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Article

# New Triterpenoids with Cytotoxic Activity from Actinidia Valvata

# Li-Ping Qu<sup>†</sup>, Guo-Yin Zheng<sup>†</sup>, Yong-Hua Su, Hui-Qing Zhang, Yan-Long Yang, Hai-Liang Xin \* and Chang-Quan Ling \*

Department of Traditional Chinese Medicine, Changhai Hospital of Second Military Medical University, Shanghai 200433, China; E-Mails: doudou0586@yahoo.com.cn (L.-P.Q.); herbzheng@163.com (G.-Y.Z.); suyh2001@126.com (Y.-H.S.); newdew628@yahoo.com.cn (H.-Q.Z.); yanlongyangzy@163.com (Y.-L.Y.)

- <sup>†</sup> These authors contributed equally to this work.
- \* Authors to whom correspondence should be addressed; E-Mails: hailiangxin@163.com (H.-L.X.); lingchangquan@gmail.com (C.-Q.L.); Tel.: +86-21-8187-1580 (H.-L.X.); +86-21-8187-1551 (C.-Q.L.); Fax: +86-21-8187-1559 (H.-L.X. & C.-Q.L.).

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**Abstract:** Two new triterpenoids,  $30-O-\beta$ -D-glucopyranosyloxy- $2\alpha$ , $3\alpha$ ,24-trihydroxyurs-12, 18-diene-28-oic acid  $O-\beta$ -D-glucopyranosyl ester (1) and  $2\alpha$ , $3\beta$ ,3,30-tetrahydroxyurs-12, 18-diene-28-oic acid  $O-\beta$ -D-glucopyranosyl ester (2) were isolated from roots of *Actinidia valvata* Dunn. Their structures were elucidated by means of extensive spectroscopic studies. Both these two new compounds showed moderate cytotoxic activity *in vitro* against BEL-7402 and SMMC-7721 tumor cell line.

Keywords: Actinidia valvata Dunn; triterpenoid; cytotoxic activity

#### 1. Introduction

*Actinidia valvata* Dunn is a shrub mainly growing in eastern China (Zhejiang and Jiangxi province) [1]. Its roots known as "mao ren shen" in traditional Chinese medicine exhibit anti-tumor and anti-inflammatory activity, and have been used for many years in the treatment of hepatoma, lung carcinoma and myeloma [2,3]. As we all know, hepatoma is very difficult to treat, and active components from medicinal herbs may be effective for research and development of new drugs [4].

In a previous study, we have carried out screening for cytotoxic activity of "mao ren shen", and two new polyoxygenated triterpenoids,  $2\beta$ , $3\alpha$ , $6\alpha$ , $20\alpha$ ,24,30-hexahydroxyurs-12-en-28-oic acid and  $2\beta$ , $3\alpha$ , $20\beta$ ,23,24,30-hexahydroxyurs-12-en-28-oic acid *O*- $\beta$ -D-glucopyranosyl ester were separated [5]. In this paper, two new triterpenoid saponins with cytotoxic activity against BEL-7402 and SMMC-7721 tumor cell line are reported.

#### 2. Results and Discussion

The roots of *Actinidia valvata* Dunn were extracted with 80% EtOH. The concentrated extract was suspended in H<sub>2</sub>O and successively extracted with petroleum ether ( $60^{\circ}-90^{\circ}$ ), AcOEt, and *n*-BuOH. The *n*-BuOH-soluble extract was repeatedly subjected to column chromatography to yield compound 1,2 Figure 1. Both these compounds were triterpenoid saponins with 12,18-diene-urs skeleton. Their structures were elucidated by detailed spectroscopic analysis.

Compound 1 was a white amorphous powder, displayed positive *Liebermann–Burchard* test, was optically active with  $[\alpha]_D^{25} = 5.89$  (c = 0.1, MeOH), and had the molecular formula  $C_{42}H_{66}O_{16}$ , with ten degrees of unsaturation, as determined according to a *pseudo*-molecular-ion peaks at 849.4240 ( $[M + Na]^+$ ; calc. 849.4249) in the HR-ESI-MS.

