



Article

Synthesis and Transformation of (-)-Isopulegol-Based Chiral β -Aminolactones and β -Aminoamides

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Abstract: A library of isopulegol-based β -amino acid derivatives has been developed from commercially-available (-)-isopulegol. Michael addition of primary and secondary amines towards α , β -unsaturated γ -lactones was accomplished resulting in β -aminolactones in highly-stereoselective reactions. Ring-opening of β -aminolactones with different amines furnished excellent yields of β -aminoamides. Moreover, the applicability of aminolactones in peptide synthesis was examined by opening the lactone ring with α - and β -aminoesters, providing dipeptides as promising chiral substrates for the synthesis of foldamers. The antiproliferative activities of β -aminolactones and β -aminoamides were explored, and the structure-activity relationships were studied from the aspects of the stereochemistry of the monoterpene ring and the substituent effects on the β -aminoamide ring system. The N-unsubstituted (-)-isopulegol-based β -aminoamides exhibited considerable antiproliferative activity against a panel of human adherent cancer cell lines (HeLa, MCF7 and MDA-MB-231).

Keywords: terpenoid; β-aminolactones; β-aminoamides; dipeptide; antiproliferative activity

1. Introduction

Sesquiterpene lactones containing the α -methylene- γ -lactone moiety are natural products occurring in many plant families. These compounds are known for their various biological activities, including cytotoxicity to tumor cells, anti-bacterial, antifungal, and anti-protozoan activities, as well as activity against human and animal parasites or inhibition of plant growth [1–3].

Conjugate addition of nucleophiles to α -methylene- γ -lactones provides β -aminolactones, which increase the proportion of cells in the G2/M and S phase [3] and serve as water-soluble derivatives that might retain cytotoxicity through a prodrug mechanism [4]. Additionally, the transformation of β -aminolactones, formally β -amino esters, to their derivatives such as 1,3-aminalcohols, proved to use those chiral auxiliaries in the enantioselective synthesis of secondary alcohols or other pharmacons, e.g., esomeprazole [5–9]. Besides their value in enantioselective catalysis, 1,3-aminoalcohols are also excellent building blocks for the synthesis of various heterocyclic ring systems, such as 1,3-oxazines, 1,3-thiazines or 1,4-oxazepams [10,11]. 2-Imino-1,3-thiazines and 2-iminothiazolidines can be found as structural units in biologically-relevant compounds, including antifungal and antimicrobial agents [12], BACE1 inhibitors [13], or cannabinoid receptor agonists [14–16].

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In addition, ring-opening of β -aminolactones with different amines may provide β -aminoamides, which are well-known subunits of biologically-important compounds such as bestatin, a potent aminopeptidase B. Its usefulness in the treatment of cancer through its ability to enhance the cytotoxic activity of known antitumor agents is well-known [17,18]. β -Aminoamides exhibit other biological activities as well, such as antidiabetic [19], HIV-protease, or renin inhibitor effects [20]. Besides interest in the synthesis of β -aminoamides, the opening of β -aminolactones with β -aminoesters is a useful method for the synthesis of dipeptides containing β -alanine moiety. β -Alanine is a precursor of the antioxidant dipeptide carnosine (β -alanine-L-histidine), which is thought to increase cell viability via an anti-senescence mechanism [21]. β -Ala-Gln has been applied in medical fields as a component of patient infusions [22]. Furthermore, β -alanine transporters were found to be highly upregulated in antibody-producing cell lines, indicating the cell's requirement for this amino acid [21].

Herein, our aim was to develop a library of monoterpene-based β -aminolactones and β -aminoamides by applying commercially-available natural (-)-isopulegol as an inexpensive chiral source, and to study their antiproliferative activity on multiple cancer cell lines. Moreover, we also report the synthesis of (-)-isopulegol-based dipeptides, which might serve as promising chiral substrates for the synthesis of chiral foldamers.

2. Results

2.1. Synthesis of α -Methylene- γ -Butyrolactones

The key intermediate (+)- α -methylene- γ -butyrolactone **2** was prepared from commercially-available (-)-isopulegol **1** with regioselective hydroxylation, followed by two-step oxidation and ring closure of the obtained γ -hydroxy-substituted α , β -unsaturated carboxylic acid by applying literature methods [23–28] (Figure 1). The diastereoisomeric (-)- α -methylene- γ -butyrolactone **4** was prepared by starting similarly from (-)-isopulegol **1**. In the first step, the hydroxy group of **1** was oxidized, followed by stereoselective reduction of the resulting carbonyl group providing (+)-neoisopulegol **3** [23–29] (Figure 1).

Figure 1. Synthesis of (-)-isopulegol-based α -methylene- γ -butyrolactones **2** and **4**.

2.2. Synthesis of β -Aminolactones

Nucleophilic addition of primary and secondary amines to α -methylene- γ -butyrolactones 2 and 4 has proven to be an efficient method for the preparation of a highly-diversified library of β -aminolactones [3,30]. When the addition of one equivalent of benzylamine to 2 was performed as a model reaction, the formation of N-benzyl aminolactone 5a and N-benzyl methylene amide 5b (the latter could not be isolated in pure form) was observed. The effect of the solvent was also studied, and it was found that the applied solvent strongly affected the yield of 5a and the ratio of the two products (Scheme 1, Table 1).

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Scheme 1. (i) Benzylamine (1 equiv.), dry EtOH, 25 °C, 20 h.

When alcohols as protic solvents were used, formation of **5a** was observed as the main product. Among of three protic solvents applied, EtOH gave target **5a** with the best chemoselectivity (entry 7). The ratio of **5a** and **5b** also depended on temperature. In alcohols, in turn, product ratios were similar at low (0 °C) and high (25 °C) temperature (compare entries 6 and 7). Furthermore, the yield of **5a** increased with temperature. At higher temperatures, however, the yield of **5a** dropped, and the products were formed in a ratio of 4:1, even with decreasing reaction time (Table 1).

Entry	Solvent	Temperature (°C)	Reaction Time (h)	Products Ratios (%)		Yield of 5a (%)
	Solvent		reaction time (ii)	5a	5b	11014 01 04 (70)
1	DCM	0	20	44	56	10
2	-	0	20	50	50	30
3	DMF	0	20	67	33	44
4	MeOH	0	20	92	8	30
5	<i>i</i> -PrOH	0	20	94	6	55
6	EtOH	0	20	94	6	60
7	EtOH	25	20	94	6	65
8	EtOH	40	15	80	20	55
9	EtOH	50	10	80	20	55
10	EtOH	60	7	80	20	50
11	EtOH	70	5	80	20	45

Table 1. Nucleophilic addition reaction of **2** with benzylamine.

After optimizing the condition for nucleophilic addition with benzylamine, amine adducts 6–10 were synthesized from 2 under these conditions (one equivalent of appropriate amine, EtOH, 25 °C) (Scheme 2). Surprisingly, when (R)-and (S)- α -methylbenzylamine and secondary amines were applied, only the formation of aminolactones was observed (Table 2). This is probably due to the steric hindrance of these amines. Besides amines, the best conditions were also successful for the addition of L- or β -aminoesters as amine sources to prepare some β -aminolactones containing aminoester moiety 11–12 (Table 2).

Scheme 2. (i) R¹R²NH (1 equiv.), dry EtOH, 25 °C, 20 h, for 6–10; (ii) aminoester. HCl (2 equiv.), Et₃N (2 equiv.), dry EtOH, 25 °C, 20 h, for 11 and 12.

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Entry	Compound	\mathbb{R}^1	R ²	Yield (%)
1	6	Н	(R)-α-Methylbenzyl	75
2	7	Н	(S)- α -Methylbenzyl	71
3	8	C_2H_5	C_2H_5	50
4	9		-(CH ₂) ₅ -	47
5	10	Benzyl	Benzyl	59
6	11	Н	β -Alanine ethyl ester	60
7	12	Н	L-Alanine ethyl ester	40

Table 2. Nucleophilic addition reaction of amines with 2.

The optimized conditions were also applied for the preparation of (+)-neoisopulegol-based β -aminolactones 13–18 starting from 4 (Scheme 3). Interestingly, under the applied conditions, exclusive formation of the amine adducts was observed. This may be due to the *cis* configuration of 4, which makes the lactone more hindered for nucleophilc attack (Table 3). The reaction of 4 with some aminoesters was effective at an elevated temperature to achieve aminoester-based β -aminolactone derivatives 19–20 (Table 3).

Scheme 3. (i) R^1R^2NH (1 equiv.), dry EtOH, 25 °C, 20 h for 13–18; (ii) aminoester. HCl (2 equiv.), Et₃N (2 equiv.), dry EtOH, 70 °C, 20 h, for 19 and 20.

Table 3. Nucleophilic addition of amines with 4.

Entry	Compo	und R ¹	R ²	Temperature (°C)	Yield (%)
1	13	Н	Benzyl	25	60
2	14	Н	(R)-α-Methylbenzyl	25	65
3	15	Н	(S)- α -Methylbenzyl	25	70
4	16	C_2H_5	C_2H_5	25	50
5	17	-	-(CH ₂) ₅ -	25	53
6	18	Benzyl	Benzyl	25	50
7	19	Н	β -Alanine ethyl ester	70	60
8	20	Н	L-Alanine ethyl ester	70	44

The relative configuration of compounds 5a–12 and 13–20 was determined by means of NOESY experiments. Clear NOE signals were observed between the H-1 and H-3, as well as the H-3 and H-7 protons in the case of 5a–12, while significant NOE signals were shown between the H-3 and H-7, as well as the H-4 and H-7 protons in the case of 13–20 (Figure 2).

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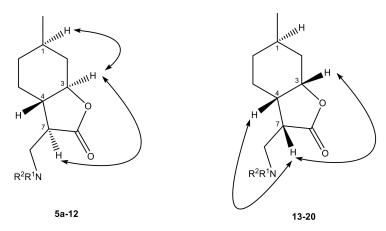


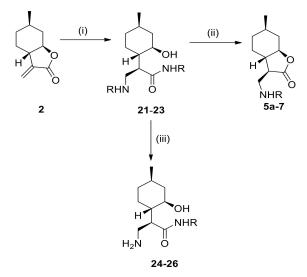
Figure 2. Determination of the relative configuration of β-aminolactones by NOESY.

