol	-	-	40,000	-	-	-	[165]
<i>Salmonella</i> sp.	Ethanol	-	-	60,000	-	-	-	[165]
<i>P. syringae</i>	Methanol	-	-	-	>2000	-	^{4P} 2.50	[96]
	Rosmarinic acid	-	-	-	>2000	-	^{4P} 2.50	[96]
	Eugenol	-	-	-	500	-	^{4P} 2.50	[96]
<i>S. flexneri</i>	(–)- β -elememe	-	-	-	-	100	-	[159]
<i>S. typhi</i>	Essential oil	-	9	-	-	-	-	[153,159]
	Methyl eugenol	-	-	-	-	50	-	[153,159]
<i>E. herbicola</i>	Essential oil	-	-	-	-	2 $\mu\text{L}/\text{mL}$	^{5k} 8	[166]
<i>P. putida</i>	Essential oil	-	-	-	-	1 $\mu\text{L}/\text{mL}$	^{5k} 4	[166]
<i>Aeromonas hydrophila</i>	Ethanol	-	-	-	-	3820	^{5e} 5	[166]
<i>Vibrio harveyi</i>	Ethanol	-	-	-	-	4460	^{5e} 7	[135]
<i>Vibrio vulnificus</i>	Ethanol	-	-	-	-	580	^{5e} 6	[135]

(-) = Test not performed; 1 = *O. americanum*, 2 = *O. basilicum*, 3 = *O. gratissimum*, 4 = *O. campechianum*, 5 = *O. sanctum*; Positive control: ^b(Ciprofloxacin), ^c(Tetracycline), ^k(Streptomycin), ^o(Imipenem), ^P(Cloramphenicol).

The MBC data were used to show the ability of the compound to kill bacterial growth. Based on the MBC data in Table 8, the species from *Ocimum* that had the best activity in

killing bacterial growth was *O. sanctum* which could be seen in the MBC values against *B. cereus*, *E. herbicola*, and *P. putida*. Therefore, based on the data of inhibition zone, MIC, and MBC, the *Ocimum* species that have more potential as natural antibacterials are *O. basilicum* and *O. sanctum*.

Table 8. Data of minimum bactericidal concentration (MBC) of *Ocimum* species.

Microorganism	Extract	MBC ($\mu\text{g/mL}$)					Reference
		1	2	3	4	5	
Gram-Positive Bacteria							
<i>S. mutans</i>	Essential oil	0.08% <i>v/v</i>	-	-	-	-	
	Lauric acid	-	2500	-	-	-	[126,129,158]
	β -Sitosterol	-	50,000	-	-	-	
<i>L. casei</i>	Essential oil	0.30% <i>v/v</i>	-	-	-	-	[126]
<i>E. faecalis</i>	β -Sitosterol	-	50,000	-	-	-	[129]
<i>S. aureus</i>	Essential oil	-	-	1000	-	-	
	70%	7340	-	-	-	-	[24,55,133,155]
	hydroethanolic						
	Linalool	-	>1024	-	-	-	
	Caryophyllene	-	-	-	-	>200	
	70%						
<i>B. cereus</i>	hydroethanolic	6150	-	-	-	-	[137,159]
	Eugenol	-	-	-	-	50	
<i>S. phylogenes</i>	Essential oil	-	100	4.2	-	-	[139,140]
<i>Listeria mono-</i> <i>cytogenes</i>	Essential oil	-	>20,000	-	-	-	[160,164]
<i>S. epidermidis</i>	Ethanol	-	-	2150	-	-	
<i>B. subtilis</i>	Bark	-	2000	-	-	-	[145]
<i>S. sanguinis</i>	Methanol	-	625	-	-	-	[143]
	Lauric acid	-	1250	-	-	-	
	Nevadensin	-	15,000	-	-	-	[158,161]
Gram-Negative Bacteria							
<i>P. gingivalis</i>	Essential oil	700	-	-	-	-	
<i>P. intermedia</i>	Essential oil	700	-	-	-	-	[146]
<i>F. nucleatum</i>	Essential oil	1400	-	-	-	-	
<i>A. baumannii</i>	Essential oil	8000	-	-	-	-	[151]
<i>E. coli</i>	Essential oil	-	-	1000	-	-	
	(-)- β -elememe	-	-	-	-	200	[133,159]
<i>S. typhimurium</i>	Essential oil	-	3200	100–300			[139,159,163]
	(-)- β -elememe	-	-	-	-	>200	
<i>S. dysenteriae</i>	Bark	-	20,000	-	-	-	[145]
<i>P. aeruginosa</i>	Linalool	-	>1024	-	-	-	[55]
<i>K. pneumoniae</i>	Hydroethanolic	-	>20,000	-	-	-	
<i>M. morganii</i>	Hydroethanolic	-	>20,000	-	-	-	[164]
<i>P. mirabilis</i>	Hydroethanolic	-	>20,000	-	-	-	
<i>S. flexneri</i>	(-)- β -elememe	-	-	-	-	200	
<i>S. typhi</i>	Methyl eugenol	-	-	-	-	100	[159]
<i>E. herbicola</i>	Essential oil	-	-	-	-	8	
<i>P. putida</i>	Essential oil	-	-	-	-	4	[166]

(-) = Test not performed; 1 = *O. americanum*, 2 = *O. basilicum*, 3 = *O. gratissimum*, 4 = *O. campechianum*, 5 = *O. sanctum*.

4. Interaction of Essential Oils to Bacteria

Essential oils usually contain chemical components in the terpenoid and phenylpropanoid groups. The five *Ocimum* species were proven to contain many essential oil components, especially in the monoterpenoid and sesquiterpenoid groups, as well as some phenylpropanoids [167,168]. Therefore, studies on antibacterial activity have also been carried out on the essential oil components. Essential oils of plants can inhibit both Gram-positive and negative bacteria. However, some studies reported that essential oils

were more sensitive to Gram-positive bacteria, but others reported more sensitive to Gram-negative bacteria [169]. Based on El-Shenaway et al. [170], essential oils were more sensitive against Gram-positive bacteria. This was because of differences in cell wall structures. In Gram-negative bacteria, there was an external capsule which prevented penetration of essential oils into microbial cells. The capsule is composed of a more complex bacterial cell wall with a 2–3 nm thick peptidoglycan layer, where there is an outer membrane on the outer layer of peptidoglycan. The presence of the outer membrane is one of the differences between the cell walls of Gram-negative and Gram-positive bacteria. Peptidoglycan and this outer membrane have strong covalent bonds to Braun lipoproteins. This causes the hydrophobic structure of the essential oil to more easily penetrate the cell walls of Gram-positive bacteria [169].

The ability of essential oils to inhibit bacterial growth is due to their hydrophobicity. It increases cell permeability and leak cell constituents [171]. Essential oils will cross cell wall and cytoplasmic membrane which arrange a lot of polysaccharide layers, fatty acids, and phospholipids. As a result, the interaction between lipophilic compounds from essential oils and various structures found in cell walls and membranes causes a cytotoxic effect [172]. Furthermore, essential oils can also agglomerate in the cytoplasm and cause damage to the protein and lipid layers [173].

In the cell membrane, there is a process of ATP production. The action of essential oils can affect changes in intracellular and external ATP balance. This results in a disturbance with ion loss and a decrease in membrane potential, a decrease in the amount of ATP, and a collapse of the proton pump. [174,175]. This will compromise vital functions such as energy systems, synthesis of structural macromolecules, and secretion of enzymes for growth [171]. In addition, essential oils can affect the pH conditions of bacteria. The presence of essential oils on the membrane can disrupt pH homeostasis and cause a significant decrease in pH. Then, the membrane will lose its capacity to block protons. This is because at low pH, the hydroxyl groups in essential oils do not dissociate so that their hydrophobicity will increase, resulting in easier interaction with bacterial cell membrane lipids [176,177].

There are several methods for extracting essential oils from *Ocimum* species, such as hydrodistillation, steam distillation, solvent extraction, enfleurage, cohobation, and maceration [178]. However, this method requires a fairly long process if it is to be consumed simply in the community. As an alternative, the *Ocimum* plant has been widely used traditionally to cure bacterial infections. *Ocimum* can be processed into juice to relieve toothache and treat otitis. This plant can also be made into an infusion for mouthwash. Decoction of this plant can provide an anesthetic effect and act as an antiseptic. Decoction of the leaves and stems can treat diarrhea, fever, and inflammation of the mucous membranes of the nose [179].

