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Cascading *cis*-Cleavage on Transcript from *trans*-Acting siRNA-Producing Locus 3

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Abstract: The production of small RNAs (sRNAs) from phased positions set by microRNA-directed cleavage of *trans*-acting-siRNA-producing locus (TAS) transcript has been characterized extensively; however, the production of sRNAs from non-phased positions remains unknown. We report three *cis*-cleavages that occurred in *TAS3* transcripts in *Vitis vinifera*, by combining high-throughput sRNA deep sequencing information with evolutionary conservation and genome-wide RNA degradome analysis. The three *cis*-cleavages can be deciphered to generate an orderly cleavage cascade, and can also produce distinct phasing patterns. Each of the patterns, either upstream or downstream of the *cis*-cleaved position, had a set of sRNAs arranged in 21-nucleotide increments. Part of the cascading *cis*-cleavages was also conserved in *Arabidopsis thaliana*. Our results will enhance the understanding of the production of sRNAs from non-phased positions that are not set by microRNA-directed cleavage.

Keywords: *trans*-acting siRNA; *cis*-cleavage; ta-siRNA-producing locus; miRNA

1. Introduction

In plants, many endogenous small RNAs (sRNAs), including microRNAs (miRNAs), heterochromatic small interfering RNAs (siRNAs), natural antisense siRNAs, and *trans*-acting siRNAs (ta-siRNAs), play important roles in regulating gene expression networks [1,2]. The sRNAs are also valuable tools for functional genomics studies. Usually, the sRNAs silence gene expression by either degrading mRNA or repressing translation but, in a few cases, they also generate a population of secondary siRNAs. The ta-siRNAs are secondary siRNAs that are produced by a miRNA-targeted trigger that bridges the pathways of miRNA and siRNA regulation. ta-siRNAs can regulate plant development, metabolism, and responses to biotic and abiotic stresses, and thus have received more attention in the recent decade [3–5].

During the biogenesis of ta-siRNA, a single-stranded RNA is transcribed from a ta-siRNA-producing locus (TAS) and then cleaved by a phase-initiator (a miRNA or, in some cases, a ta-siRNA). Then, RNA-dependent RNA polymerase 6 (RDR6)-dependent conversion of the resulting fragments into double-stranded RNA and its subsequent cleavage by dicer-like 4 (DCL4) at every ~21 nucleotide (nt) relative to the phase-initiator cleavage site generates ~21-nt phased sRNAs. Some of the phased sRNAs become ta-siRNAs by binding argonaute (AGO) proteins to direct a *trans*-cleavage of targeted mRNAs [5–7]. Plant TASs can be classified into at least eight families, based on initiator-dependence, sequence similarity, and target gene identity. *TAS1* and *TAS2* are targets of miR173 and their ta-siRNAs can target the pentatricopeptide repeat genes [8]. *TAS3* is a target of miR390 and its ta-siRNA can target the auxin response factor gene family [8]. The initiator of *TAS4* is miR828, and the *TAS4* ta-siRNA can target the MYB transcription factor gene family [9]. *TAS5* is triggered by miR482 and its ta-siRNA can target the Bs4 resistance gene [10]. miR156 and miR529 initiate *TAS6*, which targets an mRNA that encodes a zinc finger protein [11]. miR828 initiates *TAS7*, which can target 13 genes, including genes that encode the leucine-rich receptor protein kinase-like protein and a calcium-transporting ATPase [12]. *Atlg63130* is a pentatricopeptide repeat gene that was reported to be cleaved by *TAS2*-derived ta-siR2140 [13]. *TAS3* is flanked by two miR390 binding sites; one of which can be cleaved by the interaction of miR390 and AGO7, and another that is non-cleavable. Both binding sites are critical for the biogenesis of the *TAS3* ta-siRNAs. In contrast, other TASs have only a single miRNA binding site and are cleaved by the interaction of miRNA/ta-siRNA and AGO1. Recently, AGO2 has been reported to mediate *cis*-cleavage of *TAS1c*, although its slicer activity has not been demonstrated so far [14]. Taken together, it might be expected that many of the sequenced sRNAs could be mapped onto the phased positions set by the phase-initiator. Yet, while many of the sRNAs were successfully mapped, unexpectedly, many were mapped onto non-phased positions; that is, the intervals between the phased positions [12,13]. These sRNAs have been called “non-phased sRNAs” and how they are produced remains unclear.