2.3. Synthesis of β -Aminoamides and Dipeptides

Nucleophilic addition and ring-opening of lactones were simultaneously performed from **2** using excess amines to form β -aminoamides **21–23** in one step (Scheme 4). It is interesting that benzylamine reacted at room temperature, while (R)- and (S)- α -methylbenzylamine required a higher temperature and longer reactions (Table 4). This is probably due to steric hindrance exerted by the α -methyl group. Our efforts in the opening of lactones with secondary amines failed. Hydrolysis of β -aminoamides under acidic conditions resulted in the original starting material β -aminolactones **5a–7** (Scheme 4).

Table 4. Preparation of β-aminoamides from **2** and **13–15**.

Entry	Compo	ound R	Temperature (°C)	Reaction Time (h)	Yield (%)
1	21	Benzyl	25	20	90
2	22	(R) - α -Meth	ylbenzy170	48	58
3	23	(S) - α -Meth	ylbenzyl70	48	54
4	32	Benzyl	70	24	70
5	33	(R) - α -Meth	ylbenzy170	72	42
6	34	(S)- α -Meth	ylbenzyl70	72	45



Scheme 4. (i) RNH₂ (4 equiv.), dry EtOH; (ii) 10% aqueous HCl solution, 25 °C, 24 h, 5a: 70%, 6, 7: 65%; (iii) catalyst, H₂ (1 atm.), MeOH, 25 °C, 62–80%.

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Debenzylation via hydrogenolysis of compounds **21–23** over appropriate catalysts in MeOH gave primary aminoamides **24–26** in moderate yields (Table 5).

Entry	Compo	ound R	Catalyst	Reaction Time (h)	Yield (%)
1	24	Benzyl	5% Pd/C	96	80
2	25	(R) - α -Methy	lbe 52 /yIPd/C	168	62
3	26	(S) - α -Methyl	behdyOH) ₂ /C	200	65
4	35	Benzyl	5% Pd/C	96	70
5	36	(R) - α -Methy	lbe 52 /yIPd/C	240	70
6	37	(S)- α -Methyl	behdyOH) ₂ /C	300	52

Table 5. Debenzylation of aminoamides 21–23 and 32–34.

In further studies starting from **2**, the addition and ring-opening reaction with β -aminoester successfully gave dipeptide **27**. The application of α -aminoesters failed despite using long reaction times and elevated temperatures. The probable reason is steric hindrance exerted by the α -methyl group of the aminoesters. In addition, the opening of N-benzyl aminolactone **5a** with both the α -and β -aminoester proceeded smoothly to give N-benzyl dipeptides **28–29**. Debenzylation through hydrogenolysis over Pd/C and purification of the crude products gave dipeptides **30–31**, i.e., suitable starting compounds in peptide synthesis (Scheme **5**).

Scheme 5. (i) NH₂C₂H₅COOC₂H₅ (3 equiv.), dry EtOH, 25 °C, 48 h, 63%; (ii) Benzylamine (1 equiv.), dry EtOH, 25 °C, 20 h, 65%; then amino esters (3 equiv.), dry EtOH, 70 °C, 48 h, 40–45%; (iii) 5% Pd/C, dry EtOH, 25 °C, 24 h, 50–55%.

Our effort to prepare β -aminoamides 32–34 starting from 4 failed. Fortunately, the synthesis was achieved by reacting β -aminolactones 13–15 with primary amines under reflux conditions in anhydrous THF [31] (Table 4). Again, opening the lactone ring with secondary amines was unsuccessful. Acidic hydrolysis of β -aminoamides 26–28 led to the original starting material β -aminolactones 13–15 instead of the expected β -aminoacids (Scheme 6). Debenzylation with appropriate catalysts gave primary β -aminoamides 35–37 in moderate yields (Table 5). The attempted nucleophilic addition and ring-opening of 6 with α - or β -aminoesters failed.

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Scheme 6. (i) RNH₂ (4 equiv.), dry THF; 70 °C; (ii) 10% aqueous HCl solution, 25 °C, 24 h, **13**: 60%, **14**, **15**: 70%; (iii) catalyst, H₂ (1 atm), MeOH, 25 °C, 52–70%.

2.4. Antiproliferative Activities

Since several sesquiterpene-based α -methylene- γ -lactones, as well as their derivatives containing β -aminolactone moiety, exerted an antiproliferative action on adherent human cancer cell lines [3,30], antiproliferative activities of the prepared β -aminolactone and β -aminoamide analogues were also tested against a panel of human malignant cell lines isolated from cervical (HeLa) and breast (MCF7 and MDA-MB-231) cancers (Table 6). While the β -aminolactone-typed monoterpene derivatives proved to be ineffective against the utilized cell lines, the N-(S)- α -methylbenzyl-substituted β -aminoamide analogues (23, 34) exhibited modest growth inhibitory activities. The most potent newly-prepared monoterpene analogue was compound 23, exerting antiproliferative activity comparable to those of reference agent cisplatin.

Table 6. Antipr	oliferative	activities o	f the tested	l monoterpene	analogs.

Analog	Conc. (µM)	Growth Inhibition (%) \pm SEM *			
Allalog		HeLa	MCF7	MDA-MB-231	
	10	_	_	_	
2	30	_	18.33 ± 2.90	_	
4	10	_	_	_	
4	30	21.10 ± 2.44	_	_	
_	10	16.90 ± 2.60	18.76 ± 2.50	_	
5	30	28.74 ± 2.30	31.25 ± 3.01	_	
-	10	_	_	_	
7	30	_	17.29 ± 2.90	_	
01	10				
21	30	31.79 ± 1.95	24.36 ± 2.42	22.75 ± 1.84	
22	10	_	_	_	
22	30	27.09 ± 1.66	_	_	
22	10	41.25 ± 2.60	33.96 ± 1.84	24.71 ± 1.86	
23	30	94.83 ± 0.73	87.93 ± 1.47	70.56 ± 3.51	
29	10	_	17.00 ± 2.51	_	
29	30	_	36.45 ± 1.00	23.43 ± 2.03	
22	10	24.36 ± 2.70	17.06 ± 1.46	_	
32	30	32.43 ± 0.52	40.40 ± 2.88	_	
2.4	10	_	22.70 ± 1.82	19.51 ± 2.35	
34	30	36.04 ± 0.51	45.41 ± 2.92	34.61 ± 2.22	
ciculatin	10	42.61 ± 2.33	53.03 ± 2.29	67.51 ± 1.01	
cisplatin	30	99.93 ± 0.26	86.90 ± 1.24	87.75 ± 1.10	

^{*} Growth inhibition values less than 15% are considered negligible and are not given numerically. Compounds 6, 8, 9, 13, 26, 27, and 28 were also examined but did not elicit 15% growth inhibition even at 30 μ M.

 α -Methylene- γ -lactone is generally believed to be a pharmacophore acting as an alkylating agent on DNA and proteins [32]. In the present set of (-)-isopulegol analogs, the γ -lactone-type derivatives (2, 4, 5 and 7) exerted weak antiproliferative activities, while the most active member of the presented library (23) is not a typical sesquiterpene lactone, but a β -aminoamide. Based on our results, the stereochemistry of the N-substituent on the amide function ((S)- α -methylbenzyl substituent), as well as the *trans* position of the bulky β -aminoamide substituent and the hydroxy group on the cyclohexane ring, are proposed as crucial conditions accounting for the activity. The antiproliferative activity of dipeptides 27, 28, and 29 was also tested on adherent human cancer cell lines. While in case of 29 a week antiproliferative activity was observed, on MCF7 and MDA-MB-231, 27 and 28 were uneffective.

3. Discussion

Starting from commercially-available (-)-isopugeol, a new family of isopulegol- and neoisopulegol-based chiral β -aminolactone and β -aminoamide libraries has been prepared through chiral α -methylene- γ -lactones as key intermediates. Moreover, isopulegol-based chiral dipeptides, promising chiral substrates for the synthesis of chiral foldamers, were synthesized. The resulting β -aminoamides exert marked antiproliferative action on a panel of human cancer cell lines. In vitro pharmacological studies have clearly shown that the N-(S)- α -methylbenzyl substituent on the β -aminoamide function is essential. The stereochemistry of the β -aminoamides has no influence on the antiproliferative effect.

4. Materials and Methods

4.1. General Methods

Commercially-available compounds were used as obtained from suppliers (Molar Chemicals Ltd., Halásztelek, Hungary; Merck Ltd., Budapest, Hungary and VWR International Ltd., Debrecen, Hungary), while applied solvents were dried according to standard procedures. Optical rotations were measured in MeOH at 20 °C, with a Perkin-Elmer 341 polarimeter (PerkinElmer Inc., Shelton, CT, USA). Chromatographic separations and monitoring of reactions were carried out on Merck Kieselgel 60 (Merck Ltd., Budapest, Hungary). Elemental analyses for all prepared compounds were performed on a Perkin-Elmer 2400 Elemental Analyzer (PerkinElmer Inc., Waltham, MA, USA). GC measurements for direct separation of commercially-available enantiomers of isopulegol to determine the enantiomeric purity of starting material 1 were performed on a Chirasil-DEX CB column (2500 \times 0.25 mm I.D.) on a Perkin-Elmer Autosystem XL GC equipped with a Flame Ionization Detector (Perkin-Elmer Corporation, Norwalk, CT, USA) and a Turbochrom Workstation data system (Perkin-Elmer Corp., Norwalk, CT, USA). Melting points were determined on a Kofler apparatus (Nagema, Dresden, Germany) and are uncorrected. ¹H- and ¹³C-NMR spectra were recorded on BruckerAvance DRX 500 spectrometer [500 MHz (1 H) and 125 MHz (13 C), $\delta = 0$ (TMS)]. Chemical shifts are expressed in ppm (δ) relative to TMS as the internal reference. I values are given by Hz. The structures were confirmed by 1H-NMR, 13C-NMR and 2D-NMR technics (see Supplementary Materials).

