



Review

# Strengthening the AntiTumor NK Cell Function for the Treatment of Ovarian Cancer

Marco Greppi <sup>1</sup>, Giovanna Tabellini <sup>2</sup>, Ornella Patrizi <sup>2</sup>, Simona Candiani <sup>3</sup> , Andrea Decensi <sup>4,5</sup>, Silvia Parolini <sup>2</sup>, Simona Sivori <sup>1,\*</sup>, Silvia Pesce <sup>6,†</sup>, Laura Paleari <sup>7,†</sup> and Emanuela Marcenaro <sup>1,\*</sup>

<sup>1</sup> Department of Experimental Medicine (DIMES) and Centre of Excellence for Biomedical Research (CEBR), University of Genoa, 16132 Genoa, Italy; marcogreppi92@gmail.com

<sup>2</sup> Department of Molecular and Translational Medicine, University of Brescia, 25123 Brescia, Italy; giovanna.tabellini@unibs.it (G.T.); ornella.patrizi@unibs.it (O.P.); silvia.parolini@unibs.it (S.P.)

<sup>3</sup> Department of Earth Science, Environment and Life (DISTAV), University of Genoa, 16132 Genoa, Italy; candiani@unige.it

<sup>4</sup> Division of Medical Oncology, Galliera Hospital, 16126 Genoa, Italy; andrea.decensi@galliera.it

<sup>5</sup> Wolfson Institute of Preventive Medicine, Queen Mary University of London, London E1 4NS, UK

<sup>6</sup> Department of Experimental Medicine (DIMES), University of Genoa, 16132 Genoa, Italy; silvia.pesce@unige.it

<sup>7</sup> A.Li.Sa., Liguria Health Authority, 16121 Genoa, Italy; laura.paleari@regione.liguria.it

\* Correspondence: simona.sivori@unige.it (S.S.); emanuela.marcenaro@unige.it (E.M.); Tel.: +39-010-3537888 (S.S. & E.M.)

† These authors share senior authorship.

Received: 31 January 2019; Accepted: 15 February 2019; Published: 19 February 2019



**Abstract:** The crosstalk between cancer cells and host cells is a crucial prerequisite for tumor growth and progression. The cells from both the innate and adaptive immune systems enter into a perverse relationship with tumor cells to create a tumor-promoting and immunosuppressive tumor microenvironment (TME). Epithelial ovarian cancer (EOC), the most lethal of all gynecological malignancies, is characterized by a unique TME that paves the way to the formation of metastasis and mediates therapy resistance through the deregulation of immune surveillance. A characteristic feature of the ovarian cancer TME is the ascites/peritoneal fluid, a malignancy-associated effusion occurring at more advanced stages, which enables the peritoneal dissemination of tumor cells and the formation of metastasis. The standard therapy for EOC involves a combination of debulking surgery and platinum-based chemotherapy. However, most patients experience disease recurrence. New therapeutic strategies are needed to improve the prognosis of patients with advanced EOC. Harnessing the body's natural immune defenses against cancer in the form of immunotherapy is emerging as an innovative treatment strategy. NK cells have attracted attention as a promising cancer immunotherapeutic target due to their ability to kill malignant cells and avoid healthy cells. Here, we will discuss the recent advances in the clinical application of NK cell immunotherapy in EOC.

**Keywords:** ovarian cancer; NK cells; immunotherapy; immune checkpoint; PD-1; activating receptors; B7-H6; antitumor activity; hormone therapy; adoptive therapy

## 1. Overview on Epithelial Ovarian Cancer

Epithelial ovarian cancer (EOC) is an endocrine-related neoplasm and it is classified as a rare cancer both in the portal for rare diseases and orphan drugs Orphanet (<http://www.orpha.net/ORPHA398934>) and in the National Institutes of Health (NIH) Register, Genetic and Rare Diseases (GARD) (<https://rarediseases.info.nih.gov/>). Most EOCs are diagnosed at an advanced stage, which

accounts for the high mortality rate associated with this disease. The new World Health Organization (WHO) Classification of Ovarian Cancer takes the recent findings on the origin, pathogenesis and prognosis of different ovarian cancer subtypes into account. The tubal origin of hereditary and some non-hereditary high-grade serous cancers is mentioned in contrast to the previous theory of mesothelial origin of tumors while seromucinous tumors represent a new entity [1]. Several studies over the past decade have demonstrated that histological grade is one of the most important prognostic factors in EOC, having found important differences in the molecular and clinical characteristics of a low-grade serous carcinoma of the ovary (LGSCO) compared with a high-grade serous carcinoma of the ovary [2–13]. LGSCO represents approximately 10% of all serous ovarian carcinomas that are diagnosed at a younger age and individuals diagnosed with LGSCO experience a longer overall survival (OS) than those with high-grade disease [14]. Despite these differences, most women with EOC have been treated identically in the last three decades independently of the histological grade of their tumors. A growing body of research has questioned this strategy. Recent advances in the understanding of the tumor heterogeneity, including refinement in pathologic criteria, elucidation of molecular and genetic tumoral differences as well as disparate responses to treatment with chemotherapy, have led to the initiation of separate clinical trials according to epithelial histology subtypes through the NRG Oncology (Gynecologic Oncology Group) Rare Tumor Committee [14–16]. Surgery is effective in most cases of early stage EOC (Federation Internationale des Gynaecologistes et Obstetristes—FIGO stage I-IIA) with a 5-year survival rate of around 90% [17]. After surgery, the treatment of choice for advanced EOC (stage IIB-IV) is platinum-based chemotherapy (CT) [18]. Conversely, women with LGSCO exhibit poor response rates to conventional chemotherapy and remain at high risk of recurrence and cancer-related death, especially in the setting of advanced stage disease [2,14–16,19–21].

## 2. Epithelial Ovarian Cancer: A Focus on Tumor Escape Mechanisms Impairing NK Cell Function

EOC is the 7th cause of death among women with malignancies worldwide and the leading cause of death from gynecological cancers (<https://www.uicc.org/new-global-cancer-data-globocan-2018>) [22]. EOC spreads predominantly in the peritoneal cavity and is often accompanied by a massive production of (malignant) ascites. This increasing volume of ascites can generate a favorable tumor microenvironment, enabling the characteristic patterns of transcoelomic tumor spread in ovarian cancer [23]. This ascites is rich in tumor-promoting soluble factors [24,25], extracellular vesicles [26] and detached cancer cells [27] as well as large numbers of immune cells, such as Natural Killer (NK) cells, T cells and tumor-associated macrophages (TAMs). After being influenced by the tumor microenvironment, these cells are unable to defend our body against tumors but instead cooperate with resident host cells to support tumor progression and immune evasion [25,28].

The malignant transformation of normal cells comes from a multifactorial process, resulting in genomic instability [29] and a modification of immunosurveillance mechanisms that induces tolerance. To reach this state, tumors develop different strategies during their evolution to escape the immune response: (i) the secretion of immunosuppressive cytokines or soluble tumor-derived inhibitory factors [24,25]; (ii) the induction of coinhibitory receptors (e.g., immune checkpoints) [30,31]; or the dampening of costimulatory receptors [25,32,33] on infiltrated lymphocytes.

NK lymphocytes represent one of the most efficient cellular mechanisms by which the immune system can recognize and kill tumors. With an array of receptors evolved to sense cellular alterations, NK cells provide early protection against cancer cells by producing cytokines and chemokines in addition to collaborating with other immune cells and exerting direct cytolytic activity [34–36]. In particular, NK cells express a repertoire of activating and inhibitory receptors. The integration of signaling generated by these receptors will determine the activation status of these cells [37]. No single receptor dominates as synergistic signals from combinations of receptors are instead integrated to activate or inhibit natural cytotoxicity and cytokine production. A prevalence of inhibitory signals induces the blocking of NK cell functions while a prevalence of activating signals leads to the activation of NK cells and consequently the killing of NK-susceptible target cells. Despite the fact that NK cells

display potent cytolytic activity against tumor cells *in vitro*, this functional capability may be strongly impaired by the TME. Indeed, ascites not only contain large numbers of growth factors and cytokines that are able to promote the proliferation of tumor cells but they can also suppress the function of otherwise normal immune effectors, including NK cells [25,30,32,33].

