



Copinion Role of Leukotriene B₄ **Receptor-2 in Mast Cells in Allergic Airway Inflammation**

Sun-Young Kwon[®] and Jae-Hong Kim *[®]

Department of Biotechnology, College of Life Sciences, Korea University, 5-1 Anam-dong, Sungbuk-gu, Seoul 02841, Korea; kwonsy1992@naver.com

* Correspondence: jhongkim@korea.ac.kr; Tel.: +82-2-3290-3452

Received: 30 May 2019; Accepted: 12 June 2019; Published: 14 June 2019



Abstract: Mast cells are effector cells in the immune system that play an important role in the allergic airway inflammation. Recently, it was reported that BLT2, a low-affinity leukotriene (LT) B₄ receptor, plays a pivotal role in the pathogenesis of allergic airway inflammation through its action in mast cells. We observed that highly elevated expression levels of BLT2 are critical for the pathogenesis leading to allergic airway inflammation, and that if BLT2 expression is downregulated by siBLT2-mediated knockdown, allergic inflammation is dramatically alleviated. Furthermore, we demonstrated that BLT2 mediates the synthesis of vascular endothelial growth factor (VEGF) and Th2 cytokines, such as interleukin (IL)-13, in mast cells during allergic inflammation. Based on the critical roles of BLT2 in mast cells in allergic inflammation, anti-BLT2 strategies could contribute to the development of new therapies for allergic airway inflammation.

Keywords: allergic airway inflammation; mast cells; leukotriene B4 receptor-2; asthma

1. Introduction

Mast cells are multifunctional cells in the immune system that were first identified in 1878 by Paul Ehrlich, who was awarded the Nobel Prize in Physiology or Medicine in 1908 [1,2]. Mast cells, which are key components of both innate and adaptive immunity, are involved in many biological responses, such as immune response and inflammation reactions [1,3]. Mast cells are also well known for their roles in allergic diseases, such as asthma, allergic rhinitis, atopic dermatitis, and allergic conjunctivitis [4–6]. In particular, mast cells play roles in the pathophysiology of the allergic airway inflammation in asthma [7,8]. In addition, previous studies suggested that mast cells play a role in neoplastic angiogenesis through the expression of vascular endothelial growth factors (VEGFs) and their receptors [9,10]. Indeed, human mast cells were shown to be both a source and a target of angiogenic and lymphangiogenic factors [9,10]. Further, mast cells were reported to participate in systemic autoimmune disorders (e.g., rheumatoid arthritis) and cancer [11,12]. Mast cells express a wide variety of membrane receptors on their cell surfaces, including high-affinity receptor for IgE (FcɛRI), Toll-like receptors (TLRs), and complement receptors [13]. The cross-linking of allergen-specific IgE immune complexes and FcɛRI leads to the rapid release of inflammatory mediators and cytokines (e.g., histamine, leukotrienes (LTs), and interleukin (IL)-13) by mast cells [14].

Arachidonic acid is metabolized to biologically active mediators like, thromboxanes, prostaglandins, thromboxanes, and leukotrienes (LTs) [15]. Recent work has demonstrated the importance of LTs in the pathogenesis of allergic inflammation. The LTs are formed by transformation of arachidonic acid into an unstable epoxide intermediate, LTA₄, which can be converted enzymatically by hydration to LTB₄, and by addition of glutathione to LTC₄, a member of cysteinyl LTs [15]. Among LTs, LTB₄ is a potent proinflammatory lipid mediator derived from arachidonic acid by the action of 5-lipoxygenase (5-LO) [16]. LTB₄ is one of the most potent known chemoattractants, acting primarily

on neutrophils, eosinophils, T cells, and mast cells [5,16–18]. LTB₄ has been widely implicated in the pathogenesis of several inflammatory diseases, including asthma, psoriasis, rheumatoid arthritis, and inflammatory bowel disease [19]. LTB₄ also exerts its biological functions via two types of G protein-coupled receptors (GPCRs): BLT1, which is predominantly expressed in peripheral blood leukocytes, and BLT2, which is expressed ubiquitously [19]. Previous studies have reported that both BLT1 and BLT2 are expressed in human and murine mast cells, and that both receptors contribute to the chemotactic migration of mast cells toward LTB₄ [18,19]. LTB₄ activates the BLT1 and BLT2, whereas additional lipid mediators (12(*S*)-hydroxyeicosatetraenoic acid (HETE) and 12-hydroxyheptadecatrienoic acid (12-HHT)) are also agonists to the BLT2 [18,19]. Previous studies have suggested that BLT2 plays a mediatory role in the pathogenesis of the allergic airway inflammation in asthma through its action in mast cells [20–23]. This review focuses on the recently discovered novel roles of BLT2 in mast cells during allergic airway inflammation.