<sup>13</sup>C-NMR (DEPT) spectra of compound 1 revealed 42 carbon signals, including five CH<sub>3</sub>, two C=C bonds (tri-, four-substituted) and one C=O group. Assuming compound 1 has skeleton of urs-triterpenoid, the assignment of the two C=C bonds should be highly concerned. In <sup>1</sup>H-NMR spectra of compound 1, five singlets at  $\delta(H) 0.80-1.89$  consisting of five CH<sub>3</sub>, and proton signal at  $\delta(H) 5.53$  (br) was assigned to 12-position. In HMBC spectra, clear correlation of 19-CH<sub>3</sub> with two quaternary C-atoms at  $\delta(C)$  126.17 (s) and  $\delta(C)$  136.92 (s) was observed, and the correlation signals of 12-H with these two quaternary C-atoms were also observed. Thus, the two C=C bonds can be rightly assigned to 18-, 19-position, respectively. The HMBC correlation signal of 20-H with C-atom at  $\delta(C)$  69.68(t), was observed, then the oxygenation of 30-C may be deduced successfully. In NOSEY spectra, as the correlations of 2-H with 3-H, 2-H with 10-Me, 8-Me with 10-Me were observed, and by comparing with reference data [6,7], the configuration of  $2\alpha$ -OH,  $3\alpha$ -OH, and the 24-OH can be confirmed. <sup>1</sup>H-NMR and <sup>13</sup>C-NMR data of glycon moiety of compound **1** indicated it featured two glucopyranosyls. The *C*-atom at  $\delta(C)$  95.82(d) and H (5.39, d, J = 8.0 Hz), *C*-atom at  $\delta(C)$  101.85(d) and H (4.20, d, J = 8.0 Hz) were assigned as anomeric C-atoms and prontons, respectively. The linkage of two glucopyranosyls with aglycone maybe deduced by correlation of anomeric pronton with glycosidated C-atoms in HMBC spectra [H (5.39, d, J = 8.0 Hz) to C=O, H (4.20, d, J = 8.0 Hz) to C-atom at 69.68(t)].

Basing on above analysis, in combination with the <sup>1</sup>H-, <sup>13</sup>C-NMR spectra (Table 1), HMQC, HMBC, and NOSEY data (Figure 2), established the structure of compound 1 as  $30-O-\beta$ -D-glucopyranosyloxy- $2\alpha$ , $3\alpha$ ,24-trihydroxyurs-12,18-diene-28-oic acid *O*- $\beta$ -D-glucopyranosyl ester (1).

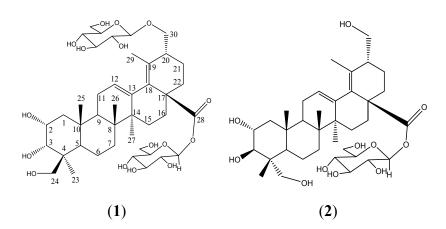
The compound **2** was obtained as white amorphous powder, displayed positive *Liebermann–Burchard* test, was optically active with  $[\alpha]_{D}^{25} = 7.35$  (c = 0.1, MeOH), and had the molecular formula  $C_{36}H_{56}O_{11}$  with nine degrees of unsaturation, as determined according to a *pseudo*-molecular-ion peaks at 687.3717 ( $[M + Na]^+$ ; calc. 687.3720) in the HR-ESI-MS.

