

Article

The ERK1/2 Signaling Pathway Is Involved in Sulfur Dioxide Preconditioning-Induced Protection against Cardiac Dysfunction in Isolated Perfused Rat Heart Subjected to Myocardial Ischemia/Reperfusion

Pan Huang^{1,†}, Yan Sun^{1,†}, Jinyan Yang¹, Siyao Chen¹, Angie Dong Liu², Lukas Holmberg², Xiaomei Huang¹, Chaoshu Tang^{3,4}, Junbao Du^{1,3} and Hongfang Jin^{1,3,*}

¹ Department of Pediatrics, Peking University First Hospital, Xi-An Men Str. No. 1, West District, Beijing 100034, China; E-Mails: huangpan036@163.com (P.H.); yansun2008@gmail.com (Y.S.); yangjinyan_tg@163.com (J.Y.); siyaochen_nymaz@163.com (S.C.);

hxm111@163.com (X.H.); junbaodu1@126.com (J.D.)

² Department of Medical and Health Sciences, Linköping University, Linköping 58183, Sweden; E-Mails: angie.dongliu@hotmail.com (A.D.L.); lukas.holmberg20@gmail.com (L.H.)

³ Key Laboratory of Molecular Cardiology, Ministry of Education, Beijing 100191, China; E-Mail: tangchaoshu@263.net.cn

⁴ Department of Physiology and Pathophysiology, Health Sciences Center, Peking University, Beijing 100191, China

† These authors contributed equally to this work.

* Author to whom correspondence should be addressed; E-Mail: jinhongfang51@126.com; Tel.: +86-10-8357-3209; Fax: +86-10-6653-0532.

Received: 12 August 2013; in revised form: 31 October 2013 / Accepted: 1 November 2013 /

Published: 8 November 2013

Abstract: Ischemia/reperfusion injury (IRI) occurs frequently during reperfusion of ischemic myocardium, and preconditioning has been regarded as one of the best strategies to prevent myocardial injury during the ischemia/reperfusion process. Our previous studies indicated that a small dose of sulfur dioxide (SO₂) used as preconditioning exerts cardioprotection. However, the mechanisms underlying the cardioprotection remain unclear. The present study was designed to examine if the extracellular regulated protein kinases 1/2 (ERK1/2) signaling pathway mediated protection against cardiac dysfunction after SO₂ preconditioning in isolated rat hearts subjected to ischemia/reperfusion (I/R). Langendorff heart perfusion was performed *in vitro*, where 56 male Wistar rats were

randomly divided into seven groups: control group, 5 $\mu\text{mol/L}$ SO_2 group (S5), 2-(2-Amino-3-methoxyphenyl)-4*H*-1-benzopyran-4-one (PD98059) + 5 $\mu\text{mol/L}$ SO_2 (PD98059 + S5) group, PD98059 group, I/R group, 5 $\mu\text{mol/L}$ SO_2 + I/R (S5 + I/R) group and PD98059 + 5 $\mu\text{mol/L}$ SO_2 + I/R (PD98059 + S5 + I/R) group. Cardiac function and myocardial phosphorylated ERK1/2 protein were measured. We found that I/R in isolated rat heart resulted in cardiac dysfunction with a significant increase in phosphorylated ERK1/2 protein. SO_2 preconditioning markedly suppressed phosphorylated ERK1/2 protein and improved cardiac function in isolated rat heart with I/R ($p < 0.05$). However, pre-treatment with PD98059 could prevent the above effects of SO_2 preconditioning. In conclusion, SO_2 preconditioning protected against cardiac dysfunction in isolated rat heart subjected to I/R via suppression of the over-activation of the ERK1/2 signaling pathway.

Keywords: myocardial ischemia/reperfusion injury; sulfur dioxide; preconditioning; mitogen activated protein kinase

1. Introduction

Myocardial ischemia/reperfusion injury (IRI) is common in clinical practice. Thus, exploring a way for myocardial protection and investigating its mechanisms are important issues in clinical science. Preconditioning has been regarded as the best strategy to prevent myocardial injury during ischemic reperfusion processes. Previous studies on preconditioning have mainly been focusing on preconditioning using ischemia, hypoxia, metabolic suppression, endotoxin, adenosine and so forth [1–3]. Interestingly, SO_2 has, during recent years, been discovered to have important biological effects on the cardiovascular system [4–11]. Zhang, *et al.* [12] demonstrated that exogenous SO_2 could damage the myocardium. The outcome indicated that a small dose of SO_2 as preconditioning exerted a significantly protective effect on myocardial injury [13]. However, the mechanisms underlying this protective effect remain unclear.

