Two New Labdane Diterpene Glycoside from Flowers of *Bacchris Medulosa* DC

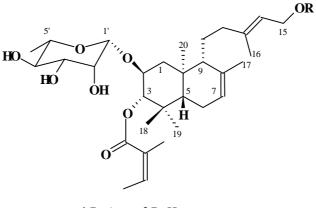
D.A. Cifuente, C.E. Tonn and O.S. Giordano

INTEQUI-CONICET-Facultad de Química, Bioquímica y Farmacia. Chacabuco y Pedernera-5700-San Luis, Argentina E-mail: cifuente@unsl.edu.ar

Abstract: Two new labdane-type diterpene glycoside, were isolated from the flowers of *Baccharis medulosa* DC (Asteraceae). Structures of these compounds were established by application of various spectroscopic techniques.

Introduction

In continuation of our studies on diterpenic compounds of *Baccharis* [1] genus (Compositae, tribe Astereae), we have investigated *B. medulosa* DC. In the present work, we described the isolation, characterization and structural determination of two new labdane-type diterpene glycoside [2] (1 and 2).



1 R=Ac **2** R=H

Experimental

Plant material. B. medulosa DC, was collected in Juana Koslay, San Luis, Province of Argentina in March 1998. Voucher N° 986. UNSL.

Extraction and isolation. Fresh flowers (2 Kg) of *B. medulosa* were extracted with Me₂CO at room temp. The Me₂CO extract was dissolved in MeOH: H₂O (8:2) and the soln was successively partitioned against, *n*-hexane, CCl₄, CHCl₃ and EtOAc. The CCl₄ and CHCl₃ extracts were subjected to several

several C.C. purifications on Si gel eluted with *n*-hexane, *n*-hexane:EtOAc increasing polarity mixtures and EtOAc -MeOH (97:3). The more polar fractions were purified by Sephadex LH-20 and RP-18 C.C., eluted with MeOH-H₂O (90:10 and 85:15) to yield 1 (300 mg) and 2 (250 mg). The sugar residues as TMS derivative were identified by GC analysis using suitable sugar standard after acid hydrolysis [3] of the natural products.

Results and Discussion

The NMR spectroscopical data for these compounds suggested nearly structural relationship according with a labdane-type glycoside framework. The ¹³C NMR spectrum of **1**, gave 33 carbon signals, which were coupled with DEPT experiments. Signals attributable to seven quaternary carbons, nine methyl carbons, five methylenes and twelve methine groups, were observed. The ¹H NMR spectral data showed the presence of three tertiary methyl groups at δ 0.85 s, 0.97 s and 0.82 s attributable each one to H-18, H-19, and H-20 on the decaline moiety. Two overlapping olefinic protons at δ 5.35 brt and δ 5.40 brs, both allylically coupled with methyl groups (δ 1.69 brs and 1.71 brs) were assigned to H-7 and H-14, respectively. From the COSY spectrum cross peaks observed between signals at δ 3.87 (ddd, J=12.0, 11.0, 3.8 Hz) and δ 4.72 (d, J=10.5Hz), were associated with H-2 and H-3, both on oxygenated carbons. Additional signals at δ 2.08 s and δ 4.59 (brd, J=7.3 Hz) indicated the presence of an acetate group on the allylic hydroxymethyl function at C-15. On the other hand, signals at $\delta_{\rm C}$ 103, $\delta_{\rm H}$ 4.19 (d, J=7.1 Hz) were agreeable with an anomeric proton; whose coupling constant indicated that the glycosidic linkage had β -configuration. One signal at δ_H 1.25 d (J=7.0 Hz) suggested that the sugar moiety was a methylpentose (L-rhamnose). Typical signals at $\delta_{\rm H}$ 6.15 qq, 1.99 dq, 1.92 dq were in agreement with the presence of an angelate group. The site of attachment of the saccharide residue as well as the position of the angelate group were established on the basis of long range HMBC experiments.

Except for the acetoxymethylene group signals, NMR spectral data of compound 2 were closely related with the spectral data observed for compound 1. In place of the signals at δ_H 4.59, one signal at δ_H 4.12 brd for an hydroxymethyl group, was observed.

Acknowledgements: Financial support of CONICET and UNSL. We thank Ing. L.A. del Vitto for plant identification, Professors E. Manta, P.C. Rossomando and E. Garcia for NMR measurements, Professor O. Varela for sugar standards.

References and Notes

- 1. Ceñal, J.P.; Giordano, O.S.; Rossomando, P.C.; Tonn, C.E. J. Nat. Prod. 1997, 60 (5), 490.
- 2. Zdero, C.; Bholmann, F.; King, R.M.; Robinson, H. Phytochemistry 1986, 25 (12), 2841.
- 3. Jahan, N.; Ahmed, W.; Malik, A. J. Nat. Prod. 1995, 58 (8), 1244.