4.2. Starting Materials

(-)-Isopulegol (1) is available commercially from Merck Co (Darmstadt, Germany) with ee = 95%. (+)- α -Methylene- γ -butyrolactone (2), (+)-neoisopulegol (3) and (-)- α -methylene- γ -butyrolactone (4) were prepared according to literature procedures. All spectroscopic data were similar to those described therein [25,27].

4.3. General Procedure for Nucleophilic Addition of α -Methylene- γ -Butyrolactone to Amines

Amines (1.2 mmol) were added to the solution of α -methylene- γ -butyrolactone **2** or **4** (1.2 mmol) in dry EtOH (2.0 mL). The reaction mixture was stirred at appropriate temperatures for 20–72 h. When

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the reaction was complete (indicated by TLC), EtOH was removed under reduced pressure. The crude residue was purified by column chromatography on silica gel with an appropriate solvent mixture. The crude products after solvent evaporation were purified as HCl salts by recrystallization in diethyl ether resulting in compounds **5a–10** and **13–18**.

4.3.1.~(3R,3aS,6R,7aR)-3-((Benzylamino)methyl)-6-methylhexahydrobenzofuran-2(3H)-one hydrochloride (5a)

Prepared from **2** with benzylamine at 25 °C for 20 h. Compound **5a** was purified by column chromatography on silica gel (CHCl₃/MeOH = 19:1). Yield: 65%, white crystals, m.p.: 190–198 °C. $[\alpha]_D^{20}$ = -8.0 (c 0.23, MeOH). 1 H NMR (500 MHz, DMSO- d_6): δ = 0.94–1.00 (1H, m), 0.95 (3H, d, J = 8.1 Hz), 1.09–1.18 (1H, m), 1.24–1.32 (1H, m), 1.59–1.69 (2H, m), 1.78–1.86 (1H, m), 2.06–2.13 (2H, m), 3.05–3.19 (3H, m), 3.93 (1H, td, J = 4.4, 14.1 Hz), 7.41–7.63 (5H, m), 9.67 (2H, s). 13 C NMR (125 MHz, DMSO- d_6): δ = 21.9, 26.1, 30.5, 33.4, 37.6, 42.7, 44.6, 47.0, 50.5, 81.9, 128.5, 128.8, 130.2, 131.7, 176.1. Anal. Calcd for C₁₇H₂₄ClNO₂: C, 65.90; H, 7.81; N, 4.52. Found: C, 65.85; H, 7.85; N, 4.52.

4.3.2. (3R,3aS,6R,7aR)-6-Methyl-3-((((R)-1-phenylethyl)amino)methyl)hexahydrobenzofuran-2(3H)-one hydrochloride (6)

Prepared from **2** with (*R*)-*α*-methylbenzylamine at 25 °C for 20 h. Compound **6** was purified by column chromatography on silica gel (CHCl₃/MeOH = 19:1). Yield: 75%, white crystals, m.p.: 170–180 °C. [α]_D²⁰ = +21.0 (c 0.23, MeOH). ¹H NMR (500 MHz, DMSO- d_6): δ = 0.91–1.00 (1H, m), 0.95 (3H, d, J = 6.5 Hz), 1.08–1.15 (1H, m), 1.23–1.28 (1H, m), 1.53–1.62 (1H, m), 1.61 (3H, d, J = 6.4 Hz), 1.68 (1H, d, J = 13.2 Hz), 1.75–1.82 (1H, m), 2.03–2.12 (2H, m), 2.78–2.86 (1H, m), 2.94 (1H, t, J = 6.3 Hz), 3.05–3.15 (1H, m), 3.92 (1H, td, J = 3.4, 11.3 Hz), 4.38 (1H, s), 7.39–7.61 (5H, m), 9.17 (1H, br s), 9.83 (1H, br s). ¹³C NMR (125 MHz, DMSO- d_6): δ = 19.4, 21.9, 26.0, 30.5, 33.4, 37.5, 46.4, 58.2, 81.9, 128.0, 128.8, 176.0. Anal. Calcd for C₁₈H₂₆ClNO₂: C, 66.76; H, 8.09; N, 4.32. Found: C, 66.75; H, 8.04; N, 4.30.

4.3.3. (3R,3aS,6R,7aR)-6-Methyl-3-((((S)-1-phenylethyl)amino)methyl)hexahydrobenzofuran-2(3H)-one hydrochloride (7)

Prepared from **2** with (*S*)- α -methylbenzylamine at 25 °C for 20 h. Compound 7 was purified by column chromatography on silica gel (CHCl₃/MeOH = 19:1). Yield: 71%, white crystals, m.p.: 170–180 °C. [α]_D²⁰ = -26.0 (c 0.24, MeOH). ¹H NMR (500 MHz, DMSO- d_6): δ = 0.91–0.97 (1H, m), 0.94 (1H, d, J = 6.6 Hz), 1.08–1.15 (1H, m), 1.19–1.28 (1H, m), 1.57–1.67 (2H, m), 1.62 (3H, d, J = 6.7 Hz), 1.74–1.81 (1H, m), 1.90–1.93 (1H, m), 2.09–2.11 (1H, m), 2.85–2.94 (1H, m), 2.95–3.02 (1H, m), 3.93 (1H, td, J = 3.5, 11.3 Hz), 4.36 (1H, br s), 7.39–7.64 (5H, m), 9.25 (1H, br s), 9.82 (1H, br s). ¹³C NMR (125 MHz, DMSO- d_6): δ = 19.4, 21.9, 26.0, 30.5, 33.4, 37.5, 42.8, 43.5, 46.8, 58.3, 82.0, 127.9, 128.9, 137.1, 176.3. Anal. Calcd for C₁₄H₂₆ClNO₂: C, 66.76; H, 8.09; N, 4.32. Found: C, 66.78; H, 8.10; N, 4.35.

4.3.4.~(3R,3aS,6R,7aR)-3-((Diethylamino)methyl)-6-methylhexahydrobenzofuran-2(3H)-one hydrochloride (8)

Prepared from **2** with diethylamine at 25 °C for 20 h. Compound **8** was purified by column chromatography on silica gel (CHCl₃/MeOH = 19:1). Yield: 50%, colorless oil. $[\alpha]_D^{20} = -7.0$ (c 0.27, MeOH). ¹H NMR (500 MHz, DMSO- d_6): $\delta = 0.91$ –0.99 (1H, m), 0.96 (3H, d, J = 6.6 Hz), 1.05 (1H, t, J = 7.0 Hz), 1.14 (1H, q, J = 11.5 Hz), 1.24 (6H, td, J = 2.3, 7.1 Hz), 1.31–1.39 (1H, m), 1.58–1.63 (1H, m), 1.71–1.82 (2H, m), 1.97–2.01 (1H, m), 2.12–2.15 (1H, m), 3.12–3.23 (6H, m), 3.38–3.44 (2H, m), 3.97 (1H, td, J = 3.6, 11.4 Hz), 9.94 (1H, br s). ¹³C NMR (125 MHz, DMSO- d_6): $\delta = 8.4$, 8.5, 21.8, 25.6, 30.6, 33.4, 37.5, 41.3, 46.8, 47.2, 47.5, 49.1, 81.9, 176.8. Anal. Calcd for C₁₄H₂₆ClNO₂: C, 60.96; H, 9.50; N, 5.08. Found: C, 60.70; H, 9.45; N, 5.10.

4.3.5. (3R,3aS,6R,7aR)-6-Methyl-3-(piperidin-1-ylmethyl)hexahydrobenzofuran-2(3*H*)-one hydrochloride (9)

Prepared from **2** with pyridine at 25 °C for 20 h. Compound **9** was purified by column chromatography on silica gel (CHCl₃/MeOH = 19:1). Yield: 47%, white crystals, m.p.: 180–190 °C. $[\alpha]_D^{20} = -3.3$ (c 0.31, MeOH). ¹H NMR (500 MHz, DMSO- d_6): $\delta = 0.92-1.00$ (1H, m), 0.96 (1H, d, J = 6.6 Hz), 1.14 (1H, q, J = 11.5 Hz), 1.30–1.40 (1H, m), 1.58–1.64 (1H, m), 1.67–1.83 (6H, m), 2.02–2.05 (1H, m), 2.11–2.13 (1H, m), 2.88–2.96 (2H, m), 3.16–3.21 (2H, m), 3.26–3.29 (1H, m), 3.42 (1H, d, J = 12.0 Hz), 3.94 (1H, td, J = 3.6, 11.3 Hz), 10.5 (1H, br s). ¹³C NMR (125 MHz, DMSO- d_6): $\delta = 21.1$, 21.8, 22.2, 25.8, 30.6, 33.4, 37.5, 41.4, 47.7, 51.7, 52.8, 53.8, 81.8, 176.7. Anal. Calcd for C₁₅H₂₆ClNO₂: C, 62.59; H, 9.10; N, 4.87. Found: C, 62.60; H, 9.15; N, 4.90.

4.3.6. (3R,3aS,6R,7aR)-3-((Dibenzylamino)methyl)-6-methylhexahydrobenzofuran-2(3*H*)-one hydrochloride (**10**)

Prepared from **2** with dibenzylamine at 70 °C for 72 h. Compound **10** was purified by column chromatography on silica gel (n-hexane/ethyl acetate = 9:1). Yield: 59%, white crystals, m.p.: 120–125 °C. [α] $_D^{20} = -27.0$ (c 0.27, MeOH). 1 H NMR (500 MHz, DMSO- d_6): δ = 0.80–0.87 (1H, m), 0.93 (3H, d, J = 6.5 Hz), 1.10 (1H, q, J = 11.4 Hz), 1.23–1.31 (1H, m), 1.55–1.72 (4H, m), 2.09 (1H, d, J = 11.0 Hz), 3.09 (1H, d, J = 12.9 Hz), 3.33–3.40 (1H, m), 3.92 (1H, t, J = 9.6 Hz), 4.30–4.33 (2H, m), 4.44 (1H, d, J = 9.3 Hz), 7.75–7.70 (10H, m), 11.1 (1H, br s). 13 C NMR (125 MHz, DMSO- d_6): δ = 21.8, 25.6, 30.5, 33.4, 37.4, 41.6, 47.4, 49.8, 56.7, 56.8, 82.0, 128.8, 129.6, 131.3, 131.4, 131.6, 176.9 Anal. Calcd for C₂₄H₃₀ClNO₂: C, 72.07; H, 7.56; N, 3.50. Found: C, 72.07; H, 7.53; N, 3.55.