Fresh NK cells (CD56<sup>+</sup>CD3<sup>-</sup>) isolated from the ascites fluid are found in relatively high concentrations compared to peripheral blood and in particular, they result enriched in CD56<sup>bright</sup> NK cells. However, although they are present in a large number, they display functional impairment [38,39]. In this regard, it has been shown that the presence of IL-18 and TGFβ in ascites can induce a strong downregulation of CD16 on NK cells, resulting in diminished antibody-dependent cell-mediated cytotoxicity (ADCC) against autologous tumor cells [25,32,40,41]. In addition, TGFβ can also contribute to the downmodulation of the NKp30 and NKG2D NK cell activating receptors [42] thus impairing NK cell-mediated natural cytotoxicity. NKG2D expression may be also impaired by macrophage migration inhibitory factor (MIF), which is another soluble factor that is detectable in ascites [43]. Recently, additional mechanisms of tumor escape have been described, including the ability of tumor cells to release soluble forms of activating NK cell receptor ligands. The chronic receptor–ligand interaction may dampen the surface expression of the activating NK receptors, thus affecting the ability of NK cells to kill tumor cells that express ligands for those receptors. In particular, ovarian cancer cells may release a soluble form of B7-H6, the main NKp30 ligand, leading to the loss of NKp30 expression on NK cells in the TME. A high amount of soluble B7-H6 is correlated with a greater downmodulation of NKp30. These NK cells display impaired IFN-γ production and cytolytic function, thereby showing poor NK cell-mediated elimination of B7-H6<sup>+</sup> ovarian cancer cells [25]. Consistent with these observations, a lower level of B7-H6 expression is correlated with a better OS and reduced metastasis and cancer progression in ovarian cancer patients [44]. Similar results have also been shown for NKG2D. Indeed, soluble MIC-A and MIC-B, two NKG2D ligands that are released by tumor cells in the TME, can downmodulate the expression of this activating receptor, which is an event associated with an adverse clinical outcome [33]. Finally, the activating NK cell receptor DNAM-1 can also be downmodulated by the chronic exposure to the ligand expressed on the surface of ovarian tumor cells [32].

Recently, the attention of researchers has been focused on other molecules expressed on immune cells, which are the inhibitory receptors that are defined as immune checkpoints. Under healthy conditions, these receptors maintain self-tolerance and modulate the duration and amplitude of immune responses upon the recognition of specific ligands on normal cells in order to prevent collateral tissue damage. However, it is now known that tumor cells may coopt these inhibitory receptors in order to avoid immune surveillance [45]. For this reason, immune checkpoint expression and engagement are now known as one of the major mechanisms related to tumor resistance or escape from the immune system-mediated attack. In fact, during tumorigenesis, cancer cells often express ligands to bind these receptors and induce immune suppression. NK cells are equipped with several inhibitory receptors. In particular, NK cell immune checkpoints include the well-known HLA-class I-specific receptors KIR, LIR-1 and NKG2A, but also non-HLA class I-specific receptors, such as PD-1. This receptor was originally discovered on cytolytic T cells and was found to exert a sharp inhibitory effect on their proliferation, cytokine production and antitumor activity upon binding to its ligands (PD-L1 and PD-L2). Thus, by expressing PD-1 ligands, tumors have evolved a remarkable mechanism to avoid T cell-mediated surveillance of cancer. A recent discovery from our research group showed that PD-1 is also expressed on a discrete fraction of NK cells in patients with EOC [30]. *In vitro* experiments showed that these cells display a strongly reduced capacity to kill PD-L<sup>+</sup> tumor cells as well as impaired release of interferon-γ (IFN-γ) and tumor necrosis factor-α (TNF-α) cytokines after stimulation with the same PD-L<sup>+</sup> tumor targets. Remarkably, these impaired effector functions induced by PD-1/PD-L interaction could be partially reverted by mAbs specific for PD-L1/PD-L2 [30].

Despite the fact that the functional capability of NK cells may be strongly impaired by the TME, NK-based immunotherapeutic approaches for treatment of tumors have garnered attention, primarily for NK cell capability of killing malignant cells without toxicity towards healthy cells.

### 3. State-of-the-Art Therapies Targeting Ovarian Cancer

Despite the prognosis of early-stage ovarian cancers being favorable with approximately 90% of patients surviving 5 years after diagnosis [46], more than 70% of patients are diagnosed with advanced disease (FIGO stage III-IV) due to the lack of sensitive screening during the early stages [47]. Many of these patients initially benefit from integrated surgery and platinum/taxane-based chemotherapy although recurrence develops in nearly 90% of cases. Furthermore, 70% of patients with advanced disease succumb to tumor relapse within less than five years [48,49]. This poor prognosis is attributed to the development of drug-resistance through the selection of chemoresistant clones, intraperitoneal spreading of tumor cells and formation of metastatic lesions and tumor relapse. In this regard, there is a clear urgent need for alternative treatments to improve the clinical outcome of patients with advanced ovarian carcinoma.

The currently available effective approaches for recurrent ovarian cancer include cytokine therapy, adoptive transfer of NK cells, hormone therapy and antibody-based immunotherapy [50].

#### 3.1. Cytokine Therapy

Early clinical trials in ovarian cancer patients aimed to improve the antitumor activity of immune cells through intraperitoneal injections of different biologic products, including *Bacillus Calmette-Guerin*, *Corynebacterium parvum* and an attenuated strain of influenza virus [51,52]. These treatments had limited clinical responses mainly due to the small number and heterogeneity of study participants.

Another immunotherapeutic approach for ovarian cancer is the intraperitoneal administration of cytokines to potentiate an autologous antitumor response in vivo. In this context, the results of several clinical trials evaluating intraperitoneal therapy with IL-2 alone or in combination with other therapies demonstrated that cytokine therapy was generally well tolerated and may improve lymphocyte and NK cell counts. However, cytokine therapy had variable levels of success and was mainly dependent on the remaining tumor burden before the start of therapy [53–57]. IL-15, which is similar to IL-2, can strongly increase NK cell numbers and may also enhance NK cell function in the ovarian cancer setting [58,59]. Currently, several clinical trials evaluating IL-15 are ongoing [60]. In this regard, it has been demonstrated that monomeric IL-15 or the IL-15 superagonist fusion complex, ALT-803, potently increases the function of ascites-derived NK cells [61,62].

#### 3.2. Adoptive Therapy of Immune Cells

An additional approach in ovarian cancer involves the adoptive transfer of immune cells isolated from the peripheral blood of patients, which was activated with various cytokines and subsequently infused back into the same patient. This aims to improve the autologous antitumor responses [63,64].

The early adoptive transfer of autologous lymphokine-activated killer (LAK) cells with a high dose of IL-2 demonstrated limited clinical responses with high rates of peritoneal fibrosis [65–67]. Cytokine-induced killer (CIK) cells (derived again from peripheral blood and stimulated with antiCD3 mAbs, IFN- $\gamma$  and IL-2) [68] demonstrated enhanced cytotoxic activity compared to LAK cells against ovarian cancer [69]. Recently, promising results were obtained by a phase III clinical trial in which the adoptive transfer of autologous CIK cells after primary debulking surgery and adjuvant carboplatin/paclitaxel chemotherapy was assessed [70]. These studies suggest that allogeneic NK cell therapy is feasible although further efforts that will generate novel strategies to increase in vivo NK cell persistence and expansion after adoptive transfer are needed.

In this regard, it has been reported that adoptive NK cells induced by different cytokines (IL-12, IL-15, IL-18) display both in vitro and in vivo enhanced functionality and persistence against ovarian cancer. Notably, this higher NK activity was detectable even upon exposure to ascitic fluid, thus suggesting its capability to circumvent the immunosuppressive nature of ovarian cancer TME [71].

In addition, the ex vivo inhibition of GSK3 kinase in peripheral blood induces an enrichment of mature adoptive NK cells from cytomegalovirus positive donors and enhances their cytokine

production and ADCC when exposed to tumor cells [72]. A phase I clinical trial using the product generated from this method has been started at the University of Minnesota (NCT03213964).

Many NK cell-adoptive therapies against malignancies are currently in clinical practice, including hematopoietic stem cell transplantation. NK cell infusions can provide safe and effective immunotherapy against tumor relapse [73]. Usually, these therapies use “adult” cell populations, such as hematopoietic stem cells (HSCs) from bone marrow (BM), peripheral blood (PB) or cord blood (CB) cells. Recent studies demonstrated the ability of “non-adult” human pluripotent stem cells (h-PSCs) to generate NK cells. The proportion of mature and functional cytolytic NK cells is higher from the hPSCs-derived progenitor cells [74,75]. This probably allows hPSC-NK cells to mediate an increased antitumor response both *in vitro* and *in vivo*, thus providing an alternative source of cells for the immunotherapy of different type of tumors, including ovarian cancer.