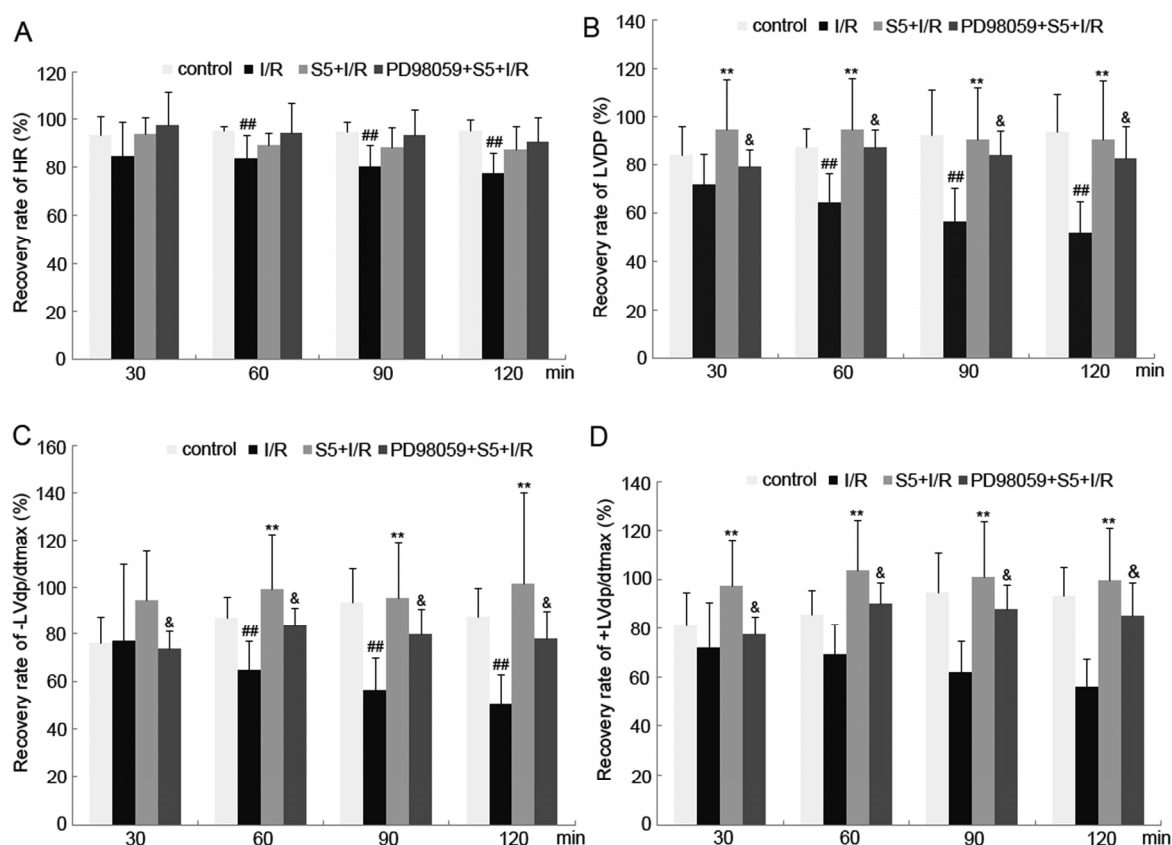
Mitogen-activated protein kinases (MAPKs) are crucial in cell signaling transduction. They contain three members: p38 MAPK, c-Jun *N*-terminal kinase (JNK) and extracellular signaling-regulated kinase (ERK). MAPKs play an important role in regulating cell function and survival [14,15]. Different family members activate various pathways by attaching to corresponding cell adhesion molecules and responding to disparate stimulation to regulate cell survival, function, growth, proliferation and differentiation. Furthermore, MAPKs are also important in regulating inflammation [16–21]. Evidence shows that the ERK1/2 signaling pathway is essential to tumorigenesis and tumor progression [22,23], and that the suppression of the excessive activation of ERK1/2 could improve vasoactivity [24]. A previous study showed a sustained activation of ERK1/2 during ischemia/reperfusion [25]. Therefore, this present study was designed to focus on this component. The phosphorylation and activation of MAP kinase (ERK1/2, p44/42MAPK) could be selectively suppressed by 2-(2-Amino-3-methoxyphenyl)-4*H*-1-benzopyran-4-one (PD98059), an organic compound inhibitor frequently used to block the activity of the ERK1/2 protein kinase [25–29]. The present study aimed at investigating whether the MAPK signaling pathway was involved in the SO_2 preconditioning-induced protection against cardiac dysfunction in isolated rat heart subjected to ischemia/reperfusion (I/R).

2. Results

2.1. Ischemia/Reperfusion Resulted in Impaired Cardiac Function in Isolated Perfused Rat Heart

Compared with the control group, the recovery rate of heart rate (HR), left ventricular developed pressure (LVDP) and maximum decreasing rate ($-LVdp/dt_{max}$) at 60, 90 and 120 min of ischemia/reperfusion were decreased significantly in the I/R group ($p < 0.01$) (Figure 1A–C).

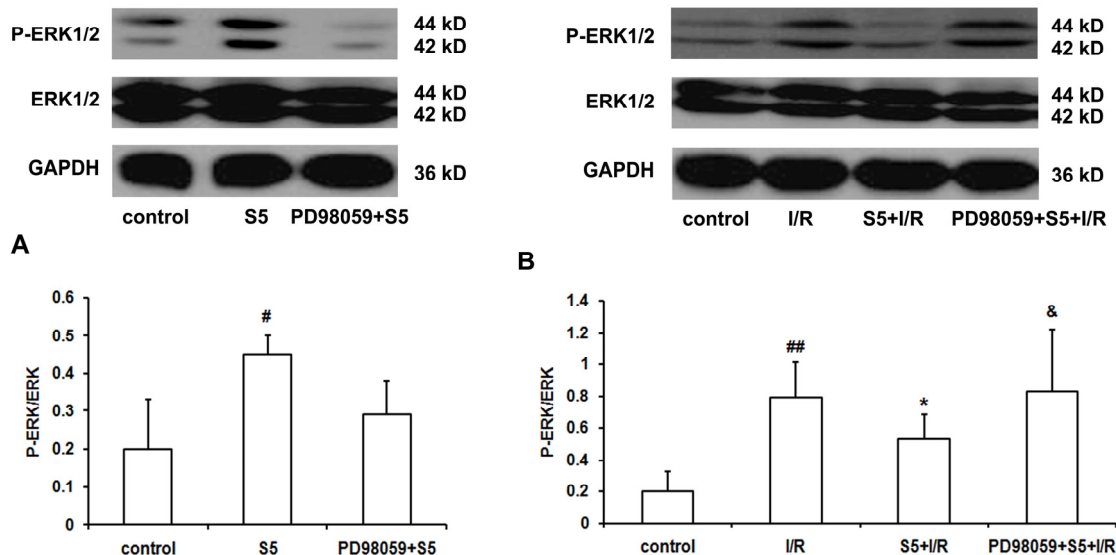
Figure 1. Recovery rate of heart rate (HR) (A); left ventricular developed pressure (LVDP) (B); left ventricular developed pressure maximum decreasing rate ($-LVdp/dt_{max}$) (C) and left ventricular developed pressure maximum increasing rate ($+LVdp/dt_{max}$) (D) of SO_2 and 2-(2-amino-3-methoxyphenyl)-4*H*-1-benzopyran-4-one (PD98059) preconditioning on ischemia/reperfusion *in vitro* at different time points (30, 60, 90 and 120 min). Ischemia/reperfusion resulted in an impaired cardiac function in isolated perfused rat heart. SO_2 preconditioning improved the myocardial function in isolated perfused rat heart subjected to ischemia/reperfusion, which could be abolished by treatment with PD98059. $## p < 0.01$ versus control group; $** p < 0.01$ versus ischemia/reperfusion (I/R) group; $\& p < 0.05$ versus 5 $\mu\text{mol/L}$ SO_2 group (S5) + I/R group.