Position	1		2	
	$\delta(C)^{a}$	δ(H) <sup>b</sup>	$\delta(C)^{a}$	$\delta(H)^{b}$
1	42.81 (t)	1.31 (dd, $J = 12,4.2$ ), 2.19 (dd, $J = 12,4.2$ )	48.03 ( <i>t</i> )	1.30 ( <i>m</i> ), 2.29 ( <i>m</i> )
2	66.98 (d)	3.88 ( <i>m</i> )	68.93 ( <i>d</i> )	4.25 ( <i>m</i> )
3	74.90 (d)	3.70 (d, J = 2.0)	78.18 (d)	4.19 ( <i>m</i> )
4	45.36 (s)		43.61 (s)	_
5	49.94 (d)	1.79 ( <i>m</i> )	48.31 ( <i>d</i> )	1.73 ( <i>m</i> )
6	19.15 (t)	1.33 ( <i>m</i> )	18.41 <i>(t)</i>	1.04 ( <i>m</i> ), 1.15 ( <i>m</i> )
7	33.23 (t)	1.28 ( <i>m</i> ), 1.30 ( <i>m</i> )	33.86 ( <i>t</i> )	1.31 ( <i>m</i> ), 1.75 ( <i>m</i> )
8	40.67 (s)		39.80 (s)	_
9	48.96 (d)	1.71 ( <i>m</i> )	48.03 ( <i>d</i> )	1.83 ( <i>m</i> )
10	39.07 (s)		38.31 (s)	_
11	24.25 (t)	1.74 ( <i>m</i> ), 1.93 ( <i>m</i> )	23.64 ( <i>t</i> )	1.74 ( <i>m</i> ), 2.01 ( <i>m</i> )
12	129.19 (d)	5.53 (br)	127.98 (d)	5.60 (br)
13	137.97 (s)		137.53 (s)	
14	44.69 (s)		43.90 (s)	_
15	29.09 (t)	1.09 ( <i>m</i> ), 1.88 ( <i>m</i> )	28.58 ( <i>t</i> )	1.06 ( <i>m</i> ), 2.25 ( <i>m</i> )
16	24.72 (t)	1.74 ( <i>m</i> ), 1.98 ( <i>m</i> )	32.97 ( <i>t</i> )	1.79 ( <i>m</i> ), 1.98 ( <i>m</i> )
17	48.83 (s)		47.25 (s)	_
18	126.17 (s)		129.59 (s)	
19	136.92 (s)		131.27 (s)	
20	51.45 (s)	3.30 ( <i>br</i> )	50.74 ( <i>d</i> )	3.55 (br)
21	24.67 (t)	1.97 ( <i>m</i> )	23.87 ( <i>t</i> )	1.99 ( <i>m</i> )
22	35.19 (t)	1.28 ( <i>m</i> ), 1.53 ( <i>m</i> )	24.41 ( <i>t</i> )	2.06 ( <i>m</i> )
23	22.56 (q)	1.07 (s)	66.52 ( <i>t</i> )	3.66 (d, J = 10.2), 4.16 (d, J = 10.2)
24	65.72 (t)	3.36 (d, J = 8.0), 3.68 (d, J = 8.0)	14.42(q)	1.03 (s)
25	17.86 (q)	0.94 (s)	17.95(q)	1.08 (s)
26	18.21 (q)	0.85 (s)	18.27(q)	1.15 (s)
27	23.17 (q)	1.89 (s)	22.23(q)	0.97(s)
28	178.07 (s)		176.24 (s)	
29	17.44 (q)	1.68 (s)	16.83 (q)	1.75(s)
30	69.68 (t)	4.09 (d, J = 20.0), 4.45 (d, J = 20.0)	62.20 ( <i>t</i> )	4.38 (d, J = 10.2)
28-glc-1	95.82 (d)	5.39 (d, 8)	95.82 ( <i>d</i> )	5.60 (d, J = 8.0)
2	73.75 (d)	3.31 ( <i>m</i> )	74.19 ( <i>d</i> )	3.31 ( <i>m</i> )
3	77.91 (d)	3.33 ( <i>m</i> )	78.84 ( <i>d</i> )	3.33 ( <i>m</i> )
4	71.46 (d)	3.33 ( <i>m</i> )	71.12 ( <i>d</i> )	3.33 ( <i>m</i> )
5	78.54 (d)	3.38 ( <i>m</i> )	79.25 (d)	3.38 ( <i>m</i> )
6	62.49 (t)	3.64 (dd, $J = 12.0, 1.8$ ), 3.77 (dd, $J = 12.0, 1.8$ )	62.99 ( <i>t</i> )	3.65 (dd, <i>J</i> = 12.0, 1.8) 3.78 (dd, <i>J</i> = 12.0, 1.8)
30-glc-1	101.85 (d)	4.20 (d, J = 8.0)		
2	74.53 (d)	3.31 ( <i>m</i> )		
3	77.83 (d)	3.33 ( <i>m</i> )		
4	70.97 (d)	3.33 (m)		
5	77.95 (d)	3.38 ( <i>m</i> )		
		3.63 (dd, J = 12.0, 1.8),		
6	62.30 (t)	$3.76 (\mathrm{dd}, J = 12.0, 1.8)$		