### 3.3. Hormone Therapy in Ovarian Cancer

A putative direct action of gonadal steroids on ovarian carcinogenesis has been suggested, which was supported by findings of mRNA transcripts and translated proteins of Estrogen receptor (ER) and Progesterone receptor (PgR) in both normal ovarian tissue and malignant ovarian tumors. A direct action of estrogen on EOC growth, metastasis and progression has been demonstrated through different pathways, including: (i) tumor production of vascular endothelial growth factor (VEGF) via ER signaling (direct pathway); and (ii) increased tumor–endothelial cell migration via mitogen-activated protein kinase (MAPK) signaling (indirect pathway) [76]. PgR activation induces apoptosis, cell cycle arrest and senescence in ovarian cancer cells, which strongly suggests the modulation of PgR levels and/or activity as a form of endocrine treatment of EOC [77]. In a recent large observational study, Sieh et al. (2013) found a high positivity for ER/PgR (60–80%) in high grade EOC, with a prognostic value that is associated with significantly improved survival [78]. Moreover, it has been shown that ER and PgR are twice as likely to be expressed in LGSCOs than in high-grade serous carcinomas of the ovary [15]. Since hormone therapy may become a viable and extremely cost-effective option for the treatment of EOC, we meta-analyzed 53 clinical trials to assess the Clinical Benefit Rates (CBRs) and deaths in EOC after hormone therapy [79]. Overall, we found a summary estimate of CBR (SCBR) of 41% (95% CI = 0.34–0.48) and 46% (95% CI = 0.34–0.57) for ER<sup>+</sup> and/or PgR<sup>+</sup> tumors while this CBR was 40% (95% CI = 0.29–0.51) in platinum resistant tumors. In particular, subgroup analyses by type of hormonal treatment showed a SCBR for aromatase inhibitors of 39% (95% CI = 0.29–0.50) and the highest clinical benefit of 43% (95% CI = 0.30–0.56) with tamoxifen, which is a selective ER modulator (SERM) that produces antiestrogen effects through competitive inhibition of the receptor itself. Explorative analyses according to line of treatment and histological grade were hampered by the low numbers although there was a tendency to find a greater effect in first-line (adjuvant setting) and LGSCO. Specifically, hormone therapy was associated with a significant reduction of mortality in LGSCO (HR = 0.66, 95% CI = 0.47–0.93) [79]. In a recent retrospective study by the MD Anderson group, 203 women with stage II–IV LGSCO who received hormonal maintenance therapy following primary treatment had a better outcome compared with those on only chemotherapy [80]. The median PFS was 26.4 months with surveillance and 64.9 months with hormonal therapy ( $p < 0.001$ ). Regarding OS, the subgroup analysis by disease at the end of adjuvant chemotherapy (disease free vs. persistent disease) showed a positive result in favor of hormonal therapy in both subgroups (in women who were disease-free, OS was 191.3 months vs. 106.8 months; in women with persistent disease, median OS was 83.3 months vs. 44.4 months). A stratified log-rank test adjusted for disease status determined that  $p = 0.014$ . A second study, performed at Johns Hopkins Hospital in conjunction with the Cleveland Clinic during the same period [81], utilized hormonal monotherapy after primary cytoreductive surgery and letrozole, which is an aromatase inhibitor that prevents estrogen synthesis, which was the predominant hormonal therapy used in 55.6%. The three-year PFS was 79.0% and three-year OS was 93.1%.

Interestingly, it has been shown that murine NK cells expressed ER $\alpha$  and ER $\beta$  and that 17 $\beta$ -estradiol elicited a significant decrease in NK activity in both wild type and ER $\alpha$ -deficient mice ( $p < 0.001$ ). This suggests that ER $\beta$  is involved in mediating the actions of estrogen on NK cell activity and increasing the potential for therapeutic modulation of NK cell activity [82]. Moreover, it has been demonstrated that the antitumor activity of NK cells in human is modulated by estrogen. The most effective treatment for breast cancer patients, the antiestrogen tamoxifen, has been shown to stimulate host NK cell activity and metastasis in xenograft models [83–85]. These data are taken into account with a view to design new therapeutic treatments for patients based on the use of NK cells in EOC therapy. In this regard, an innovative therapeutic approach against EOC could be based on the improvement in NK cell antitumor function by combining immunotherapy and hormone therapy.

### 3.4. Immunotherapy

The approaches using antibody (Ab)-based immunotherapy have also been explored and some are heading towards being used in the clinic. The mAbs generated to induce/amplify an antitumor response can function through different mechanisms, including the opsonization/activation of ADCC and blockage of immune checkpoints.

The first treatment that aims to target the previously identified ovarian cancer-associated antigens [including NY-ESO-1, CA 125 (MUC16), MUC1 and epithelial cell adhesion molecule (EpCAM)] with mAbs [55] or with new engineered bispecific antibodies and bispecific/trispecific killer engagers (BiKEs or TriKEs), which are molecules that crosslink tumor cells antigens (e.g., EpCAM) with CD16 on NK cells, thus activating/enhancing ADCC [86,87]. A fully humanized TriKE, which utilizes a modified IL-15 to crosslink the antiCD16 scFv and EpCAM scFv, not only sustains ADCC activity, but also mediates NK expansion, cytokine production and survival via IL-15 [88]. A TetraKE construct was recently engineered to simultaneously target EpCAM and CD133 bearing cells [89].

Given the results of GOG-0218 [90] and ICON7 [91] trials, bevacizumab, a mAb that functions in a non-immune-mediated manner by blocking VEGF, has recently been approved in combination with CT as a first-line treatment of advanced EOC (stage IIIb–IV) although an OS benefit has not been demonstrated [90,91]. Despite the use of bevacizumab, the disease prognosis remains poor as the European mean age-standardized 5-year OS was only 37.6% for women diagnosed between 2000 and 2007 [92] and the median OS in ICON7 in the CT plus bevacizumab arm was 58 months [91]. Despite this recent progress, not all patients may benefit from bevacizumab and the cost/benefit ratio of this drug remains unclear [93].

Another promising approach is the infusion of antitumor lymphocytes that were previously engineered with chimeric antigen receptors (CARs). These studies are mainly focused on cytotoxic T cells but recently, these technologies have also been applied to NK cells. In a recent study, the effect of the chemotherapeutic agent cisplatin in association with a CAR-based immunotherapeutic approach was evaluated to improve the clinical efficacy of ovarian cancer therapies. In particular, a lentiviral vector encoding a third-generation anti-CD133-CAR was generated and transduced in NK92 cells. This combined clinical approach led to a strong killing effect against ovarian cancer stem cells [94]. Efforts to generate additional ovarian cancer specific NK chimeric antigen receptors are ongoing [95].

In high-grade serous ovarian carcinomas, the most common histological type of ovarian cancer, almost 15% of women harbor germline BRCA mutations [96]. A recent study has shown that BRCA-mutated high-grade serous ovarian carcinomas have a high mutational load and there are more tumor-specific neoantigens [97,98] that recruit and activate tumor-infiltrating T lymphocytes. This induces a compensative upregulation of the immune checkpoint PD-1 on the surface of T cells. Immune checkpoints are inhibitory pathways that serve to prevent self-tissue damage under healthy conditions. During tumorigenesis, cancer cells often express ligands for this (PD-L1 and PD-L2) and other immune checkpoints, which downregulates T cell activity and induces immune suppression. In the case of BRCA-mutated ovarian carcinoma, the inhibition of the PD-1/PD-L1 pathway may induce cytotoxic lymphocyte activity against cancer cells. A recent clinical trial has shown antitumor activity

against tumors with mismatch repair deficiencies after treatment with immune checkpoint inhibitors (including nivolumab) [99,100]. In addition, a recent study suggests that nivolumab monotherapy in women with BRCA gene mutations, high-grade serous histology and recurrent Mullerian cancer may be an effective and well-tolerated salvage therapy [101]. Thus, targeting the PD-1 pathway with a checkpoint inhibitor is an attractive approach in hypermutated tumors. This indicates that the immune system is protective against ovarian cancer and thus adjuvant immunotherapy post-surgery and with chemotherapeutics could be effective for preventing relapse and extending survival [102,103].