4.3.7. (3S,3aS,6R,7aS)-3-((Benzylamino)methyl)-6-methylhexahydrobenzofuran-2(3*H*)-one hydrochloride (13)

Prepared from 4 with benzylamine at 25 °C for 20 h. Compound 13 was purified by column chromatography on silica gel (CHCl₃/MeOH = 19:1). Yield: 60%, white crystals, m.p.: 165–167 °C. $[\alpha]_D^{20} = -34.0$ (c 0.24, MeOH). ¹H NMR (500 MHz, DMSO- d_6): $\delta = 0.73$ –0.88 (2H, m), 0.88 (1H, d, J = 6.2 Hz), 1.24–1.30 (1H, m), 1.34–1.43 (1H, m), 1.58 (1H, d, J = 12.4 Hz), 1.81 (1H, t, J = 6.2 Hz), 2.06 (1H, d, J = 14.3 Hz), 2.56–2.59 (1H, m), 3.00 (1H, t, J = 10.7 Hz), 3.16 (1H, d, J = 12.0 Hz), 2.42–2.43 (1H, m), 4.20 (1H, q, J = 13.0 Hz), 4.58 (1H, s), 7.42–7.60 (5H, m), 9.54 (2H, s). ¹³C NMR (125 MHz, DMSO- d_6): $\delta = 21.7$, 22.4, 25.8, 31.1, 35.0, 36.2, 41.6, 44.6, 50.2, 78.1, 128.6, 129.0, 130.2, 131.8, 175.6. Anal. Calcd for C₁₇H₂₄ClNO₂: C, 65.90; H, 7.81; N, 4.52. Found: C, 65.95; H, 7.80; N, 4.55.

4.3.8. (3S,3aS,6R,7aS)-6-Methyl-3-((((R)-1-phenylethyl)amino)methyl)hexahydrobenzofuran-2(3H)-one hydrochloride (14)

Prepared from 4 with (*R*)-α-methylbenzylamine at 25 °C for 20 h. Compound 14 was purified by column chromatography on silica gel (CHCl₃/MeOH = 19:1). Yield: 65%, white crystals, m.p.: 250–252 °C. [α]_D²⁰ = -3.0 (c 0.24, MeOH). ¹H NMR (500 MHz, DMSO- d_6): δ = 0.72–0.78 (1H, m), 0.83–0.90 (1H, m), 0.87 (3H, d, J = 6.4 Hz), 1.24–1.30 (1H, m), 1.33–1.37 (1H, m), 1.57 (1H, d, J = 12.7 Hz), 1.63 (1H, d, J = 6.7 Hz), 1.82–1.85 (1H, m), 2.04 (H, d, J = 14.1 Hz), 2.61–2.66 (1H, m), 2.79–2.84 (1H, m), 2.91–2.97 (1H, m), 3.29–3.31 (1H, m), 4.44 (1H, q, J = 4.8 Hz), 4.55 (1H, d, J = 2.3 Hz), 7.40–7.64 (5H, m), 9.49 (1H, br s), 9.94 (1H, br s). ¹³C NMR (125 MHz, DMSO- d_6): δ = 19.3, 21.7, 22.4, 25.8, 31.1, 35.0, 36.3, 40.4, 44.8, 57.7, 78.1, 127.9, 128.9, 137.0, 175.6. Anal. Calcd for C₁₈H₂₆ClNO₂: C, 66.76; H, 8.09; N, 4.32. Found: C, 66.74; H, 8.13; N, 4.35.

$4.3.9.\ (3S,3aS,6R,7aS)-6-Methyl-3-((((S)-1-phenylethyl)amino)methyl) hexahydrobenzofuran-2(3H)-one hydrochloride\ (15)$

Prepared from 4 with (*S*)-α-methylbenzylamine at 25 °C for 20 h. Compound **15** was purified by column chromatography on silica gel (CHCl₃/MeOH = 19:1). Yield: 70%, white crystals, m.p.: 230–235 °C. [α]_D²⁰ = -57.0 (c 0.21, MeOH). ¹H NMR (500 MHz, DMSO- d_6): δ = 0.55–0.63 (1H, m), 0.78–0.85 (1H,

m), 0.85 (3H, d, J = 6.2 Hz), 1.22–1.30 (2H, m), 1.50 (2H, d, J = 10.9 Hz), 1.64 (1H, d, J = 6.7 Hz), 2.03 (1H, d, J = 13.4 Hz), 2.50–2.56 (1H, m), 2.59–2.63 (1H, m), 3.05 (1H, t, J = 9.7 Hz), 3.42–3.44 (1H, m), 4.47 (1H, br s), 4.57 (1H, d, J = 2.1 Hz), 7.39–7.64 (5H, m), 9.48 (1H, d, J = 7.6 Hz), 10.04 (1H, br s). ¹³C NMR (125 MHz, DMSO- d_6): δ = 19.5, 21.7, 22.1, 25.7, 31.0, 35.0, 36.0, 40.2, 44.5, 57.6, 78.0, 127.8, 128.9, 129.0, 136.9, 175.4. Anal. Calcd for C₁₈H₂₆ClNO₂: C, 66.76; H, 8.09; N, 4.32. Found: C, 66.77; H, 8.13; N, 4.29.

4.3.10. (3S,3aS,6R,7aS)-3-((Diethylamino)methyl)-6-methylhexahydrobenzofuran-2(3*H*)-one hydrochloride (**16**)

Prepared from 4 with diethylamine at 25 °C for 20 h. Compound 16 was purified by column chromatography on silica gel (CHCl₃/MeOH = 19:1). Yield: 50%, white crystals, m.p.: 158–163 °C. $[\alpha]_D^{20} = -39.0$ (c 0.24, MeOH). ¹H NMR (500 MHz, DMSO- d_6): $\delta = 0.81$ –0.93 (1H, m), 0.87 (1H, d, J = 9.0 Hz), 1.20–1.26 (6H, m), 1.26–1.29 (1H, m), 1.33–1.43 (1H, m), 1.60 (1H, d, J = 11.5 Hz), 1.82–1.85 (1H, m), 2.07 (1H, d, J = 14.6 Hz), 2.64–2.66 (1H, m), 3.14–3.22 (6H, m), 3.58 (1H, q, J = 4.2 Hz), 4.58 (1H, d, J = 2.2 Hz), 10.6 (1H, br s). ¹³C NMR (125 MHz, DMSO- d_6): $\delta = 8.0$, 8.6, 21.7, 22.6, 25.8, 31.1, 35.1, 37.1, 42.9, 45.9, 46.4, 47.1, 78.0, 176.1. Anal. Calcd for $C_{14}H_{26}ClNO_2$: C, 60.96; H, 9.50; N, 5.08. Found: C, 60.73; H, 9.53; N, 5.05.

4.3.11. (3S,3aS,6R,7aS)-6-Methyl-3-(piperidin-1-ylmethyl)hexahydrobenzofuran-2(3*H*)-one hydrochloride (17)

Prepared from 4 with pyridine at 25 °C for 20 h. Compound 17 was purified by column chromatography on silica gel (CHCl₃/MeOH = 19:1). Yield: 53%, white crystals, m.p.: 231–233 °C. [α] $_D^{20}$ = -39.0 (c 0.24, MeOH). 1 H NMR (500 MHz, DMSO- 4 6): δ = 0.76–0.81 (1H, m), 0.87–0.92 (1H, m), 0.88 (3H, d, J = 6.0 Hz), 1.22–1.28 (1H, m), 1.31–1.44 (2H, m), 1.59–1.89 (7H, m), 2.07 (1H, d, J = 14.5 Hz), 2.65 (1H, d, J = 5.6 Hz), 2.87–2.97 (2H, m), 3.17–3.22 (2H, m), 3.41 (1H, d, J = 10.9 Hz), 3.52 (1H, d, J = 11.1 Hz), 3.61 (1H, br s), 4.56 (1H, br s), 10.6 (1H, br s) 13 C NMR (125 MHz, DMSO- 4 6): δ = 21.2, 21.7, 22.1, 22.2, 22.8, 25.8, 31.1, 35.1, 37.3, 43.2, 51.2, 51.7, 52.6, 77.9, 176.0. Anal. Calcd for C₁₅H₂₆ClNO₂: C, 62.59; H, 9.10; N, 4.87. Found: C, 62.57; H, 9.05; N, 4.93.

4.3.12.~(3S,3aS,6R,7aS)-3-((Dibenzylamino)methyl)-6-methylhexahydrobenzofuran-2(3H)-one hydrochloride (18)

Prepared from 4 with dibenzylamine at 70 °C for 72 h. Compound 18 was purified by column chromatography on silica gel (n-hexane/ethyl acetate = 9:1). Yield: 50%, white crystals, m.p.: 118–120 °C. [α] $_D^{20}$ = -23.0 (c 0.21, MeOH). 1 H NMR (500 MHz, DMSO- d_6): δ = 0.45–0.52 (1H, m), 0.73–0.85 (1H, m), 0.83 (1H, d, J = 6.1 Hz), 1.17–1.30 (3H, m), 1.41 (1H, d, J = 12.4 Hz), 2.03 (1H, d, J = 13.7 Hz), 2.70 (1H, d, J = 4.9 Hz), 2.80 (1H, t, J = 10.2 Hz), 3.20–3.24 (1H, m), 3.78 (1H, br s), 4.35–4.40 (2H, m), 4.49–4.53 (2H, m), 7.33–7.70 (10H, m), 11.3 (1H, br s). 13 C NMR (125 MHz, DMSO- d_6): δ = 21.6, 22.1, 25.6, 31.0, 35.0, 36.9, 42.8, 46.0, 56.1, 56.6, 77.9, 128.8, 129.1, 129.6, 129.7, 129.9, 131.6, 131.9, 175.6. Anal. Calcd for $C_{24}H_{30}$ ClNO₂: C, 72.07; H, 7.56; N, 3.50. Found: C, 72.09; H, 7.53; N, 3.55.