5. Conclusions

O. americanum, *O. basilicum*, *O. gratissimum*, *O. campechianum*, and *O. sanctum* are five types of *Ocimum* species that have abundant chemical components and antibacterial activity. Based on data on the chemical components of the five *Ocimum* species, it is known that the most abundant compounds are terpenoids. *O. basilicum* is the most commonly used and widely available on the market. However, the activity data of the five *Ocimum* species show that this plant can be a natural product that has potential as a natural antibacterial agent.

Supplementary Materials: The following are available online at <https://www.mdpi.com/article/10.3390/molecules27196350/s1>, Figure S1: Chemical Compound Structures of *O. Americanum*, Figure S2: Chemical Compound Structures of *O. basilicum*, Figure S3: Chemical Compound Structures of *O. gratissimum*, Figure S4: Chemical Compound Structures of *O. campechianum*, Figure S5: Chemical Compound Structures of *O. sanctum*.

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References

- Wu, D.; Ding, Y.; Yao, K.; Gao, W.; Wang, Y. Antimicrobial Resistance Analysis of Clinical *Escherichia coli* Isolates in Neonatal Ward. *Front. Pediatr.* **2021**, *9*, 670470. [[CrossRef](#)] [[PubMed](#)]
- Mapanguy, C.C.M.; Adedoja, A.; Kecka, L.G.V.; Vouyoungui, J.C.; Nguimbi, E.; Velavan, T.P.; Ntoumi, F. High prevalence of antibiotic-resistant *Escherichia coli* in Congolese students. *Int. J. Infect. Dis.* **2021**, *103*, 119–123. [[CrossRef](#)] [[PubMed](#)]
- Tom, I.M.; Ibrahim, M.M.; Umoru, A.M.; Umar, J.B.; Bukar, M.A.; Haruna, A.B.; Aliyu, A. Infection of Wounds by Potential Bacterial Pathogens and Their Resistogram. *Open Access Libr. J.* **2019**, *6*, 1–13. [[CrossRef](#)]
- Sani, R.; Garba, S.; Oyewole, O. Antibiotic resistance profile of gram negative bacteria isolated from surgical wounds in Minna, Bida, Kontagora and Suleja areas of Niger state. *Am. J. Med. Med. Sci.* **2012**, *2*, 20–24.
- Chiller, K.; Selkin, B.A.; Murakawa, G.J. Skin microflora and bacterial infections of the skin. *J. Investigig. Dermatol. Symp. Proc.* **2001**, *6*, 170–174. [[CrossRef](#)]
- Huang, R.; Li, M.; Gregory, R.L. Bacterial interactions in dental biofilm. *Virulence* **2011**, *2*, 435–444. [[CrossRef](#)]
- Wang, L.; Hu, C.; Shao, L. The antimicrobial activity of nanoparticles: Present situation and prospects for the future. *Int. J. Nanomed.* **2017**, *12*, 1227–1249. [[CrossRef](#)]
- Ben, Y.; Fu, C.; Hu, M.; Liu, L.; Wong, M.H.; Zheng, C. Human health risk assessment of antibiotic resistance associated with antibiotic residues in the environment: A review. *Environ. Res.* **2019**, *169*, 483–493. [[CrossRef](#)]
- Martinez, J.L. The role of natural environments in the evolution of resistance traits in pathogenic bacteria. *Proc. R. Soc. B Biol. Sci.* **2009**, *276*, 2521–2530. [[CrossRef](#)]
- Benítez-Chao, D.F.; León-Buitimea, A.; Lerma-Escalera, J.A.; Morones-Ramírez, J.R. *Bacteriocins*: An overview of antimicrobial, toxicity, and biosafety assessment by in vivo models. *Front. Microbiol.* **2021**, *12*, 630695. [[CrossRef](#)]
- Appelbaum, P.C. Reduced glycopeptide susceptibility in methicillin-resistant *Staphylococcus aureus* (MRSA). *Int. J. Antimicrob. Agents* **2007**, *30*, 398–408. [[CrossRef](#)] [[PubMed](#)]
- Livermore, D.M.; Hope, R.; Brick, G.; Lillie, M.; Reynolds, R.; on behalf of the BSAC Working Parties on Resistance Surveillance. Non-susceptibility trends among Enterobacteriaceae from bacteraemias in the UK and Ireland, 2001–06. *J. Antimicrob. Chemother.* **2008**, *62*, ii41–ii54. [[CrossRef](#)] [[PubMed](#)]
- Jacobs, M.R. *Streptococcus pneumoniae*: Epidemiology and patterns of resistance. *Am. J. Med. Suppl.* **2004**, *117*, 3–15. [[CrossRef](#)] [[PubMed](#)]
- Stefan, M.; Zamfirache, M.; Padurariu, C.; Trută, E.; Gostin, I. The composition and antibacterial activity of essential oils in three *Ocimum* species growing in Romania. *Cent. Eur. J. Biol.* **2013**, *8*, 600–608. [[CrossRef](#)]
- Runyoro, D.; Ngassapa, O.; Vagionas, K.; Aligiannis, N.; Graikou, K.; Chinou, I. Chemical composition and antimicrobial activity of the essential oils of four *Ocimum* species growing in Tanzania. *Food Chem.* **2010**, *119*, 311–316. [[CrossRef](#)]
- Pavithra, K.H.; Narase Gowda, P.N.; Prasad, M.P. Studies on Antimicrobial Activity of *Ocimum* Species of Karnataka against Clinical Isolates. *Int. J. Pure Appl. Biosci.* **2019**, *7*, 245–254. [[CrossRef](#)]
- Joshi, B.; Lekhak, S.; Sharma, A. Antibacterial Property of Different Medicinal Plants: *Ocimum sanctum*, *Cinnamomum zeylanicum*, *Xanthoxylum armatum* and *Origanum majorana*. *Kathmandu Univ. J. Sci. Eng. Technol.* **2009**, *5*, 143–150. [[CrossRef](#)]
- Kačániová, M.; Galovičová, L.; Borotová, P.; Vukovic, N.L.; Vukic, M.; Kunová, S.; Hanus, P.; Bakay, L.; Zagrobelna, E.; Kluz, M.; et al. Assessment of *Ocimum basilicum* essential oil anti-insect activity and antimicrobial protection in fruit and vegetable quality. *Plants* **2022**, *11*, 1030. [[CrossRef](#)]
- Abd-Alla, M.; Haggag, W.M. Use of some plant essential oils as post-harvest botanical fungicides in the management of anthracnose disease of mango fruits (*Mangifera indica* L.) caused by *Colletotrichum gloeosporioides* (Penz). *Int. J. Agric. For.* **2013**, *3*, 1–6.