The recent publication of the degradome library generated from cleaved mRNA fragments and sRNA libraries generated by high-throughput deep sequencing has enabled the study of all the cleavages that occur in a TAS transcript [15,16]. Here, we studied *cis*-cleavage of grapevine *TAS3* and found a cascading *cis*-cleavage, which produce sRNAs from the so-called non-phased positions and broaden the known scope of non-phased sRNA production.

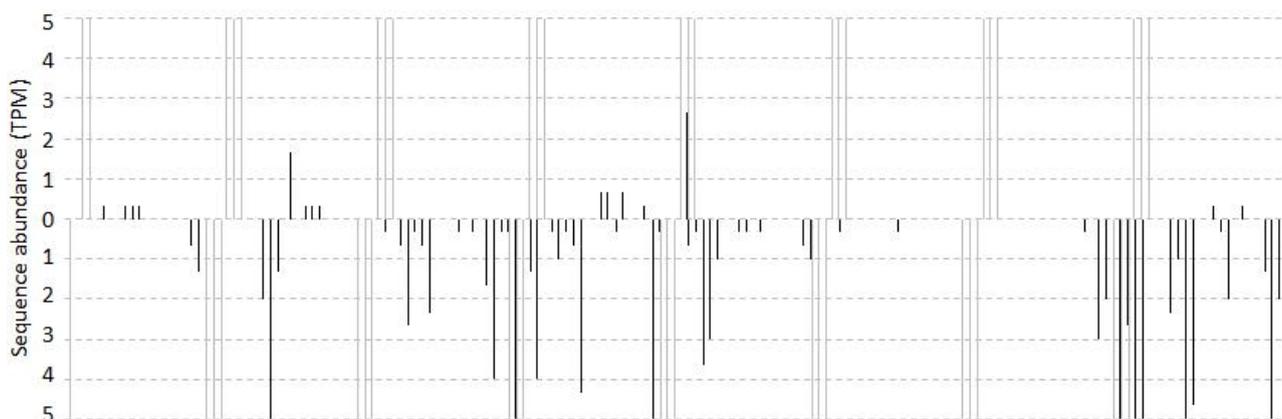
2. Results and Discussion

2.1. Overview of Small RNA Distribution on TAS3 from *Vitis vinifera*

Previously we reported that the *TAS3* from *Vitis vinifera* (*vviTAS3*) can be targeted by *vvi-miR390* to trigger ta-siRNA production in grapevine [12]. Here, to determine the distribution of sRNAs on the *vviTAS3* transcript, a sRNA library from grapevine leaves (GEO: GSM458927) was used. To improve mapping confidence, only the sRNAs that mapped to a single site on the whole *V. vinifera* genome were used, because they could be attributed with certainty to a particular locus.

As a result of the mapping, we detected 131 unique sRNAs, representing 3969 reads, which matched perfectly to *vviTAS3* (Figure 1). The 5' ends of the reads occupied 79 positions on the transcript. Only 14 (18%) of the positions were found to belong to phased positions set by *vvi-miR390* when a 1-nt offset from the phased positions was allowed. After filtering out sRNAs that had TPM (tags per million) values of five or less, some unique sRNAs that mapped to non-phased positions remained (Figure 1). The percentage of phased sRNA positions increased from 18% to 40%. Together, these results showed that some sRNAs are really generated from non-phased positions, and might even be functional because of the relatively high levels at which they are often expressed.

Figure 1. Abundance distribution of sRNAs mapped to *vviTAS3*. The number of reads with a 5' end at each position is plotted. Bars above the sequence represent sense reads; those below represent antisense reads. The cleavage site of *vvi-miR390* is marked by a vertical dotted line. Vertical gray lines indicate the miRNA-set 21-nt phased cleavage, allowing a 1-nt offset. Regions with TPMs greater than five are not shown.



2.2. Computational Prediction and Validation of cis-Cleavages

Recent reports have shown that many functional siRNAs belong to a class of 21–22-nt 5'U/A sRNAs [14]. Therefore, we filtered out the potential siRNAs from the mapped sRNAs by limiting the length of the reads to 21 nt and the 5' end to U/A. As a result, we detected 35 sRNAs that were mapped to the antisense strand that passed the rule.