4.4. General Procedure for Nucleophilic Addition of α -Methylene- γ -Butyrolactone with Amino Esters

To the solution of α -methylene- γ -butyrolactone **2** or **4** (1.2 mmol) in dry EtOH (2.0 mL) was added L- or β -alanine ethyl ester hydrochloride (2.4 mmol) and Et₃N (2.4 mmol). The reaction mixture was stirred at the appropriate temperature for 20 h. When the reaction was complete (indicated by TLC), EtOH was removed under reduced pressure. The crude residue was purified by column chromatography on silica gel with a mixture of CHCl₃ and MeOH (19:1). After solvent evaporation, the addition of a few drops of HCl/EtOH, and recrystallization in diethyl ether, compounds **11** and **12**, as well as **19** and **20**, respectively, were isolated.

4.4.1. Ethyl 3-(((3R,3aS,6R,7aR)-6-methyl-2-oxooctahydrobenzofuran-3-yl)methylamino) propanoate hydrochloride (11)

Prepared from **2** with *β*-alanine ethyl ester hydrochloride at 25 °C. Yield: 60%, white crystals, m.p.: 125–135 °C. [α]_D²⁰ = -3.7 (c 0.32, MeOH). ¹H NMR (500 MHz, CDCl₃): δ = 1.00–1.07 (1H, m), 1.02 (3H, d, J = 6.6 Hz), 1.20–1.30 (1H, m), 1.29 (3H, t, J = 7.1 Hz), 1.44–1.50 (1H, m), 1.64–1.72 (2H, m), 1.82 (1H, d, J = 13.6 Hz), 2.01 (1H, d, J = 10.7 Hz), 2.26 (1H, m), 2.89 (1H, dt, J = 4.3, 18.0 Hz), 3.10–3.38 (6H, m), 4.00 (1H, td, J = 3.6, 11.3 Hz), 4.22 (2H, q, J = 7.1 Hz), 8.70 (1H, br s), 11.30 (1H, br s). ¹³C NMR (125 MHz, CDCl₃): δ = 14.2, 22.0, 26.2, 30.2, 31.2, 33.8, 38.1, 42.8, 44.6, 47.2, 48.3, 62.2, 84.0, 171.8, 178.3. Anal. Calcd for C₁₅H₂₆ClNO₄: C, 56.33; H, 8.19; N, 4.38. Found: C, 56.35; H, 8.15; N, 4.40.

4.4.2. (S)-Ethyl 2-(((3R,3aS,6R,7aR)-6-methyl-2-oxooctahydrobenzofuran-3-yl)methylamino)propanoate hydrochloride (12)

Prepared from **2** with L-alanine ethyl ester hydrochloride at 25 °C. Yield: 40%, white crystals, m.p.: 150–160 °C. $[\alpha]_D^{20} = -3.0$ (c 0.28, MeOH). ¹H NMR (500 MHz, CDCl₃): $\delta = 0.97–1.02$ (1H, m), 1.02 (3H, d, J = 6.5 Hz), 1.22 (1H, q, J = 11.4 Hz), 1.33 (3H, t, J = 7.1 Hz), 1.49 (1H, q, J = 11.9 Hz), 1.62–1.66 (2H, m), 1.82 (6H, d, J = 6H, d, J = 6.7 Hz), 1.98 (1H, d, J = 12.4 Hz), 2.27 (1H, d, J = 11.5 Hz), 3.11 (1H, t, J = 10.0 Hz), 3.48–3.48 (1H, m), 3.54 (1H, t, J = 10.0 Hz), 3.96 (1H, q, J = 6.0 Hz), 4.03 (1H, td, J = 3.1, 11.3 Hz), 4.27–4.35 (2H, m), 8.14 (1H, br s), 12.2 (1H, br s). ¹³C NMR (125 MHz, CDCl₃): $\delta = 14.1$, 15.9, 22.0, 26.2, 31.1, 33.9, 38.1, 42.6, 46.3, 48.5, 60.0, 63.3, 83.9, 169.2, 178.6. Anal. Calcd for C₁₅H₂₆ClNO₄: C, 56.33; H, 8.19; N, 4.38. Found: C, 56.30; H, 8.20; N, 4.35.

4.4.3. Ethyl 3-(((3S,3aS,6R,7aS)-6-methyl-2-oxooctahydrobenzofuran-3-yl) methylamino)propanoate hydrochloride (19)

Prepared from 4 with *β*-alanine ethyl ester hydrochloride at 70 °C. Yield: 60%, white crystals, m.p.: 202–205 °C. [α]_D²⁰ = -37.0 (c 0.21, MeOH). ¹H NMR (500 MHz, CDCl₃): δ = 0.93 (3H, d, J = 6.4 Hz), 0.92–0.99 (1H, m), 1.09 (1H, q, J = 10.0 Hz), 1.24–1.30 (1H, m), 1.29 (3H, t, J = 6.9 Hz), 1.49–1.62 (1H, m), 1.70 (1H, d, J = 12.3 Hz), 1.78 (1H, d, J = 9.5 Hz), 2.24 (1H, d, J = 14.7 Hz), 2.66 (1H, br s), 2.92 (1H, d, J = 16.7 Hz), 3.13 (1H, d, J = 16.2 Hz), 3.26–3.30 (4H, m), 3.60–3.70 (1H, m), 4.22 (2H, q, J = 7.0 Hz), 4.65 (1H, s), 8.81 (1H, br s), 10.8 (1H, br s). ¹³C NMR (125 MHz, CDCl₃): δ = 14.2, 21.9, 23.3, 26.1, 30.4, 31.5, 35.6, 37.7, 44.4, 44.5, 44.7, 62.0, 79.7, 171.1, 177.7. Anal. Calcd for C₁₅H₂₆ClNO₄: C, 56.33; H, 8.19; N, 4.38. Found: C, 56.37; H, 8.18; N, 4.35.

4.4.4. (S)-Ethyl 2-(((3R,3aS,6R,7aR)-6-methyl-2-oxooctahydrobenzofuran-3-yl)methylamino)propanoate hydrochloride (20)

Prepared from 4 with L-alanine ethyl ester hydrochloride at 70 °C. Yield: 44%, white crystals, m.p.: 216–218 °C. [α] $_D^{20}$ = -35.0 (c 0.24, MeOH). 1 H NMR (500 MHz, CDCl₃): δ = 0.92 (3H, d, J = 6.5 Hz), 0.94–1.02 (2H, m), 1.24 (1H, t, J = 12.3 Hz), 1.33 (3H, t, J = 7.0 Hz), 1.50–1.60 (1H, m), 1.70 (1H, d, J = 11.5 Hz), 1.76 (3H, d, J = 6.9 Hz), 1.93 (1H, d, J = 10.6 Hz), 2.00–2.14 (1H, m), 2.23 (1H, d, J = 14.9 Hz), 2.78–2.85 (1H, m), 3.32 (1H, d, J = 17.2 Hz), 3.77 (1H, d, J = 4.5 Hz), 4.08–4.16 (1H, m), 4.30 (2H, q, J = 7.0 Hz), 4.61 (1H, s), 9.80 (1H, br s), 10.37 (1H, br s). 13 C NMR (125 MHz, CDCl₃): δ = 14.2, 15.0, 22.0, 23.1, 26.1, 31.6, 35.8, 37.7, 41.9, 45.2, 55.8, 63.1, 79.5, 168.7, 176.8. Anal. Calcd for C₁₅H₂₆ClNO₄: C, 56.33; H, 8.19; N, 4.38. Found: C, 56.35; H, 8.17; N, 4.40.

4.5. General Procedure for the Preparation of β -Aminoamides

To a solution of α -methylene- γ -butyrolactone, **2** (1.2 mmol) or β -aminolactones **13–15** (1.2 mmol) in an appropriate solvent (2.0 mL) was added a solution of the appropriate amine (4.8 mmol). The mixture was stirred at the appropriate temperature for 20–72 h. When the reaction was complete (indicated by TLC), the mixture was evaporated to dryness. The crude product was purified by column chromatography on silica gel with CHCl₃/MeOH (19:1), resulting in compounds **21–23** and **32–34**.

4.5.1. (R)-N-Benzyl-3-(benzylamino)-2-((1S,2R,4R)-2-hydroxy-4-methylcyclohexyl)propanamide (21)

Prepared from **2** with benzylamine at 25 °C for 20 h in dry EtOH. Yield: 90%, white crystals, m.p.: 185–195 °C. $[\alpha]_D^{20} = -24.0$ (c 0.27, MeOH). ¹H NMR (500 MHz, DMSO- d_6): $\delta = 0.66$ –0.73 (1H, m), 0.82–0.92 (2H, m), 0.85 (1H, d, J = 6.5 Hz), 1.34–1.37 (2H, m), 1.52 (1H, d, J = 12.2 Hz), 1.65 (1H, t, J = 11.1 Hz), 1.86 (1H, d, J = 12.4 Hz), 2.93–3.02 (1H, m), 3.24–3.27 (3H, m), 4.06–4.17 (3H, m), 4.44 (1H, dd, J = 6.4, 15.1 Hz), 5.02–5.09 (1H, m), 7.20–7.54 (10H, m), 8.66–8.68 (2H, m), 9.28 (1H, br s). ¹³C NMR (125 MHz, DMSO- d_6): $\delta = 22.1$, 25.1, 30.9, 33.9, 41.7, 42.3, 43.3, 44.4, 47.2, 50.4, 68.5, 126.7, 127.1, 128.2, 128.6, 128.9, 130.0, 131.8, 139.5, 172.0. Anal. Calcd for $C_{24}H_{32}N_2O_2$: C, 75.75; H, 8.48; N, 7.36. Found: C, 75.80; H, 8.45; N, 7.35.