20. Shanaida, M.; Jasicka-Misiak, I.; Makowicz, E.; Stanek, N.; Shanaida, V.; Wieczorek, P.P. Development of high-performance thin layer chromatography method for identification of phenolic compounds and quantification of rosmarinic acid content in some species of the Lamiaceae family. *J. Pharm. Bioallied Sci.* **2020**, *12*, 139–145. [[CrossRef](#)]
21. Maddi, R.; Amani, P.; Bhavitha, S.; Gayathri, T.; Lohitha, T. A review on *Ocimum* species: *Ocimum americanum* L., *Ocimum basilicum* L., *Ocimum gratissimum* L. and *Ocimum tenuiflorum* L. *Int. J. Res. Ayurveda Pharm.* **2019**, *10*, 41–48.
22. Abd El-Aziz, S.; Omer, E.; Sabra, A. Chemical composition of *Ocimum americanum* essential oil and its biological effects against, *Agrotis ipsilon*, (Lepidoptera: Noctuidae). *Res. J. Agric. Biol. Sci.* **2007**, *3*, 740–747.
23. Zahran, E.M.; Abdelmohsen, U.R.; Khalil, H.E.; Desoukey, S.Y.; Fouad, M.A.; Kamel, M.S. Diversity, phytochemical and medicinal potential of the genus *Ocimum* L. (Lamiaceae). *Phytochem. Rev.* **2020**, *19*, 907–953. [[CrossRef](#)]
24. Ali, H.M.; Nguta, J.M.; Mapenay, I.O.; Musila, F.M.; Omambia, V.M.; Matara, D.N. Ethnopharmacological uses, biological activities, chemistry and toxicological aspects of *Ocimum americanum* var. *americanum* (Lamiaceae). *J. Phytopharm.* **2021**, *10*, 56–60. [[CrossRef](#)]
25. Vidhya, E.; Vijayakumar, S.; Rajalakshmi, S.; Kalaiselvi, S.; Pandiyan, P. Antimicrobial activity and phytochemical screening of *Ocimum americanum* L extracts against pathogenic microorganisms. *Acta Ecol. Sin.* **2020**, *40*, 214–220.
26. Zengin, G.; Ferrante, C.; Gnapi, D.E.; Sinan, K.I.; Orlando, G.; Recinella, L.; Diuzheva, A.; Jekő, J.; Cziáky, Z.; Chiavaroli, A.; et al. Comprehensive approaches on the chemical constituents and pharmacological properties of flowers and leaves of American basil (*Ocimum americanum* L.). *Food Res. Int.* **2019**, *125*, 108610. [[CrossRef](#)]
27. Sutili, F.J.; Velasquez, A.; Pinheiro, C.G.; Heinzmann, B.M.; Gatlin, D.M., III; Baldisserotto, B. Evaluation of *Ocimum americanum* essential oil as an additive in red drum (*Sciaenops ocellatus*) diets. *Fish Shellfish. Immunol.* **2016**, *56*, 155–161. [[CrossRef](#)]
28. Javanmardi, J.; Stushnoff, C.; Locke, E.; Vivanco, J. Antioxidant activity and total phenolic content of Iranian *Ocimum* Accessions. *Food Chem.* **2003**, *83*, 547–550. [[CrossRef](#)]
29. Mustafa, A.A.; El-Kamali, H.H. Chemical Composition of *Ocimum americanum* In Sudan. *Res. Pharm. Health Sci.* **2019**, *5*, 172–178. [[CrossRef](#)]
30. Saha, S.; Dhar, T.; Sengupta, C.; Ghosh, P. Biological activities of essential oils and methanol extracts of five *Ocimum* species against pathogenic bacteria. *Czech J. Food Sci.* **2013**, *31*, 195–202. [[CrossRef](#)]
31. Shanaida, M.; Kernychna, I.; Shanaida, Y. Chromatographic analysis of organic acids, amino acids, and sugars in *Ocimum americanum* L. *Acta Pol. Pharm. Drug Res.* **2017**, *74*, 729–734.
32. Radha, S. A review on phytochemical and pharmacological activities of selected *Ocimum* species. *J. Pharmacogn. Phytochem.* **2021**, *10*, 100–108.
33. Barchuk, O.; Pryshlyak, A.; Shanaida, M. Chemical compositions and sedative activities of the *Dracocephalum moldavica* L. and *Ocimum americanum* L. essential oils. *Pharmacol. OnLine* **2021**, *2*, 179–187.
34. Pandey, A.K.; Singh, P.; Tripathi, N.N. Chemistry and bioactivities of essential oils of some *Ocimum* species: An overview. *Asian Pac. J. Trop. Biomed.* **2014**, *4*, 682–694. [[CrossRef](#)]
35. Farag, M.; Ezzat, S.; Salama, M.; Tadros, M. Anti-acetylcholinesterase potential and metabolome classification of 4 *Ocimum* species as determined via UPLC/qTOF/MS and chemometric tools. *J. Pharm. Biomed. Anal.* **2016**, *125*, 292–302. [[CrossRef](#)] [[PubMed](#)]
36. Chenni, M.; El Abed, D.; Rakotomanana, N.; Fernandez, X.; Chemat, F. Comparative Study of Essential Oils Extracted from Egyptian Basil Leaves (*Ocimum basilicum* L.) Using Hydro-Distillation and Solvent-Free Microwave Extraction. *Molecules* **2016**, *21*, 113. [[CrossRef](#)]
37. Altemimi, A.B.; Mohammed, M.J.; Yi-Chen, L.; Watson, D.G.; Lakhssassi, N.; Cacciola, F.; Ibrahim, S.A. Optimization of Ultrasonicated Kaempferol Extraction from *Ocimum basilicum* Using a Box–Behnken Design and Its Densitometric Validation. *Foods* **2020**, *9*, 1379. [[CrossRef](#)]
38. Calderón Bravo, H.; Vera Céspedes, N.; Zura-Bravo, L.; Muñoz, L.A. Basil Seeds as a Novel Food, Source of Nutrients and Functional Ingredients with Beneficial Properties: A Review. *Foods* **2021**, *10*, 1467. [[CrossRef](#)] [[PubMed](#)]
39. Filip, S. Basil (*Ocimum basilicum* L.) a Source of Valuable Phytonutrients. *Int. J. Clin. Nutr. Diet.* **2017**, *3*, 118. [[CrossRef](#)]
40. Bajomo, E.M.; Aing, M.S.; Ford, L.S.; Niemeyer, E.D. Chemotyping of commercially available basil (*Ocimum basilicum* L.) varieties: Cultivar and morphotype influence phenolic acid composition and antioxidant properties. *NFS J.* **2022**, *26*, 1–9. [[CrossRef](#)]
41. Carović-Stanko, K.; Šalinović, A.; Grdiša, M.; Liber, Z.; Kolak, I.; Satovic, Z. Efficiency of morphological trait descriptors in discrimination of *Ocimum basilicum* L. accessions. *Plant Biosyst. Int. J. Deal. All Asp. Plant Biol.* **2011**, *145*, 298–305.
42. Ghasemzadeh, A.; Ashkani, S.; Baghdadi, A.; Pazoki, A.; Jaafar, H.Z.E.; Rahmat, A. Improvement in Flavonoids and Phenolic Acids Production and Pharmaceutical Quality of Sweet Basil (*Ocimum basilicum* L.) by Ultraviolet-B Irradiation. *Molecules* **2016**, *21*, 1203. [[CrossRef](#)] [[PubMed](#)]
43. Osei Akoto, C.; Acheampong, A.; Boakye, Y.D.; Naazo, A.A.; Adomah, D.H. Anti-inflammatory, antioxidant, and anthelmintic activities of *Ocimum basilicum* (sweet basil) fruits. *J. Chem.* **2020**, *2020*, 2153534. [[CrossRef](#)]
44. Ch, M.A.; Naz, S.B.; Sharif, A.; Akram, M.; Saeed, M.A. Biological and Pharmacological Properties of the Sweet Basil (*Ocimum basilicum*). *Br. J. Pharm. Res.* **2015**, *7*, 330–339. [[CrossRef](#)]
45. Kaefer, C.M.; Milner, J.A. The role of herbs and spices in cancer prevention. *J. Nutr. Biochem.* **2008**, *19*, 347–361. [[CrossRef](#)]
46. Shahrajabian, M.H.; Sun, W.; Cheng, Q. Chemical components and pharmacological benefits of Basil (*Ocimum basilicum*): A review. *Int. J. Food Prop.* **2020**, *23*, 1961–1970. [[CrossRef](#)]