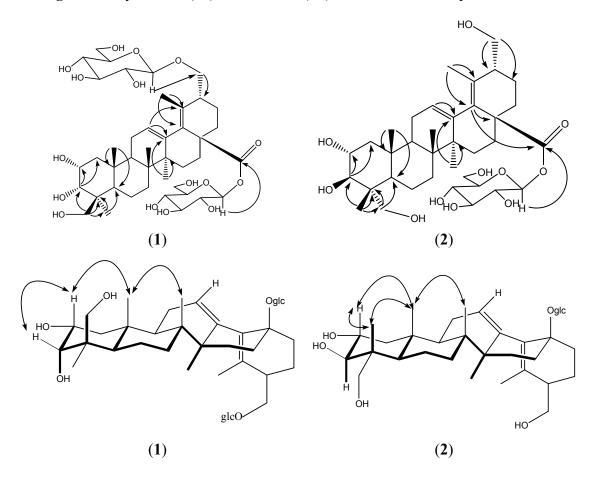
**Table 1.** <sup>1</sup>H and <sup>13</sup>C-NMR Data of 1 and 2 (in  $C_5D_5N$ ).  $\delta$  in ppm, J in Hz.

<sup>a</sup> Recorded at 150 MHz, multiplicity by DEPT; <sup>b</sup> Recorded at 600 MHz.

Figure 1. Structures of compound 1 and 2.



**Figure 2.** Key HMBC  $(\rightarrow)$  and NOESY  $(\leftrightarrow)$  correlations of compound 1 and 2.



Comparing the <sup>1</sup>H-NMR and <sup>13</sup>C-NMR (Table 1) spectra, compound **2** has only a 28-glucopyranosyl. The configuration of  $2\alpha$ -OH,  $3\beta$ -OH, and 23-OH can be confirmed by analyzing the correlations of NOSEY spectra and comparing with reference data [6,7]. In combination with the <sup>1</sup>H-, <sup>13</sup>C-NMR spectra (Table 1), HMQC, HMBC, and NOSEY data (Figure 2), established the structure of compound **2** as  $2\alpha$ , $3\beta$ ,23,30-tetrahydroxyurs-12, 18-diene-28-oic acid *O*- $\beta$ -D-glucopyranosyl ester (**2**).

Compounds 1 and 2 showed moderate *in vitro* cytotoxic activity against BEL-7402 (IC<sub>50</sub> value of 92.2 and 89.7  $\mu$ g/mL, resp.) and SMMC-7721 (IC<sub>50</sub> value of 58.1 and 89.7  $\mu$ g/mL, resp.), as determined by classical MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2*H*-tetrazolium bromide) colorimetric assay.

#### 3. Experimental Section

#### 3.1. General

Silica-gel plates (Sinopharm Chemical Reagent Co., Ltd.) were used for TLC analysis. mp: WRS-1A micro-melting-point apparatus; uncorrected. Optical rotations: JASCO P-1300 spectropolarimeter. IR: Spectra: BRUKER VECTOR-22 spectrophotometer; in cm<sup>-1</sup>. <sup>1</sup>H-, <sup>13</sup>C-, 2D-NMR spectra: BRUKER AVANCE 600 spectrometer; chemical shifts  $\delta$  in ppm rel. to (CH<sub>3</sub>)<sub>4</sub>Si, coupling constant *J* in Hz. ESI-MS: Finnigan LCQ mass spectrometer; in m/z. HR-ESI-MS: Q-Tof micro YA019 mass spectrometer.