4.5.2. (R)-2-((1S,2R,4R)-2-Hydroxy-4-methylcyclohexyl)-N-((R)-1-phenylethyl)-3-(((R)-1-phenylethyl)amino) propanamide (22)

Prepared from **2** with (*R*)-α-methylbenzylamine at 70 °C for 48 h in dry EtOH. Yield: 58%, colorless oil. [α]_D²⁰ = +54.0 (c 0.20, MeOH). ¹H NMR (500 MHz, CDCl₃): δ = 0.72–0.90 (2H, m), 0.89 (1H, 6.4 Hz), 1.04–1.11 (1H, m), 1.25–1.28 (3H, m), 1.36 (3H, d, J = 6.6 Hz), 1.47–1.62 (2H, m), 1.53 (3H, d, J = 6.9 Hz), 1.78–1.87 (2H, m), 1.96 (1H, 1H, J = 13.6 Hz), 2.70 (1H, d, J = 11.8 Hz), 3.00–3.06 (2H, m), 3.42 (1H, d, J = 8.7 Hz), 3.62–3.70 (1H, m), 5.02 (1H, t, J = 7.1 Hz), 7.20–7.49 (10H, m), 8.38 (1H, d, J = 7.1 Hz). ¹³C NMR (125 MHz, CDCl₃): δ = 20.9, 22.0, 22.2, 25.8, 31.7, 34.1, 42.3, 43.5, 44.4, 47.7, 49.8, 58.7, 70.8, 126.7, 127.1, 127.6, 128.5, 128.9, 129.1, 144.4, 172.4. Anal. Calcd for C₂₆H₃₆N₂O₂: C, 76.43; H, 8.88; N, 6.86. Found: C, 76.45; H, 8.90; N, 6.83.

4.5.3. (R)-2-((1S,2R,4R)-2-Hydroxy-4-methylcyclohexyl)-N-((S)-1-phenylethyl)-3-(((S)-1-phenylethyl)amino) propanamide (23)

Prepared from **2** with (*S*)-α-methylbenzylamine at 70 °C for 48 h in dry EtOH. Yield: 54%, white crystals, m.p.: 137–148 °C [α]_D²⁰ = -55.0 (c 0.26, MeOH). ¹H NMR (500 MHz, DMSO- d_6): δ = 0.52–0.58 (1H, m), 0.82 (3H, d, J = 6.4 Hz), 0.86–0.91 (2H, m), 1.23–1.37 (2H, m), 1.36 (3H, d, J = 7.1 Hz), 1.56 (3H, d, J = 6.8 Hz), 1.69–1.74 (1H, m), 1.84–1.88 (1H, m), 2.78–2.82 (1H, m), 2.87–2.93 (1H, m), 3.24 (2H, d, J = 10.0 Hz), 4.29 (1H, d, J = 2.3 Hz), 4.91 (1H, quin, J = 7.1 Hz), 5.10 (1H, d, J = 4.9 Hz), 7.15–7.53 (10H, m), 8.30 (1H, d, J = 9.0 Hz), 8.64 (1H, d, J = 7.7 Hz), 9.60 (1H, br s). ¹³C NMR (125 MHz, DMSO- d_6): δ = 19.8, 22.1, 22.3, 24.5, 30.8, 33.7, 41.3, 41.7, 44.2, 47.2, 48.4, 58.0, 68.4, 125.6, 126.4, 127.6, 128.0, 128.7, 128.8, 137.1, 145.4, 171.0. Anal. Calcd for C₂₆H₃₆N₂O₂: C, 76.43; H, 8.88; N, 6.86. Found: C, 76.40; H, 8.85; N, 6.90.

4.5.4. (S)-N-Benzyl-3-(benzylamino)-2-((1S,2S,4R)-2-hydroxy-4-methylcyclohexyl)propanamide (32)

Prepared from **13** with benzylamine at 70 °C for 24 h in dry THF. Yield: 70%, white crystals, m.p.: 251–253 °C. $[\alpha]_D^{20} = +21.0$ (c = 0.20, MeOH). ¹H NMR (500 MHz, DMSO- d_6): δ = 0.76–0.80 (1H, m), 0.79 (3H, d, J = 6.3 Hz), 1.00 (1H, t, J = 13.0 Hz), 1.18 (1H, d, J = 10.1 Hz), 1.38–1.43 (1H, m), 1.51–1.58 (2H, m), 1.68 (1H, d, J = 11.2 Hz), 2.75 (1H, t, J = 6.7 Hz), 3.05 (1H, d, J = 11.8 Hz), 3.12–3.22 (1H, m), 3.79 (1H, s), 4.11 (2H, s), 4.18 (1H, dd, J = 5.2, 14.9 Hz), 4.29 (1H, dd, J = 6.1, 14.9 Hz), 4.74 (1H, s), 7.22–7.54 (10H, m), 8.75 (1H, t, J = 5.6 Hz), 9.06 (1H, br s), 9.17 (1H, br s). ¹³C NMR (125 MHz, DMSO- d_6): δ = 22.2, 24.3, 25.1, 34.3, 41.2, 41.8, 42.5, 44.3, 45.3, 50.1, 64.0, 126.8, 127.5, 128.2, 128.6, 128.9, 130.1, 131.7, 139.0, 172.3. Anal. Calcd for $C_{24}H_{32}N_2O_2$: C_7 75.75; H_7 8.48; N_7 7.36. Found: C_7 75.76; H_7 8.50; N_7 7.32.

4.5.5. (S)-2-((1S,2S,4R)-2-Hydroxy-4-methylcyclohexyl)-N-((R)-1-phenylethyl)-3-(((R)-1-phenylethyl)amino) propanamide (33)

Prepared from **14** with (*R*)- α -methylbenzylamine at 70 °C for 72 h in dry THF. Yield: 42%, white crystals, m.p.: 190–192 °C. [α]_D²⁰ = +46.0 (c 0.21, MeOH). ¹H NMR (500 MHz, CDCl₃): δ = 0.64–0.72 (1H, m), 0.79 (3H, 6.5 Hz), 0.99 (1H, t, J = 12.5 Hz), 1.14 (1H, d, J = 9.8 Hz), 1.39–1.54 (3H, m), 1.62–1.64 (1H, m), 1.77–1.81 (1H, m), 1.78 (3H, d, J = 6.7 Hz), 2.96 (2H, d, J = 8.6 Hz), 3.27 (1H, d, J = 8.9 Hz), 3.97 (1H, s), 4.36 (1H, q, J = 7.3 Hz), 4.80 (1H, quin, J = 7.3 Hz), 7.17–7.52 (10H, m), 8.23 (1H, d, J = 7.7 Hz). ¹³C

NMR (125 MHz, CDCl₃): δ = 20.2, 22.3, 25.1, 26.0, 34.3, 42.1, 42.6, 45.8, 46.1, 49.7, 59.8, 65.7, 126.6, 127.1, 127.6, 128.5, 129.6, 129.7, 135.7, 143.8, 171.6. Anal. Calcd for C₂₆H₃₆N₂O₂: C, 76.43; H, 8.88; N, 6.86. Found: C, 76.41; H, 8.85; N, 6.90.

4.5.6. (S)-2-((1S,2S,4R)-2-Hydroxy-4-methylcyclohexyl)-N-((S)-1-phenylethyl)-3-(((S)-1-phenylethyl)amino) propanamide (34)

Prepared by **15** with (*S*)- α -methylbenzylamine at 70 °C for 72 h in dry THF. Yield: 45%, colorless oil. [α]_D²⁰ = -36.0 (c 0.23, MeOH). ¹H NMR (500 MHz, CDCl₃): δ = 0.84 (3H, d, J = 6.3 Hz), 0.83–0.89 (2H, m), 1.01 (1H, t, J = 12.5 Hz), 1.22–1.29 (3H, m), 1.37–1.43 (1H, m), 1.42 (3H, d, J = 6.6 Hz), 1.49 (3H, d, J = 6.9 Hz), 1.51–1.57 (2H, m), 1.66–1.80 (4H, m), 2.61 (1H, br s), 2.77 (2H, d, J = 5.0 Hz), 3.70 (1H, t, J = 6.9 Hz), 3.77 (1H, s), 5.08 (1H, d, J = 7.1 Hz), 7.23–7.41 (10H, m), 7.65 (1H, br s). ¹³C NMR (125 MHz, CDCl₃): δ = 22.0, 22.1, 22.3, 25.2, 25.9, 29.8, 34.7, 42.0, 42.8, 46.5, 47.8, 49.2, 58.9, 66.3, 126.3, 126.5, 127.3, 128.5, 128.7, 128.8, 129.0, 143.7, 172.9. Anal. Calcd for C₂₆H₃₆N₂O₂: C, 76.43; H, 8.88; N, 6.86. Found: C, 76.45; H, 8.83; N, 6.87.

4.6. General Procedure for the Hydrolysis of β -Aminoamides

The solution of β -aminoamides 21–23 or 32–34 (0.5 mmol) in EtOH (2 mL) mixed with 10% aqueous HCl (10 mL) was stirred at room temperature. After completion of the reaction (as monitored by TLC, 24 h), the mixture was extracted with CH₂Cl₂ (3 × 10 mL), dried over Na₂SO₄, filtered, and evaporated to dryness. The crude product was purified by recrystallization with diethyl ether, resulting in compounds 5a–7 or 13–15, respectively. All spectroscopic data are listed above.

4.7. General Procedure for Preparation of Dipeptides

To the solution of α -methylene- γ -butyrolactone **2** or *N*-benzyl aminolactone **5a** (1.2 mmol) in dry EtOH (2.0 mL) was added L- or β -alanine ethyl ester (3.6 mmol). The mixture was stirred at the appropriate temperature for 48 h. When the reaction was complete (monitored by TLC), the mixture was evaporated to dryness, then purified by column chromatography on silica gel (CHCl₃/MeOH = 19:1), affording compounds **27–29**.