2.2. SO_2 Induced Phosphorylation of ERK1/2 Protein and Depressed Cardiac Function in Isolated Perfused Rat Heart without Ischemia/Reperfusion

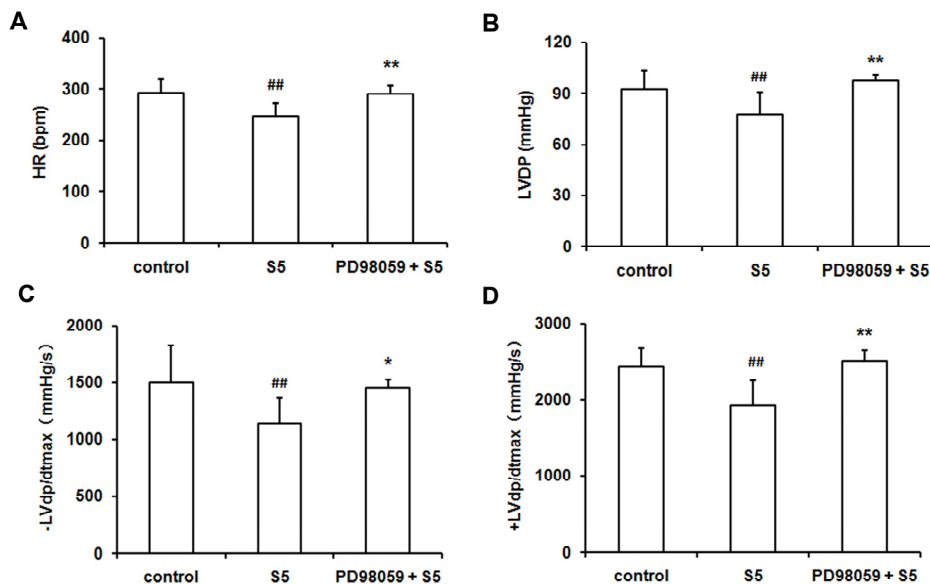
The SO_2 donor elevated the phosphorylation of ERK1/2 protein in rat myocardium ($p < 0.05$), which was successfully prevented by PD98059 (Figure 2A).

Figure 2. Phosphorylated ERK1/2 protein expressions of myocardium *in vitro*. SO₂ induced phosphorylation of ERK1/2 protein in rat myocardium (A); SO₂ preconditioning inhibited the excessively-induced myocardial phosphorylation of ERK1/2 protein induced by ischemia/reperfusion. Pretreatment with PD98059 successfully abolished the above inhibitory effect (B). # *p* < 0.05; ## *p* < 0.01 versus control; * *p* < 0.05 versus I/R; & *p* < 0.05 versus S5 + I/R.



The SO₂ donor depressed the cardiac function in isolated rat heart presented by the decreases in the HR, LVDP, -LVdp/dt_{max} and +LVdp/dt_{max}, which was prevented by PD98059 pretreatment (Figure 3). However, PD98059 alone did not change the cardiac function in isolated rat heart (data was not shown).

Figure 3. The change in HR (A); LVDP (B); -LVdp/dt_{max} (C) and +LVdp/dt_{max} (D) in the isolated rat heart without ischemia/reperfusion. The 5 μmol/L SO₂ donor depressed the cardiac function, while PD98059 could prevent the inhibitory effect of the SO₂ donor. ## *p* < 0.01 versus control group * *p* < 0.05; ** *p* < 0.01 versus S5 group.



2.3. SO₂ Preconditioning Inhibited the Excessively-Induced Myocardial Phosphorylation of ERK1/2 Protein Induced by Ischemia/Reperfusion

In the ischemia/reperfusion group, the phosphorylation of myocardial ERK1/2 protein was significantly higher than that of the control group ($p < 0.01$). However, SO₂ preconditioning markedly inhibited the excessively-induced myocardial phosphorylation of the ERK1/2 protein caused by ischemia/reperfusion ($p < 0.05$). However, pretreatment with PD98059 successfully abolished the above inhibitory effect (Figure 2B, $p < 0.05$).

