

Supplementary Materials: A Different Microbiome Gene Repertoire in the Airways of Cystic Fibrosis Patients with Severe Lung Disease

S1. Library preparation and NGS sequencing

Qualified DNA samples were used to prepare sequence library with an insert size of 180 bp. First of all, purified DNA samples were sheared into smaller fragments by Nebulization technique. Then the overhangs resulting from fragmentation were converted into blunt ends by using T4 DNA polymerase, Klenow Fragment and T4 Polynucleotide Kinase. After adding an 'A' base to the 3' end of the blunt phosphorylated DNA fragments, adapters were ligated to the ends of the DNA fragments. After that, too short fragments were removed using Ampure beads. The qualified library was used for sequencing with 2G raw data output per sample and a length of 100bp per read. After raw data generation, low quality reads were removed. Data filtration was done by BGI custom scripts, and listed as follows:

1. removing reads with 3 N;
2. removing reads contaminated by adapter;
3. removing reads with a certain proportion of low quality bases (40% as default, parameter setting at 36 bp);
4. removing duplication contamination.

Finally, the clean data obtained were used for subsequent bioinformatic analysis. The library preparation procedures, as well as the sequencing pipeline described here were implemented and performed by the Beijing Genomics Institute (BGI, Shenzhen, Guangdong, China). Additional details about the bioinformatics pipeline used in this work including information about raw data control were reported in the main text (Material and Methods section) and in S2 Table.

S2. File to be uploaded to iPath site (<http://pathways.embl.de/>)

Supplementary Figure Legends and Tables

Figure S1. Metabolic and regulatory pathway distribution as obtained from HUMAnN analysis. Only pathways reporting a coverage values higher than 80% were reported.

Figure S2. Metabolic map drawn with iPath. Equally distributed pathways were reported in green, whereas pathways with a higher abundance in normal/mild and severe groups were reported in blue and red. In particular, pathways reporting higher values in patients belonging to the normal/mild group were reported in blue and pathways more represented in patients belonging to the severe group were reported in red. An interactive version of this plot can be generated using Supporting Information S2 directly from iPath site (<http://pathways.embl.de/>).

Figure S3. Regulatory distribution derived from the metabolic map as reported in S2 Fig. Different colors refer to different distributions as reported in S2 Fig legend. An interactive version of this plot can be generated using S2 Supporting Information directly from iPath site (<http://pathways.embl.de/>).

Figure S4. Contig length distribution. Boxplots were drawn reporting the interquartile range (IQR) between the 25th and the 75th percentile (first and third quartiles), whereas the inner line represents the median. Whiskers represent the lowest and highest values within 1.5 times IQR from the first and third quartiles Outliers were not reported.

Figure S5. Distribution patterns of transporter proteins as reported by TCDB. On the left part of the panel, the top 30 transporter categories were enlarged for easy reading. "m" and "s" correspond to "normal/mild" and "severe" groups.

Figure S6. Virulence factor distribution for each patient. Only factors displaying a significant (Student's t-test p-value ≤ 0.05) diverging distribution in the two groups of patients were reported.

Figure S7. Distribution patterns of virulence factors classified with MvirDB. The top 20 sub-categories were reported for each main-category.

Figure S8. Distribution of antibiotic resistance genes for each category detected at least in one sample. The percentage of resistance genes found has been reported for each patient.

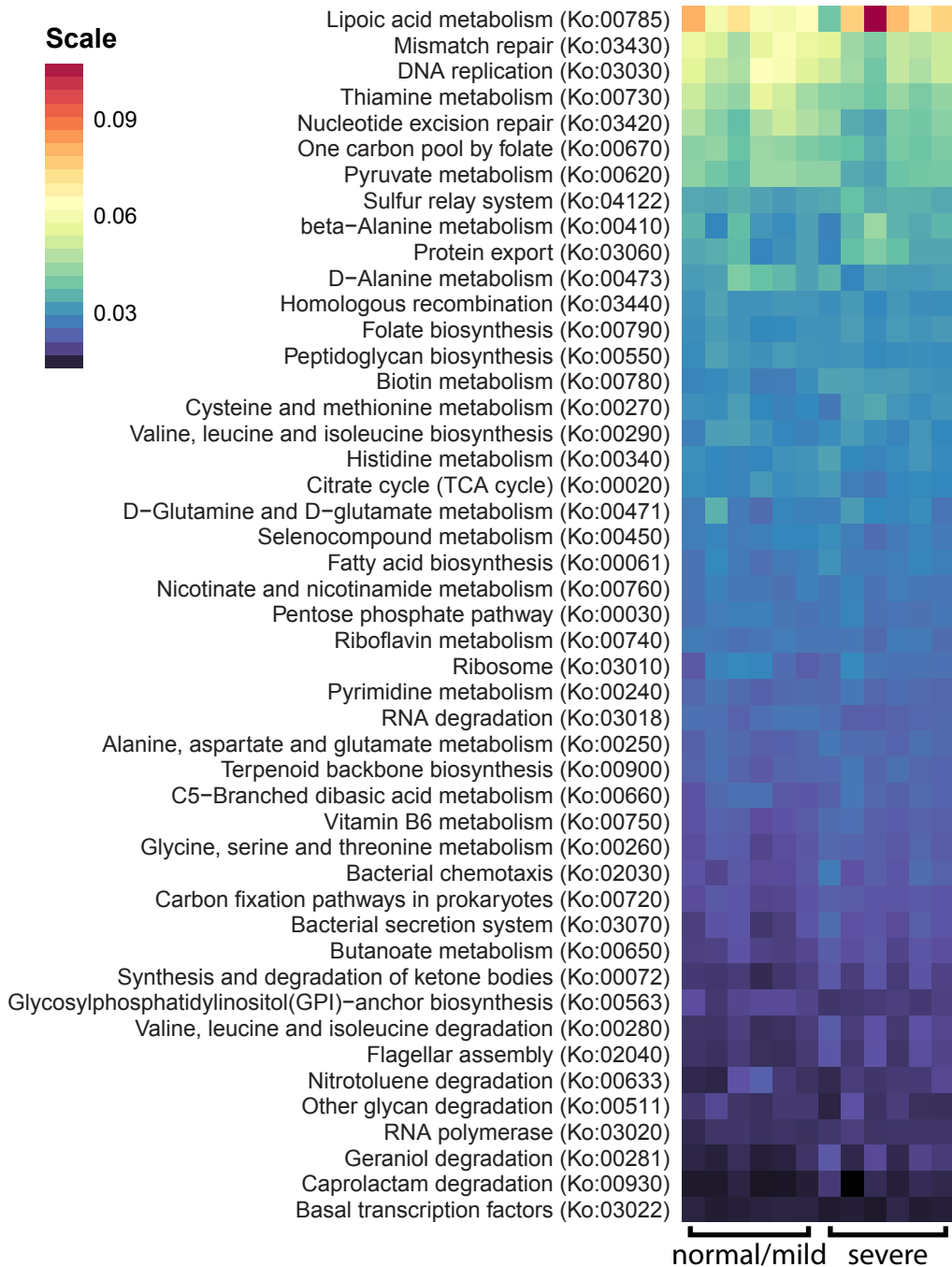
Figure S9. Top-20 taxa detected with both NGS and culture