47. Zhan, Y.; An, X.; Wang, S.; Sun, M.; Zhou, H. Basil polysaccharides: A review on extraction, bioactivities and pharmacological applications. *Bioorganic Med. Chem.* **2020**, *28*, 115179. [[CrossRef](#)]
48. Pedro, A.C.; Moreira, F.; Granato, D.; Rosso, N.D. Extraction of bioactive compounds and free radical scavenging activity of purple basil (*Ocimum basilicum* L.) leaf extracts as affected by temperature and time. *An. Acad. Bras. Ciências* **2016**, *88*, 1055–1068. [[CrossRef](#)]
49. Gebrehiwot, H.; Bachetti, R.; Dekebo, A. Chemical composition and antimicrobial activities of leaves of sweet basil (*Ocimum basilicum* L.) herb. *Int. J. Basic Clin. Pharmacol.* **2015**, *4*, 869–875. [[CrossRef](#)]
50. Nguyen, V.T.; Nguyen, N.Q.; Thi, N.Q.N.; Thi, C.Q.N.; Truc, T.T.; Nghi, P.T.B. Studies on chemical, polyphenol content, flavonoid content, and antioxidant activity of sweet basil leaves (*Ocimum basilicum* L.). *IOP Conf. Ser. Mater. Sci. Eng.* **2021**, *1092*, 012083. [[CrossRef](#)]
51. De Martino, L.; Amato, G.; Caputo, L.; Nazzaro, F.; Scognamiglio, M.R.; De Feo, V. Variations in composition and bioactivity of *Ocimum basilicum* cv 'Aroma 2' essential oils. *Ind. Crops Prod.* **2021**, *172*, 114068. [[CrossRef](#)]
52. Eftekhar, N.; Moghimi, A.; Mohammadian Roshan, N.; Saadat, S.; Boskabady, M.H. Immunomodulatory and anti-inflammatory effects of hydro-ethanolic extract of *Ocimum basilicum* leaves and its effect on lung pathological changes in an ovalbumin-induced rat model of asthma. *BMC Complement. Altern. Med.* **2019**, *19*, 349. [[CrossRef](#)] [[PubMed](#)]
53. Nadeem, H.R.; Akhtar, S.; Sestili, P.; Ismail, T.; Neugart, S.; Qamar, M.; Esatbeyoglu, T. Toxicity, antioxidant activity, and phytochemicals of basil (*Ocimum basilicum* L.) leaves cultivated in southern Punjab, Pakistan. *Foods* **2022**, *11*, 1239. [[CrossRef](#)] [[PubMed](#)]
54. Prinsi, B.; Morgutti, S.; Negrini, N.; Faoro, F.; Espen, L. Insight into Composition of Bioactive Phenolic Compounds in Leaves and Flowers of Green and Purple Basil. *Plants* **2019**, *9*, 22. [[CrossRef](#)]
55. Da Silva, L.A.L.; Pezzini, B.R.; Soares, L. Spectrophotometric determination of the total flavonoid content in *Ocimum basilicum* L. (Lamiaceae) leaves. *Pharmacogn. Mag.* **2015**, *11*, 96–101. [[CrossRef](#)]
56. Falowo, A.B.; Mukumbo, F.E.; Idamokoro, E.M.; Afolayan, A.J.; Muchenje, V. Phytochemical constituents and antioxidant activity of sweet basil (*Ocimum Basilicum* L.) essential oil on ground beef from Boran and Nguni cattle. *Int. J. Food Sci.* **2019**, *2019*, 2628747. [[CrossRef](#)]
57. Ion, V.A.; Nicolau, F.; Petre, A.; Bujor, O.-C.; Badulescu, L. Variation of bioactive compounds in organic *Ocimum basilicum* L. during freeze-drying processing. *Sci. Pap. Ser. B Hortic.* **2020**, *64*, 397–404.
58. Tavallali, V.; Rowshan, V.; Gholami, H.; Hojati, S. Iron-urea nano-complex improves bioactive compounds in essential oils of *Ocimum basilicum* L. *Sci. Hortic.* **2020**, *265*, 109222. [[CrossRef](#)]
59. Amaral-Baroli, A.; Lago, J.H.G.; de Almeida, C.V.; de Almeida, M.; Scotti, M.T.; Leone, G.F.; Soares, M.G.; Cavalari, A.A.; Sartorelli, P. Variability in essential oil composition produced by micropropagated (in vitro), acclimated (ex vitro) and in-field plants of *Ocimum basilicum* (Lamiaceae). *Ind. Crop. Prod.* **2016**, *86*, 180–185. [[CrossRef](#)]
60. Avetisyan, A.; Markosian, A.; Petrosyan, M.; Sahakyan, N.; Babayan, A.; Aloyan, S.; Trchounian, A. Chemical composition and some biological activities of the essential oils from basil *Ocimum* different cultivars. *BMC Complement. Altern. Med.* **2017**, *17*, 2–8. [[CrossRef](#)]
61. Pirbalouti, A.G.; Mahdad, E.; Craker, L. Effects of drying methods on qualitative and quantitative properties of essential oil of two basil landraces. *Food Chem.* **2013**, *141*, 2440–2449. [[CrossRef](#)] [[PubMed](#)]
62. Tshilanda, D.D.; Babady, P.B.; Onyamboko, D.N.V.; Tshiongo, C.M.T.; Tshibangu, D.S.-T.; Ngoboua, K.-T.; Tsalu, P.; Mpiana, P.T. Chemo-type of essential oil of *Ocimum basilicum* L. from DR Congo and relative in vitro antioxidant potential to the polarity of crude extracts. *Asian Pac. J. Trop. Biomed.* **2016**, *6*, 1022–1028. [[CrossRef](#)]
63. Koroch, A.R.; Simon, J.E.; Juliani, H.R. Essential oil composition of purple basil, their reverted green varieties (*Ocimum basilicum*) and their associated biological activity. *Ind. Crop. Prod.* **2017**, *107*, 526–530. [[CrossRef](#)]
64. Sonmezdag, A.S.; Amanpour, A.; Kelebek, H.; Sellı, S. The most aroma-active compounds in shade-dried aerial parts of basil obtained from Iran and Turkey. *Ind. Crop. Prod.* **2018**, *124*, 692–698. [[CrossRef](#)]
65. Milenković, L.; Stanojević, J.; Cvetković, D.; Stanojević, L.; Lalević, D.; Šunić, L.; Fallik, E.; Ilić, Z. New technology in basil production with high essential oil yield and quality. *Ind. Crop. Prod.* **2019**, *140*, 111718. [[CrossRef](#)]
66. Ahmed, A.F.; Attia, F.A.K.; Liu, Z.; Li, C.; Wei, J.; Kang, W. Antioxidant activity and total phenolic content of essential oils and extracts of sweet basil (*Ocimum basilicum* L.) plants. *Food Sci. Hum. Wellness* **2019**, *8*, 299–305. [[CrossRef](#)]
67. Pistelli, L.; Ascrizzi, R.; Giuliani, C.; Cervelli, C.; Ruffoni, B.; Princi, E.; Fontanesi, G.; Flamini, G.; Pistelli, L. Growing basil in the underwater biospheres of nemo's garden®: Phytochemical, physiological and micromorphological analyses. *Sci. Hortic.* **2020**, *259*, 108851. [[CrossRef](#)]
68. Purushothaman, B.; Srinivasan, R.P.; Suganthi, P.; Ranganathan, B.; Gimbu, J.; Shanmugam, K. A Comprehensive Review on *Ocimum basilicum*. *J. Nat. Remedies* **2018**, *18*, 71–85. [[CrossRef](#)]
69. Maurya, S.; Chandra, M.; Yadav, R.K.; Narnoliya, L.K.; Sangwan, R.S.; Bansal, S.; Sandhu, P.; Singh, U.; Kumar, D.; Sangwan, N.S. Interspecies comparative features of trichomes in *Ocimum* reveal insights for biosynthesis of specialized essential oil metabolites. *Protoplasma* **2019**, *256*, 893–907. [[CrossRef](#)]
70. Tirillini, B.; Maggi, F. Volatile Organic Compounds of the Glandular Trichomes of *Ocimum basilicum* and Artifacts during the Distillation of the Leaves. *Appl. Sci.* **2021**, *11*, 7312. [[CrossRef](#)]