2.4. SO₂ Preconditioning Improved the Myocardial Function in Isolated Perfused Rat Heart Subjected to Ischemia/Reperfusion, Which Could Be Abolished by Treatment with PD98059

Compared with the I/R group, the recovery rate of LVDP ($p < 0.01$) and +LVdp/dt_{max} ($p < 0.01$) were increased significantly at 30, 60, 90 and 120 min of ischemia/reperfusion, respectively, for the S5 + I/R group. Furthermore, at 60, 90 and 120 min, the recovery rate of -LVdp/dt_{max} was also increased significantly ($p < 0.01$). However, the above effects could be successfully abolished by pretreatment with PD98059 ($p < 0.05$) (Figure 1B–D). The measurement of creatine kinase (CK) and glutamic-oxaloacetic transaminase (GOT) activities in the coronary perfusion fluid (CPF) showed that the increase in the CK and GOT activities of the CPF of the I/R group was alleviated by SO₂ preconditioning (Table 1).

Table 1. The activities of creatine kinase (CK) and glutamic-oxaloacetic transaminase (GOT) in the coronary perfusion fluid.

Group	n	CK (U/L)	GOT (U/L)
Control	8	3.50 ± 1.85	3.25 ± 0.71
I/R	8	14.33 ± 8.34 ^{##}	6.50 ± 1.88 ^{##}
S5 + I/R	8	6.71 ± 4.23 [*]	3.71 ± 1.25 ^{**}

^{##} $p < 0.01$, vs. control group; ^{**} $p < 0.01$; ^{*} $p < 0.05$ vs. I/R group.

3. Discussion

I/R is an important pathophysiologic process in clinical practice and was first described by Jennings in 1960. It not only exists in different species and organs, but is involved in various pathological processes, such as multi-organ failure, shock and heart failure [30–32]. Currently, cardiovascular disease is one of the most serious threats to people's life and health. There are now various treatments for cardiovascular diseases, such as thrombolytic therapy and coronary artery bypass grafting for coronary heart disease. Although they have decreased the mortality dramatically, the I/R injuries that come with these treatments make the results less satisfying. Therefore, the prevention and treatment of I/R are now the hot spots in medical research. The phenomenon of ischemic preconditioning was first brought out by Murry in 1986 [33], and it opened up a new field in I/R research. Thereafter, studies on IPC were conducted worldwide. However, studies about IPC have certain ethical and clinical limitations.

It has been shown that SO₂ which is considered one of the endothelium-derived hyperpolarizing factors [34,35] can be produced endogenously from coronary arteries. Our research group previously proved that a small dose of exogenous SO₂ could induce myocardial injury [12], and as a result, we

further demonstrated that SO₂ preconditioning could protect myocardium by antagonizing I/R *in vivo* [13]. However, the mechanisms responsible for the protection of cardiac function provided by SO₂ preconditioning have not yet been fully understood.

MAPKs are one of the important signaling molecules in cell signal transduction. Previous studies have showed that the MEK1-ERK2 signaling pathway is involved in the regulation of cell survival [36–39]. Therefore, for the purposes of exploring the mechanisms responsible for SO₂ preconditioning against I/R, we investigated whether the ERK/MAPK signaling pathway mediated the cardioprotection by SO₂ preconditioning in isolated perfused rat heart subjected to I/R *in vitro*.

The results of the present study showed that the phosphorylation of ERK1/2 protein in myocardium was increased, and at the same time, cardiac function was impaired after I/R. However, SO₂ pretreatment could elevate the phosphorylation of ERK1/2 protein in myocardium in isolated perfused rat heart without exposure to I/R, and its preconditioning markedly inhibited the increased myocardial phosphorylation of ERK1/2 protein induced by I/R and protected impaired cardiac function and cardiac injury.

To further explore the significance of the ERK/MAPK signaling pathway in cardioprotection by SO₂ preconditioning during I/R, we used PD98059, a MAPKK inhibitor, and observed if the cardioprotective effect of SO₂ preconditioning could be prevented in isolated perfused rat hearts subjected to I/R. The molecular weight of PD98059 is 267.28, and its molecular formula is C₁₆H₁₃NO₃. It can penetrate cells and inhibit MEK1 (one of the MAPK kinases) selectively to suppress the phosphorylation and activation of MAP kinase (ERK1/2, p44/42MAPK) [25–29]. Of note, our results showed that pretreatment with PD98059 successfully abolished the inhibitory effect of SO₂ preconditioning on increased phosphorylation of ERK1/2 protein in the myocardium and the protective effect of SO₂ preconditioning on the cardiac function of isolated perfused rat heart subjected to I/R. The above evidence proved that SO₂ preconditioning could inhibit the over-activation of the ERK1/2 signaling pathway to protect the myocardium in isolated perfused rat heart during I/R.

Regarding why there was an increased ERK1/2 protein phosphorylation in the PD98059 + S5 + I/R group, we supposed that pretreatment with SO₂ could stimulate a moderate ERK1/2 protein phosphorylation, as shown in Figure 2A. Then, in the S5 + I/R group, I/R-induced over-activation of ERK1/2 protein would be inhibited, while PD98059 inhibited ERK1/2 protein phosphorylation induced by SO₂ pretreatment before I/R challenge, as shown in Figure 2A. Therefore, in the PD98059 + S5 + I/R group, the isolated perfused rat heart would not experience the sufficiently activated ERK1/2 status before I/R challenge. Therefore, I/R-induced over-activation of ERK1/2 protein could not be inhibited.

Our study indicated that SO₂ preconditioning could protect myocardial function. This is possibly due to the suppression of induced ERK phosphorylation during I/R in isolated perfused rat hearts. However, the exact mechanisms by which SO₂ preconditioning protects myocardial function from I/R need further investigation.