Additionally, it has been reported that many functional cleaved positions tend to be conserved through evolution, and they have been found to be highly conserved in alignments of genomic sequences from different species [8]. To identify conserved positions in the *vviTAS3* transcript,

Table 1. Validated *cis*-cleaved positions on *vviTAS3* transcript and their ca-siRNAs.

<i>cis</i> -cleaved position	ca-siRNA	
	Name	Sequence
63	ca-siRM72	UGGAAAACGAAGAAGAAAUGC
85	ca-siRM94	UGGUAUAGAGUUCAUGACAAG
138	ca-siRM147	UGAGUUGGGCGGAAACGGGGA UGAGUUGGGCGGAAACGGGGA

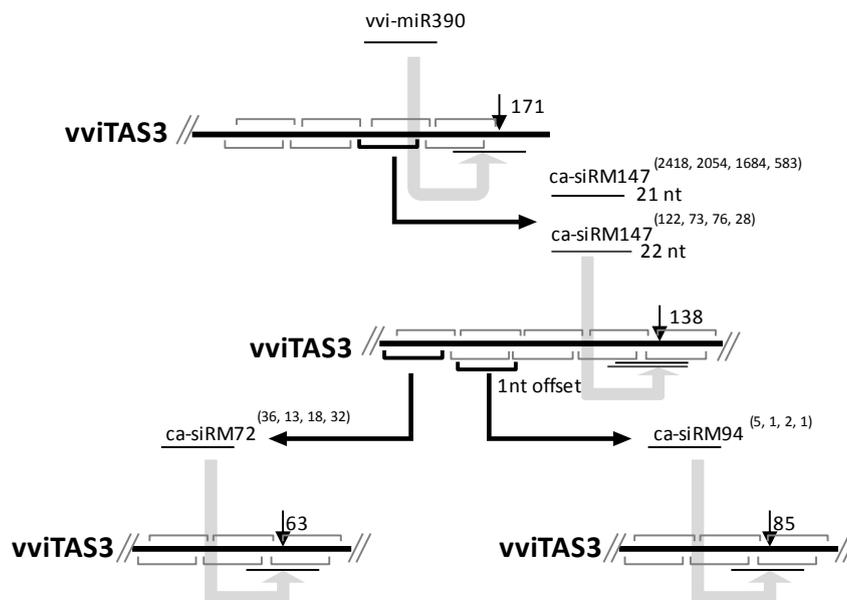
It has been demonstrated that two miR390 binding sites located on each side of *TAS3* are critical for *TAS3* ta-siRNAs biogenesis [8]. Therefore, we looked for ca-siRNA targeting sites on *vviTAS3* and the flanking 300 bp upstream and downstream of the gene where the two miR390 binding sites were located. We found one targeted site on *vviTAS3* for each of the ca-siRNAs. This finding suggested that *cis*-cleavage might use a different mechanism from the mechanism used by miR390 to initiate the cleavage of *vviTAS3*.

We used the same method and criteria to examine the antisense strand that was not targeted by miR390. Surprisingly, no ca-siRNA targeting sites were detected on the antisense strand of *vviTAS3*. The asymmetrical distribution between the targeted and non-targeted strand might imply that the non-targeted strand is readily in a double-stranded RNA form and is constantly processed by DCL4 and, therefore, protected from *cis*-cleavage. In addition, it might support the hypothetically biological function of *cis*-cleavage, *i.e.*, inactivating *TAS* transcription to feedback control ta-siRNA's production, as the sense strand was a template strand synthesizing antisense strand, so it would be more effective when the *cis*-cleavages preferred to occur in sense strand rather than in antisense strand.