Table S1. Culture-based diagnostic microbiology of sputum from patients with CF.

Table S2. Number of metagenomic sequences obtained and processed for each patient.

Table S3. Mantel test P values and R statistics obtained comparing all biotic features. In the right part of the table, the values of the R statistics were reported whereas the P values were reported in the left part. T=taxonomic; K=KEGG pathways; E=gene frequencies (eggNOG database); V=virulence factors (MvirDB database); A=antibiotic resistance genes (Resfam database).

Table S4. Variable loadings on the PCs across samples. The top-50% variables were reported in bold.



Glycan Biosynthesis and Metabolism

Nucleotide Metabolism

Metabolism of Cofactors and Vitamins

Biosynthesis of Other Secondary Metabolites

Lipid Metabolism

Carbohydrate Metabolism

Amino Acid Metabolism

Energy Metabolism

Metabolism of Other Amino Acid

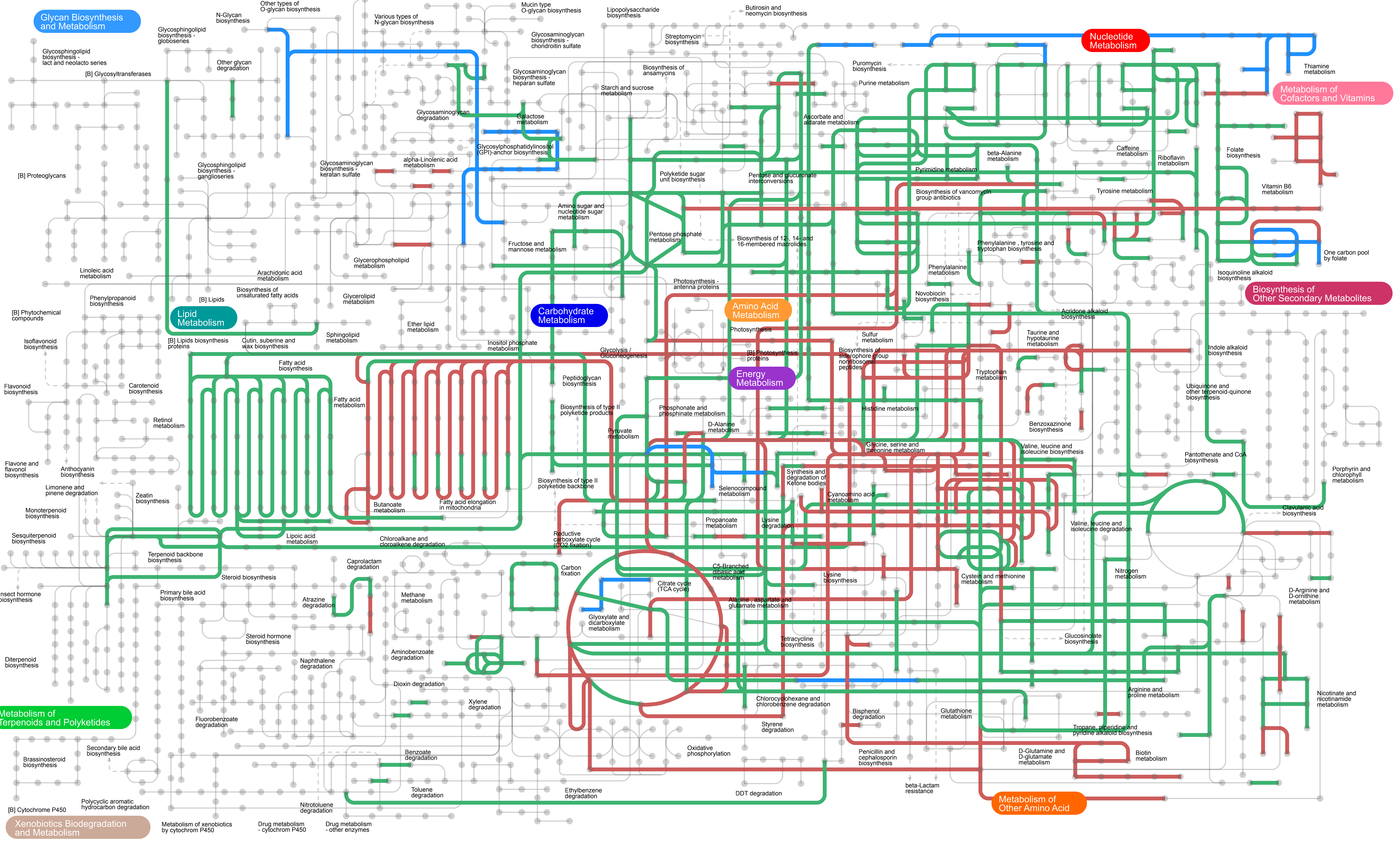
Metabolism of Terpenoids and Polyketides

Xenobiotics Biodegradation and Metabolism

Metabolism of xenobiotics by cytochrom P450

Drug metabolism - cytochrom P450

Drug metabolism - other enzymes



Transcription

RNA polymerase

DNA replication

Mismatch repair

The diagram illustrates the mismatch repair (MMR) process in a heteroduplex region of DNA. The top strand is the template (blue) and the bottom strand is the newly synthesized strand (red). A mismatch is present in the red strand. The MMR process involves identifying the mismatch, excising the segment, and resynthesizing the correct sequence.