4. Experimental Section

4.1. Animals

Fifty-six male adult Wistar rats, weighing 250–300 g, were provided by the Experimental Animal Center, Peking University Health Science Center (Beijing, China), and had free access to water and

standard rat chow. All studies were performed with the approval of the Experimental Animal Committee at Peking University, and the animals were cared for in a manner that complied with the Animal Management Rules of the Ministry of Health of the People's Republic of China (documentation number 19890503).

4.2. Reagents

PD98059 was purchased from Promega (Madison, WI, USA), and p-p44/42 MAPK and p44/42 MAPK polyclonal antibodies were purchased from Cell Signaling Technology (Beverly, MA, USA). SO₂ derivatives and a mixture of sulfite and bisulfite (Na₂SO₃/NaHSO₃) in a molar ratio of 3:1 were purchased from Sigma (St. Louis, MO, USA).

4.3. Animal Grouping

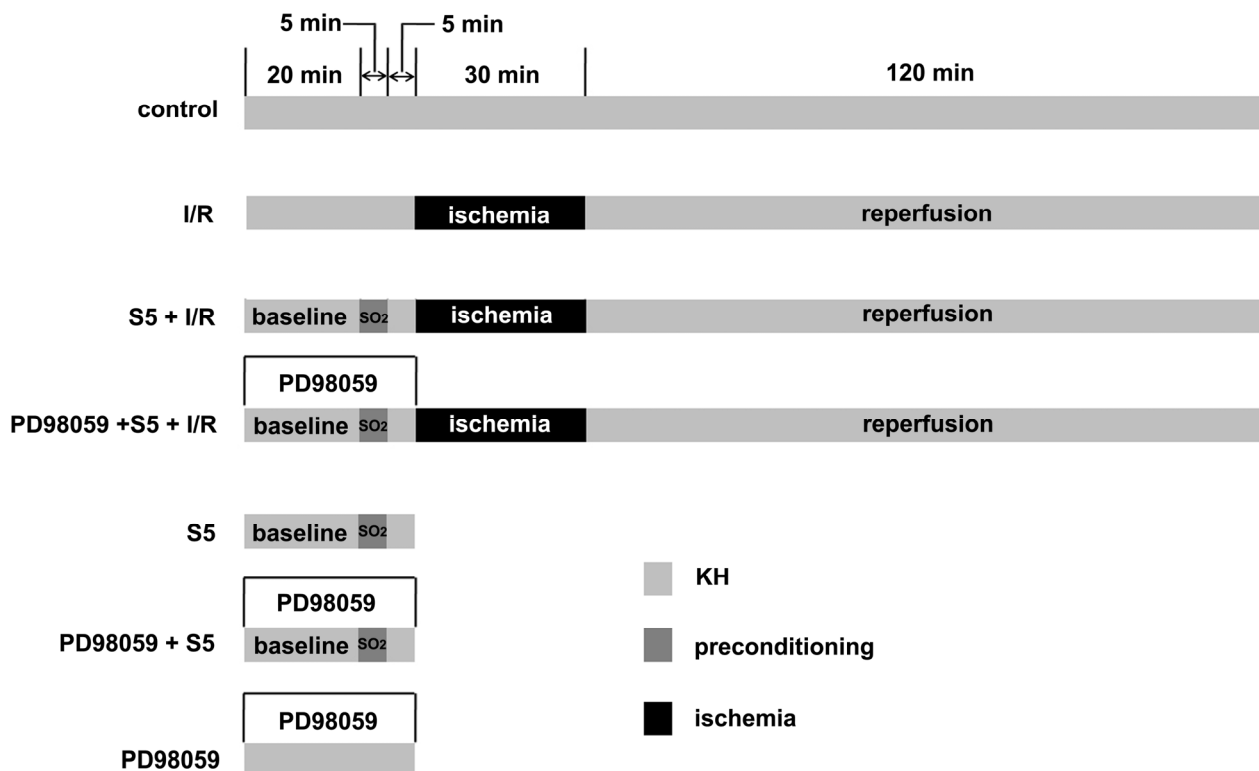
Fifty-six male Wistar rats were divided into the following 7 groups: the control group without I/R ($n = 8$), the 5 $\mu\text{mol/L}$ SO₂ pretreated group without I/R (S5 group, $n = 8$), the PD98059 + 5 $\mu\text{mol/L}$ SO₂ pretreated group without I/R (PD98059 + S5 group, $n = 8$), the PD98059 without I/R group (PD98059 group, $n = 8$), the ischemia/reperfusion group (I/R group, $n = 8$), the 5 $\mu\text{mol/L}$ SO₂ pretreated group + I/R (S5 + I/R group, $n = 8$) and the PD98059 + 5 $\mu\text{mol/L}$ SO₂ pretreated + I/R group (PD98059 + S5 + I/R group, $n = 8$). The control group received perfusion of Krebs-Henseleit (KH) solution during the whole experiment. In the S5 group, after 20 min of stabilization, the perfusion was pretreated with KH containing 5 $\mu\text{mol/L}$ SO₂ derivatives for 5 min, and then, the rat hearts were perfused with KH solution for 5 min. In the PD98059 + S5 group, perfusion was given with PD98059 10 $\mu\text{mol/L}$ for 30 min on the basis of the S5 group. In the PD98059 group, perfusion was given with PD98059 10 $\mu\text{mol/L}$ for 30 min. In the I/R group, after 30 min of stabilization, perfusion was stopped for 30 min (ischemia), and the heart was then reperfused for 120 min with KH solution (37 °C). In the S5 + I/R group, 10 min before ischemia, perfusion was preconditioned with KH containing 5 $\mu\text{mol/L}$ SO₂ derivatives for 5 min. Then, the perfusion was stopped for 30 min, and then, the heart was reperfused for 120 min with KH solution (37 °C). Finally, in the PD98059 + S5 + I/R group, perfusion was given with PD98059 for 30 min before ischemia and was preconditioned with 5 $\mu\text{mol/L}$ SO₂ derivatives for 5 min, 10 min before ischemia. The heart was then reperfused for 120 min with KH solution (37 °C).