2.3. Cascading *cis*-Cleavages

After identifying the *cis*-cleaved positions and their ca-siRNAs, we investigated how ca-siRNA production is triggered. It has been shown that ca-siRM147 can be triggered by miR390 [12], but for other ca-siRNAs the triggers remain unclear because they are out of the register set by miR390. To identify possible initiators, we investigated the distribution of *cis*-cleaved position and the locations of ca-siRNA 5' ends on the *vviTAS3* transcript. We found that the 5' end of ca-siRM72 occurred precisely at the register set by ca-siRM147 in which the cleaved site of ca-siRM147 located on sense strand was 65 nt away from the 5' end of ca-siRM72 on antisense strand. The 5' end of ca-siRM94 was offset by 1 nt from the register set by ca-siRM147. Because the cleavage of phasing sRNAs often occurs within 1–2 nt of the phased position [8,13], we propose that the production of ca-siRM147, ca-siRM72, and ca-siRM94 is triggered by miR390, ca-siRM147, and ca-siRM147, respectively. The cascading *cis*-cleavage that we have proposed is shown schematically in Figure 3.

Figure 3. Cleavage cascades generated by *cis*-cleavages of siRNA on the sense strand of *vviTAS3*. The four numbers in the brackets indicate the raw abundance of *ca*-siRNA in grapevine berries, leaves, inflorescences, and tendrils, respectively.



2.4. The Accumulated Levels of *ca*-siRNAs

When we analyzed the accumulated levels of the *ca*-siRNAs in the grapevine leaf, berry (GEO: GSM458930), inflorescence (GEO: GSM458929), and tendril (GEO: GSM458928) libraries, we found that the levels were in agreement with the cleavage cascades. The abundance of the *ca*-siRNAs that were located upstream of the cascade was always higher than the abundance of the *ca*-siRNAs that were downstream. For example, in the grapevine leaf library, the 21-nt *ca*-siRM147 located upstream had 2054 sequenced reads, while *ca*-siRM72, located downstream of *ca*-siRM147 cleavage, had only 13 sequenced reads. Moreover, the *ca*-siRNAs that occurred precisely at the register were always more abundant than those that occurred out of the register. For example, in the grapevine leaf library, *ca*-siRM72 had 13 sequenced reads, while *ca*-siRM94, which was shifted by 1 nt from the phased positions set by *ca*-siRM147, had only one sequenced read. Similar results were obtained for the other three tissues (Figure 3).

2.5. *cis*-Cleavages Produced sRNAs in Increments of Approximately 21 nt

To test whether or not *cis*-cleavages can also produce phased sRNAs in increments of approximately 21 nt, we searched for sRNAs with 5' ends that overlapped the predicted phased and non-phased positions by allowing an offset of 1 nt. As expected, each *cis*-cleavage had a set of corresponding phased sRNAs arranged in ~21-nt increments upstream and downstream of the cleavage position (Table 2).

Table 2. Phased patterns set by different *cis*-cleavages.

<i>cis</i> -cleaved position	Upstream of <i>cis</i> -cleaved position			Downstream of <i>cis</i> -cleaved position		
	Number of phased sRNAs	Number of non-phased sRNAs	<i>p</i> value *	Number of phased sRNAs	Number of non-phased sRNAs	<i>p</i> -value
63	2	12	1.11×10^{-1}	3	47	4.92×10^{-1}
85	5	27	7.54×10^{-3}	4	30	1.00×10^{-1}
138	8	41	4.71×10^{-4}	3	16	7.43×10^{-2}

* *p*-values less than 0.01 are in bold font.

To test whether or not the phased patterns produced from *cis*-cleavages were statistically significant, we developed an improved equation (see Experimental Section) by modifying previous algorithms [12,13,19] to evaluate the phasing pattern set by *cis*-cleavage. First, the new equation is not constrained by the previous 231-bp length requirement [12,13,19], but requires only a multiple of 21 nt, which provides a more accurate TAS evaluation, especially for TASs that are longer or shorter than 231 bp. Second, our equation uses a variable *s* to reflect the maximum offset from a phase position [12,19], making the evaluation more flexible. This equation could be applied to TAS identification in the future. Using our improved algorithm [12,13,19], we determined that two *cis*-cleavages had significant phasing patterns (*p*-value < 0.01). When the number of sRNAs located in the phased positions set by ca-siRNAs was counted, we found that 54 unique sRNA located in non-phased positions set by miRNA390 were included in the phasing patterns. These results suggested that some common processes might be used for both miRNA-mediated ta-siRNA production and the *cis*-cleavage of siRNA.