2. Role of BLT2 in Mast Cells

2.1. Role of BLT2 in Allergen-Induced Th2 Cytokine Synthesis in Mast Cells

Mast cells are one of the major effector cells in allergic asthma and other allergic inflammatory diseases [24,25]. Mast cell-derived mediators are critical for the initiation of inflammatory responses and are associated with allergic diseases [20]. Mast cells express the high-affinity IgE receptor (F $c\epsilon$ RI) on their surface. The cross-linking of cell surface-bound IgE to $Fc\epsilon RI$ by an allergen (Ag) leads to the rapid release of inflammatory mediators, including histamines, leukotrienes, and cytokines [24,25]. Among these mediators, Th2 cytokines (e.g., IL-4 and IL-13) produced by mast cells have been reported to regulate important allergic inflammatory responses, such as asthma, atopic dermatitis, allergic rhinitis, and anaphylaxis, as well as allergies to drugs, toxins, and food [26–28]. Previous studies have reported that the activation of mast cells through $Fc \in RI$ leads to the production of intracellular reactive oxygen species (ROS) [29], which plays roles in the pathogenesis of various inflammatory diseases, including asthma and cancer [30,31]. Our previous study showed that BLT2 mediates the synthesis of Th2 cytokines through ROS generation in Ag-stimulated mast cells [20]. In addition, BLT2 blockade by siBLT2-mediated knockdown or antagonist treatment suppressed IL-4 and IL-13 production in allergic asthma [20]. Quite importantly, the protein levels of IL-4 and IL-13 were also modestly increased by BLT2 overexpression alone; the addition of BLT2 ligands further enhanced cytokine synthesis [20], suggesting a mediatory role of BLT2 in the allergen-induced Th2 cytokine synthesis. In contrast to BLT2, BLT1 overexpression had no significant effect on cytokine production in bone marrow-derived mast cells (BMMC) (data not shown). In summary, BLT2 may play roles in the pathogenesis of the allergic airway inflammatory response and Ag-stimulated Th2 cytokine synthesis in mast cells. In addition to the Ag–IgE action to $Fc\epsilon RI$, TLR4 activation by lipopolysaccharide (LPS) or ST2 activation by IL-33 were also reported to stimulate the release of BLT2 agonist lipid mediators (e.g., LTB₄ and 12(S)-HETE) that can activate BLT2, thus leading to the synthesis of Th2 cytokines, such as IL-13, as will be discussed later.

2.2. Role of BLT2 in Ag-Induced VEGF Synthesis in Mast Cells

Ag-induced cross-linking of cell surface-bound IgE to Fc ϵ RI activates mast cells to release the following three classes of mediators: (1) preformed mediators (e.g., histamines and proteases); (2) lipid mediators (e.g., prostaglandin D₂ (PGD₂), LTB₄, and LTC₄); and (3) a variety of chemokines and cytokines as well as VEGF [9,10,32]. VEGF is one of the regulators of vascular angiogenesis, permeability, and remodeling [33–35]. Angiogenesis, the process by which new capillaries develop from the preexisting vasculature, plays a key role in various pathogenesis [9]. And, mast cells were shown to express VEGFs and their receptors to mediate vascular and neoplastic angiogenesis [9,10]. A previous study reported that ROS regulate the activity of nuclear factor kappa-light-chain-enhancer of activated B cells (NF- κ B) [36], a primary transcription factor involved in VEGF expression in mast