Non-homologous end-joining

The diagram illustrates the non-homologous end-joining (NHEJ) process. A DNA double-strand break is shown, and the broken ends are recognized and ligated together, resulting in a deletion or insertion.

Homologous recombination

A 10x10 grid of green squares representing homologous recombination events. The grid shows a complex pattern of horizontal and vertical connections between columns, indicating recombination events across the genome.

Signal
Transduction

The diagram illustrates a two-component system with two parallel horizontal tracks. The left track contains several rectangular blocks of varying widths and heights, connected by horizontal lines. The right track also contains similar blocks and lines, with some blocks appearing to be stacked or overlapping. The overall structure suggests a complex interaction or assembly of components within a two-dimensional space.

[illegible]

Basal transcription factors

Folding, Sorting
and Degradation

Replication
and Repair

Cell Motility

Translation

Bacterial chemotaxis

Ribosome

Ubiquitin mediated proteolysis

Base excision repair

Nucleotide excision repair

The diagram illustrates the hierarchical assembly of a flagellum. At the base is the basal body, which is embedded in the cell envelope. Above the basal body is the hook, which is composed of flagellin subunits. The hook is connected to the flagellin filament, which is the long, whip-like structure that extends from the cell. The diagram shows the assembly of these components, with the basal body and hook being the most complex structures, and the filament being the simplest.

The diagram illustrates the steps of aminoacyl-tRNA biosynthesis. It shows a tRNA molecule (represented by a line with a loop) and an amino acid (represented by a circle). The steps are numbered 1 through 10, showing the process of aminoacylation and the release of pyrophosphate (PPi).

1. The tRNA molecule is shown with a loop.
2. The tRNA molecule is shown with a loop.
3. The tRNA molecule is shown with a loop.
4. The tRNA molecule is shown with a loop.
5. The tRNA molecule is shown with a loop.
6. The tRNA molecule is shown with a loop.
7. The tRNA molecule is shown with a loop.
8. The tRNA molecule is shown with a loop.
9. The tRNA molecule is shown with a loop.
10. The tRNA molecule is shown with a loop.

Protein export

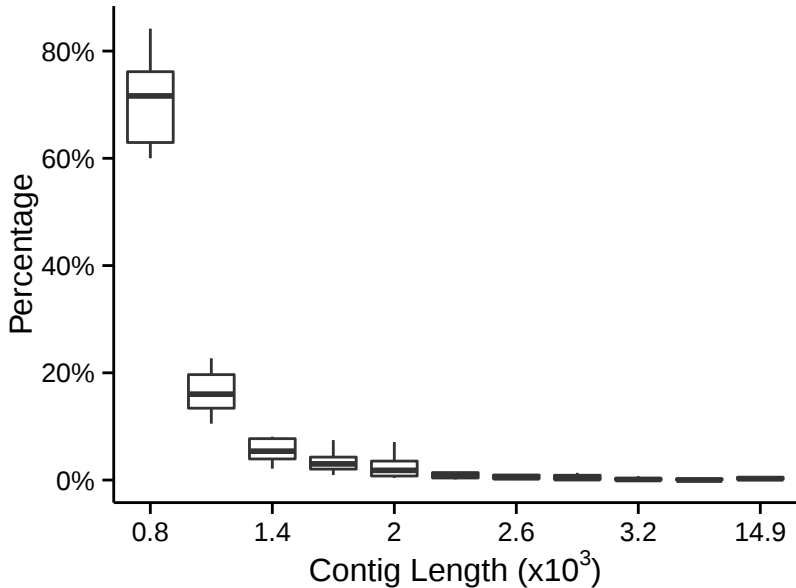
The diagram illustrates the cycle of an ABC transporter. It shows a membrane (indicated by a horizontal line) and a transport protein (represented by a vertical line with a grey oval at the top). The cycle is as follows:

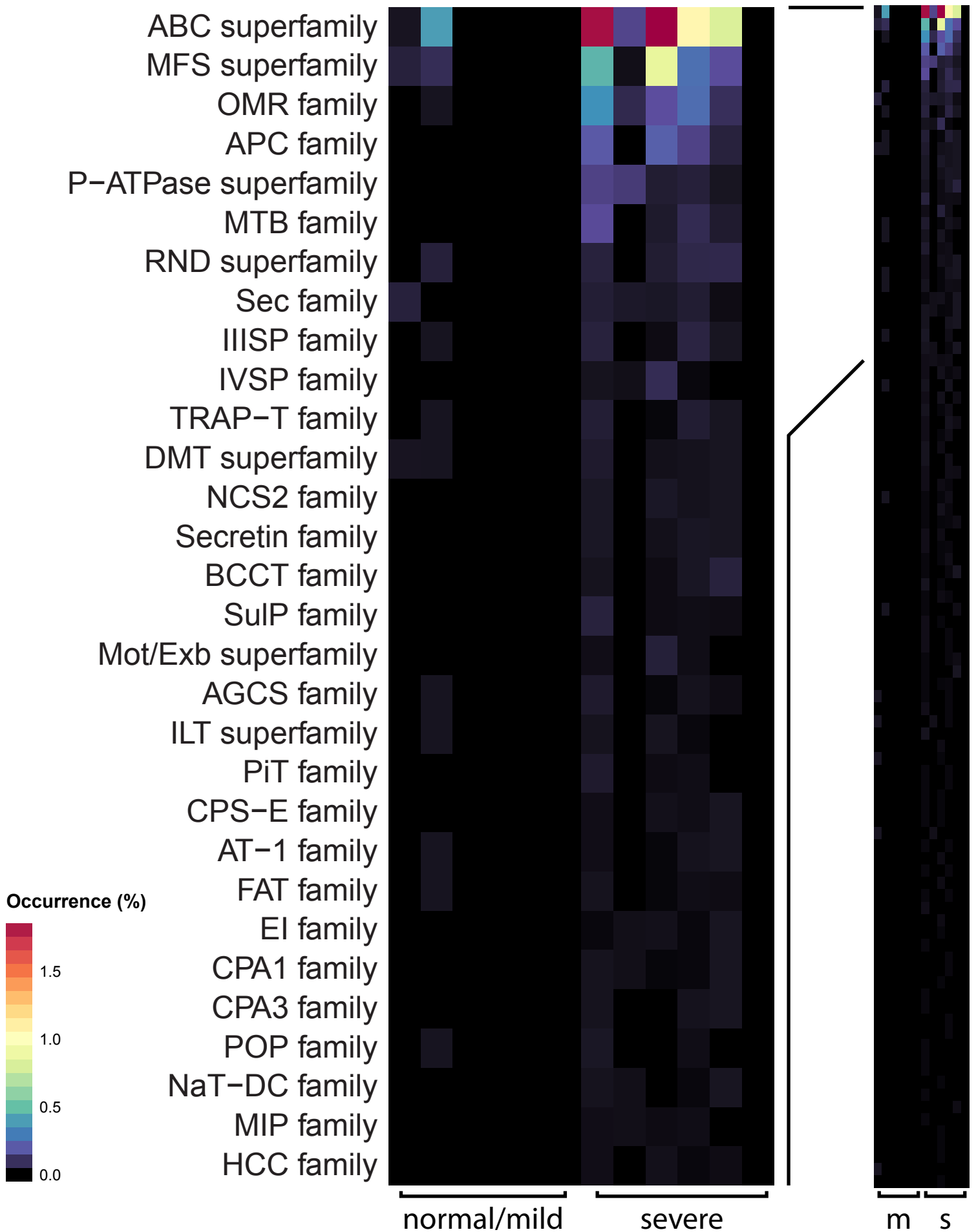
- ATP Binding:** ATP (represented by a grey oval) binds to the transporter protein.
- Substrate Binding:** The substrate (represented by a grey oval) binds to the transporter protein.
- ATP Hydrolysis:** ATP is hydrolyzed to ADP (represented by a grey oval) and inorganic phosphate (represented by a grey oval).
- Substrate Release:** The substrate is released into the extracellular space.
- ADP Release:** ADP is released from the transporter protein.
- ATP Binding:** The cycle repeats with a new ATP molecule binding to the transporter protein.