Current efforts for a NK cell based therapy are mainly based on strategies that manipulate the function of inhibitory receptors. In this context, it has been recently demonstrated that ascites-associated NK cells can also express high levels of PD-1, with this expression potentially impairing the antitumor activity of these innate effectors [30]. These findings may extend the therapeutic use of anti-PD-1 mAbs to unleash the cytotoxic potential of NK cells against NK susceptible malignancies, including ovarian cancer cells. Moreover, the combined antibody-mediated blocking of multiple inhibitory checkpoints on NK cells, including NKG2A, KIR and PD-1, by triggering their ability to kill tumor cells is likely to facilitate the uptake of novel/additional tumor antigens by antigen presenting cells and subsequent massive recruitment of antigen-specific T lymphocytes [104]. In this context, a phase-1 dose-ranging study of anti-NKG2A (IPH2201) in patients with gynecologic malignancy, including high-grade serous ovarian/fallopian tube or peritoneal carcinoma, cervical cancer (squamous cell carcinoma) or endometrial cancer (adenocarcinoma), that is advanced/metastatic/recurrent or unresectable and for which no curative therapy exists is ongoing (NCT02459301, Canadian Cancer Trials Group, Collaborator: Innate Pharma).

Future research is needed to clarify the effects that checkpoint inhibitors have on the NK cell response and the potential to enhance adoptive NK cell immunotherapy in ovarian cancer [71].

A combination of these different techniques of NK cell-based immunotherapy hold great potential and may represent an effective weapon against ovarian cancer after primary cytoreductive surgery and adjuvant chemotherapy.

#### 4. Conclusions

Based on the evidence of the high frequency of persistent tumors at the completion of primary postoperative chemotherapy and the high relapse rate for women with advanced EOCs treated with standard surgery/chemotherapy, there is a clear medical need for identifying a management strategy that results in improved outcomes. Further studies are needed to provide new insights on the immunological mechanisms involved in the development of certain forms of ovarian cancer, which will be crucial for the development of new immunotherapeutic strategies (e.g., simultaneous blockage of different immune checkpoints, including hormone receptors, or infusion of antitumor lymphocytes previously engineered with CARs).

The recent evidence for a strong prognostic effect of ER in a large proportion of newly diagnosed EOC offers a therapeutic target for a very cost-effective precise medical approach.

Increasing our understanding of the mechanisms of NK cell activation, including those regulated by different molecular checkpoints and hormone receptors, and the identification of new tumor biomarkers will provide a firm basis for how to optimize NK cell reactivity against cancer. This knowledge can be applied to the development of an optimal design for cancer immunotherapy by targeting NK cells either alone or in combination with other therapies.

**Funding:** Supported by grants awarded by Associazione Italiana per la Ricerca sul Cancro (AIRC)-Special Program Metastatic disease: the key unmet need in oncology 5 per mille 2018 Id. 21147 (S.S., EM, S.Pe., S.P., M.G., S.C., A.D.); AIRC IG 2017 Id. 20312 (S.S., E.M., S.Pe., S.P., M.G., S.C.); AIRC IG 2018 Id. 21534 (A.D., L.P.); Progetto Roche per la Ricerca 2017 (S.Pe., E.M.). S.Pe. is recipient of a fellowship awarded by Fondazione Umberto Veronesi.

**Acknowledgments:** We dedicate this contribution to Alessandro Moretta who sadly passed away in mid-February 2018. We mourn his invaluable scientific insight and mentorship and even more, his humanity, irony and smile.

**Conflicts of Interest:** The authors declare no competing financial interests.

## References

1. Meinhold-Heerlein, I.; Fotopoulou, C.; Harter, P.; Kurzeder, C.; Mustea, A.; Wimberger, P.; Hauptmann, S.; Sehouli, J. The new WHO classification of ovarian, fallopian tube and primary peritoneal cancer and its clinical implications. *Arch. Gynecol. Obstet.* **2016**, *293*, 695–700. [[CrossRef](#)] [[PubMed](#)]
2. Gershenson, D.M.; Sun, C.C.; Lu, K.H.; Coleman, R.L.; Sood, A.K.; Malpica, A.; Deavers, M.T.; Silva, E.G.; Bodurka, D.C. Clinical behavior of stage II-IV low-grade serous carcinoma of the ovary. *Obstet. Gynecol.* **2006**, *108*, 361–368. [[CrossRef](#)] [[PubMed](#)]
3. Ozols, R.F. Maintenance therapy in advanced ovarian cancer: Progression-free survival and clinical benefit. *J. Clin. Oncol.* **2003**, *21*, 2451–2453. [[CrossRef](#)] [[PubMed](#)]
4. Gourley, C.; Farley, J.; Provencher, D.M.; Pignata, S.; Mileskin, L.; Harter, P.; Maenpaa, J.; Kim, J.W.; Pujade-Lauraine, E.; Glasspool, R.M.; et al. Gynecologic Cancer InterGroup (GCIG) consensus review for ovarian and primary peritoneal low-grade serous carcinomas. *Int. J. Gynecol. Cancer* **2014**, *24*, S9–S13. [[CrossRef](#)] [[PubMed](#)]
5. Baak, J.P.; Delemarre, J.F.; Langley, F.A.; Talerman, A. Grading ovarian tumors. Evaluation of decision making by different pathologists. *Anal. Quant. Cytol. Histol.* **1986**, *8*, 349–353. [[PubMed](#)]
6. Stalsberg, H.; Abeler, V.; Blom, G.P.; Bostad, L.; Skarland, E.; Westgaard, G. Observer variation in histologic classification of malignant and borderline ovarian tumors. *Hum. Pathol.* **1988**, *19*, 1030–1035. [[CrossRef](#)]
7. Bertelsen, K.; Holund, B.; Andersen, E. Reproducibility and prognostic value of histologic type and grade in early epithelial ovarian cancer. *Int. J. Gynecol. Cancer* **1993**, *3*, 72–79. [[CrossRef](#)] [[PubMed](#)]
8. Shimizu, Y.; Kamoi, S.; Amada, S.; Akiyama, F.; Silverberg, S.G. Toward the development of a universal grading system for ovarian epithelial carcinoma: Testing of a proposed system in a series of 461 patients with uniform treatment and follow-up. *Cancer* **1998**, *82*, 893–901. [[CrossRef](#)]
9. Ishioka, S.; Sagae, S.; Terasawa, K.; Sugimura, M.; Nishioka, Y.; Tsukada, K.; Kudo, R. Comparison of the usefulness between a new universal grading system for epithelial ovarian cancer and the FIGO grading system. *Gynecol. Oncol.* **2003**, *89*, 447–452. [[CrossRef](#)]
10. Seidman, J.D.; Mehrotra, A. Benign ovarian serous tumors: A re-evaluation and proposed reclassification of serous “cystadenomas” and “cystadenofibromas”. *Gynecol. Oncol.* **2005**, *96*, 395–401. [[CrossRef](#)]
11. Bonome, T.; Lee, J.Y.; Park, D.C.; Radonovich, M.; Pise-Masison, C.; Brady, J.; Gardner, G.J.; Hao, K.; Wong, W.H.; Barrett, J.C.; et al. Expression profiling of serous low malignant potential, low-grade and high-grade tumors of the ovary. *Cancer Res.* **2005**, *65*, 10602–10612. [[CrossRef](#)] [[PubMed](#)]
12. Jazaeri, A.A.; Lu, K.; Schmandt, R.; Harris, C.P.; Rao, P.H.; Sotiriou, C.; Chandramouli, G.V.; Gershenson, D.M.; Liu, E.T. Molecular determinants of tumor differentiation in papillary serous ovarian carcinoma. *Mol. Carcinog.* **2003**, *36*, 53–59. [[CrossRef](#)] [[PubMed](#)]
13. Meinhold-Heerlein, I.; Bauerschlag, D.; Hilpert, F.; Dimitrov, P.; Sapinoso, L.M.; Orłowska-Volk, M.; Bauknecht, T.; Park, T.W.; Jonat, W.; Jacobsen, A.; et al. Molecular and prognostic distinction between serous ovarian carcinomas of varying grade and malignant potential. *Oncogene* **2005**, *24*, 1053–1065. [[CrossRef](#)] [[PubMed](#)]
14. Gershenson, D.M.; Bodurka, D.C.; Lu, K.H.; Nathan, L.C.; Milojevic, L.; Wong, K.K.; Malpica, A.; Sun, C.C. Impact of Age and Primary Disease Site on Outcome in Women With Low-Grade Serous Carcinoma of the Ovary or Peritoneum: Results of a Large Single-Institution Registry of a Rare Tumor. *J. Clin. Oncol.* **2015**, *33*, 2675–2682. [[CrossRef](#)] [[PubMed](#)]
15. Wong, K.K.; Tsang, Y.T.; Deavers, M.T.; Mok, S.C.; Zu, Z.; Sun, C.; Malpica, A.; Wolf, J.K.; Lu, K.H.; Gershenson, D.M. BRAF mutation is rare in advanced-stage low-grade ovarian serous carcinomas. *Am. J. Pathol.* **2010**, *177*, 1611–1617. [[CrossRef](#)] [[PubMed](#)]
16. Grisham, R.N.; Iyer, G.; Garg, K.; Delair, D.; Hyman, D.M.; Zhou, Q.; Iasonos, A.; Berger, M.F.; Dao, F.; Spriggs, D.R.; et al. BRAF mutation is associated with early stage disease and improved outcome in patients with low-grade serous ovarian cancer. *Cancer* **2013**, *119*, 548–554. [[CrossRef](#)]
17. Hennessy, B.T.; Coleman, R.L.; Markman, M. Ovarian cancer. *Lancet* **2009**, *374*, 1371–1382. [[CrossRef](#)]
18. Colombo, N.; Peiretti, M.; Parma, G.; Lapresa, M.; Mancari, R.; Carinelli, S.; Sessa, C.; Castiglione, M.; Group, E.G.W. Newly diagnosed and relapsed epithelial ovarian carcinoma: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann. Oncol.* **2010**, *21*, v23–v30. [[CrossRef](#)]

