

## Supplementary Material

### Phenolic-Rich Extracts from Circular Economy: Chemical Profile and Activity against Filamentous Fungi and Dermatophytes

**Abstract.** Fungal infections represent a relevant issue in agri-food and biomedical fields because they could compromise quality of food and humans' health. Natural extracts represent a safe alternative to synthetic fungicides and in the green chemistry and circular economy scenario, agro-industrial wastes and by-products offer an eco-friendly source of bioactive natural compounds. In this paper, phenolic-rich extracts from *Olea europaea* L. de-oiled pomace, *Castanea sativa* Mill. wood, *Punica granatum* L. peel, *Vitis vinifera* L. pomace and seeds were characterized by HPLC-MS-DAD analysis. Finally, these extracts were tested as antimicrobial agents against pathogenic filamentous fungi and dermatophytes as *Aspergillus brasiliensis*, *Alternaria* sp., *Rhizopus stolonifer* and *Trichophyton interdigitale*. The experimental results evidenced that all extracts exhibited a significant growth inhibition for *Trichophyton interdigitale*. *Punica granatum* L., *Castanea sativa* Mill., and *Vitis vinifera* L. extracts showed a high activity against *Alternaria* sp. and *Rhizopus stolonifer*. These data resulted promising for the potentiality of applications of some of these extracts as antifungal agents for food and biomedical fields.

#### Figures and Tables

**Figure S1.** Chromatographic profile of OEP at 280 nm.

**Table S1.** Quali-quantitative analysis of OEP.

**Figure S2.** Chromatographic profile of CSW at 254 and 280 nm.

**Table S2.** Quali-quantitative analysis of CSW.

**Figure S3.** Chromatographic profile of PGP at 254 and 280 nm.

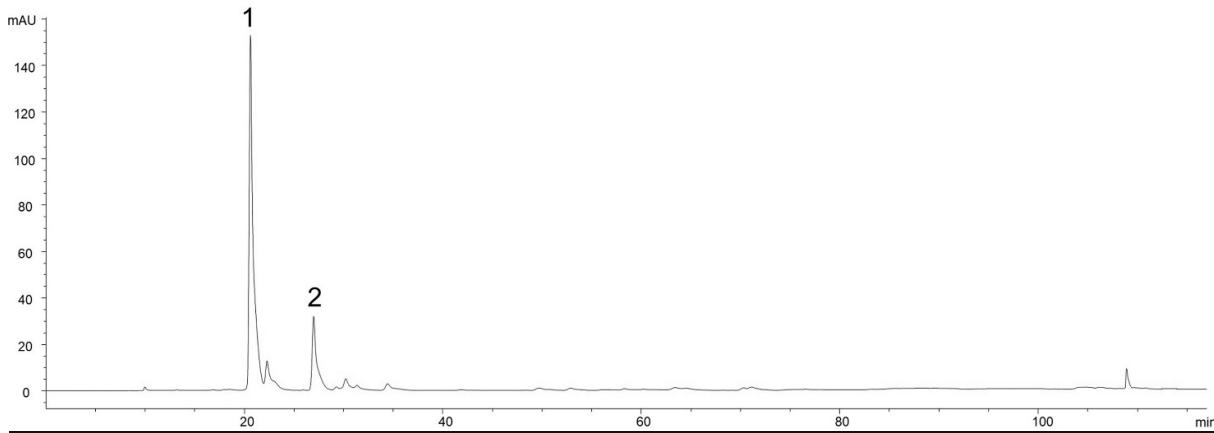
**Table S3.** Quali-quantitative analysis of PGP.