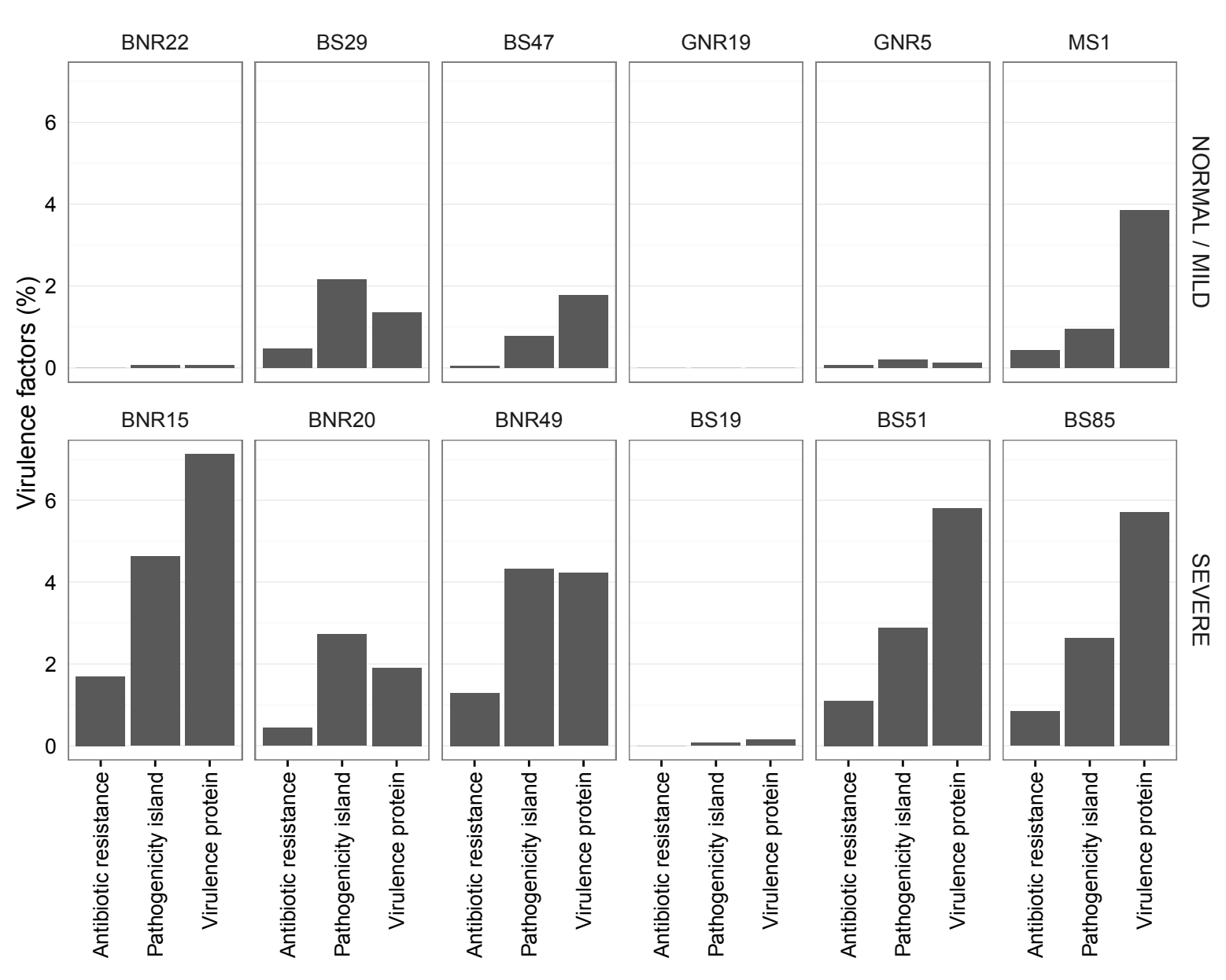
Bacterial secretion system

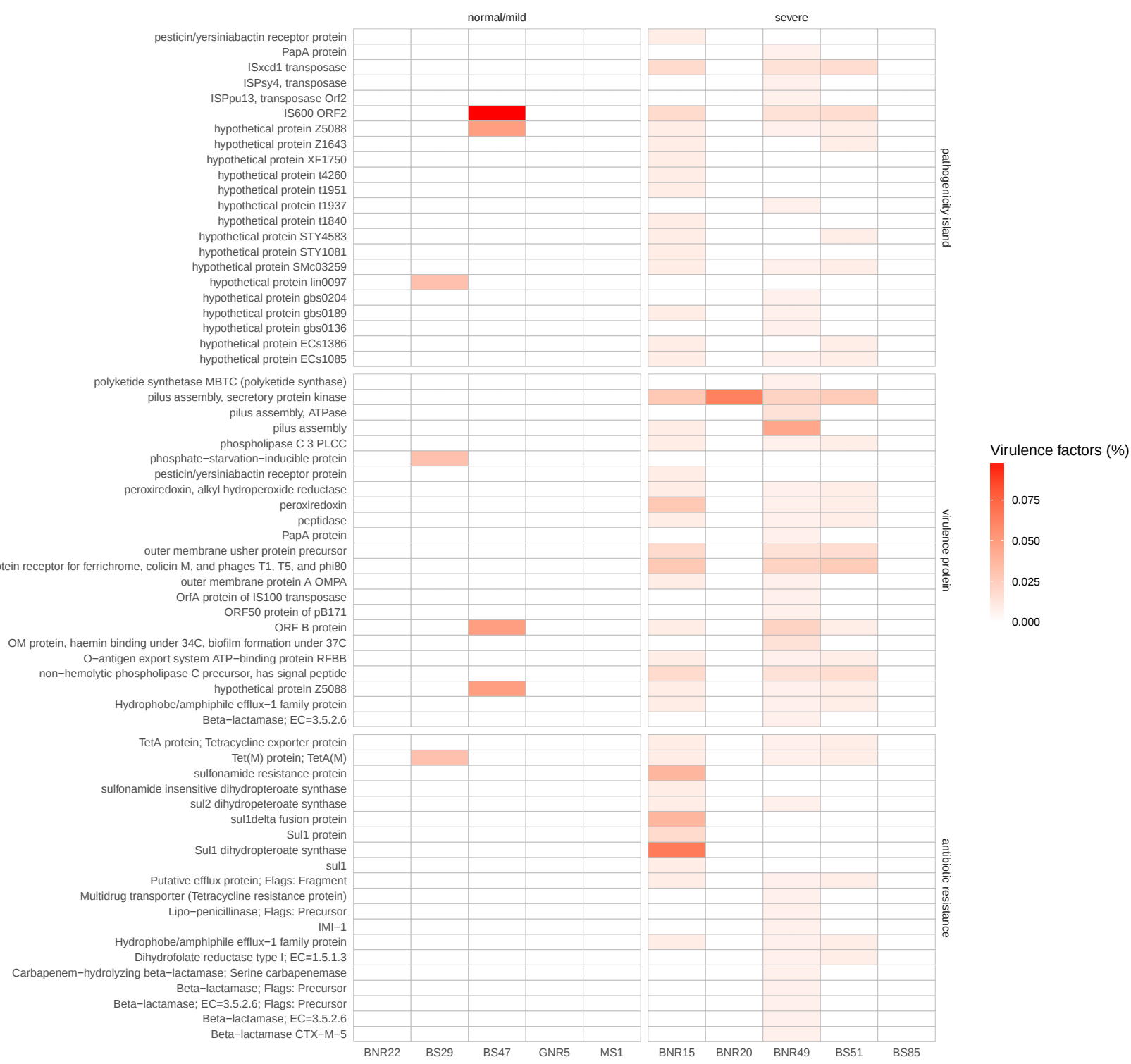
The diagram illustrates the Phosphotransferase system (PTS) pathway. It shows the transfer of a phosphate group from Phosphoenolpyruvate (PEP) to a sugar molecule (represented by a hexagon) through a series of phosphoryl carrier proteins. The proteins involved are PEP, HPr, HdcA, and HdcB. The phosphate group is transferred from PEP to HPr, then to HdcA, and finally to HdcB, which then transfers it to the sugar molecule. The diagram also shows the transfer of the phosphate group from HdcB back to PEP, completing the cycle.

