

Supplementary Materials

for

Characterization of Various Titanium-Dioxide-Based Catalysts Regarding Photocatalytic Mineralization of Carbamazepine also Combined with Ozonation

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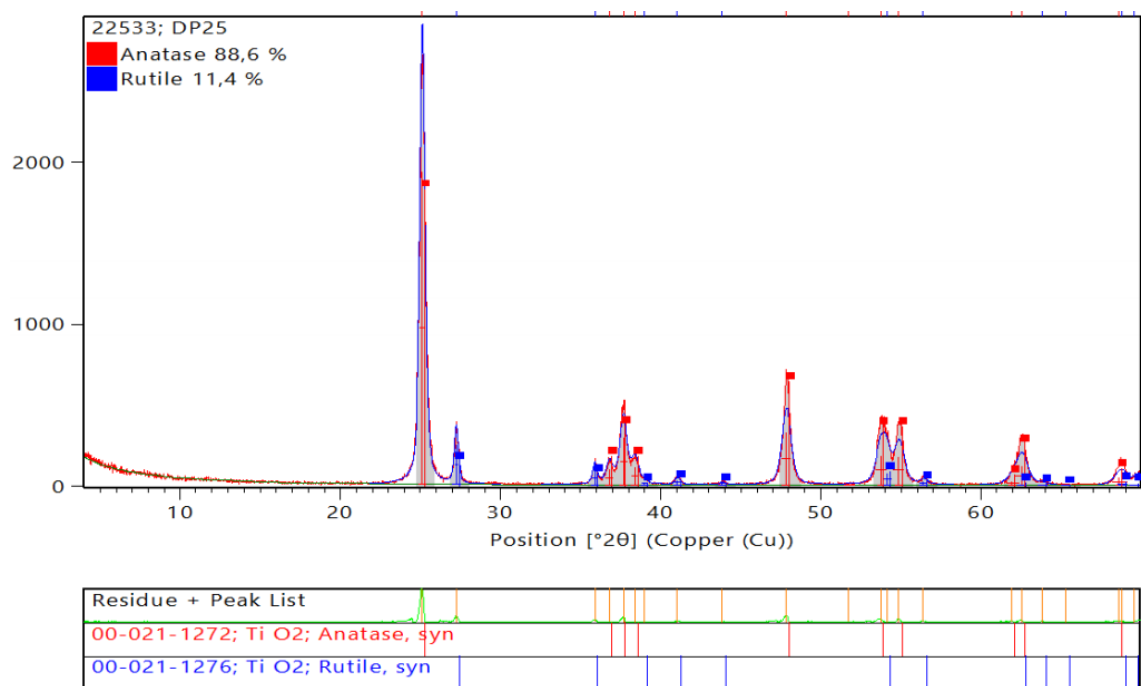


Figure S1. a) Identification and quantification of anatase and rutile in the XRD pattern of pristine DP25 TiO₂.

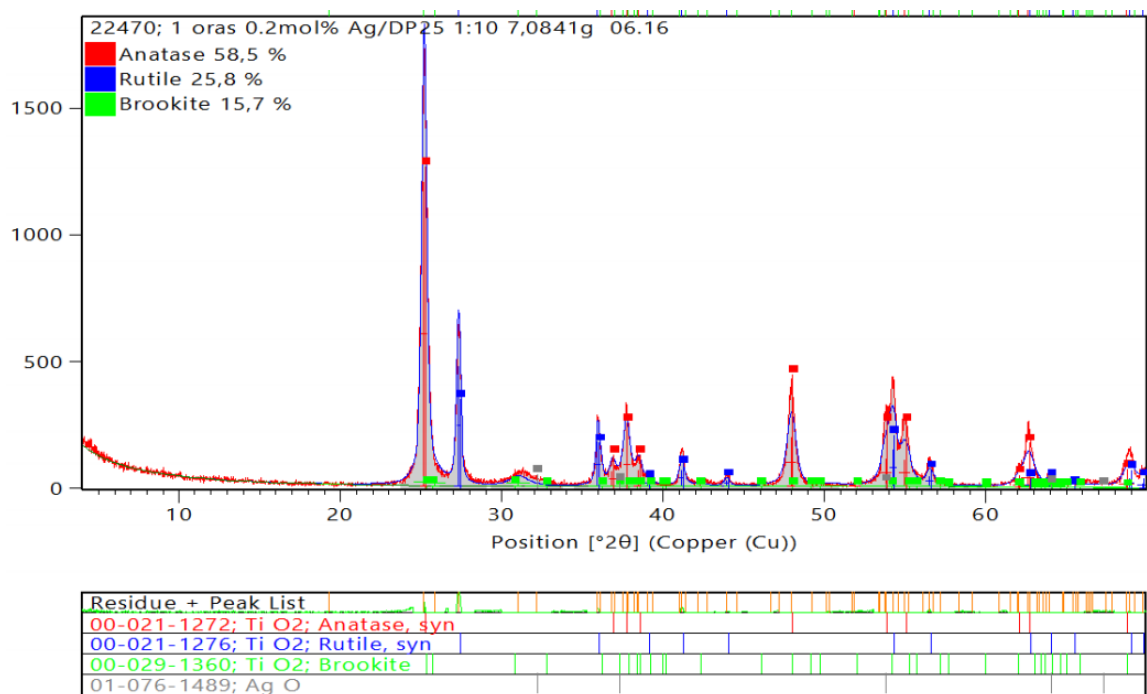


Figure S1. b) Identification and quantification of anatase, rutile, and brookite in the XRD pattern of mechanochemically silverized DP25 TiO₂.

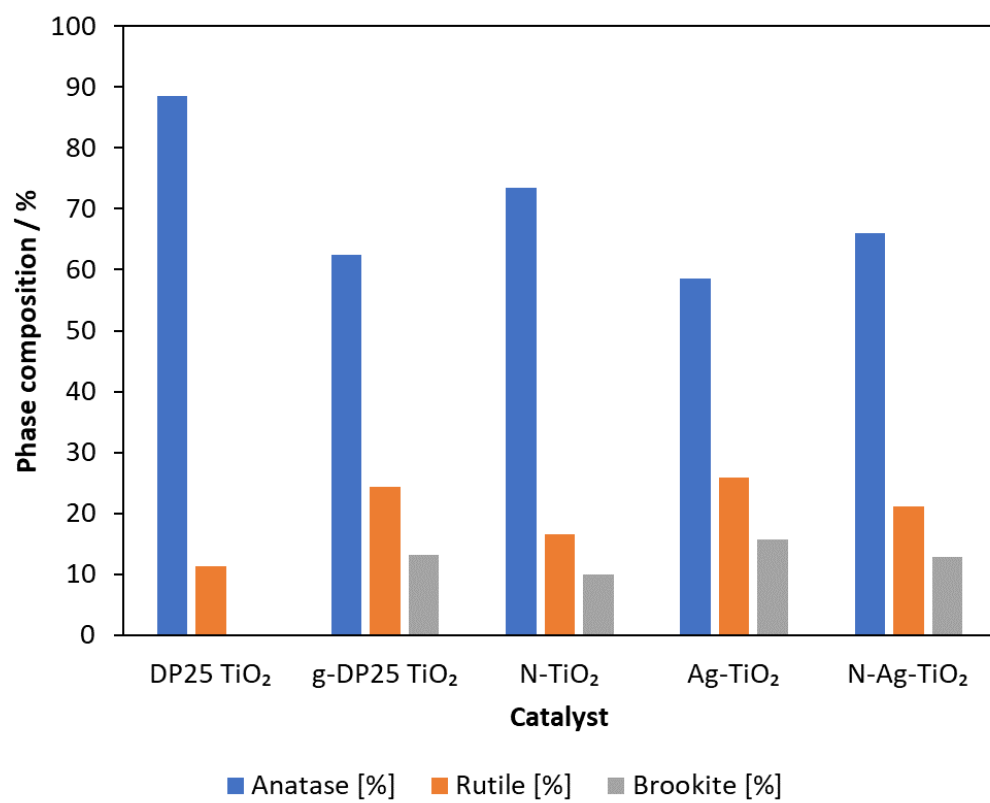


Figure S2. The phase composition of catalysts used.

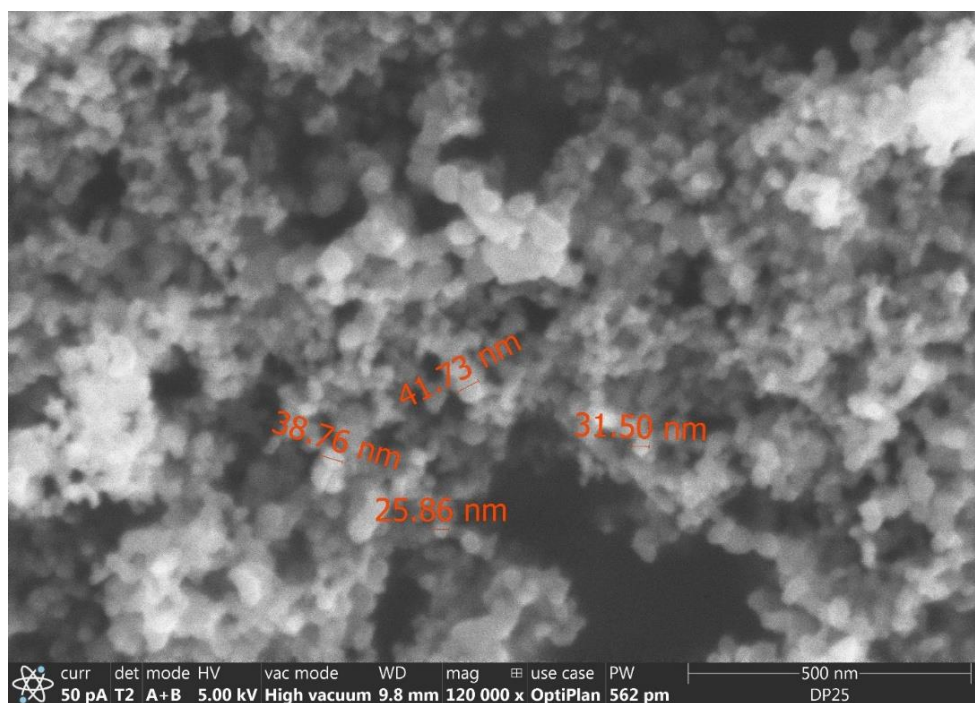


Figure S3. a) SEM morphology of pristine DP25 TiO₂ catalyst at higher magnification.