cells [37,38]. In addition, mast cells regulate airway inflammation and hyperresponsiveness through orchestrating VEGF expression in allergic asthmatic responses [39]. Thus, VEGF expression in mast cells is likely to play an essential role in the initiation and development of allergic asthmatic responses. Indeed, levels of angiogenic factors, including VEGF-A, were shown to be increased in the airways of asthmatic subjects [10,40,41]. In a previous study, we observed that blocking BLT2 completely abrogated the production of VEGF in Ag-stimulated bone marrow-derived mast cells (BMMCs) [21]. The synthesis of BLT2 ligands (e.g., LTB_4 and 12(S)-HETE) was also required for the VEGF production, suggesting a mediating role of an autocrine BLT2 ligands-BLT2 axis in the production of VEGF in mast cells [21]. Further, we demonstrated that the Nox1-reactive oxygen species (ROS)-NF-KB cascade is downstream of BLT2 during Ag signaling to VEGF synthesis in mast cells, implying that BLT2 mediates the synthesis of VEGF through the Nox1-ROS-NF-κB cascade in Ag-stimulated mast cells [21]. Additionally, we observed that VEGF levels in BMMCs isolated from BLT2-overexpressing TG mice were clearly elevated compared with BMMCs derived from WT mice [21]. Further, VEGF levels were much enhanced by treatment with BLT2 ligands in BMMCs derived from BLT2 TG mice [21]. In summary, these results suggest that BLT2 plays a mediatory role in VEGF production in Ag-stimulated mast cells, potentially contributing to the allergic airway inflammation.

2.3. Role of BLT2 in LPS-Induced IL-13 Synthesis in Mast Cells

Recently, the roles of TLRs in the pathogenesis of allergic asthma have received much attention [42]. For instance, the activation of TLR4 by lipopolysaccharide (LPS), particularly at low doses, has been reported to exacerbate allergic airway inflammation by activating mast cells and promoting Th2 responses [43,44]. In addition, it has been suggested that mast cells play a role in this response, as LPS-exacerbated airway inflammation and Th2 cytokine production are not observed in mast cell-deficient mice. Furthermore, the adoptive transfer of bone marrow-derived mast cells (BMMCs) from TLR4-deficient mice into mast cell-deficient mice does not restore allergic airway inflammation and Th2 cytokine production, while the transfer of BMMCs from wild-type mice into mast cell-deficient mice restores allergic airway inflammation [44,45]. These results suggest that TLR4 in mast cells plays a role in bacterial infection-induced exacerbation of allergic responses. Previous studies have demonstrated that Th2 cytokines induced by TLR4 activation in mast cells are critical for allergic inflammation. For example, Th2 cytokine production induced via the activation of TLR4 in mast cells by LPS was shown to be dependent on the MyD88-NF- κ B signaling pathway [46,47]. Similarly, our recent study showed that BLT2 plays a pivotal role in LPS-induced Th2 cytokine production, especially IL-13 synthesis, in mast cells via a TLR4-MyD88-BLT2-ROS-NF-κB-linked cascade [22]. Moreover, we suggested that BLT2 likely acts as a potential connection between the innate and adaptive immune systems, leading to a Th2 response in mast cells [21,22].

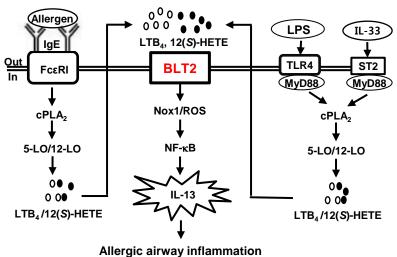
2.4. Role of BLT2 in IL-33-Induced IL-13 Synthesis in Mast Cells

IL-33, a member of the IL-1 cytokine family, acts as an alarmin that is immediately released in response to epithelial cell damage [48–50]. Previously, IL-33 was suggested to be closely associated with asthmatic development and exacerbation in animal model study, as IL-33 levels are increased when the severity of asthma is increased in mouse [51]. The relationship between IL-33 and asthma has been demonstrated using an IL-33-overexpressing transgenic mouse, which showed asthmatic airway inflammation with an influx of inflammatory cells and increased expression levels of Th2 cytokines [52]. Consistent with the proposed role of IL-33 in asthma, earlier studies showed that blockade of IL-33 or its cell surface receptor ST2 clearly reduces disease severity, as evidenced by reduced Th2 cytokine production, airway inflammation, airway hyperresponsiveness, and mucus production [53]. Although mast cells are known to respond to exogenous danger signals [50], recent studies have suggested that mast cells play a critical role in the recognition of endogenous danger signals such as IL-33 [50]. Indeed, several studies have demonstrated that mast cells are a major ST2-expressing cell type and are associated with the IL-33-related development of asthma [53–55]. For example, the activation of mast cells by IL-33