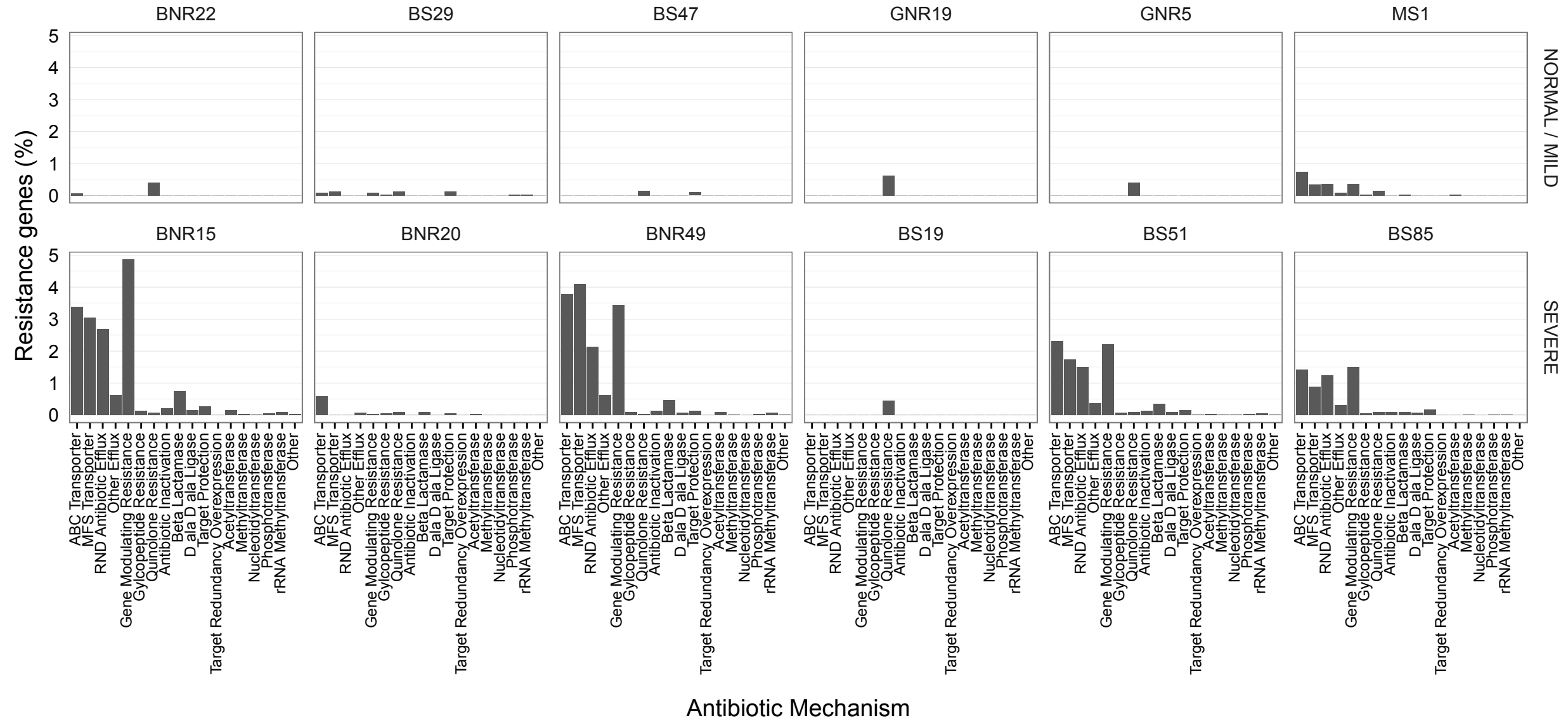
Membrane Transport











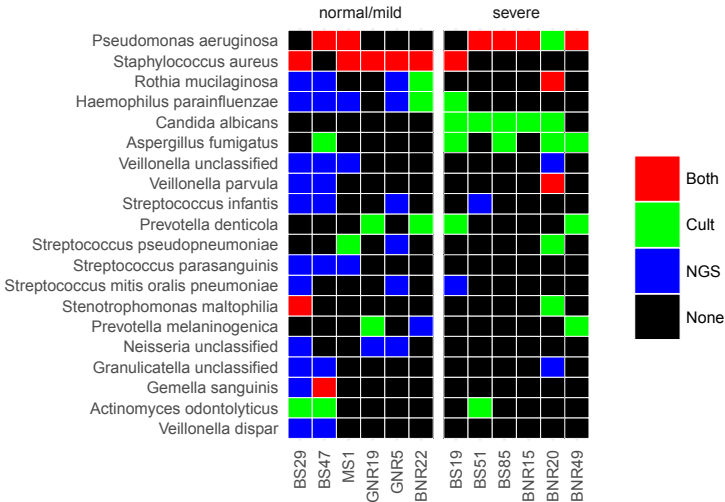


Table S1. Culture-based diagnostic microbiology of sputum from patients with CF.

Study ID	Lung disease status ¹	CF-related pathogens	<i>P. aeruginosa</i> / MSSA/MRSA colonization ²
BS29	normal/mild	<i>Stenotrophomonas maltophilia</i> , MSSA ³	Intermittent colonization with <i>P. aeruginosa</i> (from 2010)
BS47	normal/mild	<i>Pseudomonas aeruginosa</i>	Chronic colonization with multi-drug resistance (MDR) <i>P. aeruginosa</i> (before 2000)
MS1	normal/mild	<i>P. aeruginosa</i> (mucoid phenotype), <i>Staphylococcus aureus</i>	Chronic colonization with <i>P. aeruginosa</i> and intermittent colonization with <i>S. aureus</i> (from 2011)
GNR19	normal/mild	MSSA, <i>Ralstonia</i> spp.	Intermittent colonization with MSSA (before 2005)
GNR5	normal/mild	MSSA, <i>Pseudomonas</i> spp.	Intermittent colonization with MSSA (before 2005) and <i>P. aeruginosa</i> (from 2007)
BNR22	normal/mild	MSSA	Chronic colonization with MSSA (before 2000)
BS19	severe	MRSA ⁴ , MSSA	Chronic colonization with MRSA; (before 2000) Intermittent colonization with MSSA (from 2003)
BS51	severe	<i>P. aeruginosa</i> (mucoid phenotype), <i>Achromobacter xylosoxidans</i>	Chronic colonization with MDR <i>P. aeruginosa</i> (before 2000)
BS85	severe	<i>P. aeruginosa</i> (mucoid phenotype), <i>P. aeruginosa</i> (small phenotype)	Chronic colonization with MDR <i>P. aeruginosa</i> (before 2000)
BNR15	severe	MRSA, <i>P. aeruginosa</i> (mucoid phenotype), <i>P. aeruginosa</i> (frayed phenotype), <i>P. aeruginosa</i> (small colony), <i>P. aeruginosa</i> (round phenotype)	Chronic colonization with MDR <i>P. aeruginosa</i> (before 2000) and MRSA (before 2000)
BNR20	severe	<i>P. aeruginosa</i> (mucoid phenotype), <i>P. aeruginosa</i> (punctiform phenotype), <i>P. aeruginosa</i> (frayed phenotype), <i>S. maltophilia</i>	Chronic colonization with MDR <i>P. aeruginosa</i> (from 2011) and <i>S. maltophilia</i> (from 2003)
BNR49	severe	<i>P. aeruginosa</i> (mucoid phenotype), <i>P. aeruginosa</i> (frayed phenotype)	Chronic colonization with MDR <i>P. aeruginosa</i> (from 2010)