19. Schmeler, K.M.; Sun, C.C.; Bodurka, D.C.; Deavers, M.T.; Malpica, A.; Coleman, R.L.; Ramirez, P.T.; Gershenson, D.M. Neoadjuvant chemotherapy for low-grade serous carcinoma of the ovary or peritoneum. *Gynecol. Oncol.* **2008**, *108*, 510–514. [[CrossRef](#)]
20. Gershenson, D.M.; Sun, C.C.; Bodurka, D.; Coleman, R.L.; Lu, K.H.; Sood, A.K.; Deavers, M.; Malpica, A.L.; Kavanagh, J.J. Recurrent low-grade serous ovarian carcinoma is relatively chemoresistant. *Gynecol. Oncol.* **2009**, *114*, 48–52. [[CrossRef](#)]
21. Grabowski, J.P.; Harter, P.; Heitz, F.; Pujade-Lauraine, E.; Reuss, A.; Kristensen, G.; Ray-Coquard, I.; Heitz, J.; Traut, A.; Pfisterer, J.; et al. Operability and chemotherapy responsiveness in advanced low-grade serous ovarian cancer. An analysis of the AGO Study Group metadatabase. *Gynecol. Oncol.* **2016**, *140*, 457–462. [[CrossRef](#)] [[PubMed](#)]
22. Jemal, A.; Bray, F.; Center, M.M.; Ferlay, J.; Ward, E.; Forman, D. Global cancer statistics. *CA Cancer J. Clin.* **2011**, *61*, 69–90. [[CrossRef](#)] [[PubMed](#)]
23. Kipps, E.; Tan, D.S.; Kaye, S.B. Meeting the challenge of ascites in ovarian cancer: New avenues for therapy and research. *Nat. Rev. Cancer* **2013**, *13*, 273–282. [[CrossRef](#)] [[PubMed](#)]
24. Kulbe, H.; Chakravarty, P.; Leinster, D.A.; Charles, K.A.; Kwong, J.; Thompson, R.G.; Coward, J.I.; Schioppa, T.; Robinson, S.C.; Gallagher, W.M.; et al. A dynamic inflammatory cytokine network in the human ovarian cancer microenvironment. *Cancer Res.* **2012**, *72*, 66–75. [[CrossRef](#)] [[PubMed](#)]
25. Pesce, S.; Tabellini, G.; Cantoni, C.; Patrizi, O.; Coltrini, D.; Rampinelli, F.; Matta, J.; Vivier, E.; Moretta, A.; Parolini, S.; et al. B7-H6-mediated downregulation of NKG2D in NK cells contributes to ovarian carcinoma immune escape. *Oncoimmunology* **2015**, *4*, e1001224. [[CrossRef](#)]
26. Peng, P.; Yan, Y.; Keng, S. Exosomes in the ascites of ovarian cancer patients: Origin and effects on antitumor immunity. *Oncol. Rep.* **2011**, *25*, 749–762. [[PubMed](#)]
27. Latifi, A.; Luwor, R.B.; Bilandzic, M.; Nazaretian, S.; Stenvers, K.; Pyman, J.; Zhu, H.; Thompson, E.W.; Quinn, M.A.; Findlay, J.K.; et al. Isolation and characterization of tumor cells from the ascites of ovarian cancer patients: Molecular phenotype of chemoresistant ovarian tumors. *PLoS ONE* **2012**, *7*, e46858. [[CrossRef](#)] [[PubMed](#)]
28. Preston, C.C.; Goode, E.L.; Hartmann, L.C.; Kalli, K.R.; Knutson, K.L. Immunity and immune suppression in human ovarian cancer. *Immunotherapy* **2011**, *3*, 539–556. [[CrossRef](#)] [[PubMed](#)]
29. Cancer Genome Atlas Research Network. Integrated genomic analyses of ovarian carcinoma. *Nature* **2011**, *474*, 609–615.
30. Pesce, S.; Greppi, M.; Tabellini, G.; Rampinelli, F.; Parolini, S.; Olive, D.; Moretta, L.; Moretta, A.; Marcenaro, E. Identification of a subset of human natural killer cells expressing high levels of programmed death 1: A phenotypic and functional characterization. *J. Allergy Clin. Immunol.* **2017**, *139*, 335–346. [[CrossRef](#)] [[PubMed](#)]
31. Worzfeld, T.; Pogge von Strandmann, E.; Huber, M.; Adhikary, T.; Wagner, U.; Reinartz, S.; Muller, R. The Unique Molecular and Cellular Microenvironment of Ovarian Cancer. *Front. Oncol.* **2017**, *7*, 24. [[CrossRef](#)] [[PubMed](#)]
32. Carlsten, M.; Norell, H.; Bryceson, Y.T.; Poschke, I.; Schedvins, K.; Ljunggren, H.G.; Kiessling, R.; Malmberg, K.J. Primary human tumor cells expressing CD155 impair tumor targeting by down-regulating DNAM-1 on NK cells. *J. Immunol.* **2009**, *183*, 4921–4930. [[CrossRef](#)] [[PubMed](#)]
33. Vyas, M.; Reinartz, S.; Hoffmann, N.; Reiners, K.S.; Lieber, S.; Jansen, J.M.; Wagner, U.; Muller, R.; von Strandmann, E.P. Soluble NKG2D ligands in the ovarian cancer microenvironment are associated with an adverse clinical outcome and decreased memory effector T cells independent of NKG2D downregulation. *Oncoimmunology* **2017**, *6*, e1339854. [[CrossRef](#)] [[PubMed](#)]
34. Marcenaro, E.; Dondero, A.; Moretta, A. Multi-directional crossregulation of NK cell function during innate immune responses. *Transpl. Immunol.* **2006**, *17*, 16–19. [[CrossRef](#)] [[PubMed](#)]
35. Marcenaro, E.; Ferranti, B.; Moretta, A. NK-DC interaction: On the usefulness of auto-aggression. *Autoimmun. Rev.* **2005**, *4*, 520–525. [[CrossRef](#)] [[PubMed](#)]
36. Vivier, E.; Raulet, D.H.; Moretta, A.; Caligiuri, M.A.; Zitvogel, L.; Lanier, L.L.; Yokoyama, W.M.; Ugolini, S. Innate or adaptive immunity? The example of natural killer cells. *Science* **2011**, *331*, 44–49. [[CrossRef](#)] [[PubMed](#)]
37. Sivori, S.; Carlomagno, S.; Pesce, S.; Moretta, A.; Vitale, M.; Marcenaro, E. TLR/NCR/KIR: Which One to Use and When? *Front. Immunol.* **2014**, *5*, 105. [[CrossRef](#)]