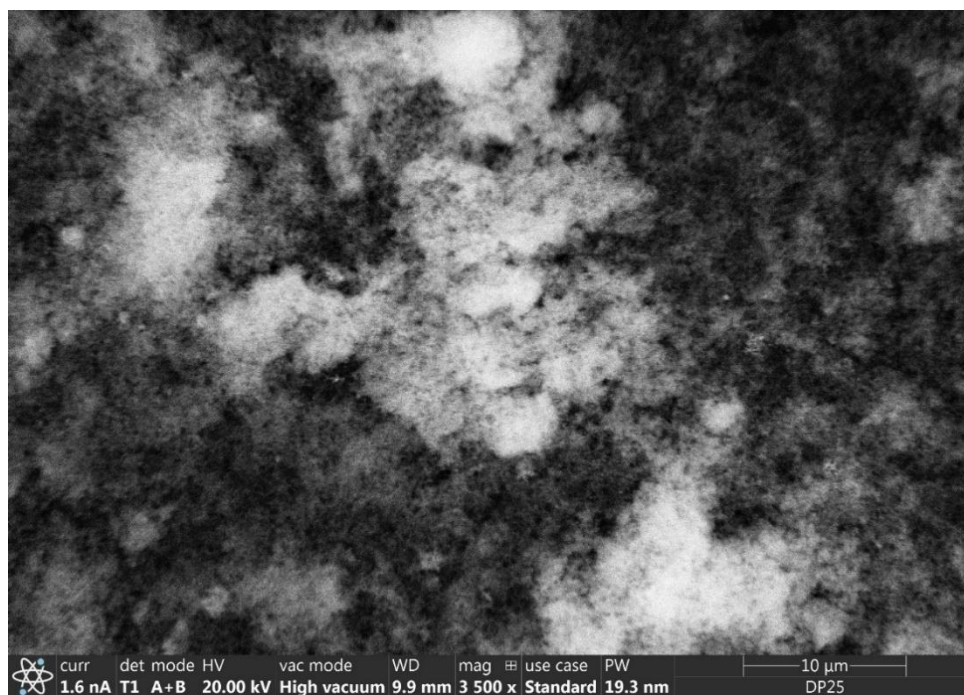


Figure S3. b) SEM morphology of pristine DP25 TiO₂ catalyst at lower magnification.

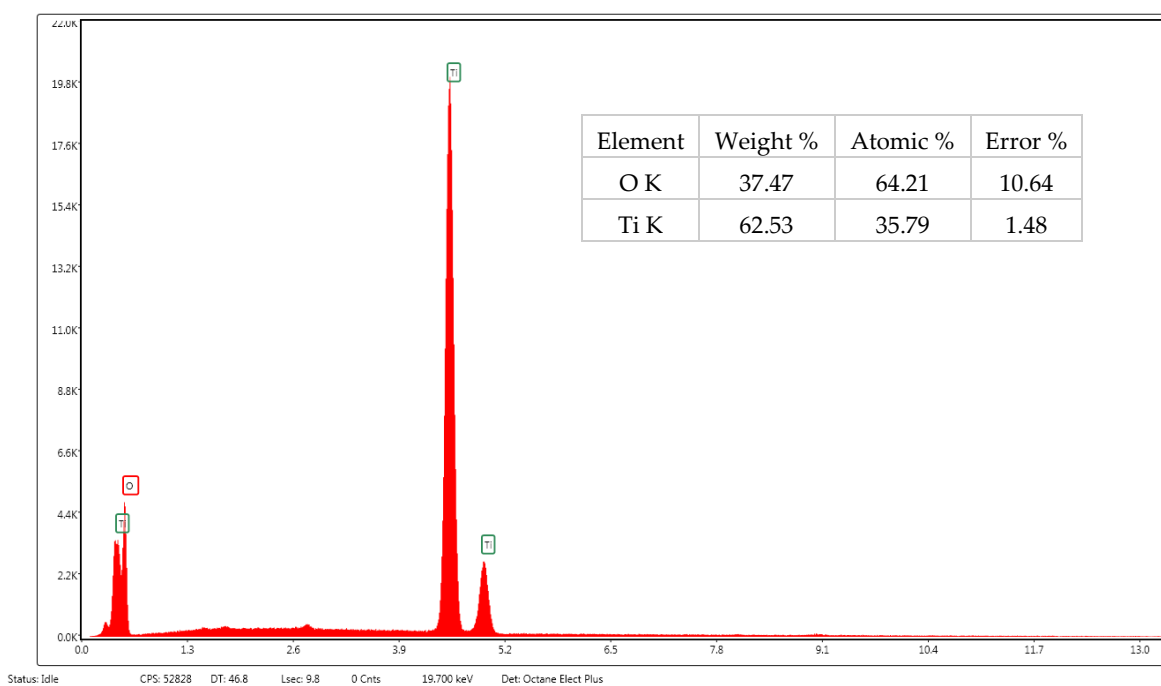
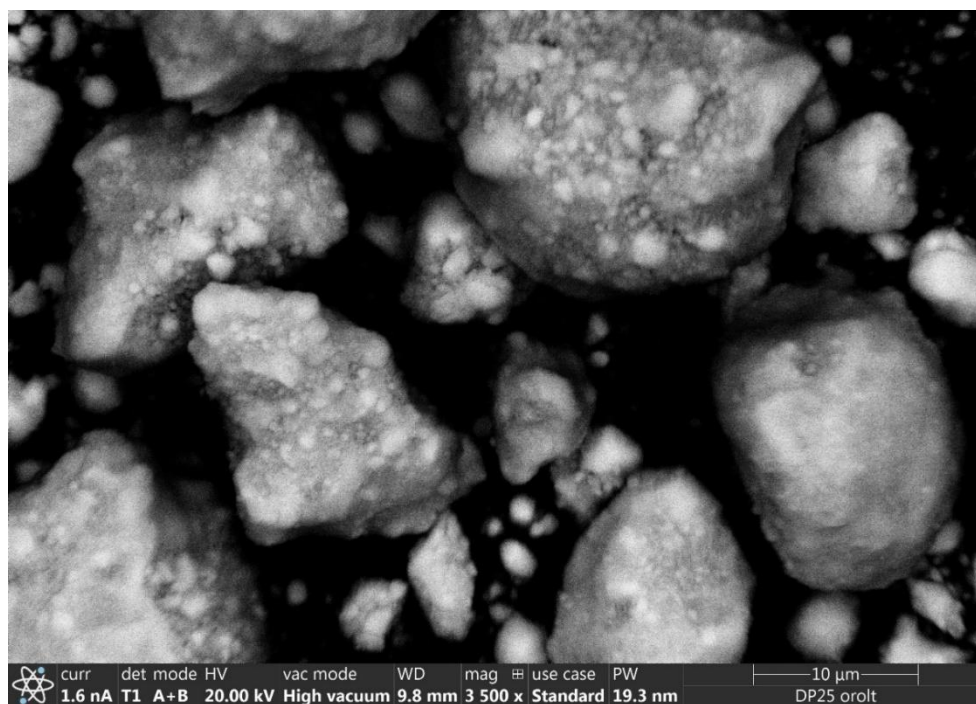


Figure S3. c) SEM morphology and elemental composition of ground DP25 TiO₂ catalyst.