#### 3.2. Material

The roots of *Actinidia valvata* Dunn were collected in Changshan County, Zhejiang Province, China, in October 2006, and identified by Zheng Han-Chen, Department of pharmacognosy, School of pharmacy, Second military medical university. A voucher specimen (No. 20061005) was deposited at Department of pharmacognosy, School of pharmacy, Second military medical university.

#### 3.3. Extraction and Isolation

The powdered plant material of roots of *Actinidia valvata* Dunn 30 kg was refluxed with 8 times of 80% EtOH solution for 3 times, 1.5 h each time. The extract was concentrated under reduced pressure to brown syrup, which was partitioned between H<sub>2</sub>O and petroleum ether (PE), AcOEt, and BuOH, successively. The *n*-BuOH soluble fraction (280.6 g) was subjected to column chromatography (CC) on silica gel (SiO<sub>2</sub>), eluting with CHCl<sub>3</sub>/MeOH/H<sub>2</sub>O (10:1:0.1 to 2:1 0.1) to afford 9 fraction 1–9. Fraction 5 were repeatedly subjected to CC (Pharmadex LH-20 and RP C-18) to yield compound **1** (10.4 mg) and 2 (14.6 mg).

30-*O*-β-D-glucopyranosyloxy-2α,3α,24-trihydroxyurs-12, 18-diene-28-oic acid *O*-β-D-glucopyranosyl ester (**1**): white amorphous powder, mp 140°–142°,  $[\alpha]_D^{25} = 5.89$  (*c* = 0.1, MeOH). IR (KBr): 3432, 2920, 2852, 1641, 1380, 1038. <sup>1</sup>H-NMR (600 MHz, C<sub>5</sub>D<sub>5</sub>N) and <sup>13</sup>C-NMR (150 MHz, C<sub>5</sub>D<sub>5</sub>N): Table 1. ESI-MS: 849.48 ( $[M + Na]^+$ ), HR-ESI-MS: 849.4240 ( $[M + Na]^+$ , C<sub>42</sub>H<sub>66</sub>N<sub>a</sub>O<sub>16</sub><sup>+</sup>, calc. 849.4249).

 $2\alpha,3\beta,23,30$ -tetrahydroxyurs-12, 18-diene-28-oic acid *O*- $\beta$ -D-glucopyranosyl ester (**2**): white amorphous powder. mp 220° (carbonification),  $[\alpha]_{D}^{25} = 7.35$  (c = 0.1, MeOH). IR (KBr): 3448, 2963, 1681, 1644, 1381, 1278, 1080. <sup>1</sup>H-NMR (600 MHz, C<sub>5</sub>D<sub>5</sub>N) and <sup>13</sup>C-NMR (150 MHz, C<sub>5</sub>D<sub>5</sub>N): Table 1 ESI-MS: 687.89 ( $[M + Na]^+$ ), HR-ESI-MS: 687.3717 ( $[M + Na]^+$ , C<sub>36</sub>H<sub>56</sub>N<sub>a</sub>O<sub>11</sub><sup>+</sup>, calc. 687.3720).

#### 4. Conclusions

In the present research, two new triterpenoids,  $30-O-\beta$ -D-glucopyranosyloxy- $2\alpha$ , $3\alpha$ ,24-trihydroxyurs-12, 18-diene-28-oic acid  $O-\beta$ -D-glucopyranosyl ester and  $2\alpha$ , $3\beta$ ,23,30-tetrahydroxyurs-12, 18-diene-28-oic acid  $O-\beta$ -D-glucopyranosyl ester were isolated from roots of *Actinidia valvata* Dunn, and their structures were elucidated by means of extensive spectroscopic studies. Both these new compounds showed moderate cytotoxic activity *in vitro* against BEL-7402 and SMMC-7721 tumor cell line.

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