2.6. The Conservation of *ca*-siRNAs and Cascading *cis*-Cleavages

To examine the conservation of *cis*-cleavages further, we first looked for the presence of ca-siRNAs in the sRNA datasets of *V. vinifera*, *A. thaliana*, *M. domestica*, and *P. persica* downloaded from the Gene Expression Omnibus (GEO) or the plant MPSS databases. We found that although the accumulation levels of the four ca-siRNAs varied in the different species, they were expressed in all four species, except for 22-nt ca-siRNA147, which was not detected in *P. persica* (peach) (Table 3).

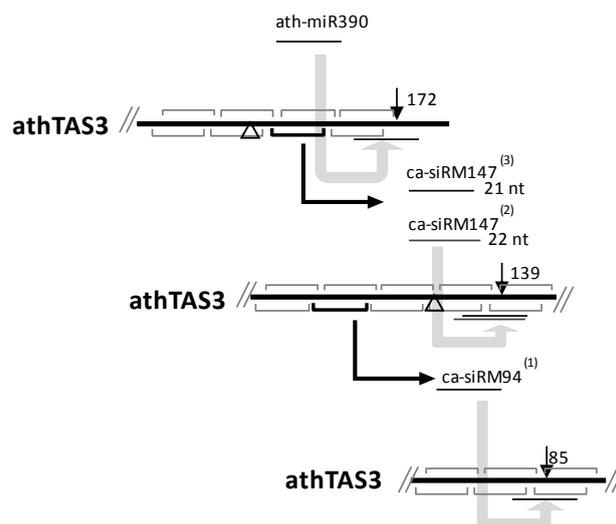
Table 3. Conservation of ca-siRNAs in the sRNA datasets of four species.

ca-siRNAs		Average of normalized abundance (TPM)			
Name	Length	Grapevine	Apple	Peach	Arabidopsis
ca-siRM72	21 nt	6	2	1	1
ca-siRM94	21 nt	1	3	14	1
ca-siRM147	21 nt	419	32	9	2
	22 nt	18	1	0	2

We then evaluated the corresponding *cis*-cleavages based on the Col7d samples (GEO: GSE20197) and the degradome library (GEO: GSM280227) from Arabidopsis, and found that, two *cis*-cleavages occurred in positions 85 and 139 (equivalent to position 138 in *vviTAS3* because of the nucleotide deletion in the *vviTAS3* sequence) were also validated on *athTAS3* (Figure 4). The accumulated levels of ca-siRNAs in Col7d sample were also in agreement with the cleavage cascades. In which, the 21-nt and

22-nt ca-siRM147 located upstream had, respectively, three and two sequenced reads, while ca-siRM94, located downstream of ca-siRM147 cleavage, had one sequenced reads. In a previous study [5], it was suggested that *cis*-cleavage occurred on position 139 in *athTAS3*, although the ca-siRNA and functional sRNAs were not found. Here, we detected the ca-siRNA and a secondary ca-siRNA product (ca-siRNA94), which we believe provides enough evidence to establish the *cis*-cleavage on *TAS3*. The findings reported here for *athTAS3* indicate that cascading *cis*-cleavage is conserved.

Figure 4. Cleavage cascades generated by *cis*-cleavages of siRNA on the sense strand of *athTAS3*. The triangle indicates the position of the nucleotide deletion in *vviTAS3*. The number in the bracket indicates the raw abundance of ca-siRNA in Col7d (GEO: GSE20197).



3. Experimental Section

3.1. Sources of sRNA Libraries

In this study, we used four deep sequencing sRNA datasets; namely, two degradome library and two sRNA libraries. All the datasets were downloaded from the Gene Expression Omnibus (GEO). The GEO accession numbers for these libraries are given in the Results section.