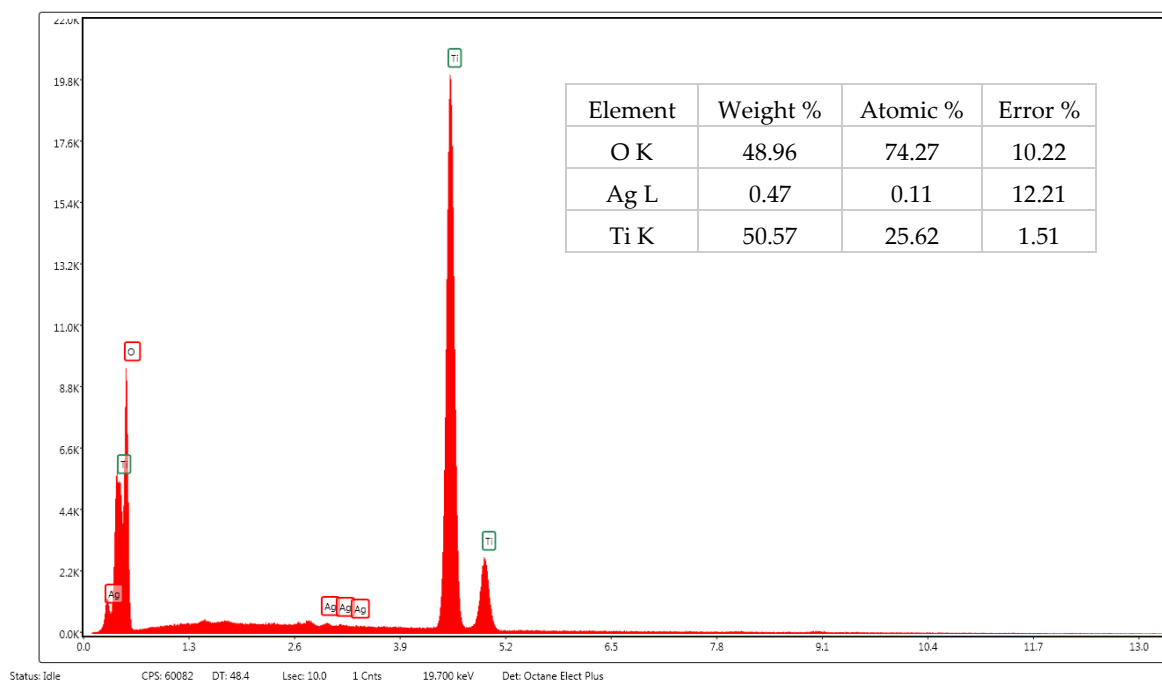
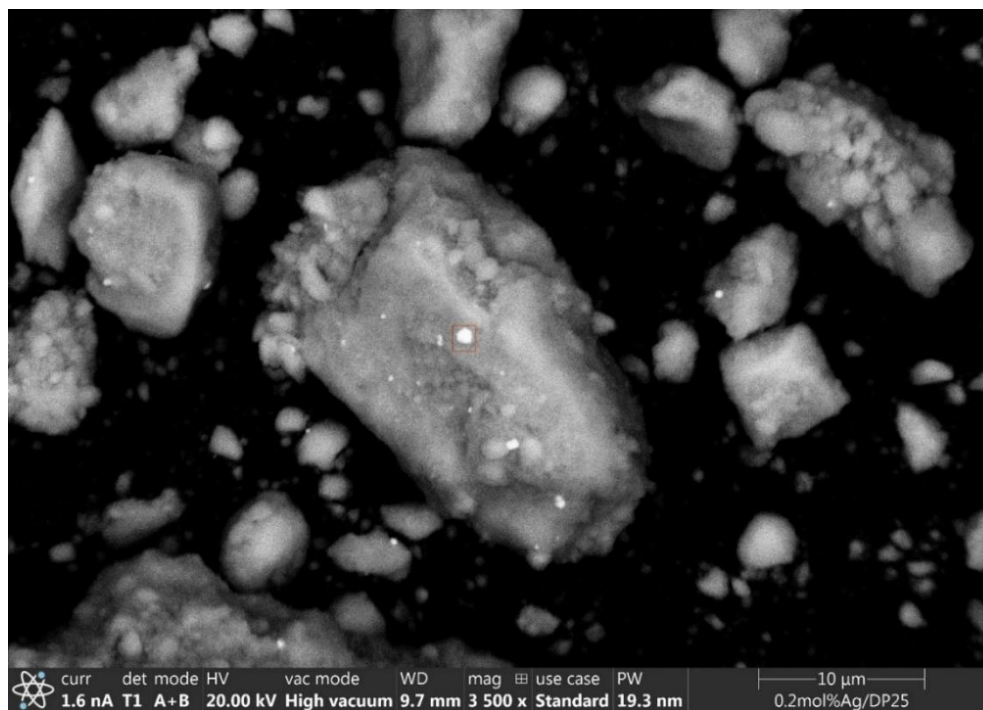


Figure S3. d) SEM morphology and elemental composition of Ag-TiO₂ catalyst.

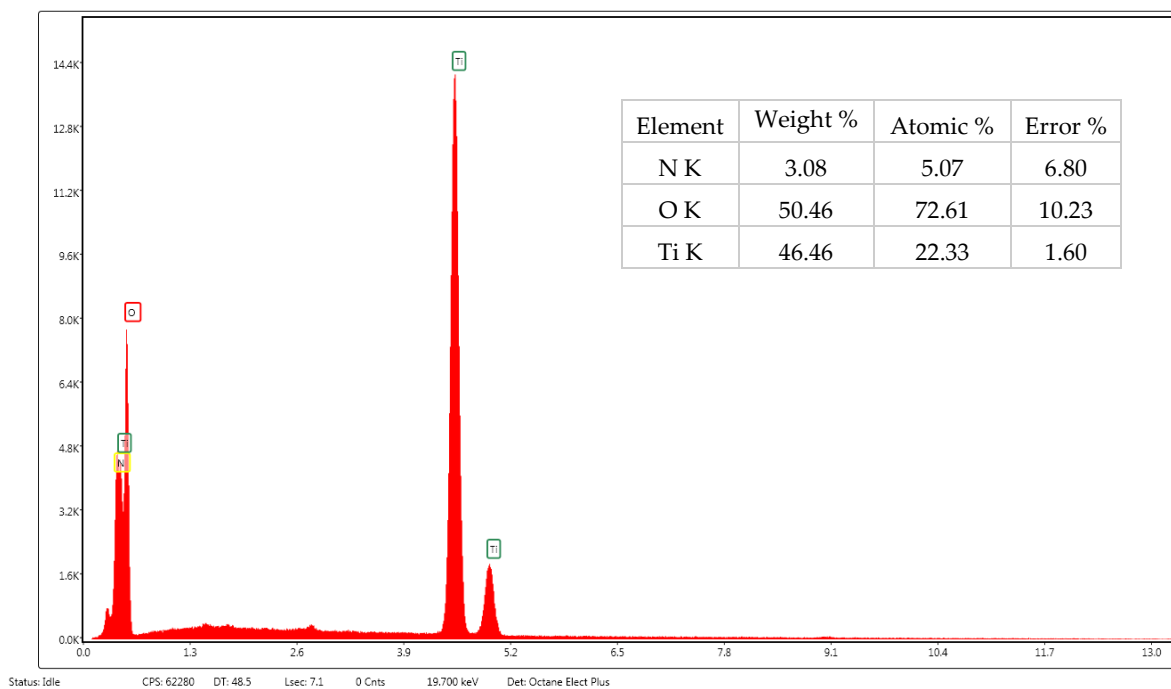
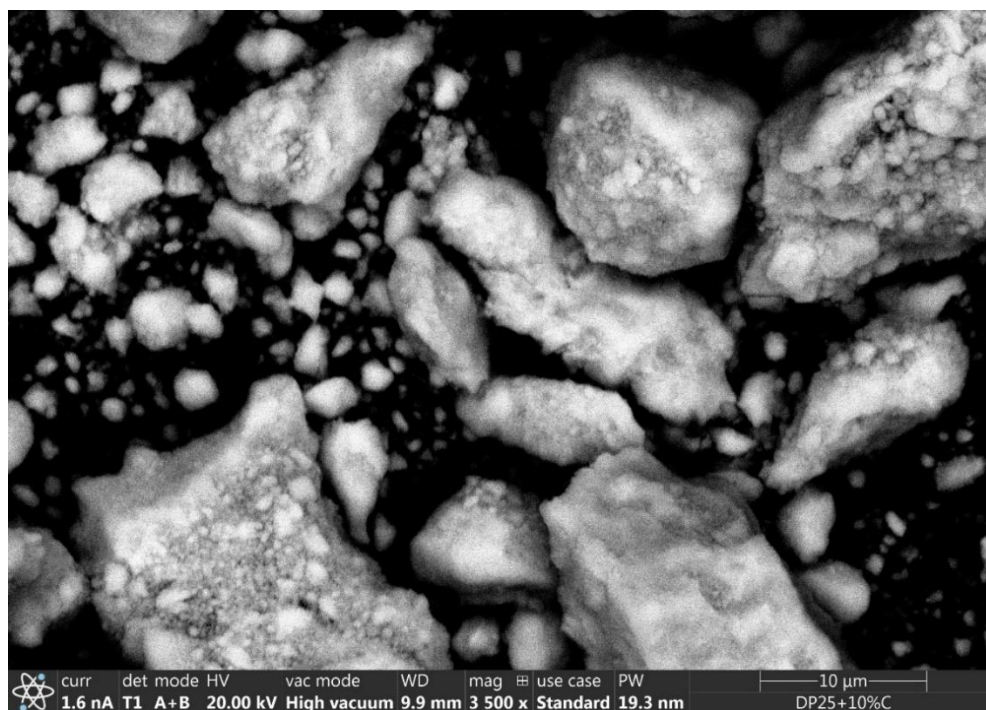


Figure S3. e) SEM morphology and elemental composition of N-TiO₂ catalyst.