¹ Normal/mild, FEV₁ > 70%; severe, FEV₁ < 40%.

² In parentheses, the year of first acquisition of CF pathogen.

³ MSSA, methicillin-susceptible *Staphylococcus aureus*

⁴ MRSA, methicillin-resistant *Staphylococcus aureus*

Table S2. Number of metagenomic sequences obtained and processed for each patient.

Sample ID	Lung disease status ¹	# of reads	bacterial reads, %	# of bacterial reads	# of contigs	# of contigs, $\geq 500\text{bp}$	filtered contigs, %	N50	coverage (avg. $\pm$ st. error)	# of ORF
BS29	normal/ mild	15005000	2.87	430546	19297	959	4.97	697	3.81 ± 0.63	3153
BS47	normal/ mild	15005000	2.89	433770	12778	420	3.29	714	14.01 ± 3.42	2035
MS1	normal/ mild	15005000	2.62	393657	18859	800	4.24	652	3.2 ± 0.36	3252
GNR19	normal/ mild	15005000	1.48	222641	4532	252	5.56	741	11.83 ± 2.71	1127
GNR5	normal/ mild	15005000	2.1	315463	8356	305	3.65	817	13.77 ± 2.71	1487
BNR22	normal/ mild	15005000	2.4	360642	8895	329	3.7	797	6.07 ± 0.92	1471
BS19	severe	15005000	3.36	503666	7440	271	3.64	908	11.91 ± 2.19	1352
BS51	severe	15005000	3.53	529435	23062	4223	18.31	806	2.45 ± 0.21	11326
BS85	severe	15005000	2.74	411701	14291	1922	13.45	686	2.48 ± 0.33	6170
BNR15	severe	15005000	2.87	430430	12245	4085	33.36	1655	2.46 ± 0.19	10710
BNR20	severe	15005000	4.55	682833	17545	995	5.67	645	4.04 ± 0.61	4774
BNR49	severe	15005000	4.87	731105	52370	5302	10.12	1199	2.91 ± 0.16	13318

Table S3. Mantel test P values and R statistics obtained comparing all biotic features. In the right part of the table, the values of the R statistics were reported whereas the P values were reported in the left part. T=taxonomic; K=KEGG pathways; E=gene frequencies (eggNOG database); V=virulence factors (MvirDB database); A=antibiotic resistance genes (Resfam database).

P value	R statistic					
		T	K	E	V	A
	T		0.285	0.571	0.510	0.345
	K	0.026		0.159	0.175	0.219
	E	0.003	0.163		0.873	0.619
	V	0.001	0.169	0.001		0.478
	A	0.008	0.025	0.001	0.002	

Table S4. Variable loadings on the PCs across samples. The top-50% variables were reported in bold.

Antibiotic Resistance	A1	A2				
exp*	0.808	0.072				
ABC Transporter	0.248	-0.126				
Acetyltransferase	0.237	-0.220				
Antibiotic Inactivation	0.259	0.024				
Beta Lactamase	0.256	-0.049				
D ala D ala Ligase	0.254	0.109				
Gene Modulating Resistance	0.259	-0.107				
Glycopeptide Resistance	0.254	-0.026				
Methyltransferase	0.252	0.025				
MFS Transporter	0.237	-0.210				
Nucleotidyltransferase	0.207	0.471				
Other	0.238	0.01				
Other Efflux	0.251	-0.149				
Phosphotransferase	0.234	0.131				
Quinolone Resistance	-0.163	0.075				
RND Antibiotic Efflux	0.256	-0.088				
rRNA Methyltransferase	0.254	0.024				
Target Protection	0.229	0.043				
Target Redundancy Overexpression	0.086	0.766				
Functional Categories	F1	F2	F3	F4	F5	F6
exp*	0.417	0.146	0.127	0.081	0.075	0.065
Translation ribosomal structure and biogenesis	-0.068	-0.078	0.512	-0.262	-0.138	-0.016
RNA processing and modification	-0.192	0.056	-0.032	-0.024	0.09	0.617
Transcription	-0.237	-0.186	-0.072	0.331	0.222	0.110
Replication recombination and repair	0.288	-0.095	0.172	0.145	-0.039	-0.110
Chromatin structure and dynamics	-0.161	0.166	-0.256	-0.263	0.062	-0.462
Cell cycle control cell division chromosome partitioning	-0.132	-0.222	0.313	0.099	-0.391	-0.145
Defense mechanisms	-0.193	-0.009	0.234	-0.228	0.129	0.200
Signal transduction mechanisms	-0.217	0.132	0.09	0.474	-0.136	0.165
Cell wall membrane envelope biogenesis	0.314	0.117	-0.053	-0.036	0.075	0.024
Cell motility	-0.297	0.007	0.051	0.159	-0.096	-0.123
Cytoskeleton	-0.09	-0.054	-0.075	0.146	0.527	-0.085
Extracellular structures	-0.063	-0.300	0.208	0.277	0.350	-0.327
Intracellular trafficking secretion and vesicular transport	-0.191	-0.372	-0.119	-0.198	0.019	-0.120
Posttranslational modification protein turnover chaperones	-0.310	0.021	0.003	-0.068	-0.144	-0.152
Energy production and conversion	-0.219	0.293	-0.265	-0.09	0.053	-0.124
Carbohydrate transport and metabolism	-0.029	-0.468	-0.276	-0.074	0.029	0.022
Amino acid transport and metabolism	-0.189	0.289	0.195	-0.307	0.230	0.068
Nucleotide transport and metabolism	0.038	-0.273	-0.433	-0.093	-0.250	0.219
Coenzyme transport and metabolism	-0.298	-0.101	0.027	-0.230	0.014	-0.063
Lipid transport and metabolism	-0.311	-0.045	0.026	0.062	0.147	0.163
Inorganic ion transport and metabolism	-0.273	-0.024	-0.097	0.047	-0.358	-0.014