**Figure S4.** Chromatographic profile of VVP at 520 and 280 nm.

**Table S4.** Quali-quantitative analysis of VVP.

**Figure S5.** Chromatographic profile of VVS acquired at 280 nm.

**Table S5.** Quali-quantitative analysis of VVS.

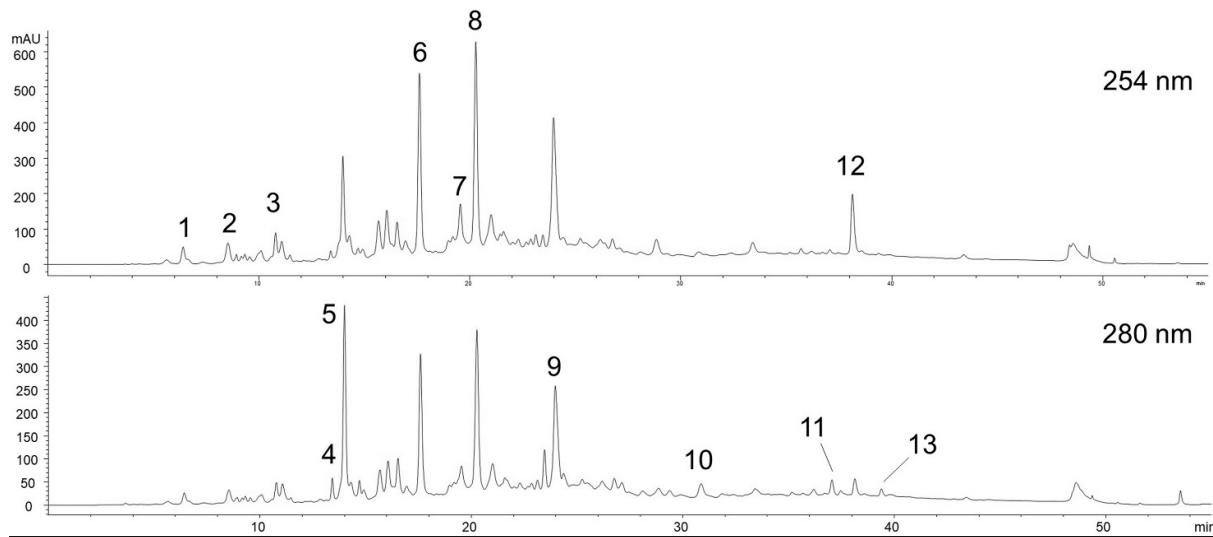


**Figure S1.** Chromatographic profile of OEP at 280 nm.

**Table S1.** Quali-quantitative analysis of OEP. The individual compounds are numbered as in **Figure S1**.

Identification	RT (min)	$\lambda_{\max}$ (nm)	[M-H] <sup>-</sup> (m/z)	mg/g
1. Hydroxytyrosol	20.6	280	153	138 ± 4.0
2. Tyrosol	27.0	276	137	35.0 ± 0.8
Total polyphenols				173 ± 5

Results are expressed as mg of each compound per g of extract. Retention times (RT), wavelengths of maximum UV absorbance ( $\lambda_{\max}$ ) and the m/z values for the ESI-MS molecular ions after negative ionization of each compound are reported.