71. Prabhu, K.S.; Lobo, R.; Shirwaikar, A.A.; Shirwaikar, A. *Ocimum gratissimum*: A Review of its Chemical, Pharmacological and Ethnomedicinal Properties. *Open Complement. Med. J.* **2009**, *1*, 1–15. [CrossRef]
72. Tellu, J.O.; Oseni, S.O. Comparative Profiling of Solvent-mediated Phytochemical Expressions in *Ocimum gratissimum* and *Vernonia amygdalina* Leaf Tissues via FTIR Spectroscopy and Colorimetric Assays. *J. Adv. Med. Pharm. Sci.* **2019**, *19*, 1–25. [CrossRef]
73. Monga, S.; Dhanwal, P.; Kumar, R.; Kumar, A.; Chhokar, V. Pharmacological and physico-chemical properties of Tulsi (*Ocimum gratissimum* L.): An updated review. *Pharma Innov.* **2017**, *6*, 181–186.
74. Chowdhury, T.; Mandal, A.; Roy, S.C.; De Sarker, D. Diversity of the genus *Ocimum* (Lamiaceae) through morpho-molecular (RAPD) and chemical (GC–MS) analysis. *J. Genet. Eng. Biotechnol.* **2017**, *15*, 275–286. [CrossRef]
75. Ogendo, J.; Kostyukovsky, M.; Ravid, U.; Matasyoh, J.; Deng, A.; Omolo, E.; Kariuki, S.; Shaaya, E. Bioactivity of *Ocimum gratissimum* L. oil and two of its constituents against five insect pests attacking stored food products. *J. Stored Prod. Res.* **2008**, *44*, 328–334. [CrossRef]
76. Akara, E.U.; Emmanuel, O.; Ude, V.C.; Uche-Ikonne, C.; Eke, G.; Ugbogu, E.A. *Ocimum gratissimum* leaf extract ameliorates phenylhydrazine-induced anaemia and toxicity in wistar rats. *Drug Metab. Pers. Ther.* **2021**, *36*, 311–320. [CrossRef] [PubMed]
77. Usunomera, U.; Eseosa, U. In vitro medicinal studies on *Ocimum gratissimum* leaves. *ARC J. Pharm. Sci.* **2016**, *2*, 1–5.
78. Dambolena, J.S.; Zunino, M.P.; López, A.G.; Rubinstein, H.R.; Zygaldo, J.A.; Mwangi, J.W.; Thoithi, G.N.; Kibwage, I.O.; Mwalukumbi, J.M.; Kariuki, S.T. Essential oils composition of *Ocimum basilicum* L. and *Ocimum gratissimum* L. from Kenya and their inhibitory effects on growth and fumonisin production by *Fusarium verticillioides*. *Innov. Food Sci. Emerg. Technol.* **2010**, *11*, 410–414. [CrossRef]
79. Lee, S.J.; Umano, K.; Shibamoto, T.; Lee, K.G. Identification of volatile components in basil (*Ocimum basilicum* L.) and thyme leaves (*Thymus vulgaris* L.) and their antioxidant properties. *Food Chem.* **2005**, *91*, 131–137. [CrossRef]
80. Costa, R.S.; Carneiro, T.C.B.; Cerqueira-Lima, A.T.; Queiroz, N.V.; Alcântara-Neves, N.M.; Pontes-De-Carvalho, L.C.; da Silva Velozo, E.; Oliveira, E.J.; Figueiredo, C.A. *Ocimum gratissimum* Linn. and rosmarinic acid, attenuate eosinophilic airway inflammation in an experimental model of respiratory allergy to *Blomia tropicalis*. *Int. Immunopharmacol.* **2012**, *13*, 126–134. [CrossRef]
81. Olamilosoye, K.P.; Akomolafe, R.O.; Akinsomisoye, O.S.; Adefisayo, M.A.; Alabi, Q.K. The aqueous extract of *Ocimum gratissimum* leaves ameliorates acetic acid-induced colitis via improving antioxidant status and hematological parameters in male Wistar rats. *Egypt. J. Basic Appl. Sci.* **2018**, *5*, 220–227. [CrossRef]
82. Talabi, J.Y.; Makajuola, S. Proximate, Phytochemical, and In Vitro Antimicrobial Properties of Dried Leaves from *Ocimum gratissimum*. *Prev. Nutr. Food Sci.* **2017**, *22*, 191–194. [CrossRef] [PubMed]
83. Naveed, M.; Hejazi, V.; Abbas, M.; Kamboh, A.A.; Khan, G.J.; Shumzaid, M.; Ahmad, F.; Babazadeh, D.; FangFang, X.; Modarresi-Ghazani, F.; et al. Chlorogenic acid (CGA): A pharmacological review and call for further research. *Biomed. Pharmacother.* **2018**, *97*, 67–74. [CrossRef] [PubMed]
84. Noori, S.; Hassan, Z.M.; Yaghmaei, B.; Dolatkhah, M. Antitumor and immunomodulatory effects of salvigenin on tumor bearing mice. *Cell. Immunol.* **2013**, *286*, 16–21. [CrossRef] [PubMed]
85. Lin, H.-H.; Chan, K.-C.; Sheu, J.-Y.; Hsuan, S.-W.; Wang, C.-J.; Chen, J.-H. *Hibiscus sabdariffa* leaf induces apoptosis of human prostate cancer cells in vitro and in vivo. *Food Chem.* **2012**, *132*, 880–891. [CrossRef]
86. Salehi, B.; Machin, L.; Monzote, L.; Sharifi-Rad, J.; Ezzat, S.M.; Salem, M.A.; Merghany, R.M.; El Mahdy, N.M.; Kılıç, C.S.; Sytar, O.; et al. Therapeutic Potential of Quercetin: New Insights and Perspectives for Human Health. *ACS Omega* **2020**, *5*, 11849–11872. [CrossRef]
87. Abdulkhaleq, L.A.; Assi, M.A.; Noor, M.H.M.; Abdullah, R.; Saad, M.Z.; Taufiq-Yap, Y.H. Therapeutic uses of epicatechin in diabetes and cancer. *Vet. World* **2017**, *10*, 869–872. [CrossRef]
88. Ajayi, A.M.; Martins, D.T.D.O.; Balogun, S.O.; de Oliveira, R.G.; Ascêncio, S.D.; Soares, I.M.; Barbosa, R.D.S.; Ademowo, O.G. *Ocimum gratissimum* L. leaf flavonoid-rich fraction suppress LPS-induced inflammatory response in RAW 264.7 macrophages and peritonitis in mice. *J. Ethnopharmacol.* **2017**, *204*, 169–178. [CrossRef]
89. Venuprasad, M.; Kandikattu, H.K.; Razack, S.; Khanum, F. Phytochemical analysis of *Ocimum gratissimum* by LC-ESI-MS/MS and its antioxidant and anxiolytic effects. *S. Afr. J. Bot.* **2014**, *92*, 151–158. [CrossRef]
90. Sahoo, D.; Kumar, A. Secondary metabolites of *Ocimum gratissimum* and their biological activities. *Int. Res. J. Pharm.* **2013**, *4*, 10–28. [CrossRef]
91. Okoye, F.B.; Obonga, W.O.; Onyebule, F.A.; Ndu, O.O.; Ihewereme, C.P. Chemical composition and anti-inflammatory activity of essential oils from the leaves of *Ocimum basilicum* L. and *Ocimum gratissimum* L. (Lamiaceae). *Int. J. Pharm. Sci. Res.* **2014**, *5*, 2174–2180.
92. Mith, H.; Yayi-Ladékan, E.; Sika Kpoviessi, S.D.; Yaou Bokossa, I.; Moudachirou, M.; Daube, G.; Clinquart, A. Chemical Composition and Antimicrobial Activity of Essential Oils of *Ocimum basilicum*, *Ocimum canum* and *Ocimum gratissimum* in Function of Harvesting Time. *J. Essent. Oil Bear. Plants* **2016**, *19*, 1413–1425. [CrossRef]
93. Sen, A. Prophylactic and therapeutic roles of oleanolic acid and its derivatives in several diseases. *World J. Clin. Cases* **2020**, *8*, 1767–1792. [CrossRef] [PubMed]
94. Vieira, A.; Beserra, F.; Souza, M.; Totti, B.; Rozza, A. Limonene: Aroma of innovation in health and disease. *Chem.-Biol. Interact.* **2018**, *283*, 97–106. [CrossRef]