has been shown to enhance airway hyperresponsiveness and asthmatic inflammation via the production of Th2 cytokines such as IL-13 [48,56,57]. In addition, IL-13 secreted from IL-33-stimulated mast cells was shown to increase airway smooth muscle contraction and contribute to the development of airway inflammation [57]. Previous studies have reported that 5-LO or 12-lipoxygenase (12-LO) is closely associated with the development of allergic asthmatic inflammation in animal model studies [58-60]. For instance, LTB_4 , a 5-LO product, has been demonstrated to perform important functions during the development of asthma. In consistent with these findings in animal studies, HETE, a 12-LO product, was detected at higher levels in the sputum and bronchoalveolar lavage fluid (BALF) of asthmatic patients than in those of healthy subjects [59]. These findings suggest that 5-/12-LO are involved in the development of asthmatic airway inflammation in both animal model and clinical subjects. Recently, we found that 5-/12-LO plays critical roles in IL-33 signaling to induce IL-13 production in mast cells [23]. In addition, we reported that the "MyD88-5-/12-LO-BLT2-NF-κB" signaling pathway contributes to IL-33-induced IL-13 synthesis in mast cells, potentially exacerbating the airway inflammation. Thus, the action of BLT2 in mast cells plays a role in IL-33 signaling, potentially contributing to IL-13 synthesis and allergic airway inflammation. Quite recently, IL-33 was reported to participate in innate and adaptive immunity and inflammation, and acting on CD34+ cells causes mast cells differentiation and maturation [61–63]. Clearly, further studies are necessary to investigate the roles of BLT2 in mast cells, especially in terms of mast cells stimulation, differentiation, and maturation.

3. Conclusions

Mast cells, which are principal effector cells in the immune system, play roles in the allergic airway inflammation. In recent studies, we demonstrated that BLT2 as well as its ligands LTB_4 and 12-HHT plays an important role in the synthesis of VEGF and Th2 cytokines in Ag-stimulated mast cells, contributing to allergic airway inflammation. In addition, we demonstrated that this autocrine-acting BLT2 cascade mediates LPS/TLR4- or IL-33-induced synthesis of Th2 cytokines in mast cells. Based on those findings, we proposed a hypothetical model as summarized in Figure 1.



Allergic all way initialititation

Figure 1. Scheme of BLT2-dependent lipopolysaccharide/Toll-like receptors 4 (LPS/TLR4), interleukin (IL)-33/ST2, or allergen/ high-affinity IgE receptor (Fc ϵ RI) immune response signaling pathways responsible for IL-13 synthesis in mast cells, contributing to the allergic airway inflammation. Mast cells activation by allergen/Fc ϵ RI, LPS/TLR4, or IL-33/ST2 stimuli activates cytosolic phospholipase A₂(cPLA₂)-5-/12-LO-BLT2 cascades and induces allergic airway inflammation by significantly increasing Th2 cytokines synthesis (e.g., IL-13) production via the reactive oxygen species-nuclear factor kappa-light-chain-enhancer of activated B cells (ROS-NF κ B) pathway.

This model proposes that BLT2 likely acts as a potential connection between the innate and adaptive immune systems, leading to a Th2 response, at least in mast cells. In this model, NOX1/ROS and NF- κ B lies downstream of BLT2, thus mediating the innate and adaptive signaling for IL-13 production. Thus, understanding the detailed roles of BLT2 in mast cells during allergic airway inflammation may contribute to the development of new therapeutic targets for allergic airway inflammation. Indeed, we are currently working on the development of BLT2 receptor antagonists as a novel therapy for allergic disorders.

Author Contributions: Writing—original draft preparation, S.-Y.K.; writing—review and editing, S.-Y.K. and J.-H.K.; and supervision, J.-H.K.

Funding: This work was supported by Bio and Medical Technology Development Program grants (2017M3A9D8063317) and a Mid-Career Researcher Program grant (2017R1A2B4002203) through the National Research Foundation (NRF) funded by the Ministry of Science, Information and Communication Technologies (ICT), and Future Planning, Republic of Korea. This work was also supported by the BK21 Plus Program (College of Life Sciences and Biotechnology, Korea University) and Korea University Grant.

Conflicts of Interest: The authors declare that they have no conflicts of interest.