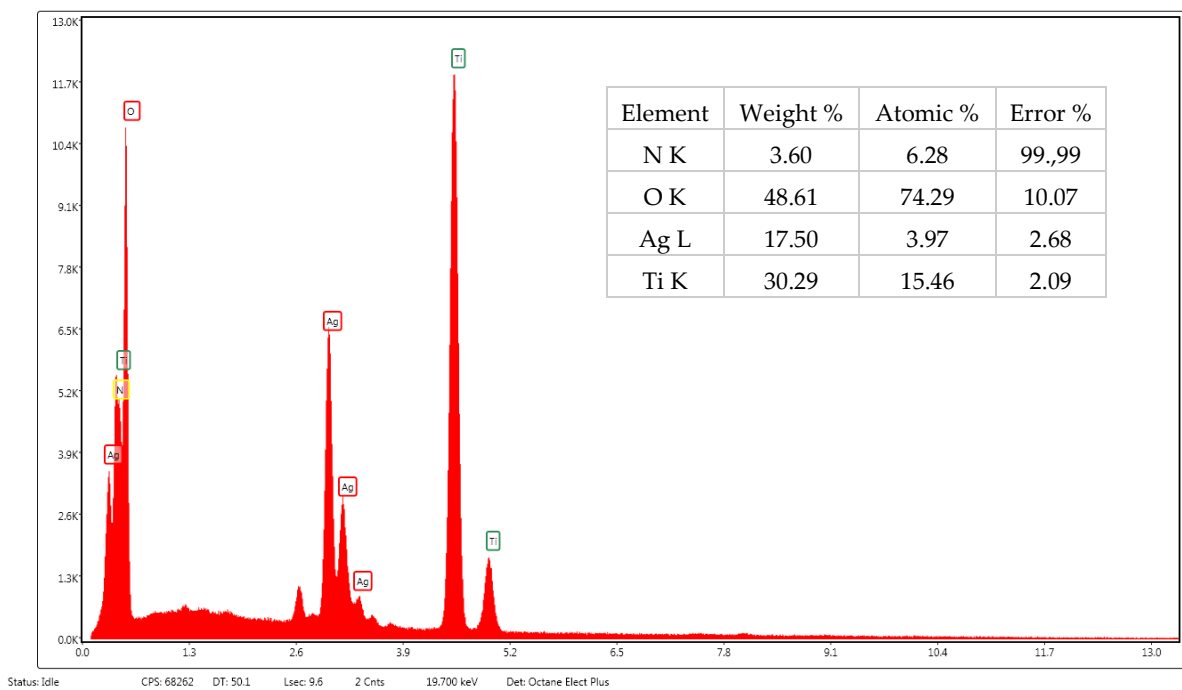
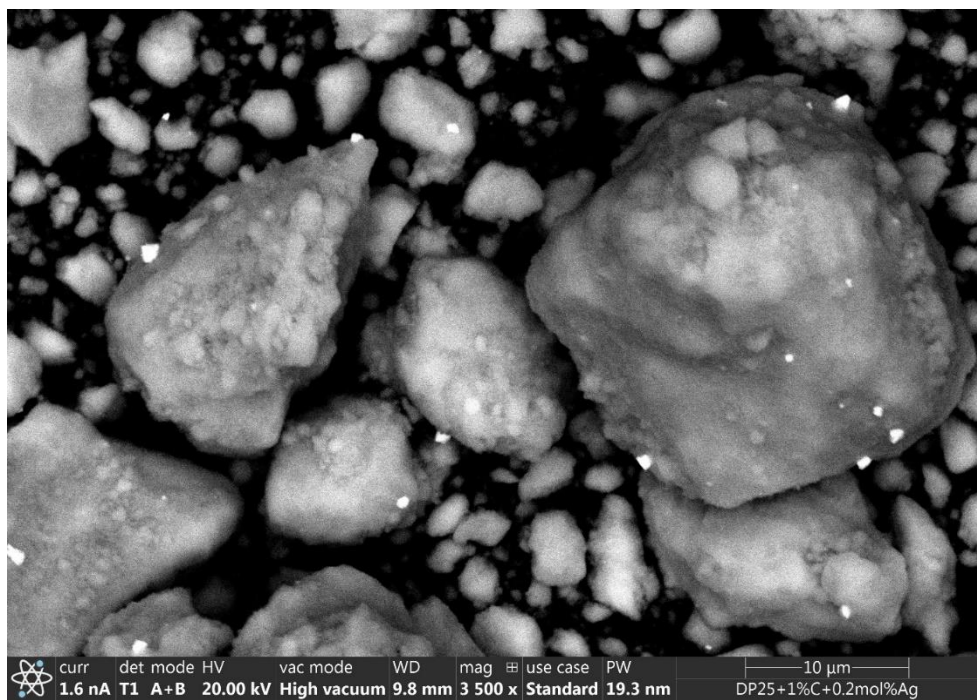


Figure S3. f) SEM morphology and elemental composition of Ag-N-TiO₂ catalyst.

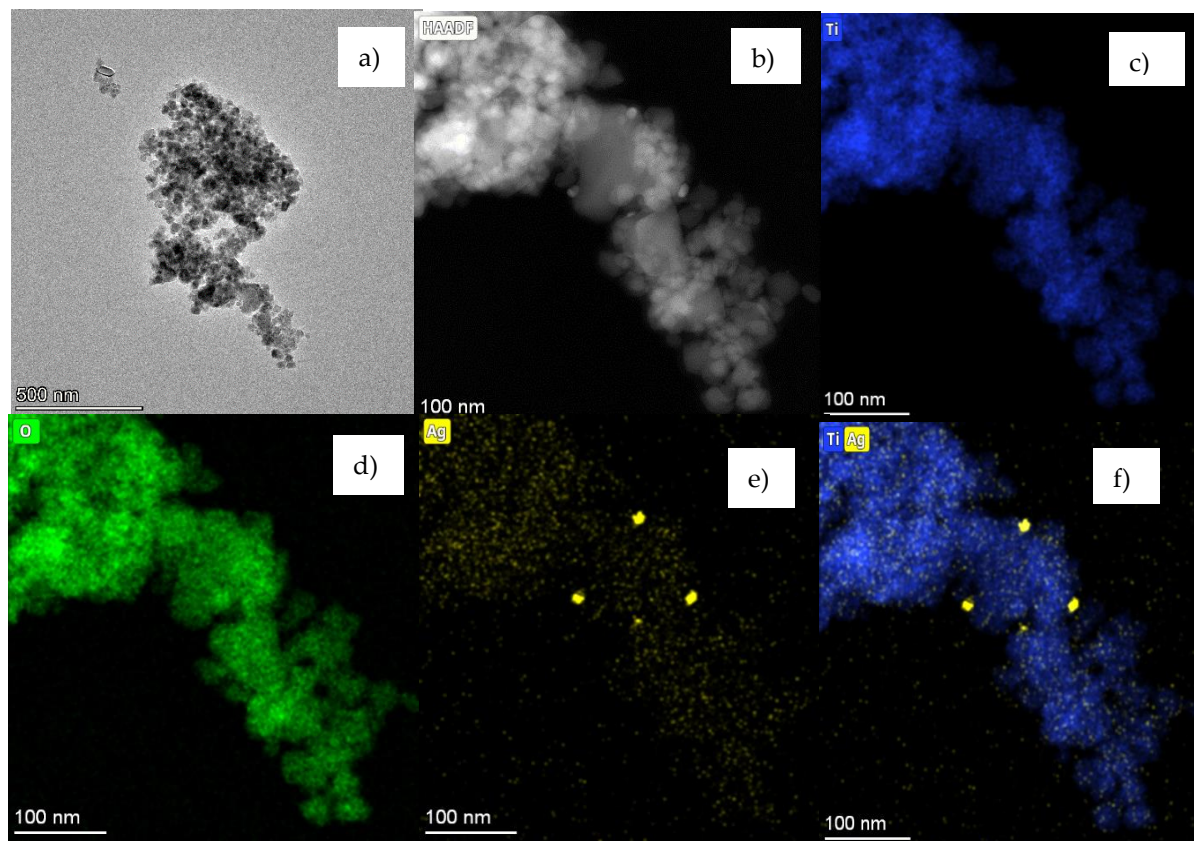


Figure S4. TEM analysis of the Ag-TiO₂ catalyst; (a, b) TEM micrographs with different magnifications; (c-f) element maps obtained in STEM-EDS mode.

Table S1. Band gaps of catalysts produced by mechanochemical treatment (grinding).

Catalyst	g-DP25 TiO ₂	N-TiO ₂	Ag-TiO ₂	N-Ag-TiO ₂
Band gap [eV]	3.029	3.007	3.028	3.017
Wavelength [nm]	409	412	409	411
Rutile content [%]	24.4	16.6	25.8	21.2

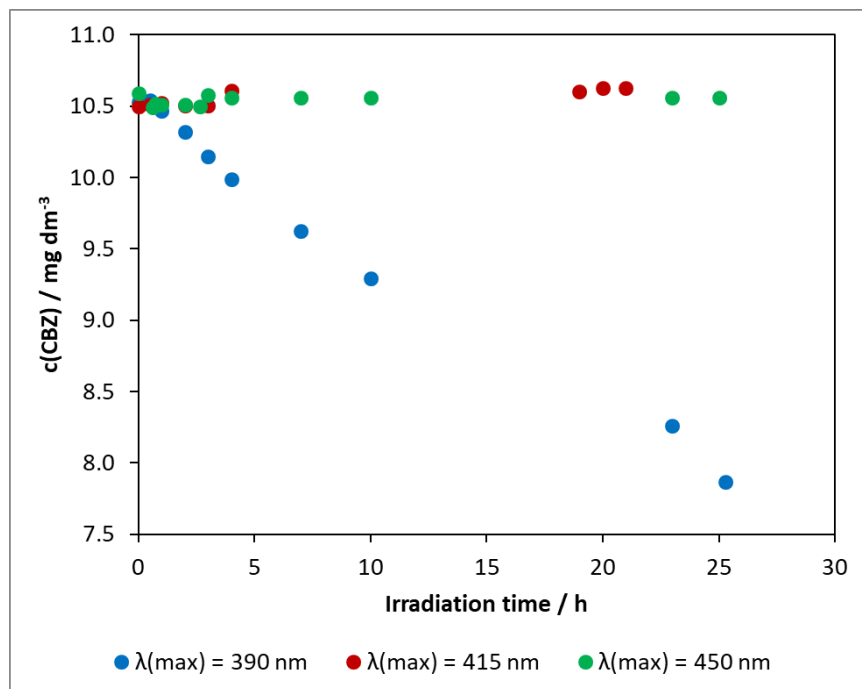


Figure S5. Changes in carbamazepine concentration during photolysis. $c(\text{CBZ})_0$: 10 mg dm^{-3} , airstream: $10 \text{ dm}^3 \text{ h}^{-1}$.

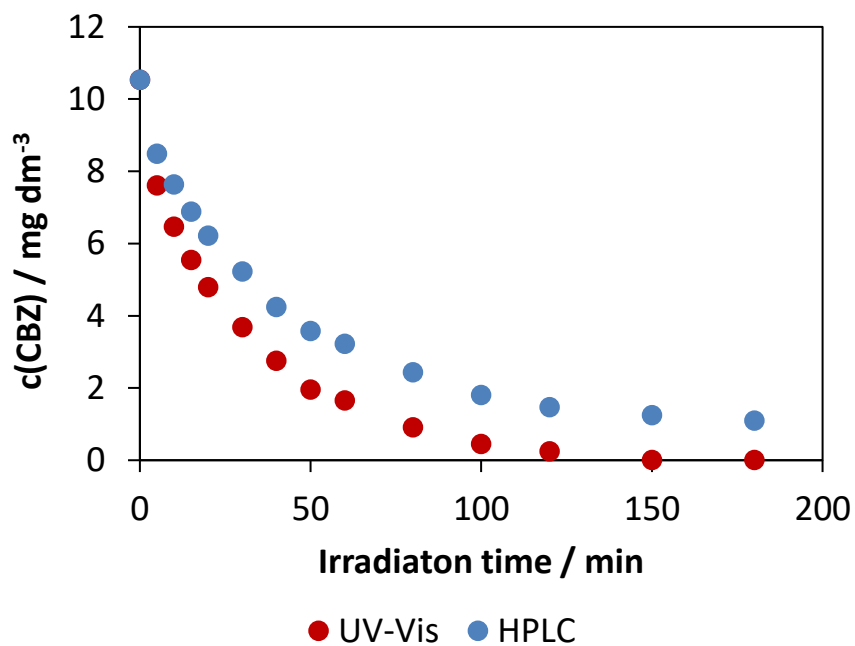


Figure S6. Changes in carbamazepine concentration, using DP25 TiO_2 photocatalyst. $c(\text{CBZ})_0$: 10 mg dm^{-3} , airstream: $10 \text{ dm}^3 \text{ h}^{-1}$, 1 g dm^{-3} DP25 TiO_2 , l : 1 cm , $\lambda(\text{det})$: 285 nm .