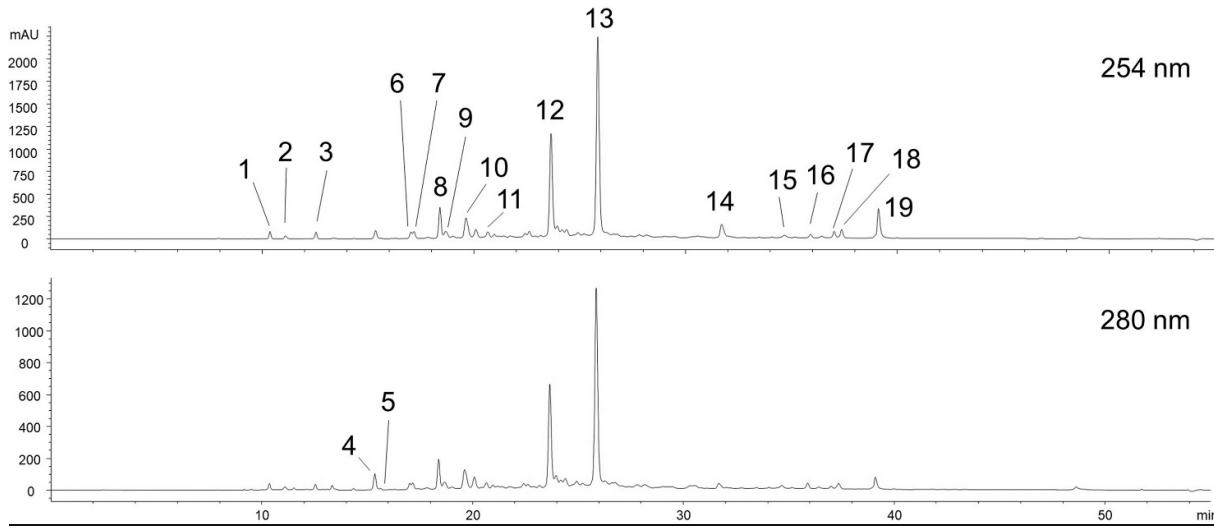


**Figure S2.** Chromatographic profile of CSW at 254 and 280 nm.

**Table S2.** Quali-quantitative analysis of CSW. The individual compounds are numbered as in **Figure S2**.

Identification	RT (min)	$\lambda_{\text{max}}$ (nm)	[M-H] <sup>-</sup> (m/z)	mg/g
1. Vescalin	6.9	246, 276sh	631	9.3 ± 0.2
2. Castalin	8.8	246, 280sh	631	8.1 ± 0.2
3. Pedunculagin I	11.5	258, 378sh	783	10.0 ± 0.2
4. Monogalloyl glucose	14.1	274	331	3.81 ± 0.08
5. Gallic acid	15.4	272	169	16.2 ± 0.3
6. Vescalagin	18.4	245, 280 sh	933	47.6 ± 0.5
7. Dehydrated tergallic-C-glucoside	20.8	250, 374	613	9.3 ± 0.2
8. Castalagin	21.9	248, 280 sh	933	97.7 ± 0.9
9. Digalloyl glucose	24.1	274	483	19.6 ± 0.2
10. Trigalloyl glucose	32.4	276	635	20.6 ± 0.2
11. Tetragalloyl glucose	38.0	276	787	7.7 ± 0.1
12. Ellagic acid	39.6	254, 370	301	6.1 ± 0.2
13. Pentagalloyl glucose	40.8	274	939	4.26 ± 0.08
Total polyphenols				260 ± 3

Results are expressed as mg of each compound per g of extract. Retention times (RT), wavelengths of maximum UV absorbance ( $\lambda_{\text{max}}$ ) and the m/z values for the ESI-MS molecular ions after negative ionization of each compound are reported.