References

- 1. Gonzalez-de-Olano, D.; Alvarez-Twose, I. Mast cells as key players in allergy and inflammation. *J. Investig. Allergol. Clin. Immunol.* **2018**, *28*, 365–378. [CrossRef] [PubMed]
- Piro, A.; Tagarelli, A.; Tagarelli, G.; Lagonia, P.; Quattrone, A. Paul Ehrlich: The Nobel Prize in physiology or medicine 1908. *Int. Rev. Immunol.* 2008, 27, 1–17. [CrossRef] [PubMed]
- 3. Beaven, M.A. Our perception of the mast cell from Paul Ehrlich to now. *Eur. J. Immunol.* 2009, 39, 11–25. [CrossRef] [PubMed]
- 4. Galli, S.J.; Tsai, M. IgE and mast cells in allergic disease. Nat. Med. 2012, 18, 693–704. [CrossRef] [PubMed]
- Ohnishi, H.; Miyahara, N.; Gelfand, E.W. The role of leukotriene B(4) in allergic diseases. *Allergol. Int.* 2008, 57, 291–298. [CrossRef] [PubMed]
- 6. Gomez, G. Current strategies to inhibit high affinity FcepsilonRI-mediated signaling for the treatment of allergic disease. *Front. Immunol.* **2019**, *10*, 175. [CrossRef] [PubMed]
- Cruse, G.; Bradding, P. Mast cells in airway diseases and interstitial lung disease. *Eur. J. Pharmacol.* 2016, 778, 125–138. [CrossRef]
- 8. Brown, J.M.; Wilson, T.M.; Metcalfe, D.D. The mast cell and allergic diseases: Role in pathogenesis and implications for therapy. *Clin. Exp. Allergy* **2008**, *38*, 4–18. [CrossRef]
- 9. Varricchi, G.; Loffredo, S.; Borriello, F.; Pecoraro, A.; Rivellese, F.; Genovese, A.; Spadaro, G.; Marone, G. Superantigenic activation of human cardiac mast cells. *Int. J. Mol. Sci.* **2019**, *20*, 1828. [CrossRef]
- Detoraki, A.; Staiano, R.I.; Granata, F.; Giannattasio, G.; Prevete, N.; de Paulis, A.; Ribatti, D.; Genovese, A.; Triggiani, M.; Marone, G. Vascular endothelial growth factors synthesized by human lung mast cells exert angiogenic effects. *J. Allergy Clin. Immunol.* 2009, 123, 1142–1149. [CrossRef]
- 11. Varricchi, G.; Loffredo, S.; Galdiero, M.R.; Marone, G.; Cristinziano, L.; Granata, F.; Marone, G. Innate effector cells in angiogenesis and lymphangiogenesis. *Curr. Opin. Immunol.* **2018**, *53*, 152–160. [CrossRef] [PubMed]
- 12. Varricchi, G.; Pecoraro, A.; Marone, G.; Criscuolo, G.; Spadaro, G.; Genovese, A.; Marone, G. Thymic Stromal Lymphopoietin Isoforms, Inflammatory Disorders, and Cancer. *Front. Immunol.* **2018**, *9*, 1595. [CrossRef] [PubMed]
- Krystel-Whittemore, M.; Dileepan, K.N.; Wood, J.G. Mast cell: A multi-functional master cell. *Front. Immunol.* 2015, 6, 620. [CrossRef] [PubMed]
- 14. Kubo, M. Mast cells and basophils in allergic inflammation. *Curr. Opin. Immunol.* **2018**, *54*, 74–79. [CrossRef] [PubMed]
- 15. Samuelsson, B. Leukotrienes: Mediators of immediate hypersensitivity reactions and inflammation. *Science* **1983**, 220, 568–575. [CrossRef] [PubMed]
- 16. Tager, A.M.; Luster, A.D. BLT1 and BLT2: The leukotriene B(4) receptors. *Prostaglandins Leukot. Essent. Fatty Acids* **2003**, *69*, 123–134. [CrossRef]
- 17. Kim, G.Y.; Lee, J.W.; Cho, S.H.; Seo, J.M.; Kim, J.H. Role of the low-affinity leukotriene B4 receptor BLT2 in VEGF-induced angiogenesis. *Arterioscler. Thromb. Vasc. Biol.* **2009**, *29*, 915–920. [CrossRef]