Table S2. Comparison of changes in COD measured during illuminations.

Catalyst	$\lambda(\text{max}) = 390 \text{ nm}$			$\lambda(\text{max}) = 415 \text{ nm}$			$\lambda(\text{max}) = 450 \text{ nm}$		
	$\text{COD}_{t=0\text{min}}$	$\text{COD}_{t=180\text{min}}$	ΔCOD	$\text{COD}_{t=0 \text{ min}}$	$\text{COD}_{t=180\text{min}}$	ΔCOD	$\text{COD}_{t=0\text{min}}$	$\text{COD}_{t=180\text{min}}$	ΔCOD
DP TiO ₂	24.67	0.80	23.87	25.17	17.50	7.67	24.78	20.23	4.55
g-DP TiO ₂	27.25	13.50	13.75	23.50	21.20	2.30	25.68	22.58	3.10
N-TiO ₂	25.11	21.50	3.61	25.56	22.00	3.56	24.12	22.87	1.25
Ag-TiO ₂	23.25	19.30	3.95	24.71	18.50	6.21	24.58	22.36	2.22
N-Ag-TiO ₂	24.22	19.89	4.33	24.44	22.67	1.77	25.24	22.54	2.70

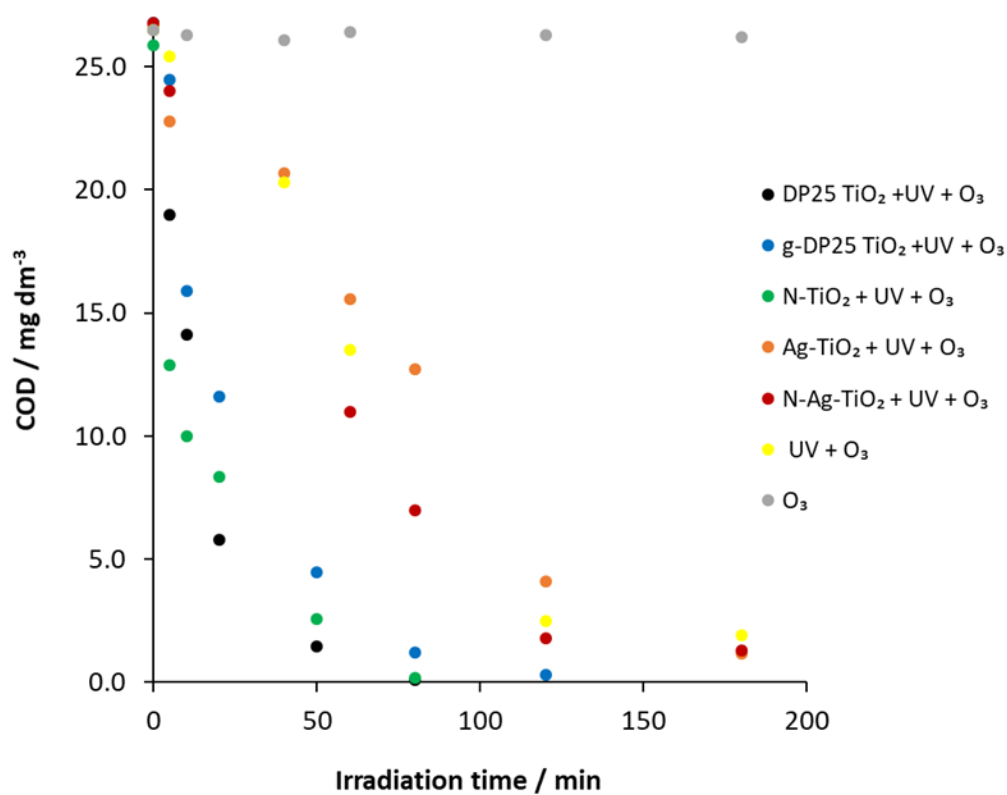


Figure S7. Changes in COD in carbamazepine solution during various treatments.

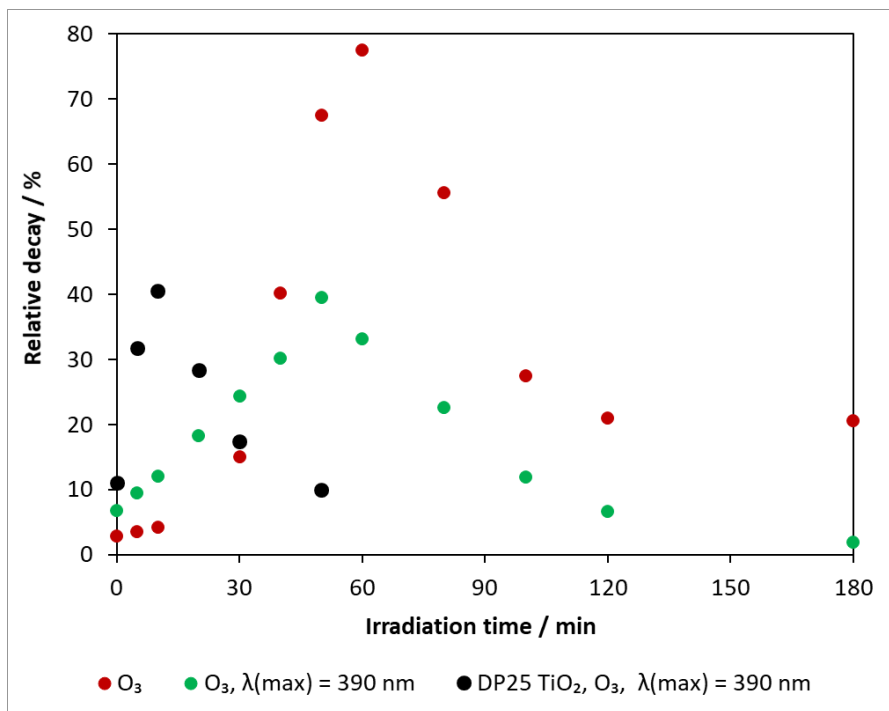


Figure S8. Changes in toxicity in carbamazepine solution, using ozone in various combinations.

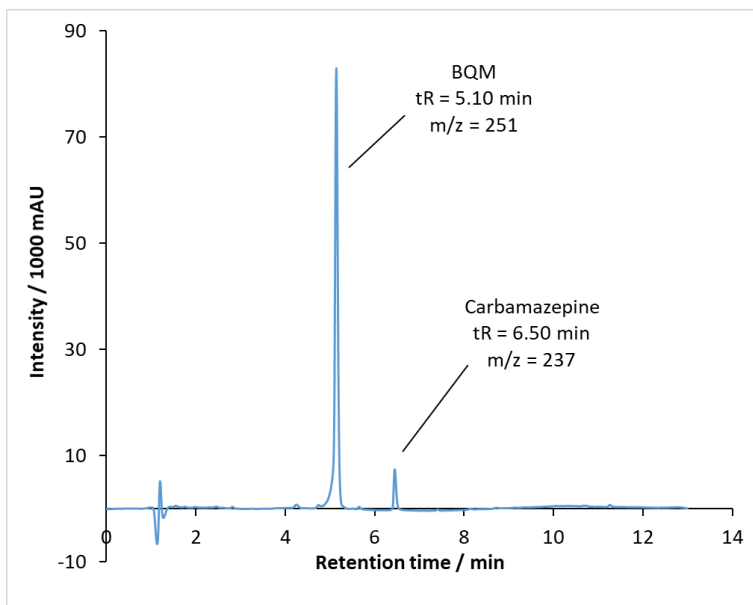


Figure S9. Chromatogram for the degradation of carbamazepine with ozone at 60 min reaction time.

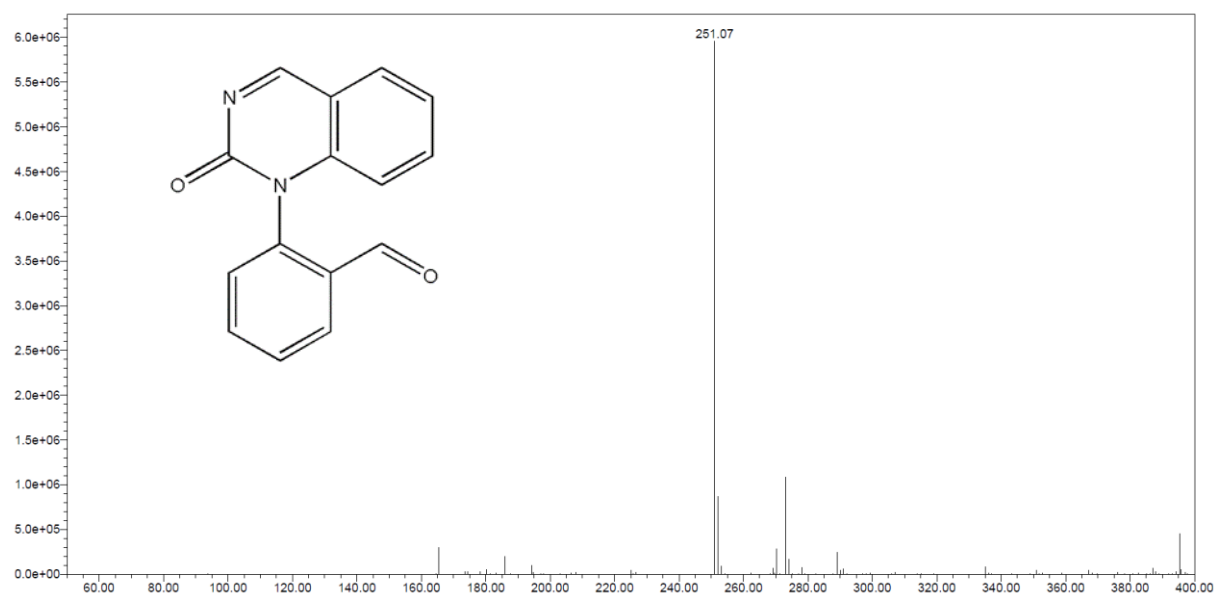


Figure S10. a) Mass spectrum of the compound with $t_R=5.1$ min in the sample of CBZ ozonized for 60 min, and the structural formula of BQM (251 m/z).