95. Sacchetti, G.; Medici, A.; Maietti, S.; Radice, M.; Muzzoli, M.; Manfredini, S.; Braccioli, E.; Bruni, R. Composition and Functional Properties of the Essential Oil of Amazonian Basil, *Ocimum micranthum* Willd., Labiateae in Comparison with Commercial Essential Oils. *J. Agric. Food Chem.* **2004**, *52*, 3486–3491. [[CrossRef](#)]
96. Tacchini, M.; Guevara, M.E.; Grandini, A.; Maresca, I.; Radice, M.; Angioletta, L.; Guerrini, A. *Ocimum campechianum* Mill. from Amazonian Ecuador: Chemical Composition and Biological Activities of Extracts and Their Main Constituents (Eugenol and Rosmarinic Acid). *Molecules* **2020**, *26*, 84. [[CrossRef](#)]
97. Silva, M.G.V.; Vieira, I.G.P.; Mendes, F.N.P.; Albuquerque, I.L.; Dos Santos, R.N.; Silva, F.O.; Morais, S.M. Variation of Ursolic Acid Content in Eight *Ocimum* Species from Northeastern Brazil. *Molecules* **2008**, *13*, 2482–2487. [[CrossRef](#)]
98. Amorim, E.L.; Nascimento, J.E.; Monteiro, J.M.; Peixoto Sobrinho, T.; Araújo, T.A.; Albuquerque, U.P. A simple and accurate procedure for the determination of tannin and flavonoid levels and some applications in ethnobotany and ethnopharmacology. *Funct. Ecosyst. Communities* **2008**, *2*, 88–94.
99. Wilson, T.M.; Murphy, B.J.; Abad, A.; Packer, C.; Poulson, A.; Carlson, R.E. Essential oil composition and stable isotope profile of cultivated *Ocimum campechianum* Mill. (Lamiaceae) from Peru. *Molecules* **2022**, *27*, 2777. [[CrossRef](#)]
100. Trevisan, M.T.S.; Silva, M.G.V.; Pfundstein, B.; Spiegelhalder, A.B.; Owen, R.W. Characterization of the Volatile Pattern and Antioxidant Capacity of Essential Oils from Different Species of the Genus *Ocimum*. *J. Agric. Food Chem.* **2006**, *54*, 4378–4382. [[CrossRef](#)]
101. Caamal-Herrera, I.O.; Carrillo-Cocom, L.M.; Escalante-Réndiz, D.Y.; Aráiz-Hernández, D.; Azamar-Barrios, J.A. Antimicrobial and antiproliferative activity of essential oil, aqueous and ethanolic extracts of *Ocimum micranthum* Willd leaves. *BMC Complement. Altern. Med.* **2018**, *18*, 55. [[CrossRef](#)] [[PubMed](#)]
102. Scalvenzi, L.; Radice, M.; Toma, L.; Severini, F.; Boccolini, D.; Bella, A.; Guerrini, A.; Tacchini, M.; Sacchetti, G.; Chiurato, M. Larvicidal activity of *Ocimum campechianum*, *Ocotea quixos* and *Piper aduncum* essential oils against *Aedes aegypti*. *Parasite* **2019**, *26*, 23. [[CrossRef](#)] [[PubMed](#)]
103. Ruiz-Vargas, J.A.; Morales-Ferra, D.L.; Ramírez-Ávila, G.; Zamilpa, A.; Negrete-León, E.; Acevedo-Fernández, J.J.; Peña-Rodríguez, L.M. α -Glucosidase inhibitory activity and in vivo antihyperglycemic effect of secondary metabolites from the leaf infusion of *Ocimum campechianum* mill. *J. Ethnopharmacol.* **2019**, *243*, 112081. [[CrossRef](#)] [[PubMed](#)]
104. Figueiredo, P.L.B.; Silva, S.G.; Nascimento, L.D.; Ramos, A.R.; Setzer, W.N.; da Silva, J.K.R.; Andrade, E.H.A. Seasonal Study of Methyleugenol Chemotype of *Ocimum campechianum* Essential Oil and Its Fungicidal and Antioxidant Activities. *Nat. Prod. Commun.* **2018**, *13*, 1934578X1801300833. [[CrossRef](#)]
105. Benitez, N.P.; León, E.M.M.; Stashenko, E.E. Eugenol and methyl eugenol chemotypes of essential oil of species *Ocimum gratissimum* L. and *Ocimum campechianum* Mill. from Colombia. *J. Chromatogr. Sci.* **2009**, *47*, 800–803. [[CrossRef](#)]
106. Vij, D.; Gupta, S. *Ocimum sanctum*: The queen of herbs. *Int. J. Multidiscip. Curr. Educ. Res. (IJMCER)* **2021**, *3*, 49–52.
107. Deshmukh, A.; Deshmukh, G.; Shirole, P. *Ocimum sanctum*: A medicinal gift from nature. *Int. J. Pharmacogn.* **2015**, *2*, 550–559.
108. Singh, E.; Sharma, S.; Dwivedi, J.; Sharma, S. Diversified potentials of *Ocimum sanctum* Linn (Tulsi): An exhaustive survey. *J. Nat. Prod. Plant Resour.* **2012**, *2*, 39–48.
109. Vishwabhan, S.; Birendra, V.; Vishal, S. A review on ethnomedical uses of *Ocimum sanctum* (Tulsi). *Int. Res. J. Pharm.* **2011**, *2*, 1–3.
110. Agarwal, S.; Kumar, V.R.; Kumar, A. Pharmacognostical characterization of *Ocimum* spp. *J. Pharmacogn. Phytochem.* **2013**, *2*, 219–224.
111. Prakash, P.; Gupta, N. Therapeutic uses of *Ocimum sanctum* Linn (Tulsi) with a note on eugenol and its pharmacological actions: A short review. *Indian J. Physiol. Pharmacol.* **2005**, *49*, 125–131. [[PubMed](#)]
112. Doss, A. Preliminary phytochemical screening of some Indian Medicinal Plants. *Anc. Sci. Life* **2009**, *29*, 12–16. [[PubMed](#)]
113. Borah, R.; Biswas, S. Tulsi (*Ocimum sanctum*), excellent source of phytochemicals. *Int. J. Environ. Agric. Biotechnol.* **2018**, *3*, 265258.
114. Pattanayak, P.; Behera, P.; Das, D.; Panda, S.K. *Ocimum sanctum* Linn. A reservoir plant for therapeutic applications: An overview. *Pharmacogn. Rev.* **2010**, *4*, 95–105. [[CrossRef](#)]
115. Hakkim, F.L.; Shankar, C.G.; Girija, S. Chemical Composition and Antioxidant Property of Holy Basil (*Ocimum sanctum* L.) Leaves, Stems, and Inflorescence and Their in Vitro Callus Cultures. *J. Agric. Food Chem.* **2007**, *55*, 9109–9117. [[CrossRef](#)]
116. Pandey, R.; Chandra, P.; Srivastava, M.; Mishra, D.; Kumar, B. Simultaneous quantitative determination of multiple bioactive markers in *Ocimum sanctum* obtained from different locations and its marketed herbal formulations using UPLC-ESI-MS/MS combined with principal component analysis. *Phytochem. Anal.* **2015**, *26*, 383–394. [[CrossRef](#)]
117. Anbarasu, K.; Vijayalakshmi, G. Improved Shelf Life of Protein-Rich Tofu Using *Ocimum sanctum* (Tulsi) Extracts to Benefit Indian Rural Population. *J. Food Sci.* **2007**, *72*, M300–M305. [[CrossRef](#)]
118. Rahman, S.; Islam, R.; Kamruzzaman, M.; Alam, K.; Jamal, A. *Ocimum sanctum* L.: A review of phytochemical and pharmacological profile. *Am. J. Drug Discov. Dev.* **2011**, *1*, 15.
119. Singh, D.; Chaudhuri, P.K. A review on phytochemical and pharmacological properties of Holy basil (*Ocimum sanctum* L.). *Ind. Crop. Prod.* **2018**, *118*, 367–382. [[CrossRef](#)]
120. Desai, S.; Daunde, J.; Potphode, N.; Walvekar, M. GC-MS analysis of *Ocimum sanctum* seeds. *BIOINFOLET Q. J. Life Sci.* **2021**, *18*, 163–164.
121. Saharkhiz, M.J.; Alam Kamyab, A.; Kazerani, N.K.; Zomorodian, K.; Pakshir, K.; Rahimi, M.J. Chemical Compositions and Antimicrobial Activities of *Ocimum sanctum* L. Essential Oils at Different Harvest Stages. *Jundishapur J. Microbiol.* **2015**, *8*, e13720. [[CrossRef](#)] [[PubMed](#)]