3.2. Evaluation of Phasing Patterns Set by *cis*-Cleavage

Once a *cis*-cleaved position was determined, the numbers of phased and non-phased positions were counted upstream and downstream of the cleavage sites respectively. Phased positions refer to positions arranged in 21-nt increments relative to the cleavage position as well as to positions shifted by s nt relative to the positions of 21-nt increments. Non-phased positions are all the other positions. The p -value of each detected phasing pattern was calculated based on a random hyper-geometric distribution using an improved equation based on previously used algorithms [12,13,19,20]:

$$P(k_1) = \sum_{x=k_1}^{\min(k_1+k_2, \frac{2L}{21}-1)} \frac{\binom{(2L-1) - (\frac{2L}{21}-1) \times (2s-1)}{k_2} \binom{\frac{2L}{21}-1}{k_1}}{\binom{(2L-1) - (\frac{2L}{21}-1) \times 2s}{k_1+k_2}} \quad (1)$$

Where L is the length of the detected pattern and is a multiple of 21, K_1 is the number of phased positions having sRNA hits, K_2 is the number of non-phased positions having sRNA hits, and s is the maximum allowed offset from the phase position.

3.3. Expressional Conservation of *ca*-siRNAs

The expressional conservation of the grapevine *ca*-siRNAs was investigated by performing a search against 84 sRNA libraries from grapevine, apple, Arabidopsis, and peach. The sRNA libraries of grapevine (GEO: GSE18405) and apple (GEO: GSE36065) were downloaded from the GEO and the sRNA libraries of Arabidopsis and peach were used from the MPSS databases [21]. The normalized abundance is the raw expression value divided by the total number of signatures and multiplied by 1,000,000.

4. Conclusions

In this work, we reexamined the distribution of sRNAs on *vviTAS3* using a stringent threshold that used only the sRNAs that mapped to a single site on the whole *V. vinifera* genome and that had normalized abundant values of one or more TPM. Our results showed that the non-phased positions were indeed located by some of the uniquely mapped sRNAs. We identified three *cis*-cleavages that directed by four *ca*-siRNAs at positions 63, 85, and 138 on *vviTAS3* by combining computational predictions and validation. We found that three *cis*-cleavages, together with their *ca*-siRNAs, formed a cascading *cis*-cleavage. The accumulated levels of four *ca*-siRNAs in the berry, leaf, inflorescence, and tendril libraries of *V. vinifera* also agreed with the cascade. A comparative analysis showed that the expression levels of the four *ca*-siRNAs were conserved among grapevine, apple, peach, and Arabidopsis, and part of the *cis*-cascade was also identified in Arabidopsis. We also found that sRNAs were located at the phased positions set by *ca*-siRNA. These results broaden the known scope of non-phased sRNA production. We also developed an improved equation by modifying previous algorithms to evaluate the phasing pattern set by *cis*-cleavage. It could be applied to TAS identification in the future.

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Conflict of Interest

The authors declare no conflict of interest.