38. Lai, P.; Rabinowich, H.; Crowley-Nowick, P.A.; Bell, M.C.; Mantovani, G.; Whiteside, T.L. Alterations in expression and function of signal-transducing proteins in tumor-associated T and natural killer cells in patients with ovarian carcinoma. *Clin. Cancer Res.* **1996**, *2*, 161–173. [[PubMed](#)]
39. Lukesova, S.; Vroblova, V.; Tosner, J.; Kopecky, J.; Sedlakova, I.; Cermakova, E.; Vokurkova, D.; Kopecky, O. Comparative study of various subpopulations of cytotoxic cells in blood and ascites from patients with ovarian carcinoma. *Contemp. Oncol.* **2015**, *19*, 290–299. [[CrossRef](#)] [[PubMed](#)]
40. Mailliard, R.B.; Alber, S.M.; Shen, H.; Watkins, S.C.; Kirkwood, J.M.; Herberman, R.B.; Kalinski, P. IL-18-induced CD83+CCR7+ NK helper cells. *J. Exp. Med.* **2005**, *202*, 941–953. [[CrossRef](#)] [[PubMed](#)]
41. Otegbeye, F.; Ojo, E.; Moreton, S.; Mackowski, N.; Lee, D.A.; de Lima, M.; Wald, D.N. Inhibiting TGF-beta signaling preserves the function of highly activated, in vitro expanded natural killer cells in AML and colon cancer models. *PLoS ONE* **2018**, *13*, e0191358.
42. Castriconi, R.; Cantoni, C.; Della Chiesa, M.; Vitale, M.; Marcenaro, E.; Conte, R.; Biassoni, R.; Bottino, C.; Moretta, L.; Moretta, A. Transforming growth factor beta 1 inhibits expression of NKp30 and NKG2D receptors: Consequences for the NK-mediated killing of dendritic cells. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 4120–4125. [[CrossRef](#)] [[PubMed](#)]
43. Krockenberger, M.; Dombrowski, Y.; Weidler, C.; Ossadnik, M.; Honig, A.; Hausler, S.; Voigt, H.; Becker, J.C.; Leng, L.; Steinle, A.; et al. Macrophage migration inhibitory factor contributes to the immune escape of ovarian cancer by down-regulating NKG2D. *J. Immunol.* **2008**, *180*, 7338–7348. [[CrossRef](#)] [[PubMed](#)]
44. Zhou, Y.; Xu, Y.; Chen, L.; Xu, B.; Wu, C.; Jiang, J. B7-H6 expression correlates with cancer progression and patient's survival in human ovarian cancer. *Int. J. Clin. Exp. Pathol.* **2015**, *8*, 9428–9433. [[PubMed](#)]
45. Zhang, M.; Yang, J.; Hua, W.; Li, Z.; Xu, Z.; Qian, Q. Monitoring checkpoint inhibitors: Predictive biomarkers in immunotherapy. *Front. Med.* **2019**, *12*, 1–13. [[CrossRef](#)] [[PubMed](#)]
46. Hung, C.F.; Wu, T.C.; Monie, A.; Roden, R. Antigen-specific immunotherapy of cervical and ovarian cancer. *Immunol. Rev.* **2008**, *222*, 43–69. [[CrossRef](#)] [[PubMed](#)]
47. Sundar, S.; Neal, R.D.; Kehoe, S. Diagnosis of ovarian cancer. *BMJ* **2015**, *351*, h4443. [[CrossRef](#)] [[PubMed](#)]
48. Leitao, M.M., Jr.; Chi, D.S. Surgical management of recurrent ovarian cancer. *Semin. Oncol.* **2009**, *36*, 106–111. [[CrossRef](#)] [[PubMed](#)]
49. Ushijima, K. Treatment for recurrent ovarian cancer-at first relapse. *J. Oncol.* **2010**, *2010*, 497429. [[CrossRef](#)] [[PubMed](#)]
50. Uppendahl, L.D.; Dahl, C.M.; Miller, J.S.; Felices, M.; Geller, M.A. Natural Killer Cell-Based Immunotherapy in Gynecologic Malignancy: A Review. *Front. Immunol.* **2017**, *8*, 1825. [[CrossRef](#)] [[PubMed](#)]
51. Freedman, R.S. Recent immunologic advances affecting the management of ovarian cancer. *Clin. Obstet. Gynecol.* **1985**, *28*, 853–871. [[CrossRef](#)]
52. Freedman, R.S.; Edwards, C.L.; Bowen, J.M.; Lotzova, E.; Katz, R.; Lewis, E.; Atkinson, N.; Carsetti, R. Viral oncolysates in patients with advanced ovarian cancer. *Gynecol. Oncol.* **1988**, *29*, 337–347. [[CrossRef](#)]
53. Berek, J.S.; Bast, R.C., Jr.; Lichtenstein, A.; Hacker, N.F.; Spina, C.A.; Lagasse, L.D.; Knapp, R.C.; Zigelboim, J. Lymphocyte cytotoxicity in the peritoneal cavity and blood of patients with ovarian cancer. *Obstet. Gynecol.* **1984**, *64*, 708–714.
54. Recchia, F.; Saggio, G.; Cesta, A.; Candeloro, G.; Nuzzo, A.; Lombardo, M.; Carta, G.; Rea, S. Interleukin-2 and 13-cis retinoic acid as maintenance therapy in advanced ovarian cancer. *Int. J. Oncol.* **2005**, *27*, 1039–1046. [[CrossRef](#)] [[PubMed](#)]
55. Mantia-Smaldone, G.M.; Corr, B.; Chu, C.S. Immunotherapy in ovarian cancer. *Hum. Vaccin Immunother.* **2012**, *8*, 1179–1191. [[CrossRef](#)] [[PubMed](#)]
56. Pandey, V.; Oyer, J.L.; Igarashi, R.Y.; Gitto, S.B.; Copik, A.J.; Altomare, D.A. Antiovarian tumor response of donor peripheral blood mononuclear cells is due to infiltrating cytotoxic NK cells. *Oncotarget* **2016**, *7*, 7318–7328. [[CrossRef](#)] [[PubMed](#)]
57. da Silva, R.F.; Yoshida, A.; Cardozo, D.M.; Jales, R.M.; Paust, S.; Derchain, S.; Guimaraes, F. Natural Killer Cells Response to IL-2 Stimulation Is Distinct between Ascites with the Presence or Absence of Malignant Cells in Ovarian Cancer Patients. *Int. J. Mol. Sci.* **2017**, *18*, 856. [[CrossRef](#)]
58. Pillet, A.H.; Bugault, F.; Theze, J.; Chakrabarti, L.A.; Rose, T. A programmed switch from IL-15- to IL-2-dependent activation in human NK cells. *J. Immunol.* **2009**, *182*, 6267–6277. [[CrossRef](#)] [[PubMed](#)]
59. Leclercq, G.; Debacker, V.; de Smedt, M.; Plum, J. Differential effects of interleukin-15 and interleukin-2 on differentiation of bipotential T/natural killer progenitor cells. *J. Exp. Med.* **1996**, *184*, 325–336. [[CrossRef](#)]