- 18. Park, J.; Jang, J.H.; Park, G.S.; Chung, Y.; You, H.J.; Kim, J.H. BLT2, a leukotriene B4 receptor 2, as a novel prognostic biomarker of triple-negative breast cancer. *BMB Rep.* **2018**, *51*, 373–377. [CrossRef]
- 19. Okuno, T.; Yokomizo, T. Biological functions of 12(S)-hydroxyheptadecatrienoic acid as a ligand of leukotriene B4 receptor 2. *Inflamm. Regen.* **2018**, *38*, 29. [CrossRef]
- 20. Cho, K.J.; Seo, J.M.; Lee, M.G.; Kim, J.H. BLT2 is upregulated in allergen-stimulated mast cells and mediates the synthesis of Th2 cytokines. *J. Immunol.* **2010**, *185*, 6329–6337. [CrossRef]
- 21. Lee, A.J.; Ro, M.; Kim, J.H. Leukotriene B4 Receptor 2 is critical for the synthesis of vascular endothelial growth factor in allergen-stimulated mast cells. *J. Immunol.* **2016**, *197*, 2069–2078. [CrossRef] [PubMed]
- Lee, A.J.; Ro, M.; Cho, K.J.; Kim, J.H. Lipopolysaccharide/TLR4 stimulates IL-13 production through a MyD88-BLT2-linked cascade in mast cells, potentially contributing to the allergic response. *J. Immunol.* 2017, 199, 409–417. [CrossRef] [PubMed]
- 23. Ro, M.; Lee, A.J.; Kim, J.H. 5-/12-lipoxygenase-linked cascade contributes to the IL-33-induced synthesis of IL-13 in mast cells, thus promoting asthma development. *Allergy* **2018**, *73*, 350–360. [CrossRef] [PubMed]
- 24. Elieh Ali Komi, D.; Bjermer, L. Mast cell-mediated orchestration of the immune responses in human allergic asthma: Current insights. *Clin. Rev. Allergy Immunol.* **2018**. [CrossRef] [PubMed]
- Da Silva, E.Z.; Jamur, M.C.; Oliver, C. Mast cell function: A new vision of an old cell. *J. Histochem. Cytochem.* 2014, 62, 698–738. [CrossRef] [PubMed]
- 26. Kubo, M. Innate and adaptive type 2 immunity in lung allergic inflammation. *Immunol. Rev.* 2017, 278, 162–172. [CrossRef]
- Burd, P.R.; Thompson, W.C.; Max, E.E.; Mills, F.C. Activated mast cells produce interleukin 13. J. Exp. Med. 1995, 181, 1373–1380. [CrossRef]
- 28. Barnes, P.J. Th2 cytokines and asthma: An introduction. Respir. Res. 2001, 2, 64-65. [CrossRef]
- 29. Swindle, E.J.; Metcalfe, D.D. The role of reactive oxygen species and nitric oxide in mast cell-dependent inflammatory processes. *Immunol. Rev.* 2007, 217, 186–205. [CrossRef]
- 30. Henricks, P.A.; Nijkamp, F.P. Reactive oxygen species as mediators in asthma. *Pulm. Pharmacol. Ther.* **2001**, 14, 409–420. [CrossRef]
- 31. Liou, G.Y.; Storz, P. Reactive oxygen species in cancer. Free Radic. Res. 2010, 44, 479–496. [CrossRef] [PubMed]
- 32. Marshall, J.S. Mast-cell responses to pathogens. Nat. Rev. Immunol. 2004, 4, 787–799. [CrossRef] [PubMed]
- 33. Meyer, N.; Akdis, C.A. Vascular endothelial growth factor as a key inducer of angiogenesis in the asthmatic airways. *Curr. Allergy Asthma Rep.* **2013**, *13*, 1–9. [CrossRef] [PubMed]
- Shaik-Dasthagirisaheb, Y.B.; Varvara, G.; Murmura, G.; Saggini, A.; Potalivo, G.; Caraffa, A.; Antinolfi, P.; Tete, S.; Tripodi, D.; Conti, F.; et al. Vascular endothelial growth factor (VEGF), mast cells and inflammation. *Int. J. Immunopathol. Pharmacol.* 2013, 26, 327–335. [CrossRef] [PubMed]
- Lee, C.G.; Ma, B.; Takyar, S.; Ahangari, F.; Delacruz, C.; He, C.H.; Elias, J.A. Studies of vascular endothelial growth factor in asthma and chronic obstructive pulmonary disease. *Proc. Am. Thorac. Soc.* 2011, *8*, 512–515. [CrossRef] [PubMed]
- Gloire, G.; Legrand-Poels, S.; Piette, J. NF-kappaB activation by reactive oxygen species: Fifteen years later. Biochem. Pharmacol. 2006, 72, 1493–1505. [CrossRef] [PubMed]
- 37. Xie, T.X.; Xia, Z.; Zhang, N.; Gong, W.; Huang, S. Constitutive NF-kappaB activity regulates the expression of VEGF and IL-8 and tumor angiogenesis of human glioblastoma. *Oncol. Rep.* **2010**, *23*, 725–732.
- Huang, S.; Pettaway, C.A.; Uehara, H.; Bucana, C.D.; Fidler, I.J. Blockade of NF-kappaB activity in human prostate cancer cells is associated with suppression of angiogenesis, invasion, and metastasis. *Oncogene* 2001, 20, 4188–4197. [CrossRef]
- 39. Lee, K.S.; Kim, S.R.; Park, S.J.; Min, K.H.; Lee, K.Y.; Choe, Y.H.; Park, S.Y.; Chai, O.H.; Zhang, X.; Song, C.H.; et al. Mast cells can mediate vascular permeability through regulation of the PI3K-HIF-1alpha-VEGF axis. *Am. J. Respir. Crit. Care Med.* **2008**, *178*, 787–797. [CrossRef]
- Chetta, A.; Zanini, A.; Foresi, A.; D'Ippolito, R.; Tipa, A.; Castagnaro, A.; Baraldo, S.; Neri, M.; Saetta, M.; Olivieri, D. Vascular endothelial growth factor up-regulation and bronchial wall remodeling in asthma. *Clin. Exp. Allergy* 2005, *35*, 1437–1442. [CrossRef]
- 41. Hoshina, M.; Takahashi, M.; Aoike, N. Expression of vascular endothelial growth factor, basic fibroblast growth factor, and angiogenin immunoreactivity in asthmatic airways and its relationship to angiogenesis. *J. Allergy Clin. Immunol.* **2001**, *107*, 295–301. [CrossRef] [PubMed]