4.4. Heart Perfusion In Vitro and Cardiac Function Measurement

After anesthetization with pentobarbital sodium via intraperitoneal injection at 40 mg/kg, rat hearts were taken out and placed on a HV-4 Langendorff Perfusion Apparatus (Taimeng Science and Technology Ltd., Chengdu, China) after the left auricle was cut off. Then, the rat heart was perfused with Krebs-Henseleit (KH) solution (mmol/L) [40] through the aorta retrogradely. Krebs-Henseleit (KH) solution (mmol/L) consisted of: NaCl, 118.0; KCl, 4.7; KH₂PO₄, 0.93; MgSO₄·7H₂O, 1.2; CaCl₂, 1.5; NaHCO₃, 25; and C₆H₁₂O₆, 11.0 at pH 7.4 and 37 °C and with 100 cm H₂O (1 cm H₂O = 0.098 kPa) constant pressure and 95% O₂/5% CO₂ pre-saturated mixed gas. A cardiac catheter with balloon was inserted into the left ventricle from the atrioventricular valve. The balloon was filled and the left

ventricular end-diastolic pressure (LVEDP) maintained 0–10 mmHg (1 mmHg = 0.133 kPa). After pre-perfusion equilibrium for 10 min, HR, LVDP and $\pm LVdp/dt_{max}$ were recorded using a BL-420F Biological Function Experiment System (Taimeng Science and Technology Ltd., Chengdu, China). The perfusion procedure is shown in Figure 4.

Figure 4. Heart perfusion procedure. Baseline: cardiac function change before ischemia. Ten min before ischemia, the rat heart was preconditioned with 5 $\mu\text{mol/L}$ SO_2 derivatives for 5 min. The heart was perfused with PD98059 for 30 min before ischemia. KH: Krebs-Henseleit.



4.5. Measurement of CK and GOT Activity in CPF

The CPF was collected during reperfusion in the control group, I/R group and S5 + I/R group. The activities of CK and GOT were assayed by the enzymologic method with an Automated Biochemistry Instrument (Hitachi 7060, Hitachi Company, Tokyo, Japan).

4.6. Myocardial ERK and P-ERK Detection by Western Blotting

Myocardium stored at $-70\text{ }^\circ\text{C}$ was weighed, lysed in cell lysis solution measured in the ratio of 1:10 (mass:volume) and then homogenized with ultrasound at $4\text{ }^\circ\text{C}$, and the homogenized solution was centrifuged at $13,000 \times g$ for 10 min. The supernatant was then put through electrophoresis, and the gel was loaded with an equivalent amount of mixed protein samples in each pore and then transferred to a nitrocellulose membrane. The membrane was blocked with non-fat milk for 1 h, and primary antibodies were added (ERK1/2 and p-ERK1/2 polyclonal antibodies, diluted to 1:2000) and incubated at $4\text{ }^\circ\text{C}$ overnight. Secondary antibodies (goat anti-rabbit monoclonal antibody, diluted to 1:8000) were added to the membrane. The membrane was incubated at room temperature for 1 h, washed by

Tween/Tris-buffered salt solution (TTBS) 4 times (10 min each time) and incubated with chemiluminescence for 1 min. After film exposure, development and photographic fixing, protein stripes were scanned with a gel imaging system from AlphaImager (San Leandro, CA, USA), for testing the optical density of the protein stripes.

4.7. Statistics

Measurement data are expressed as the mean \pm standard deviation (SD). Data were processed by SPSS13.0 (Chicago, IL, USA). One-Way ANOVA followed by least-significance difference (LSD) was used to compare the differences of detected indices among the groups. A paired *t*-test was used to compare the difference between two groups. $p < 0.05$ was considered statistically significant.

5. Conclusions

Our study indicated that SO₂ preconditioning could protect myocardial function and injury by suppressing the over-activated ERK phosphorylation during I/R in isolated perfused rat hearts.

Acknowledgments

This work was supported by the Major Basic Research Program of China (2012CB517806 and 2011CB503904), the National Natural Science Foundation of China (31130030, 81070111 and 81121061), the Beijing Natural Science Foundation (7112130) and the Program for New Century Excellent Talents of the Ministry of Education, China. We thank Qiuyu Yao and Yaqian Huang for their kind help in langendorff perfusion and western blot experiments.

Conflicts of Interest

The authors declare no conflict of interest.

References

1. Zhao, H.X.; Wang, X.L.; Wang, Y.H.; Wu, Y.; Li, X.Y.; Lv, X.P.; Zhao, Z.Q.; Zhao, R.R.; Liu, H.R. Attenuation of myocardial injury by postconditioning: Role of hypoxia inducible factor-1 α . *Basic Res. Cardiol.* **2010**, *105*, 109–118.
2. Duan, Z.; Zhang, L.; Liu, J.; Xiang, X.; Lin, H. Early protective effect of total hypoxic preconditioning on rats against systemic injury from hemorrhagic shock and resuscitation. *J. Surg. Res.* **2012**, *178*, 842–850.
3. Gruner, C.; Akkaya, E.; Kretschmar, O.; Roffi, M.; Corti, R.; Jenni, R.; Eberli, F.R. Pharmacologic preconditioning therapy prior to atrial septal defect closure in patients at high risk for acute pulmonary edema. *J. Interv. Cardiol.* **2012**, *25*, 505–512.
4. Du, S.X.; Jin, H.F.; Bu, D.F.; Zhao, X.; Geng, B.; Tang, C.S.; Du, J.B. Endogenously generated sulfur dioxide and its vasorelaxant effect in rats. *Acta Pharmacol. Sin.* **2008**, *29*, 923–930.
5. Jin, H.F.; Du, S.X.; Zhao, X.; Wei, H.L.; Wang, Y.F.; Liang, Y.F.; Tang, C.S.; Du, J.B. Effects of endogenous sulfur dioxide on monocrotaline-induced pulmonary hypertension in rats. *Acta Pharmacol. Sin.* **2008**, *29*, 1157–1166.