60. Childs, R.W.; Carlsten, M. Therapeutic approaches to enhance natural killer cell cytotoxicity against cancer: The force awakens. *Nat. Rev. Drug Discov.* **2015**, *14*, 487–498. [[CrossRef](#)]
61. Felices, M.; Chu, S.; Kodala, B.; Bendzick, L.; Ryan, C.; Lenvik, A.J.; Boylan, K.L.M.; Wong, H.C.; Skubitz, A.P.N.; Miller, J.S.; et al. IL-15 super-agonist (ALT-803) enhances natural killer (NK) cell function against ovarian cancer. *Gynecol. Oncol.* **2017**, *145*, 453–461. [[CrossRef](#)] [[PubMed](#)]
62. Hoogstad-van Evert, J.S.; Maas, R.J.; van der Meer, J.; Cany, J.; van der Steen, S.; Jansen, J.H.; Miller, J.S.; Bekkers, R.; Hobo, W.; et al. Peritoneal NK cells are responsive to IL-15 and percentages are correlated with outcome in advanced ovarian cancer patients. *Oncotarget* **2018**, *9*, 34810–34820. [[PubMed](#)]
63. Kamada, M.; Sakamoto, Y.; Furumoto, H.; Mori, K.; Daitoh, T.; Irahara, M.; Aono, T.; Nii, A.; Yanagawa, H.; Sone, S.; et al. Treatment of malignant ascites with allogeneic and autologous lymphokine-activated killer cells. *Gynecol. Oncol.* **1989**, *34*, 34–37. [[CrossRef](#)]
64. Mittica, G.; Capellero, S.; Genta, S.; Cagnazzo, C.; Aglietta, M.; Sangiolo, D.; Valabrega, G. Adoptive immunotherapy against ovarian cancer. *J. Ovarian Res.* **2016**, *9*, 30. [[CrossRef](#)] [[PubMed](#)]
65. Urba, W.J.; Clark, J.W.; Steis, R.G.; Bookman, M.A.; Smith, J.W., 2nd; Beckner, S.; Maluish, A.E.; Rossio, J.L.; Rager, H.; Ortaldo, J.R.; et al. Intraperitoneal lymphokine-activated killer cell/interleukin-2 therapy in patients with intra-abdominal cancer: Immunologic considerations. *J. Natl. Cancer Inst.* **1989**, *81*, 602–611. [[CrossRef](#)] [[PubMed](#)]
66. Steis, R.G.; Urba, W.J.; VanderMolen, L.A.; Bookman, M.A.; Smith, J.W., 2nd; Clark, J.W.; Miller, R.L.; Crum, E.D.; Beckner, S.K.; McKnight, J.E.; et al. Intraperitoneal lymphokine-activated killer-cell and interleukin-2 therapy for malignancies limited to the peritoneal cavity. *J. Clin. Oncol.* **1990**, *8*, 1618–1629. [[CrossRef](#)] [[PubMed](#)]
67. Stewart, J.A.; Belinson, J.L.; Moore, A.L.; Dorigi, J.A.; Grant, B.W.; Haugh, L.D.; Roberts, J.D.; Albertini, R.J.; Branda, R.F. Phase I trial of intraperitoneal recombinant interleukin-2/lymphokine-activated killer cells in patients with ovarian cancer. *Cancer Res.* **1990**, *50*, 6302–6310. [[PubMed](#)]
68. Schmidt-Wolf, I.G.; Negrin, R.S.; Kiem, H.P.; Blume, K.G.; Weissman, I.L. Use of a SCID mouse/human lymphoma model to evaluate cytokine-induced killer cells with potent antitumor cell activity. *J. Exp. Med.* **1991**, *174*, 139–149. [[CrossRef](#)] [[PubMed](#)]
69. Kim, H.M.; Kang, J.S.; Lim, J.; Park, S.K.; Lee, K.; Yoon, Y.D.; Lee, C.W.; Lee, K.H.; Han, G.; Yang, K.H.; et al. Inhibition of human ovarian tumor growth by cytokine-induced killer cells. *Arch. Pharm. Res.* **2007**, *30*, 1464–1470. [[CrossRef](#)] [[PubMed](#)]
70. Liu, J.; Li, H.; Cao, S.; Zhang, X.; Yu, J.; Qi, J.; An, X.; Yu, W.; Ren, X.; Hao, X. Maintenance therapy with autologous cytokine-induced killer cells in patients with advanced epithelial ovarian cancer after first-line treatment. *J. Immunother.* **2014**, *37*, 115–122. [[CrossRef](#)] [[PubMed](#)]
71. Uppendahl, L.D.; Felices, M.; Bendzick, L.; Ryan, C.; Kodala, B.; Hinderlie, P.; Boylan, K.L.M.; Skubitz, A.P.N.; Miller, J.S.; Geller, M.A. Cytokine-induced memory-like natural killer cells have enhanced function, proliferation and in vivo expansion against ovarian cancer cells. *Gynecol. Oncol.* **2019**. [[CrossRef](#)] [[PubMed](#)]
72. Cichocki, F.; Valamehr, B.; Bjordahl, R.; Zhang, B.; Rezner, B.; Rogers, P.; Gaidarova, S.; Moreno, S.; Tuininga, K.; Dougherty, P.; et al. GSK3 Inhibition Drives Maturation of NK Cells and Enhances Their Antitumor Activity. *Cancer Res.* **2017**, *77*, 5664–5675. [[CrossRef](#)] [[PubMed](#)]
73. Ruggeri, L.; Capanni, M.; Urbani, E.; Perruccio, K.; Shlomchik, W.D.; Tosti, A.; Posati, S.; Rogaia, D.; Frassoni, F.; Aversa, F.; et al. Effectiveness of donor natural killer cell alloreactivity in mismatched hematopoietic transplants. *Science* **2002**, *295*, 2097–2100. [[CrossRef](#)] [[PubMed](#)]
74. Kaufman, D.S. Toward clinical therapies using hematopoietic cells derived from human pluripotent stem cells. *Blood* **2009**, *114*, 3513–3523. [[CrossRef](#)] [[PubMed](#)]
75. Woll, P.S.; Grzywacz, B.; Tian, X.; Marcus, R.K.; Knorr, D.A.; Verneris, M.R.; Kaufman, D.S. Human embryonic stem cells differentiate into a homogeneous population of natural killer cells with potent in vivo antitumor activity. *Blood* **2009**, *113*, 6094–6101. [[CrossRef](#)] [[PubMed](#)]
76. Matsuo, K.; Sheridan, T.B.; Mabuchi, S.; Yoshino, K.; Hasegawa, K.; Studeman, K.D.; Im, D.D.; Rosenshein, N.B.; Roman, L.D.; Sood, A.K. Estrogen receptor expression and increased risk of lymphovascular space invasion in high-grade serous ovarian carcinoma. *Gynecol. Oncol.* **2014**, *133*, 473–479. [[CrossRef](#)] [[PubMed](#)]