122. Saaban, K.F.; Ang, C.H.; Chuah, C.H.; Khor, S.M. Chemical constituents and antioxidant capacity of *Ocimum basilicum* and *Ocimum sanctum*. *Iran. J. Chem. Chem. Eng. (IJCCE)* **2019**, *38*, 139–152.
123. Hussain, A.I.; Chattha, S.A.S.; Kamal, G.M.; Ali, M.A.; Hanif, M.A.; Lazhari, M.I. Chemical composition and biological activities of essential oil and extracts from *Ocimum sanctum*. *Int. J. Food Prop.* **2017**, *20*, 1569–1581. [[CrossRef](#)]
124. Panchal, P.; Parvez, N. Phytochemical analysis of medicinal herb (*Ocimum sanctum*). *Int. J. Nanomater. Nanotechnol. Nanomed.* **2019**, *5*, 008–011. [[CrossRef](#)]
125. Ijaz, B.; Hanif, M.; Mushtaq, Z.; Khan, M.; Bhatti, I.; Jilani, M. Isolation of bioactive fractions from *Ocimum sanctum* essential oil. *Oxid. Commun.* **2017**, *40*, 158–167.
126. Thaweboon, S.; Thaweboon, B. In vitro antimicrobial activity of *Ocimum americanum* L. essential oil against oral microorganisms. *Southeast Asian J. Trop. Med. Public Health* **2009**, *40*, 1025–1033.
127. Verma, R.S.; Bisht, P.S.; Padalia, R.C.; Saikia, D.; Chauhan, A. Chemical composition and antibacterial activity of essential oil from two *Ocimum* spp. grown in sub-tropical India during spring-summer cropping season. *J. Tradit. Med.* **2011**, *6*, 211–217.
128. Taufiq, M.; Darah, I. Antibacterial activity of an endophytic fungus *Lasiodiplodia pseudotheobromae* IBRL OS-64 residing in leaves of a medicinal herb, *Ocimum sanctum* Linn. *J. Appl. Biol. Biotechnol.* **2019**, *7*, 35–41.
129. Evangelina, I.A.; Herdiyati, Y.; Laviana, A.; Rikmasari, R.; Zubaedah, C.; Anisah; Kurnia, D. Bio-mechanism inhibitory prediction of B-Sitosterol from Kemangi (*Ocimum basilicum* L.) as an inhibitor of MurA Enzyme of Oral bacteria: In vitro and in silico study. *Adv. Appl. Bioinform. Chem. AABC* **2021**, *14*, 103–115. [[CrossRef](#)]
130. Jesuwenu, A.D.; Michael, O.O. Ntibacterial activity of *Ocimum gratissimum* L. Against some selected human bacterial pathogens. *J. Pharm. Res. Int.* **2010**, *20*, 1–8. [[CrossRef](#)]
131. Carović-Stanko, K.; Orlić, S.; Politeo, O.; Strikić, F.; Kolak, I.; Milos, M.; Satovic, Z. Composition and antibacterial activities of essential oils of seven *Ocimum* taxa. *Food Chem.* **2010**, *119*, 196–201. [[CrossRef](#)]
132. Moghaddam, A.M.D.; Shayegh, J.; Mikaili, P.; Sharaf, J.D. Antimicrobial activity of essential oil extract of *Ocimum basilicum* L. Leaves on a variety of pathogenic bacteria. *J. Med. Plants Res.* **2011**, *5*, 3453–3456.
133. Melo, R.S.; Azevedo, M.A.; Pereira, A.M.G.; Rocha, R.R.; Cavalcante, R.M.B.; Matos, M.N.C.; Lopes, P.H.R.; Gomes, G.A.; Rodrigues, T.H.S.; dos Santos, H.S.; et al. Chemical Composition and Antimicrobial Effectiveness of *Ocimum gratissimum* L. Essential Oil Against Multidrug-Resistant Isolates of *Staphylococcus aureus* and *Escherichia coli*. *Molecules* **2019**, *24*, 3864. [[CrossRef](#)] [[PubMed](#)]
134. Rajesh, T.; Venkatanagaraju, E.; Goli, D.; Basha, S.J. Evaluation of antimicrobial activity of different herbal plant extracts. *Int. J. Pharm. Sci. Res.* **2014**, *5*, 1460–1468.
135. Harikrishnan, R.; Kim, M.-C.; Kim, J.-S.; Balasundaram, C.; Jawahar, S.; Heo, M.-S. Identification and Antimicrobial Activity of Combined Extract from *Azadirachta indica* and *Ocimum sanctum*. *Isr. J. Aquac.-Bamidgeh* **2010**, *62*, 85–95. [[CrossRef](#)]
136. Ogundare, A. Antibacterial properties of the leaf extracts of *Vernonia amygdalina*, *Ocimum gratissimum*, *Corchorous olitorius* and *Manihot palmate*. *J. Microbiol. Antimicrob.* **2011**, *3*, 77–86.
137. Ali, H.; Nguta, J.; Musila, F.; Ole-Mapenay, I.; Matara, D.; Mailu, J. Evaluation of antimicrobial activity, cytotoxicity, and phytochemical composition of *Ocimum americanum* L. (Lamiaceae). *Evid. -Based Complement. Altern. Med.* **2022**, *2022*, 6484578. [[CrossRef](#)]
138. Torpol, K.; Wiriacharee, P.; Sriwattana, S.; Sangsuwan, J.; Prinyawiwatkul, W. Antimicrobia activity of garlic (*Allium sativum* L.) and holy basil (*Ocimum sanctum* L.) essential oils applied by liquid vs. vapour phases. *Int. J. Food Sci. Technol.* **2018**, *53*, 2119–2128. [[CrossRef](#)]
139. Helal, I.M.; El-Bessoumy, A.; Al-Bataineh, E.; Joseph, M.R.P.; Rajagopalan, P.; Chandramoorthy, H.C.K.; Ahmed, S.B.H. Antimicrobial Efficiency of Essential Oils from Traditional Medicinal Plants of Asir Region, Saudi Arabia, over Drug Resistant Isolates. *BioMed. Res. Int.* **2019**, *2019*, 8928306. [[CrossRef](#)]
140. Opara, A.; Egbuobi, R.; Ndudim, J.N.D.; Onywuchi, C.; Nnodim, J. Antibacterial Activity of *Ocimum gratissimum* (Nchu-Anwu) and *Vernonia amygdalina* (Bitter-Leaf) Antibacterial Activity of *Ocimum gratissimum* (Nchu-Anwu) and *Vernonia amygdalina* (Bitter-Leaf). *Br. Biotechnol. J.* **2014**, *4*, 1115–1122. [[CrossRef](#)]
141. Hapsari, I.P.; Feroniasanti, Y.M.L. Phytochemical screening and in vitro antibacterial activity of sweet basil leaves (*Ocimum basilicum* L.) essential oil against *Cutibacterium acnes* ATCC 11827. *AIP Conf. Proc.* **2019**, *2099*, 020007.
142. Bulfon, C.; Volpatti, D.; Galeotti, M. In Vitro Antibacterial Activity of Plant Ethanolic Extracts against Fish Pathogens. *J. World Aquac. Soc.* **2014**, *45*, 545–557. [[CrossRef](#)]
143. Gupta, P.C.; Batra, R.; Chauhan, A.; Goyal, P.; Kaushik, P. Antibacterial activity and TLC bioautography of *Ocimum basilicum* L. against pathogenic bacteria. *J. Pharm. Res.* **2009**, *2*, 407–409.
144. Stanley, M.C.; Ifeanyi, O.E.; Chinedum, O.K.; Chinene, N.D. The Antibacterial Activity of Leaf Extracts of *Ocimum Gratissimum* and *Sida Acuta*. *Int. J. Microbiol. Res.* **2014**, *5*, 124–129. [[CrossRef](#)]
145. Issazadeh, K.; Majid, K.P.M.R.; Massiha, A.; Bidarigh, S.; Giahi, M.; Zulfagar, M.P. Analysis of the phytochemical contents and anti-microbial activity of *Ocimum basilicum* L. *Int. J. Mol. Clin. Microbiol.* **2012**, *2*, 141–147.
146. Thaweboon, S.; Thaweboon, B. *Ocimum americanum* L. essential oil exhibits antimicrobial activity against oral bacteria related to periodontal disease. *Adv. Mater. Res.* **2014**, *1025–1026*, 755–759. [[CrossRef](#)]
147. Mahmood, K.; Yaqoob, U.; Bajwa, R. Antibacterial activity of essential oil of *Ocimum sanctum* L. *Mycopath* **2008**, *6*, 2.