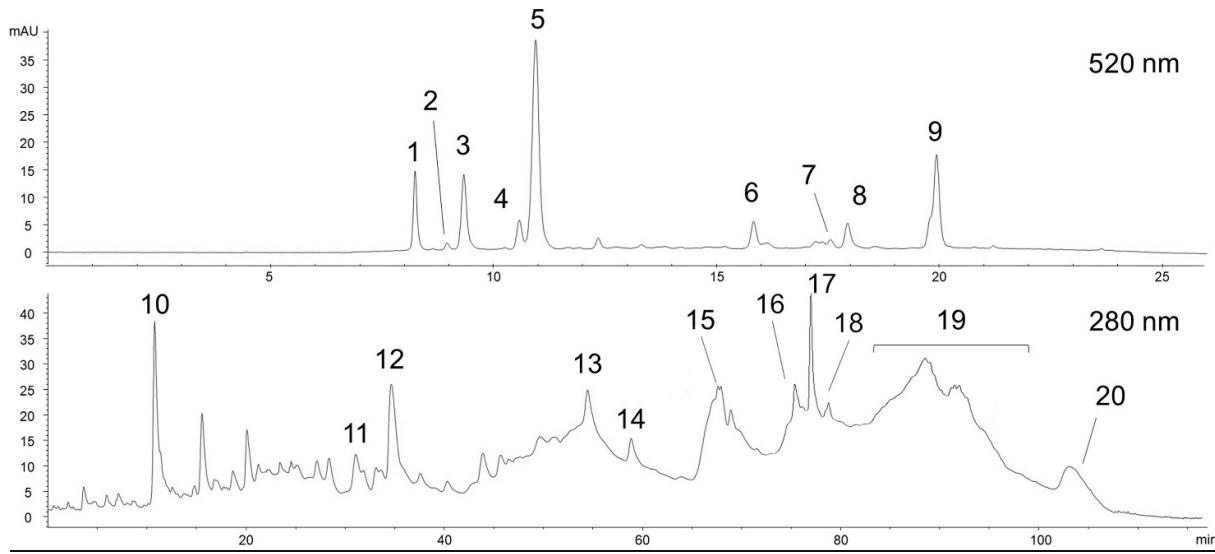


**Figure S3.** Chromatographic profile of PGP at 254 and 280 nm.

**Table S3.** Quali-quantitative analysis of PGP. The individual compounds are numbered as in **Figure S3**.

Identification	RT (min)	$\lambda_{\text{max}}$ (nm)	[M-H] <sup>-</sup> (m/z)	mg/g
1. HHDP glucose 1	10.4	slope	481	$0.75 \pm 0.01$
2. HHDP glucose 2	11.1	slope	481	$0.437 \pm 0.009$
3. HHDP glucose 3	12.5	slope	481	$0.71 \pm 0.01$
4. Gallic acid	15.4	272	169	$1.25 \pm 0.02$
5. Monogalloyl glucose	15.5	274	331	$0.106 \pm 0.005$
6. $\alpha$ -Punicalin	17.0	258, 378	781	$1.25 \pm 0.04$
7. $\beta$ -Punicalin	17.2	258, 380	781	$1.32 \pm 0.02$
8. Punicalagin isomer 1	18.4	258, 378	1083	$6.90 \pm 0.09$
9. Pedunculagin I	18.7	258, 378sh	783	$1.16 \pm 0.06$
10. Punicalagin isomer 2	19.6	258, 378	1083	$6.91 \pm 0.08$
11. Pedunculagin III	21.0	260, 378	933	$0.69 \pm 0.01$
12. $\alpha$ -Punicalagin	23.7	258, 378	1083	$27.1 \pm 0.3$
13. $\beta$ -Punicalagin	25.9	258, 380	1083	$58.5 \pm 0.6$
14. Ellagic acid hexoside	31.7	254, 362	463	$2.10 \pm 0.08$
15. Vanoleic acid bilactone	34.7	258, 366	469	$0.45 \pm 0.01$
16. Ellagitannin <i>m/z</i> 951	35.9	264, 364	951	$1.02 \pm 0.02$
17. Ellagic acid rhamnoside	37.0	254, 360	447	$0.61 \pm 0.03$
18. Ellagic acid pentoside	37.4	254, 362	433	$0.87 \pm 0.04$
19. Ellagic acid	39.1	254, 368	301	$2.60 \pm 0.08$
Total polyphenols				$115 \pm 2$

Results are expressed as mg of each compound per g of extract. Retention times (RT), wavelengths of maximum UV absorbance ( $\lambda_{\text{max}}$ ) and the m/z values for the ESI-MS molecular ions after negative ionization of each compound are reported.