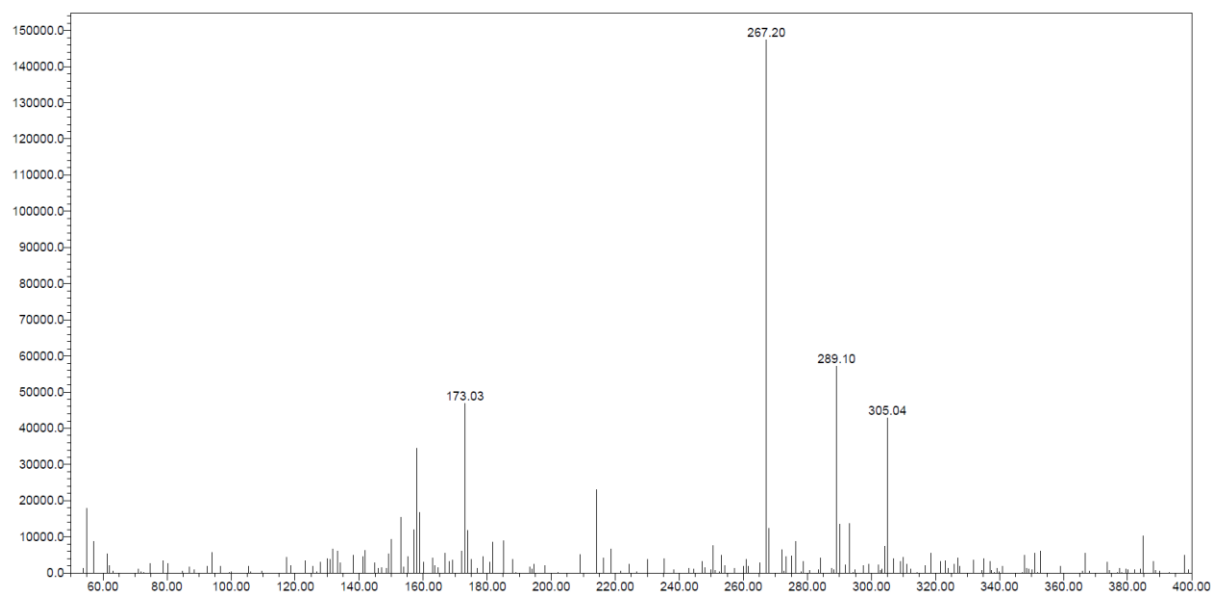


Figure S10. b) Mass spectrum of the compound (m/z 267) with $t_R=4.2$ min in the sample of CBZ ozonized and irradiated for 50 min.

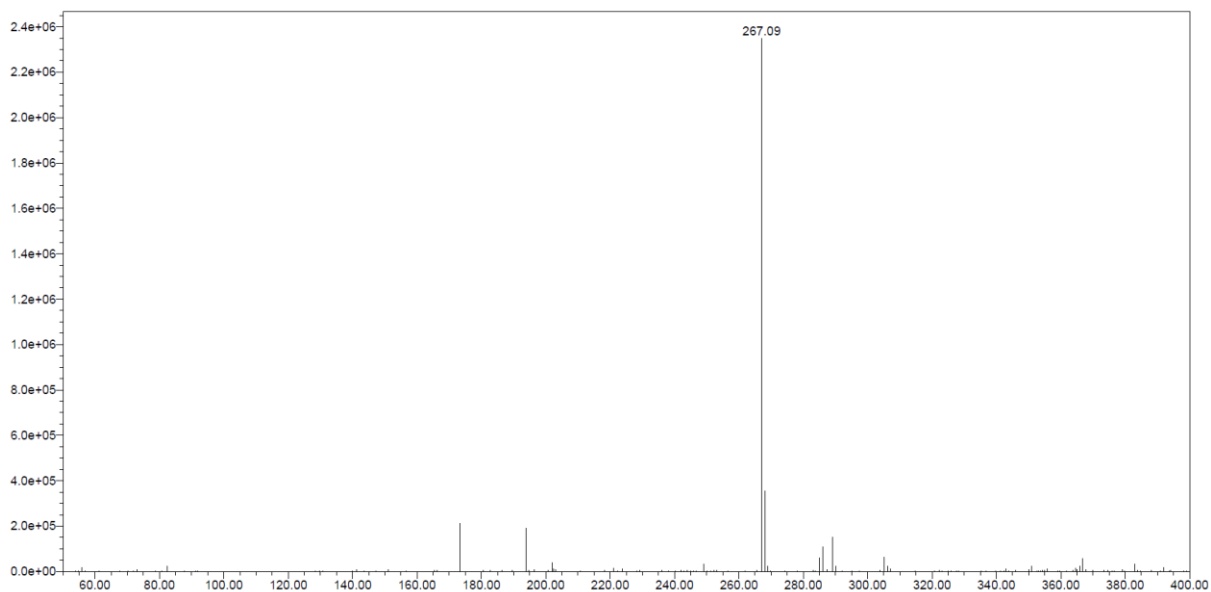
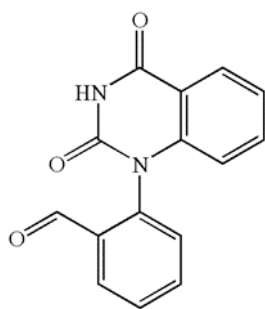
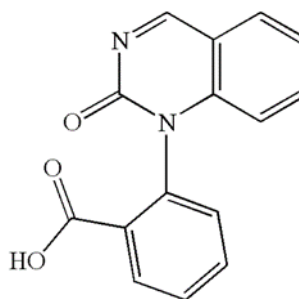


Figure S10. c) Mass spectrum of the compound (m/z 267) with tR=4.7 min in the sample of CBZ ozonized and irradiated for 50 min.



(a)



(b)

Figure S10. d) Structural formula of **(a)** BQD (m/z 267) and **(b)** BaQM (m/z 267).

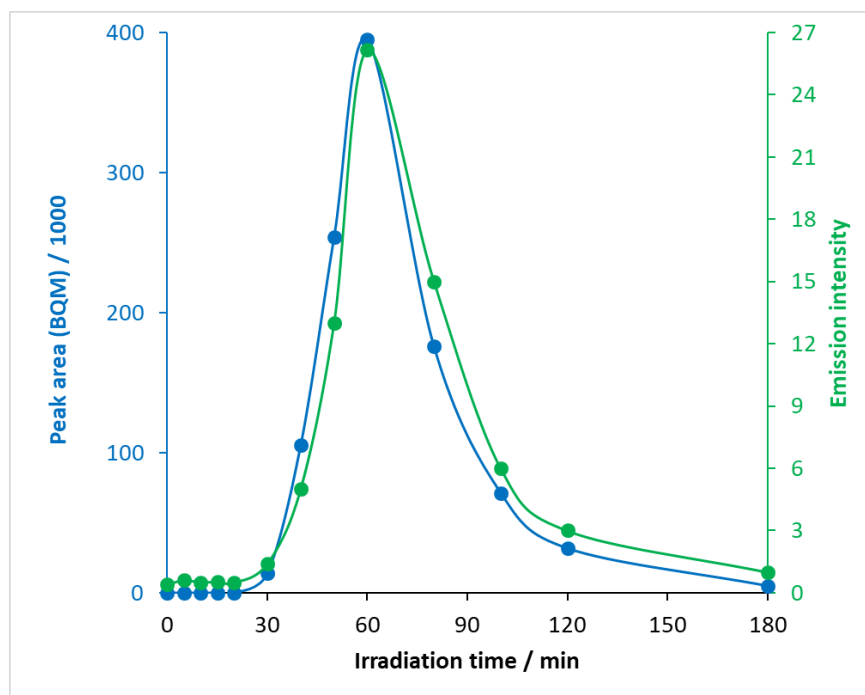


Figure S11. Peak areas and emission intensities of BQM during ozonization of carbamazepine.

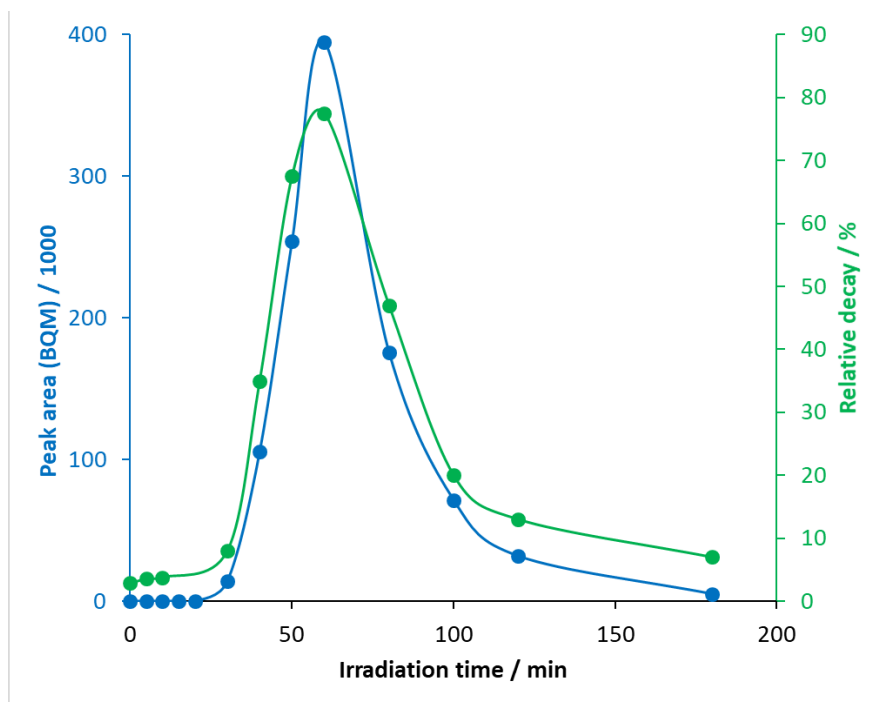


Figure S12. Peak areas of BQM and change in toxicity during ozonization of carbamazepine.