- 42. Athari, S.S.; Athari, S.M.; Beyzay, F.; Movassaghi, M.; Mortaz, E.; Taghavi, M. Critical role of toll-like receptors in pathophysiology of allergic asthma. *Eur. J. Pharmacol.* **2017**, *808*, 21–27. [CrossRef] [PubMed]
- 43. Nigo, Y.I.; Yamashita, M.; Hirahara, K.; Shinnakasu, R.; Inami, M.; Kimura, M.; Hasegawa, A.; Kohno, Y.; Nakayama, T. Regulation of allergic airway inflammation through toll-like receptor 4-mediated modification of mast cell function. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 2286–2291. [CrossRef] [PubMed]
- 44. Yamashita, M.; Nakayama, T. Progress in allergy signal research on mast cells: Regulation of allergic airway inflammation through toll-like receptor 4-mediated modification of mast cell function. *J. Pharmacol. Sci.* **2008**, *106*, 332–335. [CrossRef] [PubMed]
- 45. Murakami, D.; Yamada, H.; Yajima, T.; Masuda, A.; Komune, S.; Yoshikai, Y. Lipopolysaccharide inhalation exacerbates allergic airway inflammation by activating mast cells and promoting Th2 responses. *Clin. Exp. Allergy* **2007**, *37*, 339–347. [CrossRef] [PubMed]
- 46. Avila, M.; Gonzalez-Espinosa, C. Signaling through toll-like receptor 4 and mast cell-dependent innate immunity responses. *IUBMB Life* **2011**, *63*, 873–880. [CrossRef] [PubMed]
- 47. Gon, Y.; Nunomura, S.; Ra, C. Common and distinct signalling cascades in the production of tumour necrosis factor-alpha and interleukin-13 induced by lipopolysaccharide in RBL-2H3 cells. *Clin. Exp. Allergy* **2005**, *35*, 635–642. [CrossRef]
- 48. Jang, T.Y.; Kim, Y.H. Interleukin-33 and mast cells bridge innate and adaptive immunity: From the allergologist's perspective. *Int. Neurourol. J.* **2015**, *19*, 142–150. [CrossRef]
- Khaitov, M.R.; Gaisina, A.R.; Shilovskiy, I.P.; Smirnov, V.V.; Ramenskaia, G.V.; Nikonova, A.A.; Khaitov, R.M. The role of interleukin-33 in pathogenesis of bronchial asthma. New experimental data. *Biochemistry (MOSC)* 2018, 83, 13–25. [CrossRef]
- 50. Saluja, R.; Khan, M.; Church, M.K.; Maurer, M. The role of IL-33 and mast cells in allergy and inflammation. *Clin. Transl. Allergy* **2015**, *5*, 33. [CrossRef]
- Sjoberg, L.C.; Nilsson, A.Z.; Lei, Y.; Gregory, J.A.; Adner, M.; Nilsson, G.P. Interleukin 33 exacerbates antigen driven airway hyperresponsiveness, inflammation and remodeling in a mouse model of asthma. *Sci. Rep.* 2017, 7, 4219. [CrossRef] [PubMed]
- Zhiguang, X.; Wei, C.; Steven, R.; Wei, D.; Wei, Z.; Rong, M.; Zhanguo, L.; Lianfeng, Z. Over-expression of IL-33 leads to spontaneous pulmonary inflammation in mIL-33 transgenic mice. *Immunol. Lett.* 2010, 131, 159–165. [CrossRef] [PubMed]