148. Nweze, E.I.; Eze, E.E. Justification for the use of *Ocimum gratissimum* L in herbal medicine and its interaction with disc antibiotics. *BMC Complement. Altern. Med.* **2009**, *9*, 37. [CrossRef]
149. Londhe, A.; Kulkarni, A.; Lawand, R. In-vitro comparative study of antibacterial and antifungal activities: A case study of *Ocimum kilimandscharicum*, *Ocimum tenuiflorum* and *Ocimum gratissimum*. *Int. J. Pharmacogn. Phytochem. Res.* **2015**, *7*, 104–110.
150. Helen, M.; Raju, V.; Gomathy, S.; Nizzy, S.; Sree, S. Essential oil analysis in *Ocimum* sps. *Herb. Technol. Ind.* **2011**, *8*, 12–15.
151. Intorasoot, A.; Chornchoem, P.; Sookkhee, S.; Intorasoot, S. Bactericidal activity of herbal volatile oil extracts against multidrug-resistant *Acinetobacter baumannii*. *J. Intercult. Ethnopharmacol.* **2017**, *6*, 218–222. [CrossRef] [PubMed]
152. Niculae, M.; Spînu, M.; Sandru, C.D.; Brudaşă, F.; Cedar, D.; Szakacs, B.; Scurtu, I.; Bolfă, P.; Mateş, C. Antimicrobial potential of some Lamiaceae essential oils against animal multi-resistant bacteria. *Lucr. Științifice Med. Vet.* **2009**, *42*, 170–175.
153. Moghaddam, A.M.D.; Shayegh, J.; Sharaf, J.D. Comparison between Two Groups of Pathogenic Bacteria under Different Essential Oil Extract of *Ocimum basilicum* L. *J. Med. Plants By-Prod.* **2017**, *6*, 241–245. [CrossRef]
154. Hussain, A.I.; Anwar, F.; Sherazi, S.T.H.; Przybylski, R. Chemical composition, antioxidant and antimicrobial activities of basil (*Ocimum basilicum*) essential oils depends on seasonal variations. *Food Chem.* **2008**, *108*, 986–995. [CrossRef] [PubMed]
155. Ighodaro, O.; Agunbiade, S.; Akintobi, O. Phytotoxic and anti-microbial activities of flavonoids in *Ocimum gratissimum*. *Life Sci. J.* **2010**, *7*, 45–48.
156. Katara, A.; Pradhan, C.K.; Singh, P.; Singh, V.; Ali, M. Volatile Constituents and Antimicrobial Activity of Aerial parts of *Ocimum gratissimum* Linn. *J. Essent. Oil Bear. Plants* **2013**, *16*, 283–288. [CrossRef]
157. De Aguiar, F.C.; Solarte, A.L.; Tarradas, C.; Luque, I.; Maldonado, A.; Galán-Relaño, Á.; Huerta, B. Antimicrobial activity of selected essential oils against *Streptococcus suis* isolated from pigs. *MicrobiologyOpen* **2018**, *7*, e00613. [CrossRef]
158. Herdiyati, Y.; Astrid, Y.; Shadrina, A.A.N.; Wiani, I.; Satari, M.H.; Kurnia, D. Potential Fatty Acid as Antibacterial Agent against Oral Bacteria of *Streptococcus mutans* and *Streptococcus sanguinis* from Basil (*Ocimum americanum*): In vitro and In silico Studies. *Curr. Drug Discov. Technol.* **2021**, *18*, 532–541. [CrossRef]
159. Phanthong, P.; Lomarat, P.; Chomnawang, M.T.; Bunyaphraphatsara, N. Antibacterial activity of essential oils and their active components from Thai spices against foodborne pathogens. *ScienceAsia* **2013**, *39*, 472. [CrossRef]
160. Koche, D.; Kokate, P.; Suradkar, S.; Bhadange, D. Preliminary phytochemistry and antibacterial activity of ethanolic extract of *Ocimum gratissimum* L. *Biosci. Discov.* **2012**, *3*, 20–24.
161. Shadrina, A.A.N.; Herdiyati, Y.; Wiani, I.; Satari, M.H.; Kurnia, D. Prediction Mechanism of Nevadensin as Antibacterial Agent against *S. sanguinis*: In vitro and In silico Studies. *Comb. Chem. High Throughput Screen.* **2022**, *25*, 1488–1497. [CrossRef] [PubMed]
162. Yamani, H.A.; Pang, E.C.; Mantri, N.; Deighton, M.A. Antimicrobial Activity of Tulsi (*Ocimum tenuiflorum*) Essential Oil and Their Major Constituents against Three Species of Bacteria. *Front. Microbiol.* **2016**, *7*, 681. [CrossRef] [PubMed]
163. Onyebuchi, C.; Kavaz, D. Effect of extraction temperature and solvent type on the bioactive potential of *Ocimum gratissimum* L. extracts. *Sci. Rep.* **2020**, *10*, 21760. [CrossRef]
164. Majdi, C.; Pereira, C.; Dias, M.I.; Calhelha, R.C.; Alves, M.J.; Rhourri-Frih, B.; Charrouf, Z.; Barros, L.; Amaral, J.S.; Ferreira, I.C. Phytochemical Characterization and Bioactive Properties of Cinnamon Basil (*Ocimum basilicum* cv. ‘Cinnamon’) and Lemon Basil (*Ocimum × citriodorum*). *Antioxidants* **2020**, *9*, 369. [CrossRef] [PubMed]
165. Ejele, A.; Duru, I.; Ogukwe, C.; Iwu, I. Phytochemistry and antimicrobial potential of basic metabolites of *Piper umbellatum*, *Piper guineense*, *Ocimum gratissimum* and *Newbouldia laevis* extracts. *J. Emerg. Trends Eng. Appl. Sci.* **2012**, *3*, 309–314.
166. Pandey, A.K.; Singh, P.; Palni, U.; Tripathi, N. In-vitro antibacterial activities of the essential oils of aromatic plants against *Erwinia herbicola* (Lohnis) and *Pseudomonas putida* (Kris Hamilton). *J. Serb. Chem. Soc.* **2012**, *77*, 313–323. [CrossRef]
167. Maffei, M.E.; Gertsch, J.; Appendino, G. Plant volatiles: Production, function and pharmacology. *Nat. Prod. Rep.* **2011**, *28*, 1359–1380. [CrossRef]
168. Bhattacharjya, D.; Adhikari, S.; Biswas, A.; Bhuimali, A.; Ghosh, P.; Saha, S. Ocimum Phytochemicals and Their Potential Impact on Human Health. In *Phytochemicals in Human Health*; IntechOpen: London, UK, 2020.
169. Nazzaro, F.; Fratianni, F.; De Martino, L.; Coppola, R.; De Feo, V. Effect of Essential Oils on Pathogenic Bacteria. *Pharmaceuticals* **2013**, *6*, 1451–1474. [CrossRef]
170. El-Shenawy, M.A.; Baghdadi, H.H.; El-Hosseiny, L.S. Antibacterial activity of plants essential oils against some epidemiologically relevant food-borne pathogens. *Open Public Health J.* **2015**, *8*, 30–34. [CrossRef]
171. Faleiro, M.L. The mode of antibacterial action of essential oils. *Sci. Against Microb. Pathog. Commun. Curr. Res. Technol. Adv.* **2011**, *2*, 1143–1156.
172. Murbach Teles Andrade, B.F.; Nunes Barbosa, L.; da Silva Probst, I.; Fernandes Júnior, A. Antimicrobial activity of essential oils. *J. Essent. Oil Res.* **2014**, *26*, 34–40. [CrossRef]
173. Akthar, M.S.; Degaga, B.; Azam, T. Antimicrobial activity of essential oils extracted from medicinal plants against the pathogenic microorganisms: A review. *Issues Biol. Sci. Pharm. Res.* **2014**, *2*, 1–7.
174. Turgis, M.; Han, J.; Caillet, S.; Lacroix, M. Antimicrobial activity of mustard essential oil against *Escherichia coli* O157:H7 and *Salmonella typhi*. *Food Control* **2009**, *20*, 1073–1079. [CrossRef]
175. Di Pasqua, R.; Hoskins, N.; Betts, G.; Mauriello, G. Changes in Membrane Fatty Acids Composition of Microbial Cells Induced by Addiction of Thymol, Carvacrol, Limonene, Cinnamaldehyde, and Eugenol in the Growing Media. *J. Agric. Food Chem.* **2006**, *54*, 2745–2749. [CrossRef] [PubMed]

176. Calsamiglia, S.; Busquet, M.; Cardozo, P.; Castillejos, L.; Ferret, A. Invited Review: Essential Oils as Modifiers of Rumen Microbial Fermentation. *J. Dairy Sci.* **2007**, *90*, 2580–2595. [[CrossRef](#)] [[PubMed](#)]
177. Oussalah, M.; Caillet, S.; Saucier, L.; Lacroix, M. Antimicrobial effects of selected plant essential oils on the growth of a *Pseudomonas putida* strain isolated from meat. *Meat Sci.* **2006**, *73*, 236–244. [[CrossRef](#)]
178. Rassem, H.H.; Nour, A.H.; Yunus, R.M. Techniques for extraction of essential oils from plants: A review. *Aust. J. Basic Appl. Sci.* **2016**, *10*, 117–127.
179. Ugbogu, O.C.; Emmanuel, O.; Agi, G.O.; Ibe, C.; Ekweogu, C.N.; Ude, V.C.; Uche, M.E.; Nnanna, R.O.; Ugbogu, E.A. A review on the traditional uses, phytochemistry, and pharmacological activities of clove basil (*Ocimum gratissimum* L.). *Heliyon* **2021**, *7*, e08404. [[CrossRef](#)]