6. Sun, Y.; Tian, Y.; Prabha, M.; Liu, D.; Chen, S.; Zhang, R.; Liu, X.; Tang, C.; Tang, X.; Jin, H.; *et al.* Effects of sulfur dioxide on hypoxic pulmonary vascular structural remodeling. *Lab. Invest.* **2010**, *90*, 68–82.
7. Li, J.; Meng, Z. The role of sulfur dioxide as an endogenous gaseous vasoactive factor in synergy with nitric oxide. *Nitric Oxide* **2009**, *20*, 166–174.
8. Liang, Y.; Liu, D.; Ochs, T.; Tang, C.; Chen, S.; Zhang, S.; Geng, B.; Jin, H.; Du, J. Endogenous sulfur dioxide protects against isoproterenol-induced myocardial injury and increases myocardial antioxidant capacity in rats. *Lab. Invest.* **2011**, *91*, 12–23.
9. Meng, Z.; Zhang, H. The vasodilator effect and its mechanism of sulfur dioxide-derivatives on isolated aortic rings of rats. *Inhal. Toxicol.* **2007**, *19*, 979–986.
10. Wang, Y.K.; Ren, A.J.; Yang, X.Q.; Wang, L.G.; Rong, W.F.; Tang, C.S.; Yuan, W.J.; Lin, L. Sulfur dioxide relaxes rat aorta by endothelium-dependent and independent mechanisms. *Physiol. Res.* **2009**, *58*, 521–527.
11. Zhang, Q.; Meng, Z. The vasodilator mechanism of sulfur dioxide on isolated aortic rings of rats: Involvement of the K⁺ and Ca²⁺ channels. *Eur. J. Pharmacol.* **2009**, *602*, 117–123.
12. Zhang, S.Q.; Du, J.B.; Jin, H.F.; Li, W.; Liang, Y.F.; Geng, B.; Li, S.K.; Zhang, C.Y.; Tang, C.S. Endogenous sulfur dioxide aggravates myocardial injury in isolated rat heart with ischemia and reperfusion. *Transplantation* **2009**, *87*, 517–524.
13. Wang, X.B.; Huang, X.M.; Ochs, T.; Li, X.Y.; Jin, H.F.; Tang, C.S.; Du, J.B. Effect of sulfur dioxide preconditioning on rat myocardial ischemia/reperfusion injury by inducing endoplasmic reticulum stress. *Basic Res. Cardiol.* **2011**, *106*, 865–878.
14. Widmann, C.; Gibson, S.; Jarpe, M.B.; Johnson, G.L. Mitogen-activated protein kinase: Conservation of a three-kinase module from yeast to human. *Physiol. Rev.* **1999**, *79*, 143–180.
15. Gerits, N.; Kostenko, S.; Moens, U. *In vivo* functions of mitogen-activated protein kinases: Conclusions from knock-in and knock-out mice. *Transgenic Res.* **2007**, *16*, 281–314.
16. Cargnello, M.; Roux, P.P. Activation and function of the MAPKs and their substrates, the MAPK-activated protein kinases. *Microbiol. Mol. Biol. Rev.* **2011**, *75*, 50–83.
17. Pang, L.; Sawada, T.; Decker, S.J.; Saltiel, A.R. Inhibition of MAP kinase kinase blocks the differentiation of PC-12 cells induced by nerve growth factor. *J. Biol. Chem.* **1995**, *270*, 13585–13588.
18. Park, J.; Song, K.H.; Ha, H. Fractalkine increases mesangial cell proliferation through reactive oxygen species and mitogen-activated protein kinases. *Transpl. Proc.* **2012**, *44*, 1026–1028.
19. Chung, J.W.; Choi, R.J.; Seo, E.K.; Nam, J.W.; Dong, M.S.; Shin, E.M.; Guo, L.Y.; Kim, Y.S. Anti-inflammatory effects of *Z*-ligustilide through suppression of mitogen-activated protein kinases and nuclear factor- κ B activation pathways. *Arch. Pharm. Res.* **2012**, *35*, 723–732.
20. Himaya, S.W.; Ryu, B.; Qian, Z.J.; Kim, S.K. Paeonol from *Hippocampus kuda* Bleeler suppressed the neuro-inflammatory responses *in vitro* via NF- κ B and MAPK signaling pathways. *Toxicol. Vitro* **2012**, *26*, 878–887.
21. McCarroll, J.A.; Phillips, P.A.; Park, S.; Doherty, E.; Pirola, R.C.; Wilson, J.S.; Apte, M.V. Pancreatic stellate cell activation by ethanol and acetaldehyde: Is it mediated by the mitogen-activated protein kinase signaling pathway? *Pancreas* **2003**, *27*, 150–160.