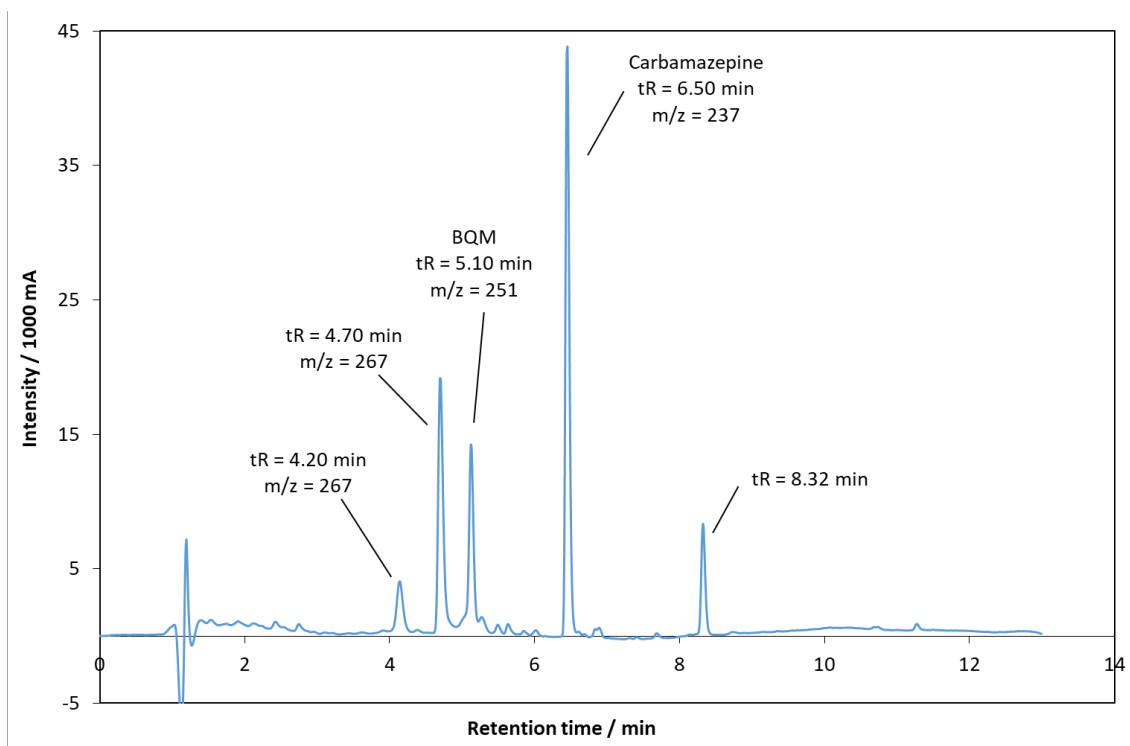


Figure S13. Chromatogram for the degradation of carbamazepine with ozone and UV light at 50 min reaction time.

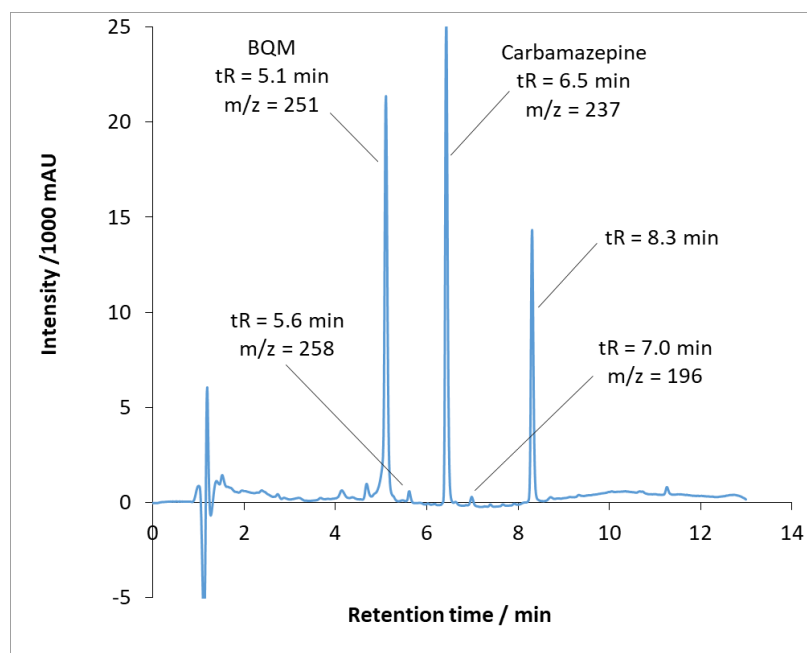


Figure S14. Fifteen min chromatogram for the degradation of carbamazepine, using N-TiO₂ + O₃ + UV.

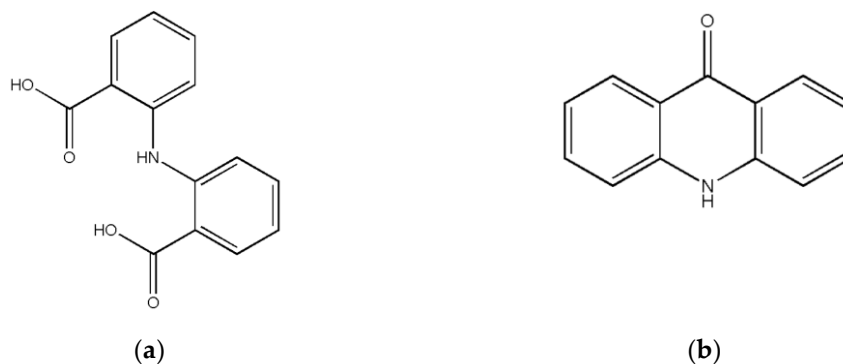


Figure S15. Structural formula of **(a)** 2,2'-imino-dibenzoic acid (m/z 258) and **(b)** acridone (m/z 196).

Table S3. Intermediates formed under irradiation at various wavelength in the presence of DP25 catalyst.

tR [min]	$\lambda(\text{max}) = 390 \text{ nm}$	$\lambda(\text{max}) = 415 \text{ nm}$	$\lambda(\text{max}) = 450 \text{ nm}$
4.70	✓	✗	✗
5.10	✓	✗	✓
5.45	✓	✓	✗
5.65	✓	✓	✓
6.00	✓	✓	✗
7.20	✓	✗	✗

✓ The intermediate product with the given retention time is formed.

✗ The intermediate with the given retention time is not formed.

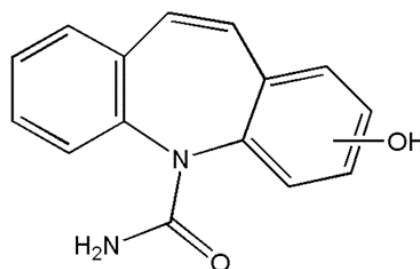


Figure S16. General structural formula of hydroxy-carbamazepine (m/z 253).

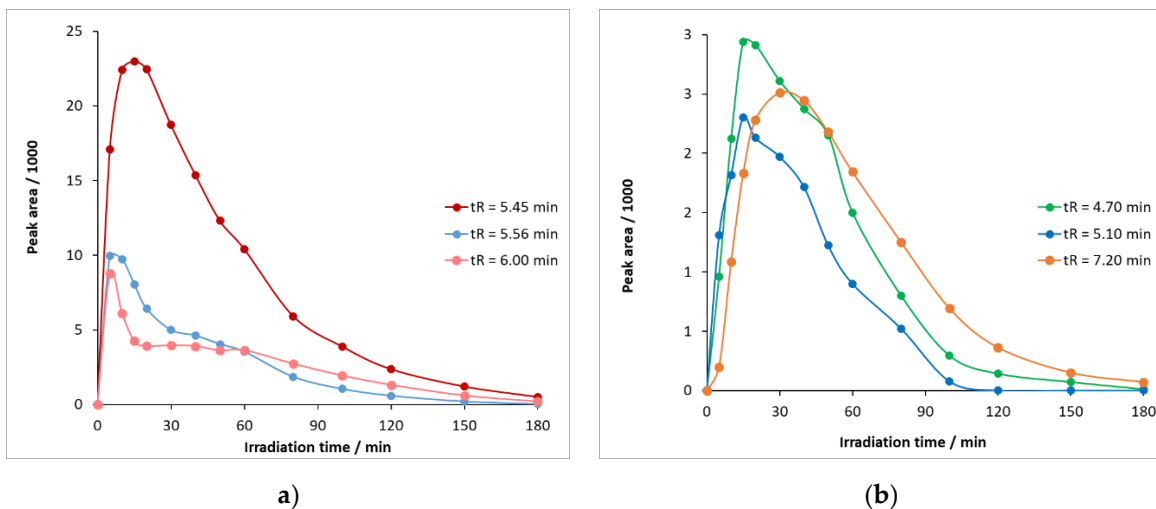


Figure S17. Temporal changes in the relative concentrations of intermediates of retention times a) 5.45, 5.56, and 6.00 min and b) 4.71, 5.10, and 7.20 min upon UV irradiation of carbamazepine solution on DP25 TiO₂ catalyst.