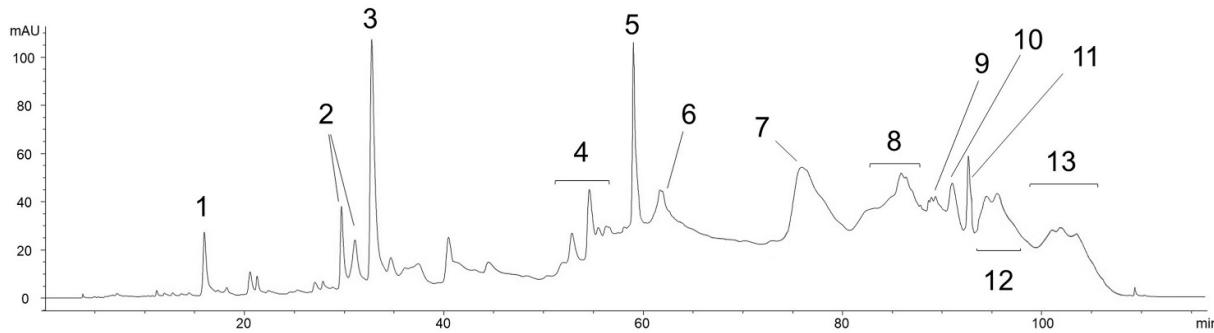


**Figure S4.** Chromatographic profile of VVP acquired at 520 and 280 nm.

**Table S4.** Quali-quantitative analysis of VVP. The individual compounds are numbered as in **Figure S4**.

Identification	RT (min)	$\lambda_{\max}$ (nm)	[M+H] <sup>+</sup> (m/z)	mg/g
1. Delphinidin-3-glucoside	8.2	522	465	0.262 ± 0.007
2. Cyanidin-3-glucoside	9.0	514	449	0.0097 ± 0.0003
3. Petunidin-3-glucoside	9.3	524	479	0.365 ± 0.008
4. Peonidin-3-glucoside	10.6	518	163	0.089 ± 0.003
5. Malvidin-3-glucoside	11.0	526	493	1.30 ± 0.02
6. Delphinidin-3-coumaroyl glucoside	15.8	530	611	0.130 ± 0.004
7. Cyanidin-3-acetyl glucoside	17.6	524	491	0.0100 ± 0.0005
8. Petunidin-3-coumaroyl glucoside	18.0	532	625	0.173 ± 0.005
9. Malvidin-3-coumaroyl glucoside	20.0	532	639	0.80 ± 0.02
10. Gallic acid	16.0	272	169 [M-H] <sup>-</sup>	2.37 ± 0.06
11. Procyanidin dimer B3	30.6	280	579	7.0 ± 0.2
12. Catechin	33.9	280	291	0.414 ± 0.008
13. Procyanidin trimers	57.4	280	867	2.01 ± 0.05
14. Procyanidin dimer B6	59.0	280	579	2.85 ± 0.08
15. Procyanidin dimer B2	64.0	280	579	10.2 ± 0.3
16. Epicatechin	76.5	280	291	0.320 ± 0.008
17. Procyanidin trimer	77.0	280	867	50 ± 2
18. Epicatechin gallate dimers	79.0	280	883	0.85 ± 0.02
19. Procyanidin tetramers	90.9	280	1155	293 ± 4
20. Epicatechin gallate dimers	104.4	280	883	53 ± 1
Total polyphenols				425 ± 8

Results are expressed as mg of each compound per g of extract. Retention times (RT), wavelengths of maximum UV absorbance ( $\lambda_{\max}$ ) and the m/z values for the ESI-MS molecular ions after positive or negative ionization of each compound are reported.



**Figure S5.** Chromatographic profile of VVS acquired at 280 nm.

**Table S5.** Quali-quantitative analysis of VVS. The individual compounds are numbered as in **Figure S5**.

Identification	RT (min)	$\lambda_{\max}$ (nm)	[M+H] <sup>+</sup> (m/z)	mg/g
1. Gallic acid	16.0	272	169 [M-H] <sup>-</sup>	1.50 ± 0.02
2. Procyanidin dimer B3	30.6	280	579	26 ± 1
3. Catechin	33.9	280	291	45 ± 1
4. Procyanidin trimer	57.4	280	867	8.8 ± 0.2
5. Procyanidin dimer B6	59.0	280	579	11.2 ± 0.3
6. Procyanidin dimer B2	64.0	280	579	13.6 ± 0.3
7. Epicatechin	76.5	280	291	30.3 ± 0.8
8. Procyanidin dimers gallate	88.3	280	731	20.1 ± 0.5
9. Procyanidin trimers digallate	89.7	280	1171	315 ± 9
10. Procyanidin tetramers (I)	90.0	280	1155	54.7 ± 0.16
11. Epicatechin gallate	92.2	280	443	6.24 ± 0.08
12. Procyanidin tetramers (II)	95.0	280	1155	11.6 ± 0.5
13. Procyanidin dimers digallate	98.5	280	883	142 ± 5
Total polyphenols				686 ± 20

Results are expressed as mg of each compound per g of extract. Retention times (RT), wavelengths of maximum UV absorbance ( $\lambda_{\max}$ ) and the m/z values for the ESI-MS molecular ions after positive or negative ionization of each compound are reported.