22. Yang, D.; Fan, X.; Yin, P.; Wen, Q.; Yan, F.; Yuan, S.; Liu, B.; Zhuang, G.; Liu, Z. Ignification of decoy receptor 3 (DcR3) and external-signal regulated kinase 1/2 (ERK1/2) in gastric cancer patients. *BMC Immunol.* **2012**, *13*, 28.
23. Fujioka, N.; Nguyen, J.; Chen, C.; Li, Y.; Pasrija, T.; Niehans, G.; Johnson, K.N.; Gupta, V.; Kratzke, R.A.; Gupta, K. Morphine-induced epidermal growth factor pathway activation in non-small cell lung cancer. *Anesth. Analg.* **2011**, *113*, 1353–1364.
24. Bhattacharya, I.; Damjanović, M.; Dominguez, A.P.; Haas, E. Inhibition of activated ERK1/2 and JNKs improves vascular function in mouse aortae in the absence of nitric oxide. *Eur. J. Pharmacol.* **2011**, *658*, 22–27.
25. Punn, A.; Mockridge, J.W.; Farooqui, S.; Marber, M.S.; Heads, R.J. Sustained activation of p42/p44 mitogen-activated protein kinase during recovery from simulated ischaemia mediates adaptive cytoprotection in cardiomyocytes. *Biochem. J.* **2000**, *350*, 891–899.
26. Crews, C.M.; Alessandrini, A.; Erikson, R.L. The primary structure of MEK, a protein kinase that phosphorylates the ERK gene product. *Science* **1992**, *258*, 478–480.
27. Cowley, S.; Paterson, H.; Kemp, P.; Marshall, C.J. Activation of MAP kinase kinase is necessary and sufficient for PC12 differentiation and for transformation of NIH 3T3 cells. *Cell* **1994**, *77*, 841–852.
28. Dudley, D.T.; Pang, L.; Decker, S.J.; Bridges, A.J.; Saltiel, A.R. A synthetic inhibitor of the mitogen-activated protein kinase cascade. *Proc. Natl. Acad. Sci. USA* **1995**, *92*, 7686–7689.
29. Alessi, D.R.; Saito, Y.; Campbell, D.G.; Cohen, P.; Sithanandam, G.; Rapp, U.; Ashworth, A.; Marshall, C.J.; Cowley, S. Identification of the sites in MAP kinase kinase-1 phosphorylated by p74raf-1. *EMBO J.* **1994**, *13*, 1610–1619.
30. Hausenloy, D.J.; Baxter, G.; Bell, R.; Botker, H.E.; Davidson, S.M.; Downey, J.; Heusch, G.; Kitakaze, M.; Lecour, S.; Mentzer, R.; *et al.* Translating novel strategies for cardioprotection: The hatter workshop recommendations. *Basic Res. Cardiol* **2010**, *105*, 677–686.
31. Koca, V.; Ari, H. Angioplasty as early revascularization in acute myocardial infarction. *Anadolu. Kardiyol. Derg.* **2008**, *8*, 77–83.
32. Yavuz, S. Surgery as early revascularization after acute myocardial infarction. *Anadolu. Kardiyol. Derg.* **2008**, *8*, 84–92.
33. Murry, C.E.; Jennings, R.B.; Reimer, K.A. Preconditioning with ischemia: A delay of lethal cell injury in ischemic myocardium. *Circulation* **1986**, *74*, 1124–1136.
34. Bai, J.; Meng, Z. Expression of apoptosis-related genes in livers from rats exposed to sulfur dioxide. *Toxicology* **2005**, *216*, 253–260.
35. Balazy, M.; Abu-Yousef, I.A.; Harpp, D.N.; Park, J. Identification of carbonyl sulfide and sulfur dioxide in porcine coronary artery by gas chromatography/mass spectrometry, possible relevance to EDHF. *Biochem. Biophys. Res. Commun.* **2003**, *311*, 728–734.
36. Yue, T.L.; Wang, C.; Gu, J.L.; Ma, X.L.; Kumar, S.; Lee, J.C.; Feuerstein, G.Z.; Thomas, H.; Maleeff, B.; Ohlstein, E.H. Inhibition of extracellular signal-regulated kinase enhances ischemia/reoxygenation-induced apoptosis in cultured cardiac myocytes and exaggerates reperfusion injury in isolated perfused heart. *Circ. Res.* **2000**, *86*, 692–699.

37. Lips, D.J.; Bueno, O.F.; Wilkins, B.J.; Purcell, N.H.; Kaiser, R.A.; Lorenz, J.N.; Voisin, L.; Saba-El-Leil, M.K.; Meloche, S.; Pouysségur, J.; *et al.* MEK1-ERK2 signaling pathway protects myocardium from ischemic injury *in vivo*. *Circulation* **2004**, *109*, 1938–1941.
38. Das, A.; Salloum, F.N.; Xi, L.; Rao, Y.J.; Kukreja, R.C. ERK phosphorylation mediates sildenafil-induced myocardial protection against ischemia-reperfusion injury in mice. *Am. J. Physiol. Heart Circ. Physiol.* **2009**, *296*, H1236–H1243.
39. Yang, X.; Liu, Y.; Yang, X.M.; Hu, F.; Cui, L.; Swingle, M.R.; Honkanen, R.E.; Soltani, P.; Tissier, R.; Cohen, M.V.; *et al.* Cardioprotection by mild hypothermia during ischemia involves preservation of ERK activity. *Basic Res. Cardiol.* **2011**, *106*, 421–430.
40. Johansen, D.; Ytrehus, K.; Baxter, G.F. Exogenous hydrogen sulfide (H₂S) protects against regional myocardial ischemia-reperfusion injury—Evidence for a role of K_{ATP} channels. *Basic Res. Cardiol.* **2006**, *101*, 53–60.

© 2013 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution license (<http://creativecommons.org/licenses/by/3.0/>).