- Lee, H.Y.; Rhee, C.K.; Kang, J.Y.; Byun, J.H.; Choi, J.Y.; Kim, S.J.; Kim, Y.K.; Kwon, S.S.; Lee, S.Y. Blockade of IL-33/ST2 ameliorates airway inflammation in a murine model of allergic asthma. *Exp. Lung Res.* 2014, 40, 66–76. [CrossRef] [PubMed]
- Enoksson, M.; Lyberg, K.; Moller-Westerberg, C.; Fallon, P.G.; Nilsson, G.; Lunderius-Andersson, C. Mast cells as sensors of cell injury through IL-33 recognition. *J. Immunol.* 2011, *186*, 2523–2528. [CrossRef] [PubMed]
- Borish, L.; Steinke, J.W. Interleukin-33 in asthma: How big of a role does it play? *Curr. Allergy Asthma Rep.* 2011, *11*, 7–11. [CrossRef] [PubMed]
- 56. Sjoberg, L.C.; Gregory, J.A.; Dahlen, S.E.; Nilsson, G.P.; Adner, M. Interleukin-33 exacerbates allergic bronchoconstriction in the mice via activation of mast cells. *Allergy* **2015**, *70*, 514–521. [CrossRef]
- 57. Kaur, D.; Gomez, E.; Doe, C.; Berair, R.; Woodman, L.; Saunders, R.; Hollins, F.; Rose, F.R.; Amrani, Y.; May, R.; et al. IL-33 drives airway hyper-responsiveness through IL-13-mediated mast cell: Airway smooth muscle crosstalk. *Allergy* **2015**, *70*, 556–567. [CrossRef]
- 58. Di Gennaro, A.; Haeggstrom, J.Z. The leukotrienes: Immune-modulating lipid mediators of disease. *Adv. Immunol.* **2012**, *116*, 51–92. [CrossRef]
- 59. Mabalirajan, U.; Rehman, R.; Ahmad, T.; Kumar, S.; Leishangthem, G.D.; Singh, S.; Dinda, A.K.; Biswal, S.; Agrawal, A.; Ghosh, B. 12/15-lipoxygenase expressed in non-epithelial cells causes airway epithelial injury in asthma. *Sci. Rep.* **2013**, *3*, 1540. [CrossRef]
- Mabalirajan, U.; Ahmad, T.; Rehman, R.; Leishangthem, G.D.; Dinda, A.K.; Agrawal, A.; Ghosh, B.; Sharma, S.K. Baicalein reduces airway injury in allergen and IL-13 induced airway inflammation. *PLoS ONE* 2013, *8*, e62916. [CrossRef]
- Tettamanti, L.; Kritas, S.K.; Gallenga, C.E.; D'Ovidio, C.; Mastrangelo, F.; Ronconi, G.; Caraffa, A.; Toniato, E.; Conti, P. IL-33 mediates allergy through mast cell activation: Potential inhibitory effect of certain cytokines. *J. Biol. Regul. Homeost. Agents* 2018, *32*, 1061–1065. [PubMed]

- 62. Varvara, G.; Tettamanti, L.; Gallenga, C.E.; Caraffa, A.; D'Ovidio, C.; Mastrangelo, F.; Ronconi, G.; Kritas, S.K.; Conti, P. Stimulated mast cells release inflammatory cytokines: Potential suppression and therapeutical aspects. *J. Biol. Regul. Homeost. Agents* **2018**, *32*, 1355–1360. [PubMed]
- 63. Gallenga, C.E.; Pandolfi, F.; Caraffa, A.; Kritas, S.K.; Varvara, G.; Ronconi, G.; Toniato, E.; Martinotti, S.; Conti, P. Interleukin-1 family cytokines and mast cells: Activation and inhibition. *J. Biol. Regul. Homeost. Agents* **2018**, 33, 1–6.



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (http://creativecommons.org/licenses/by/4.0/).