$4.7.1.\ Ethyl\ 3-((R)-2-((1S,2R,4R)-2-hydroxy-4-methylcyclohexyl)-3-((3-ethoxy-3-oxopropyl)amino) propanemido)$ propanoate (27)

Prepared from **2** with *β*-alanine ethyl ester at 25 °C. Yield: 63%, colorless oil. $[\alpha]_D^{20} = -14.2$ (c 0.33, MeOH). ¹H NMR (500 MHz, CDCl₃): $\delta = 0.81$ –1.09 (5H, m), 0.90 (3H, d, J = 6.2 Hz), 1.25–1.28 (11H, m), 1.33–1.40 (1H, m), 1.60 (2H, t, J = 14.6 Hz), 1.72 (1H, t, J = 10.2 Hz), 2.00 (1H, d, J = 13.2 Hz), 2.59 (2H, t, J = 6.3 Hz), 0.83 (1H, d, J = 17.6 Hz), 2.98–3.02 (1H, m), 3.18–3.37 (5H, m), 3.49–3.60 (3H, m), 4.14 (2H, q, J = 7.2 Hz), 4.19 (2H, q, J = 6.4 Hz). ¹³C NMR (125 MHz, CDCl₃): $\delta = 14.1$, 14.2, 22.0, 25.3, 29.7, 30.3, 31.8, 34.1, 34.2, 35.4, 42.1, 43.9, 44.0, 45.1, 47.8, 60.7, 61.6, 70.0, 170.8, 172.3, 173.2. Anal. Calcd for C₂₀H₃₆N₂O₆: C, 59.98; H, 9.06; N, 6.99. Found: C, 60.00; H, 9.05; N, 6.95.

4.7.2. Ethyl 3-((R)-3-(benzylamino)-2-((1S,2R,4R)-2-hydroxy-4-methylcyclohexyl)propanamido)propanoate (28)

Prepared from **5a** with *β*-alanine ethyl ester at 70 °C. Yield: 45%, white crystals, m.p.: 169–173 °C. [α]_D²⁰ = −24.0 (c 0.24, MeOH). ¹H NMR (500 MHz, CDCl₃): δ = 0.78–0.98 (3H, m), 0.85 (3H, d, J = 6.4 Hz), 1.24 (3H, t, J = 7.1 Hz), 1.23–1.29 (1H, m), 1.58 (2H, d, J = 10.6 Hz), 1.71 (2H, t, J = 11.5 Hz), 2.57 (2H, t, J = 5.7 Hz), 3.22–3.33 (3H, m), 3.42–3.47 (1H, m), 3.52–3.60 (2H, m), 4.08 (3H, q, J = 7.1 Hz), 4.31–4.34 (1H, m), 7.40 (3H, 6.0 Hz), 7.57 (2H, d, J = 5.3 Hz), 7.86 (1H, br s), 8.18 (1H, t, J = 5.5 Hz).10.08 (1H, br s). ¹³C NMR (125 MHz, CDCl₃): δ = 14.3, 22.0, 25.4, 31.8, 34.0, 34.3, 35.6, 42.3, 43.9, 44.7, 47.8, 52.3, 60.9, 70.2, 129.3, 129.7, 130.1, 130.7, 172.8, 173.4. Anal. Calcd for C₂₂H₃₄N₂O₄: C, 67.66; H, 8.78; N, 7.17. Found: C, 67.70; H, 8.75; N, 7.20.

4.7.3. (S)-Ethyl

2-((R)-3-(benzylamino)-2-((1S,2R,4R)-2-hydroxy-4-methylcyclohexyl)propanamido)propanoate (29)

Prepared from **5a** with L-alanine ethyl ester at 70 °C. Yield: 40%, white crystals, m.p.: 115–117 °C. [α] $_D^{20}$ = -30.0 (c 0.25, MeOH). 1 H NMR (500 MHz, CDCl $_3$): δ = 0.81–0.89 (2H, m), 0.88 (3H, d, J = 6.5 Hz), 1.04 (1H, q, J = 11.8 Hz), 1.25 (3H, t, J = 7.1 Hz), 1.28–1.38 (1H, m), 1.47 (3H, d, J = 7.2 Hz), 1.58–1.66 (2H, m), 1.81–1.90 (2H, m), 1.95–2.17 (2H, m), 3.10–3.18 (1H, m), 3.27–3.34 (2H, m), 3.64 (1H, d, J = 10.2 Hz), 4.00–4.03 (1H, m), 4.14–4.18 (3H, m), 4.42 (1H, quin, J = 6.9 Hz), 7.39 (3H, d, J = 3.6 Hz), 7.51 (2H, d, J = 3.8 Hz), 7.99 (1H, d, J = 6.1 Hz), 10.1 (br s). 13 C NMR (125 MHz, CDCl $_3$): δ = 14.3, 17.3, 22.1, 25.2, 31.8, 34.1, 42.1, 43.8, 44.2, 48.0, 49.0, 52.3, 52.3, 61.3, 70.0, 129.4, 129.8, 130.1, 130.4, 173.0, 173.2. Anal. Calcd for C $_{22}$ H $_{34}$ N $_{2}$ O $_{4}$: C, 67.66; H, 8.78; N, 7.17. Found: C, 67.65; H, 8.80; N, 7.15.

4.8. General Procedure for Debenzylation

To a suspension of 5% Pd/C or Pd(OH)₂/C (100 mg) in MeOH (10 mL) was added β -aminoamides **21–23** and **32–34** or *N*-benzyldipepetides **28–29** (0.38 mmol) in MeOH (10 mL). The mixture was stirred under H₂ at room temperature and normal pressure. When the reaction was complete (indicated by TLC), the mixture was filtered through a Celite pad, the solution was evaporated to dryness and purified by recrystallization in diethyl ether providing **24–26** and **35–37** as well as **30–31**, respectively.

4.8.1. (R)-3-Amino-N-benzyl-2-((1S,2R,4R)-2-hydroxy-4-methylcyclohexyl)propanamide (24)

Prepared from **21** with 5% Pd/C for 96 h. Yield: 80%, white crystals, m.p.: 226–230 °C. $[\alpha]_D^{20}$ = -24.0 (c 0.29, MeOH). ¹H NMR (500 MHz, DMSO- d_6): δ = 0.67–0.74 (1H, m), 0.82–0.97 (2H, m), 0.85 (3H, d, J = 6.4 Hz), 1.34–1.37 (2H, m), 1.54 (1H, d, J = 12.0 Hz), 1.69 (1H, t, J = 12.3 Hz), 1.87 (1H, d, J = 11.9 Hz), 2.81 (1H, dd, J = 1.9, 10.1 Hz), 3.08 (1H, t, J = 11.9 Hz), 3.17 (1H, d, J = 10.3 Hz), 3.28 (1H, td, J = 3.9, 10.4 Hz), 4.18 (1H, dd, J = 5.4, 15.1 Hz), 4.43 (1H, dd, J = 6.4, 15.2 Hz), 7.21–7.32 (5H, m), 7.91 (3H, br s), 8.72 (1H, t, J = 5.8 Hz). ¹³C NMR (125 MHz, DMSO- d_6): δ = 22.1, 25.0, 30.9, 34.1, 35.5, 42.2, 42.6, 44.7, 46.8, 68.7, 126.7, 127.1, 128.2, 139.6, 172.2. Anal. Calcd for $C_{17}H_{26}N_2O_2$: C, 70.31; H, 9.02; N, 9.65. Found: C, 70.35; H, 9.05; N, 9.60.

4.8.2. (R)-3-Amino-2-((1S,2R,4R)-2-hydroxy-4-methylcyclohexyl)-N-((R)-1-phenylethyl) propanamide (25)

Prepared from **22** with 5% Pd/C for 168 h. Yield: 62%, white crystals, m.p.: 145–150 °C. $\left[\alpha\right]_D^{20}$ = +28.7 (c 0.31, MeOH). ¹H NMR (500 MHz, DMSO- d_6): δ = 0.73–0.79 (1H, m), 0.87 (3H, d, J = 6.4 Hz), 0.90–1.00 (2H, m), 1.34 (3H, d, J = 6.9 Hz), 1.37–1.46 (2H, m), 1.59 (1H, d, J = 12.2 Hz), 1.81 (1H, t, J = 12.3 Hz), 1.88 (1H, d, J = 11.9 Hz), 2.81 (1H, dd, J = 2.6, 12.1 Hz), 3.05 (1H, d, J = 12.0 Hz), 3.14–3.17 (1H, m), 3.28 (1H, td, J = 3.7, 10.3 Hz), 5.01 (1H, quin, J = 7.4 Hz), 7.20–7.40 (5H, m), 8.65 (1H, d, J = 8.3 Hz). ¹³C NMR (125 MHz, DMSO- d_6): δ = 22.1, 22.6, 25.1, 31.0, 34.3, 35.5, 42.5, 44.8, 46.5, 48.0, 68.9, 126.4, 126.6, 128.2, 144.3, 171.4. Anal. Calcd for C₁₈H₂₈N₂O₂: C, 71.02; H, 9.27; N, 9.20. Found: C, 71.05; H, 9.30; N, 9.15.

4.8.3. (R)-3-Amino-2-((1S,2R,4R)-2-hydroxy-4-methylcyclohexyl)-*N*-((S)-1-phenylethyl) propanamide (**26**)

Prepared from **23** (0.16 g, 0.39 mmol) with Pd(OH)₂/C for 200 h. Yield: 65%, white crystals, m.p.: 150–160 °C. $[\alpha]_D^{20} = -37.4$ (c 0.31, MeOH). ¹H NMR (500 MHz, DMSO- d_6): $\delta = 0.59$ –0.66 (1H, m), 0.74–0.82 (1H, m), 0.84 (3H, d, J = 6.4 Hz), 0.88–0.97 (1H, m), 1.06 (1H, d, J = 10.8 Hz), 1.22–1.26 (1H, m), 1.30–1.37 (1H, m), 1.36 (3H, d, J = 7.0 Hz), 1.42 (1H, d, J = 12.1 Hz), 1.77–1.87 (2H, m), 2.75 (1H, d, J = 11.8 Hz), 3.00 (1H, d, J = 12.0 Hz), 3.16–3.25 (2H, m), 4.93 (1H, quin, J = 7.3 Hz), 7.18–7.29 (5H, m), 8.77 (1H, d, J = 7.7 Hz). ¹³C NMR (125 MHz, DMSO- d_6): $\delta = 22.1$, 22.4, 24.8, 30.9, 42.1, 44.7, 46.7, 48.3, 68.7, 125.7, 126.5, 128.1, 145.5, 171.4. Anal. Calcd for C₁₈H₂₈N₂O₂: C, 71.02; H, 9.27; N, 9.20. Found: C, 71.00; H, 9.25; N, 9.23.