Secondary metabolites biosynthesis transport and catabolism	-0.123	0.364	-0.159	0.318	-0.157	-0.164
Metabolic Pathways	M1	M2	M3	M4	M5	
exp*	0.397	0.224	0.185	0.093	0.039	
Glycosylphosphatidylinositol GPI anchor biosynthesis	0.194	-0.040	-0.140	0.008	-0.027	
Alanine aspartate and glutamate metabolism	-0.181	0.122	0.144	-0.045	0.099	
Synthesis and degradation of ketone bodies	-0.171	0.188	-0.052	0.068	0.061	
Nicotinate and nicotinamide metabolism	-0.064	-0.113	0.239	0.015	-0.153	
Other glycan degradation	-0.008	-0.257	0.153	0.128	0.022	
Selenocompound metabolism	0.068	0.164	0.258	-0.056	-0.036	
Glycine serine and threonine metabolism	-0.214	0.066	0.103	-0.005	0.004	
Homologous recombination	0.064	-0.144	0.275	0.042	0.101	
Vitamin B6 metabolism	-0.199	0.021	0.152	-0.029	-0.093	
Fatty acid biosynthesis	-0.119	0.172	0.191	0.022	0.132	
Bacterial secretion system	-0.173	0.177	0.084	-0.042	0.023	
Pyruvate metabolism	0.192	0.156	0.073	-0.021	-0.056	
Cysteine and methionine metabolism	-0.121	-0.207	-0.168	0.049	0.027	
Riboflavin metabolism	0.111	0.003	0.197	0.121	-0.189	
Protein export	-0.143	-0.186	-0.103	0.159	0.095	
Nitrotoluene degradation	0.056	-0.089	-0.133	-0.350	-0.250	
Thiamine metabolism	0.223	-0.025	0.002	-0.075	-0.017	
RNA degradation	0.17	0.136	0.115	0.061	0.139	
Geraniol degradation	-0.150	0.224	-0.068	-0.013	-0.012	
Ribosome	-0.068	-0.188	0.043	-0.336	0.026	
Carbon fixation pathways in prokaryotes	-0.200	0.064	0.146	0.021	-0.043	
Valine leucine and isoleucine biosynthesis	-0.098	-0.122	0.092	-0.324	0.192	
beta Alanine metabolism	-0.109	-0.116	-0.258	0.098	0.009	
Folate biosynthesis	-0.123	-0.099	0.174	0.109	-0.198	
Citrate cycle TCA cycle	0.147	0.188	0.101	-0.097	-0.166	
Histidine metabolism	0.008	0.262	0.129	0.093	-0.157	
Lipoic acid metabolism	-0.071	-0.156	-0.241	0.172	0.071	
Peptidoglycan biosynthesis	0.045	-0.052	0.258	-0.134	0.342	
Butanoate metabolism	-0.184	0.147	-0.062	-0.138	-0.01	
DNA replication	0.217	0.053	0.051	-0.074	-0.121	
Bacterial chemotaxis	-0.122	0.251	-0.034	-0.036	-0.132	
Biotin metabolism	-0.209	0.033	0.042	0.05	-0.223	
Pentose phosphate pathway	-0.008	-0.139	0.146	-0.335	-0.120	
Sulfur relay system	-0.162	-0.127	0.025	0.009	-0.384	
Basal transcription factors	0.168	-0.071	0.017	0.136	-0.340	
C5 Branched dibasic acid metabolism	-0.106	-0.142	0.025	-0.344	-0.083	
RNA polymerase	-0.153	-0.075	0.133	-0.150	-0.168	
Mismatch repair	0.197	0.117	0.108	0.061	0.009	
Terpenoid backbone biosynthesis	-0.125	-0.113	0.211	0.184	-0.07	
Flagellar assembly	-0.167	0.193	-0.088	-0.034	0.019	
D Glutamine and D glutamate metabolism	-0.093	-0.105	0.218	0.132	0.314	
D Alanine metabolism	0.086	0.061	-0.06	-0.336	0.172	
Valine leucine and isoleucine degradation	-0.172	0.201	-0.042	-0.032	-0.024	

Pyrimidine metabolism	-0.095	-0.149	0.244	0.120	0.051
Nucleotide excision repair	0.206	0.119	0.07	0.044	-0.003
One carbon pool by folate	0.201	-0.005	0.135	-0.026	0.024
Caprolactam degradation	-0.096	0.240	-0.092	-0.031	0.096

Virulence Factors	V1
exp*	0.852
Differential gene regulation	-0.446
Pathogenicity island	-0.416
Protein toxin	-0.447
Transcription factor	-0.448
Virulence protein	-0.478

* Proportion of variance explained