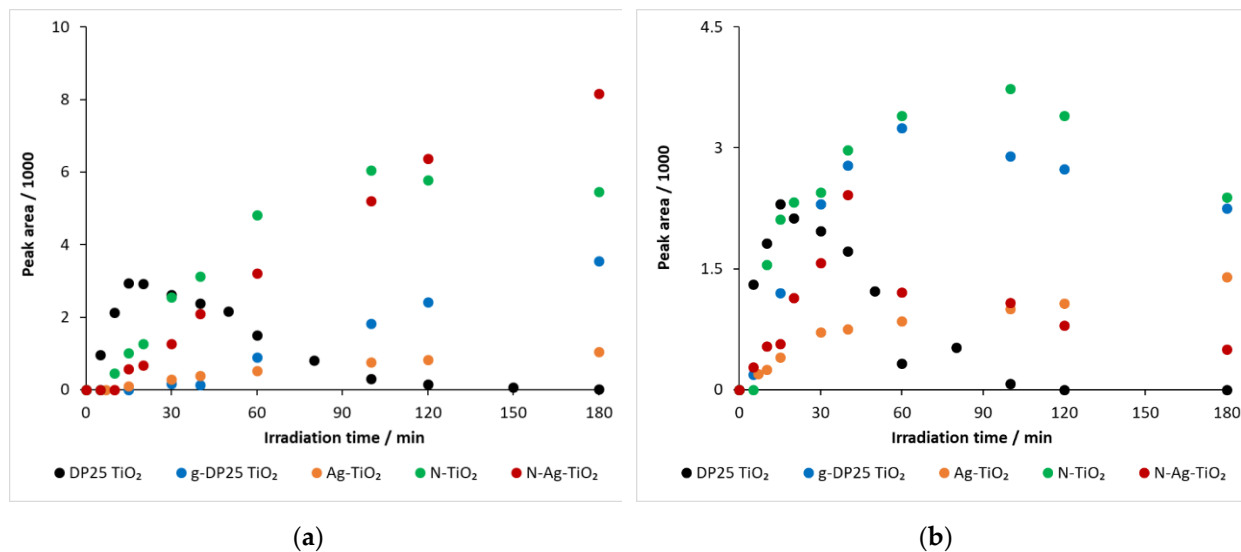


Figure S18. Temporal changes in the relative concentrations of intermediates of retention time a) 4.70 min and b) 5.10 min upon UV irradiation of carbamazepine solution on various catalysts.

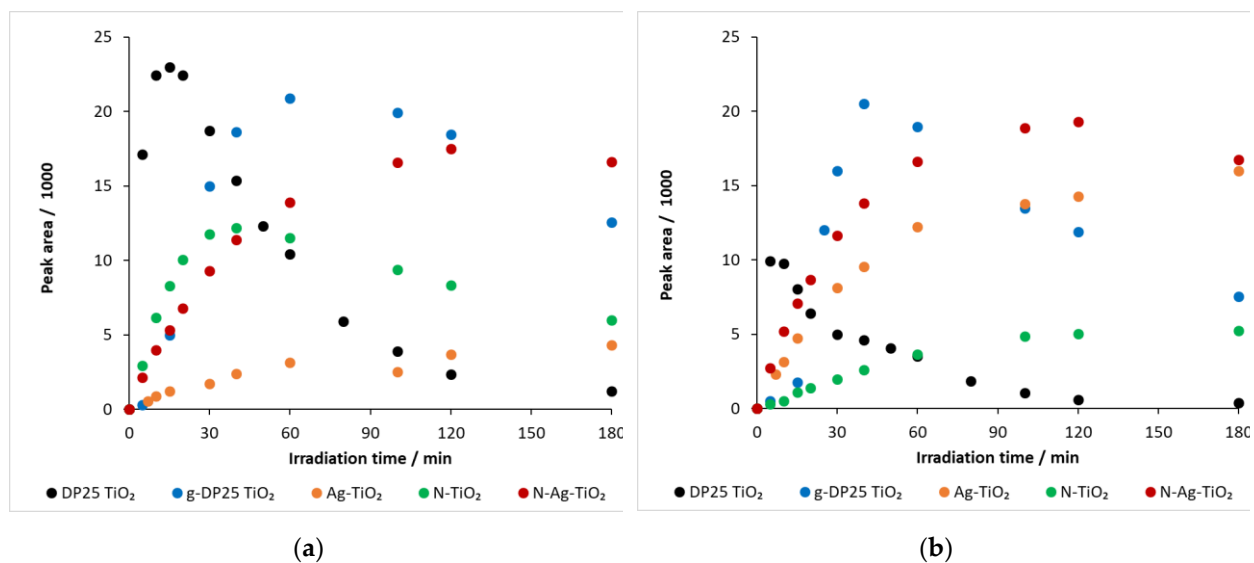


Figure S19. Temporal changes in the relative concentrations of intermediates of retention time a) 5.45 min and b) 5.75 min upon UV irradiation of carbamazepine solution on various catalysts.

Table S4. Intermediates formed under irradiation at $\lambda(\text{max}) = 450 \text{ nm}$ in the presence of various catalysts.

tR [min]	DP25 TiO ₂	g-DP25 TiO ₂	N-TiO ₂	Ag-TiO ₂	N-Ag-TiO ₂
4.57	✗	✓	✓	✗	✓
4.70	✗	✓	✓	✗	✓
5.10	✗	✓	✓	✗	✓
5.45	✓	✓	✓	✓	✓
5.65	✓	✗	✓	✓	✗

✓ The intermediate product with the given retention time is formed.

✗ The intermediate with the given retention time is not formed.

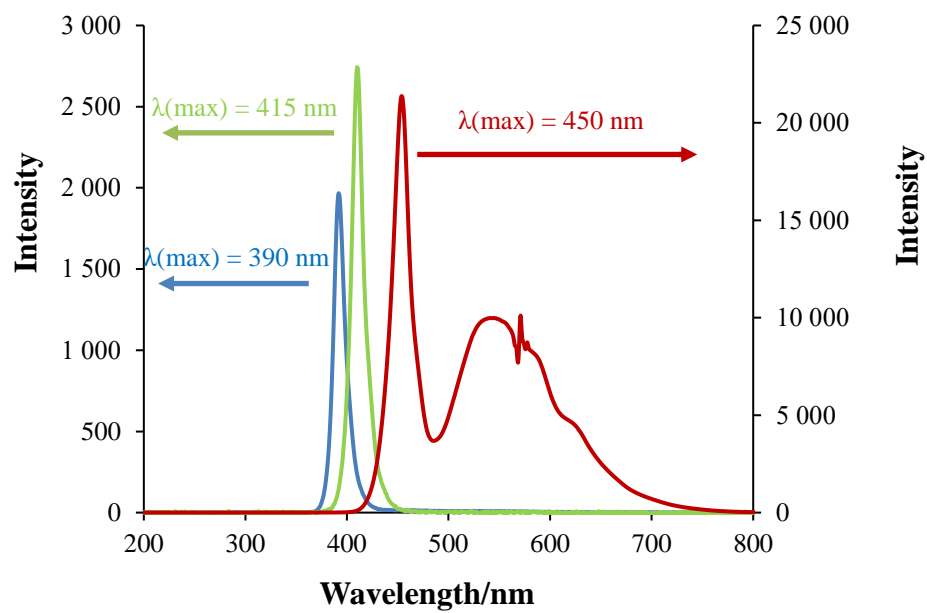


Figure S20. Emission spectra of the light sources applied.

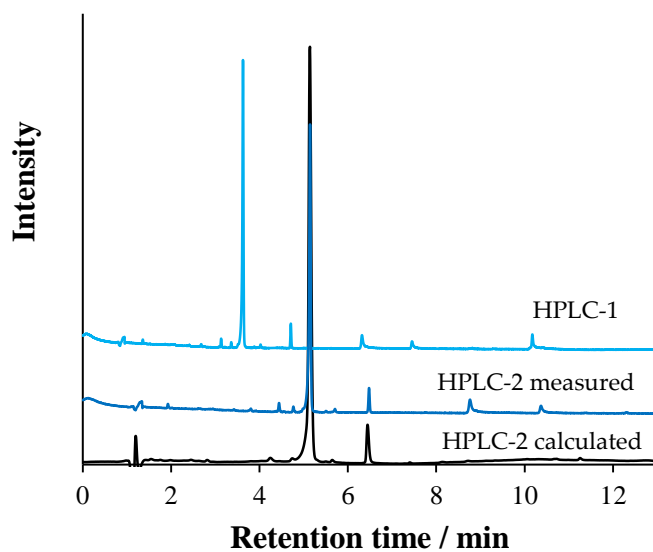


Figure S21. Relating the results of two HPLC systems to each other.

Text S1. Sample preparation for toxicity study

For measurements of the antibacterial effect, the catalyst samples were immobilized in an acrylate based polymer on a plastic surface. The catalyst concentration on the plastic surface was 485 mg/m². Discs of 1 cm diameter were cut from this plastic sheet and fixed to the bottom of small, open sample containers (glass tubes) with screw caps. The bacterial suspensions were layered on the samples. The toxicity effects of the catalysts were measured by using *Vibrio fischeri* luminescent bacteria. After reconstitution, the guaranteed life-span of bacteria was 4 h. The suspension of bacteria was incubated for 80 min at 15°C. At the end of the incubation period, 0.5 cm³ sample was measured into the sample holders, which were kept in the dark during the experiment, and the emission intensities of the samples were recorded at defined times. The reference sample was stored in a bottle, which did not contain any plastic disc. The control sample contained a disc from a commercially available plastic sheet with antibacterial surface. During the evaluation, the results obtained from 3 parallel measurements were averaged and then the relative decomposition percentage was calculated. Afterwards, the relative decomposition (%) of the actual sample was compared to the control one.