4.8.4. Ethyl 3-((R)-3-amino-2-((1S,2R,4R)-2-hydroxy-4-methylcyclohexyl)propanamido)propanoate (30)

Prepared from **28** with 5% Pd/C for 24 h. Yield: 50%, colorless oil. $[\alpha]_D^{20} = -15.5$ (c 0.31, MeOH). ¹H NMR (500 MHz, DMSO- d_6): $\delta = 0.66$ –0.73 (1H, m), 0.79–1.05 (2H, m), 0.83 (3H, d, J = 6.2 Hz), 1.17 (3H, t, J = 7.0 Hz), 1.22–1.28 (1H, m), 1.29–1.35 (1H, m) 1.41–1.53 (3H, m), 1.73–1.82 (1H, m), 2.36–2.43 (3H, m), 2.59–2.76 (3H, m), 3.16 (1H, s), 3.19–3.24 (3H, m), 4.04 (2H, q, J = 7.1 Hz). ¹³C NMR (125 MHz, DMSO- d_6): $\delta = 14.1$, 22.2, 26.0, 31.1, 34.0, 34.4, 34.7, 44.8, 46.4, 59.9, 69.3. Anal. Calcd for $C_{15}H_{28}N_2O_2$: C, 59.97; H, 9.40; N, 9.33. Found: C, 60.00; H, 9.45; N, 9.30.

4.8.5. (S)-Ethyl

2-((R)-3-amino-2-((1S,2R,4R)-2-hydroxy-4-methylcyclohexyl)propanamido)propanoate (31)

Prepared from **29** with 5% Pd/C for 24 h. Yield: 55%, colorless oil. $[\alpha]_D^{20} = -20.0$ (c 0.30, MeOH). ¹H NMR (500 MHz, DMSO- d_6): $\delta = 0.70$ –0.77 (1H, m), 0.85 (3H, d, J = 6.4 Hz), 0.84–0.93 (2H, m), 1.15 (3H, t, J = 7.1 Hz), 1.28 (2H, d, J = 7.3 H), 1.36 (1H, br s), 1.47–1.56 (2H, m), 1.74 (1H, t, J = 11.6 Hz), 1.86 (1H, d, J = 12.7 Hz), 2.78 (1H, d, J = 11.1 Hz), 3.04 (1H, t, J = 11.8 Hz), 3.13 (1H, d, J = 10.3 Hz), 3.26 (1H, td, J = 3.6, 10.2 Hz), 4.04 (2H, q, J = 6.9 Hz), 4.26 (1H, quin, J = 7.0 Hz), 8.58 (1H, d, J = 1.3 Hz). ¹³C NMR (125 MHz, DMSO- d_6): $\delta = 14.0$, 16.5, 22.2, 24.8, 30.9, 34.2, 35.4, 42.3, 44.7, 46.5, 48.0, 60.4, 68.7, 172.3, 172.6. Anal. Calcd for C₁₅H₂₈N₂O₂: C, 59.97; H, 9.40; N, 9.33. Found: C, 59.97; H, 9.38; N, 9.35.

4.8.6. (S)-3-Amino-N-benzyl-2-((1S,2S,4R)-2-hydroxy-4-methylcyclohexyl)propanamide (35)

Prepared from **32** with 5% Pd/C for 96 h. Yield: 70%, white crystals, m.p.: 270–275 °C. $[\alpha]_D^{20}$ = +36.0 (c 0.26, MeOH). ¹H NMR (500 MHz, DMSO- d_6): 0.74–0.86 (1H, m), 0.79 (3H, d, J = 6.4 Hz), 1.01 (1H, t, J = 12.8 Hz), 1.18–1.26 (2H, m), 1.34–1.40 (1H, m), 1.47 (1H, t, J = 11.4 Hz), 1.55 (1H, d, J = 12.5 Hz), 1.69 (2H, d, J = 11.0 Hz), 2.23–2.27 (1H, m), 2.67–2.71 (1H, m), 2.77–2.80 (1H, m), 3.84 (1H, s), 4.22–4.31 (1H, m), 7.20–7.32 (5H, m), 8.39 (1H, t, J = 5.6 Hz). ¹³C NMR (125 MHz, DMSO- d_6): δ = 22.5, 25.1, 25.5, 34.6, 40.8, 41.3, 41.9, 42.3, 50.6, 64.3, 126.6, 127.2, 128.2, 139.9, 174.7. Anal. Calcd for $C_{17}H_{26}N_2O_2$: C, 70.31; H, 9.02; N, 9.65. Found: C, 70.29; H, 9.03; N, 9.60.

4.8.7. (S)-3-Amino-2-((1S,2S,4R)-2-hydroxy-4-methylcyclohexyl)-N-((R)-1-phenylethyl)propanamide (36)

Prepared from **33** with 5% Pd/C for 240 h. Yield: 70%, white crystals, m.p.: 245–250 °C. $[\alpha]_D^{20}$ = +20.0 (c 0.24, MeOH). ¹H NMR (500 MHz, DMSO- d_6): δ = 0.65–0.72 (1H, m), 0.78 (3H, d, J = 6.3 Hz), 0.95–1.02 (1H, m), 1.36 (3H, d, J = 7.0 Hz), 1.35–1.39 (1H, m), 1.48–1.82 (2H, m), 1.65 (2H, d, J = 12.2 Hz), 2.58–2.64 (1H, m), 2.93 (1H, d, J = 12.1 Hz), 3.02–3.06 (1H, m), 3.74 (1H, s), 4.60 (1H, s), 4.93 (1H, quin, J = 7.3 Hz), 7.20–7.30 (5H, m), 7.87 (3H, br s), 8.68 (1H, d, J = 7.8 Hz). ¹³C NMR (125 MHz, DMSO- d_6): δ = 22.3, 22.5, 24.4, 25.2, 34.3, 37.9, 42.0, 44.9, 48.3, 63.9, 126.0, 126.6, 128.1, 144.7, 171.7. Anal. Calcd for C₁₈H₂₈N₂O₂: C, 71.02; H, 9.27; N, 9.20. Found: C, 71.00; H, 9.25; N, 9.25.

4.8.8. (S)-3-Amino-2-((1S,2S,4R)-2-hydroxy-4-methylcyclohexyl)-N-((S)-1-phenylethyl) propanamide (37)

Prepared from **34** with Pd(OH)₂/C for 300 h. Yield: 52%, white crystals, m.p.: 225–228 °C. $\left[\alpha\right]_D^{20} = -32.0$ (c 0.24, MeOH). ¹H NMR (500 MHz, DMSO- d_6): $\delta = 0.78$ –0.85 (1H, m), 0.82 (3H, d, J = 6.3 Hz), 0.96–1.19 (1H, m), 1.02 (3H, d, J = 6.3 Hz), 1.10 (3H, d, J = 6.0 Hz), 1.35 (3H, d, J = 7.0 Hz), 1.44–1.55 (2H, m), 1.61–1.73 (3H, m), 2.64 (1H, br s), 2.92 (1H, d, J = 11.4 Hz), 3.03–3.15 (2H, m), 3.89 (1H, s), 4.96 (1H, quin, J = 7.0 Hz), 7.20–7.36 (5H, m), 8.68 (1H, d, J = 7.5 Hz). ¹³C NMR (125 MHz, DMSO- d_6): $\delta = 22.3$, 22.5, 24.4, 25.3, 34.4, 41.3, 42.1, 48.0, 49.0, 64.1, 126.2, 126.7, 128.2, 144.2, 171.9. Anal. Calcd for $C_{18}H_{28}N_2O_2$: C, 71.02; H, 9.27; N, 9.20. Found: C, 70.97; H, 9.30; N, 9.17.

4.9. Determination of Antiproliferative Properties

The human cancer cell lines isolated from cervical adenocarcinoma (HeLa) and breast cancers (MCF7 and MDA-MB-231) were purchased from European Collection of Cell Cultures (Salisbury, UK). The cells were maintained in Minimum Essential Medium (MEM) supplemented with fetal calf serum (10%), non-essential amino acids (1%), and penicillin-streptomycin (1%) at 37 °C in a humidified atmosphere containing 5% CO_2 . All media and supplements for these experiments were obtained from Lonza Group Ltd. (Basel, Switzerland). The antiproliferative properties of the prepared compounds were determined by the MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] assay [33]. Briefly, cells were seeded into 96 well plates (5000 cells/well) and incubated with the tested compounds at 10 and 30 μ M under cell-culturing conditions for 72 h. Then MTT solution (5 mg/mL) was added to each sample, which were incubated for a further 4 h. The formazan crystals precipitated were dissolved in 100 μ L dimethyl sulfoxide, and the absorbance was measured at 545 nm with a microplate reader (Awareness Technology, Palm City, FL, USA). Two independent experiments were performed with five wells for each condition. Cisplatin (Ebewe GmbH, Unterach, Austria), a clinically used anticancer agent, was used as a reference agent. Calculations were performed by means of the GraphPad Prism 5.01 software (GraphPad Software Inc., San Diego, CA, USA).

Supplementary Materials: Supplementary materials can be found at http://www.mdpi.com/1422-0067/19/11/3522/s1.

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Conflicts of Interest: The authors declare no conflict of interest.

Abbreviations

DCM Dichloromethane
DMF Dimethylformamide
THF Tetrahydrofuran

EtOH Ethanol

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