References

1. Axtell, M.J. Classification and comparison of small RNAs from plants. *Annu. Rev. Plant Biol.* **2013**, *64*, 137–159.
2. Chen, X. Small RNAs and their roles in plant development. *Annu. Rev. Cell Dev. Biol.* **2009**, *25*, 21–44.
3. Peragine, A.; Yoshikawa, M.; Wu, G.; Albrecht, H.L.; Poethig, R.S. SGS3 and SGS2/SDE1/RDR6 are required for juvenile development and the production of trans-acting siRNAs in *Arabidopsis*. *Genes Dev.* **2004**, *18*, 2368–2379.
4. Vazquez, F.; Vaucheret, H.; Rajagopalan, R.; Lepers, C.; Gascioli, V.; Mallory, A.C.; Hilbert, J.L.; Bartel, D.P.; Crete, P. Endogenous trans-acting siRNAs regulate the accumulation of *Arabidopsis* mRNAs. *Mol. Cell* **2004**, *16*, 69–79.
5. Allen, E.; Xie, Z.; Gustafson, A.M.; Carrington, J.C. microRNA-directed phasing during trans-acting siRNA biogenesis in plants. *Cell* **2005**, *121*, 207–221.
6. Gascioli, V.; Mallory, A.C.; Bartel, D.P.; Vaucheret, H. Partially redundant functions of *Arabidopsis* DICER-like enzymes and a role for DCL4 in producing trans-acting siRNAs. *Curr. Biol.* **2005**, *15*, 1494–1500.
7. Yoshikawa, M.; Peragine, A.; Park, M.Y.; Poethig, R.S. A pathway for the biogenesis of trans-acting siRNAs in *Arabidopsis*. *Genes Dev.* **2005**, *19*, 2164–2175.
8. Axtell, M.J.; Jan, C.; Rajagopalan, R.; Bartel, D.P. A two-hit trigger for siRNA biogenesis in plants. *Cell* **2006**, *127*, 565–577.
9. Rajagopalan, R.; Vaucheret, H.; Trejo, J.; Bartel, D.P. A diverse and evolutionarily fluid set of microRNAs in *Arabidopsis thaliana*. *Genes Dev.* **2006**, *20*, 3407–3425.
10. Li, F.; Orban, R.; Baker, B. SoMART: A web server for plant miRNA, tasiRNA and target gene analysis. *Plant J.* **2012**, *70*, 891–901.
11. Arif, M.A.; Fattash, I.; Ma, Z.; Cho, S.H.; Beike, A.K.; Reski, R.; Axtell, M.J.; Frank, W. DICER-LIKE3 activity in *Physcomitrella patens* DICER-LIKE4 mutants causes severe developmental dysfunction and sterility. *Mol. Plant* **2012**, *5*, 1281–1294.
12. Zhang, C.; Li, G.; Wang, J.; Fang, J. Identification of trans-acting siRNAs and their regulatory cascades in grapevine. *Bioinformatics* **2012**, *28*, 2561–2568.
13. Chen, H.M.; Li, Y.H.; Wu, S.H. Bioinformatic prediction and experimental validation of a microRNA-directed tandem trans-acting siRNA cascade in *Arabidopsis*. *Proc. Natl. Acad. Sci. USA* **2007**, *104*, 3318–3323.
14. Rajeswaran, R.; Aregger, M.; Zvereva, A.S.; Borah, B.K.; Gubaeva, E.G.; Pooggin, M.M. Sequencing of RDR6-dependent double-stranded RNAs reveals novel features of plant siRNA biogenesis. *Nucleic Acids Res.* **2012**, *40*, 6241–6254.
15. Howell, M.D.; Fahlgren, N.; Chapman, E.J.; Cumbie, J.S.; Sullivan, C.M.; Givan, S.A.; Kasschau, K.D.; Carrington, J.C. Genome-wide analysis of the RNA-DEPENDENT RNA POLYMERASE6/DICER-LIKE4 pathway in *Arabidopsis* reveals dependency on miRNA- and tasiRNA-directed targeting. *Plant Cell* **2007**, *19*, 926–942.

16. German, M.A.; Pillay, M.; Jeong, D.H.; Hetawal, A.; Luo, S.; Janardhanan, P.; Kannan, V.; Rymarquis, L.A.; Nobuta, K.; German, R.; *et al.* Global identification of microRNA-target RNA pairs by parallel analysis of RNA ends. *Nat. Biotechnol.* **2008**, *26*, 941–946.
17. Larkin, M.A.; Blackshields, G.; Brown, N.P.; Chenna, R.; McGettigan, P.A.; McWilliam, H.; Valentin, F.; Wallace, I.M.; Wilm, A.; Lopez, R.; *et al.* Clustal W and Clustal X version 2.0. *Bioinformatics* **2007**, *23*, 2947–2948.
18. Pantaleo, V.; Szittyá, G.; Moxon, S.; Miozzi, L.; Moulton, V.; Dalmay, T.; Burgyan, J. Identification of grapevine microRNAs and their targets using high-throughput sequencing and degradome analysis. *Plant J. Cell Mol. Biol.* **2010**, *62*, 960–976.
19. Dai, X.; Zhao, P.X. pssRNAMiner: A plant short small RNA regulatory cascade analysis server. *Nucleic Acids Res.* **2008**, *36*, W114–W118.
20. Zhang, C.; Wang, J.; Hua, X.; Fang, J.; Zhu, H.; Gao, X. A mutation degree model for the identification of transcriptional regulatory elements. *BMC Bioinforma.* **2011**, *12*, doi:10.1186/1471-2105-12-262.
21. Nakano, M.; Nobuta, K.; Vemaraju, K.; Tej, S.S.; Skogen, J.W.; Meyers, B.C. Plant MPSS databases: Signature-based transcriptional resources for analyses of mRNA and small RNA. *Nucleic Acids Res.* **2006**, *34*, D731–D735.

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