77. Takahashi, A.; Kato, K.; Kuboyama, A.; Inoue, T.; Tanaka, Y.; Kuhara, A.; Kinoshita, K.; Takeda, S.; Wake, N. Induction of senescence by progesterone receptor-B activation in response to cAMP in ovarian cancer cells. *Gynecol. Oncol.* **2009**, *113*, 270–276. [[CrossRef](#)] [[PubMed](#)]
78. Sieh, W.; Kobel, M.; Longacre, T.A.; Bowtell, D.D.; deFazio, A.; Goodman, M.T.; Hogdall, E.; Deen, S.; Wentzensen, N.; Moysich, K.B.; et al. Hormone-receptor expression and ovarian cancer survival: An Ovarian Tumor Tissue Analysis consortium study. *Lancet. Oncol.* **2013**, *14*, 853–862. [[CrossRef](#)]
79. Paleari, L.; Gandini, S.; Provinciali, N.; Puntoni, M.; Colombo, N.; DeCensi, A. Clinical benefit and risk of death with endocrine therapy in ovarian cancer: A comprehensive review and meta-analysis. *Gynecol. Oncol.* **2017**, *146*, 504–513. [[CrossRef](#)] [[PubMed](#)]
80. Gershenson, D.M.; Bodurka, D.C.; Coleman, R.L.; Lu, K.H.; Malpica, A.; Sun, C.C. Hormonal Maintenance Therapy for Women With Low-Grade Serous Cancer of the Ovary or Peritoneum. *J. Clin. Oncol.* **2017**, *35*, 1103–1111. [[CrossRef](#)]
81. Fader, A.N.; Bergstrom, J.; Jernigan, A.; Tanner, E.J., 3rd; Roche, K.L.; Stone, R.L.; Levinson, K.L.; Ricci, S.; Wethington, S.; Wang, T.L.; et al. Primary cytoreductive surgery and adjuvant hormonal monotherapy in women with advanced low-grade serous ovarian carcinoma: Reducing overtreatment without compromising survival? *Gynecol. Oncol.* **2017**, *147*, 85–91. [[CrossRef](#)] [[PubMed](#)]
82. Curran, E.M.; Berghaus, L.J.; Vernetti, N.J.; Saporita, A.J.; Lubahn, D.B.; Estes, D.M. Natural killer cells express estrogen receptor-alpha and estrogen receptor-beta and can respond to estrogen via a non-estrogen receptor-alpha-mediated pathway. *Cell Immunol.* **2001**, *214*, 12–20. [[CrossRef](#)] [[PubMed](#)]
83. Baral, E.; Nagy, E.; Berczi, I. Modulation of natural killer cell-mediated cytotoxicity by tamoxifen and estradiol. *Cancer* **1995**, *75*, 591–599. [[CrossRef](#)]
84. Gauchez, A.S.; Riandel, J.; Fernandes-Carlos, T.; Jacrot, M.; Guiraud, P.; Coudray, C.; Calop, J.; Favier, A. Effect of oestrone on the natural killer (NK) cell activity, antioxidant status and tumour growth in athymic mice xenografted with human tumours. *Anticancer Res.* **1996**, *16*, 853–859. [[PubMed](#)]
85. Fernandes-Carlos, T.; Riandel, J.; Glise, D.; Guiraud, P.; Favier, A. Modulation of natural killer cell functional activity in athymic mice by beta-carotene, oestrone and their association. *Anticancer Res.* **1997**, *17*, 2523–2527. [[PubMed](#)]
86. Gleason, M.K.; Verneris, M.R.; Todhunter, D.A.; Zhang, B.; McCullar, V.; Zhou, S.X.; Panoskaltis-Mortari, A.; Weiner, L.M.; Vallera, D.A.; Miller, J.S. Bispecific and trispecific killer cell engagers directly activate human NK cells through CD16 signaling and induce cytotoxicity and cytokine production. *Mol. Cancer Ther.* **2012**, *11*, 2674–2684. [[CrossRef](#)] [[PubMed](#)]
87. Vallera, D.A.; Zhang, B.; Gleason, M.K.; Oh, S.; Weiner, L.M.; Kaufman, D.S.; McCullar, V.; Miller, J.S.; Verneris, M.R. Heterodimeric bispecific single-chain variable-fragment antibodies against EpCAM and CD16 induce effective antibody-dependent cellular cytotoxicity against human carcinoma cells. *Cancer Biother. Radiopharm.* **2013**, *28*, 274–282. [[CrossRef](#)] [[PubMed](#)]
88. Schmohl, J.U.; Felices, M.; Todhunter, D.; Taras, E.; Miller, J.S.; Vallera, D.A. Tetraspecific scFv construct provides NK cell mediated ADCC and self-sustaining stimuli via insertion of IL-15 as a crosslinker. *Oncotarget* **2016**, *7*, 73830–73844. [[CrossRef](#)] [[PubMed](#)]
89. Davis, Z.B.; Vallera, D.A.; Miller, J.S.; Felices, M. Natural killer cells unleashed: Checkpoint receptor blockade and BiKE/TriKE utilization in NK-mediated antitumor immunotherapy. *Semin. Immunol.* **2017**, *31*, 64–75. [[CrossRef](#)] [[PubMed](#)]
90. Burger, R.A.; Brady, M.F.; Bookman, M.A.; Fleming, G.F.; Monk, B.J.; Huang, H.; Mannel, R.S.; Homesley, H.D.; Fowler, J.; Greer, B.E.; et al. Incorporation of bevacizumab in the primary treatment of ovarian cancer. *N. Engl. J. Med.* **2011**, *365*, 2473–2483. [[CrossRef](#)]
91. Oza, A.M.; Cook, A.D.; Pfisterer, J.; Embleton, A.; Ledermann, J.A.; Pujade-Lauraine, E.; Kristensen, G.; Carey, M.S.; Beale, P.; Cervantes, A.; et al. Standard chemotherapy with or without bevacizumab for women with newly diagnosed ovarian cancer (ICON7): Overall survival results of a phase 3 randomised trial. *Lancet. Oncol.* **2015**, *16*, 928–936. [[CrossRef](#)]
92. De Angelis, R.; Sant, M.; Coleman, M.P.; Francisci, S.; Baili, P.; Pierannunzio, D.; Trama, A.; Visser, O.; Brenner, H.; Ardanaz, E.; et al. Cancer survival in Europe 1999–2007 by country and age: Results of EUROcare-5—a population-based study. *Lancet. Oncol.* **2014**, *15*, 23–34. [[CrossRef](#)]
93. Oliver, K.E.; McGuire, W.P. Ovarian cancer and antiangiogenic therapy: Caveat emptor. *J. Clin. Oncol.* **2014**, *32*, 3353–3356. [[CrossRef](#)] [[PubMed](#)]

94. Klapdor, R.; Wang, S.; Hacker, U.; Buning, H.; Morgan, M.; Dork, T.; Hillemanns, P.; Schambach, A. Improved Killing of Ovarian Cancer Stem Cells by Combining a Novel Chimeric Antigen Receptor-Based Immunotherapy and Chemotherapy. *Hum. Gene. Ther.* **2017**, *28*, 886–896. [[CrossRef](#)] [[PubMed](#)]
95. Hermanson, D.L.; Kaufman, D.S. Utilizing chimeric antigen receptors to direct natural killer cell activity. *Front. Immunol.* **2015**, *6*, 195. [[CrossRef](#)] [[PubMed](#)]
96. Suh, D.H.; Kim, M.; Lee, K.H.; Eom, K.Y.; Kjeldsen, M.K.; Mirza, M.R.; Kim, J.W. Major clinical research advances in gynecologic cancer in 2017. *J. Gynecol. Oncol.* **2018**, *29*, e31. [[CrossRef](#)]
97. Hamanishi, J.; Mandai, M.; Matsumura, N.; Abiko, K.; Baba, T.; Konishi, I. PD-1/PD-L1 blockade in cancer treatment: Perspectives and issues. *Int. J. Clin. Oncol.* **2016**, *21*, 462–473. [[CrossRef](#)]
98. Schumacher, T.N.; Schreiber, R.D. Neoantigens in cancer immunotherapy. *Science* **2015**, *348*, 69–74. [[CrossRef](#)]
99. Le, D.T.; Uram, J.N.; Wang, H.; Bartlett, B.R.; Kemberling, H.; Eyring, A.D.; Skora, A.D.; Luber, B.S.; Azad, N.S.; Laheru, D.; et al. PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. *N. Engl. J. Med.* **2015**, *372*, 2509–2520. [[CrossRef](#)]
100. Le, D.T.; Durham, J.N.; Smith, K.N.; Wang, H.; Bartlett, B.R.; Aulakh, L.K.; Lu, S.; Kemberling, H.; Wilt, C.; Luber, B.S.; et al. Mismatch repair deficiency predicts response of solid tumors to PD-1 blockade. *Science* **2017**, *357*, 409–413. [[CrossRef](#)]
101. Matsuo, K.; Spragg, S.E.; Ciccone, M.A.; Blake, E.A.; Ricker, C.; Pham, H.Q.; Roman, L.D. Nivolumab use for BRCA gene mutation carriers with recurrent epithelial ovarian cancer: A case series. *Gynecol. Oncol. Rep.* **2018**, *25*, 98–101. [[CrossRef](#)] [[PubMed](#)]
102. Braly, P.; Nicodemus, C.F.; Chu, C.; Collins, Y.; Edwards, R.; Gordon, A.; McGuire, W.; Schoonmaker, C.; Whiteside, T.; Smith, L.M.; et al. The Immune adjuvant properties of front-line carboplatin-paclitaxel: A randomized phase 2 study of alternative schedules of intravenous oregovomab chemoimmunotherapy in advanced ovarian cancer. *J. Immunother.* **2009**, *32*, 54–65. [[CrossRef](#)] [[PubMed](#)]
103. Zhang, F.R.; Liu, H.; Irwanto, A.; Fu, X.A.; Li, Y.; Yu, G.Q.; Yu, Y.X.; Chen, M.F.; Low, H.Q.; Li, J.H.; et al. HLA-B\*13:01 and the dapsone hypersensitivity syndrome. *N. Engl. J. Med.* **2013**, *369*, 1620–1628. [[PubMed](#)]
104. Andre, P.; Denis, C.; Soulas, C.; Bourbon-Caillet, C.; Lopez, J.; Arnoux, T.; Blery, M.; Bonnafous, C.; Gauthier, L.; Morel, A.; et al. AntiNKG2A mAb Is a Checkpoint Inhibitor that Promotes Antitumor Immunity by Unleashing Both T and NK Cells. *Cell* **2018**, *175*, 1731–1743. [[CrossRef](#)] [[PubMed](#)]



© 2019 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).