

Supporting Information for:

Constructing 3-Dimensional Atomic-Resolution Models of Nonsulfated Glycosaminoglycans with Arbitrary Lengths Using Conformations from Molecular Dynamics

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Table S1. Most Probable End-to-End Distances (d) in MD-Generated 20-mer Conformations with Glycosidic Linkage Conformations in Secondary Basins ¹

	Hyaluronan		Non-Sulfated Keratan	
	GlcA β 1-3GlcNAc $-\phi, +\psi$ d (Å)	GlcNAc β 1-4GlcA $-\phi, -\psi$ d (Å)	Gal β 1-4GlcNAc $-\phi, -\psi$ d (Å)	GlcNAc β 1-3Gal $-\phi, +\psi$ d (Å)
Run 1	81.5	74.0	88.5	79.5
Run 2	73.0	73.5	85.0	84.0
Run 3	61.0	74.0	84.5	88.5
Run 4	77.0	76.5	84.5	73.5
All ²	66.0	74.0	84.5	84.0

¹Probabilities were calculated for end-to-end distances sorted into 0.5 Å bins. ²All = end-to-end distance distribution aggregated across all four runs.

Table S2. Percent of Occurrences of Different IdoA Ring Puckers in
Non-Sulfated Dermatan MD Simulations

	20-mer Ensemble	10-mer Ensemble
¹ C ₄	68.727%	70.100%
⁴ C ₁	7.062%	9.025%
B _{1,4}	0.018%	0.005%
⁵ S ₁	0.740%	0.685%
^{2,5} B	3.213%	2.610%
² S _O	19.702%	17.065%
B _{3,O}	0.080%	0.070%
¹ S ₃	0.125%	0.125%
^{1,4} B	0.178%	0.115%
¹ S ₅	0.155%	0.200%

Table S3. Percent of Occurrences of Different IdoA Ring Puckers in
Non-Sulfated Heparan MD Simulations

	20-mer Ensemble	10-mer Ensemble
¹ C ₄	54.222%	64.645%
⁴ C ₁	4.410%	6.730%
³ S ₁	0.003%	0.000%
B _{1,4}	0.085%	0.035%
⁵ S ₁	4.120%	3.110%
^{2,5} B	12.510%	8.550%
² S _O	24.505%	16.520%
B _{3,O}	0.008%	0.000%
¹ S ₃	0.035%	0.085%
^{1,4} B	0.047%	0.155%
¹ S ₅	0.055%	0.170%

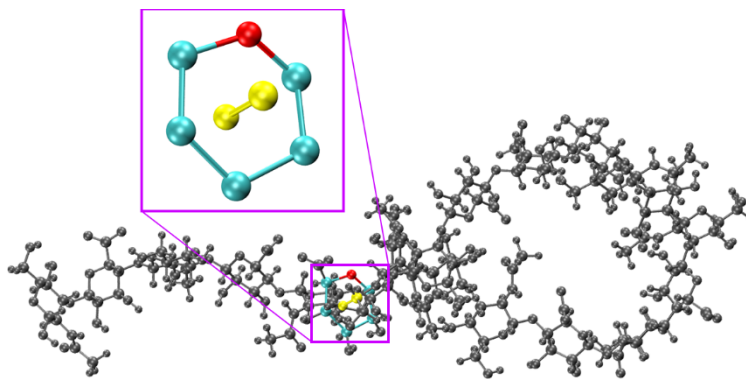


Figure S1. Excluded constructed hyaluronan 20-mer conformation with GlcA 20 ring pierced by C₁-O₃ bond in GlcA 6 β 1-3 GlcNAc 5 linkage post-minimization ($E_b = 690.5$ kcal/mol; closeup shows atoms involved in the ring pierce); $E_{b, \text{cutoff}} = 128.5$ kcal/mol.

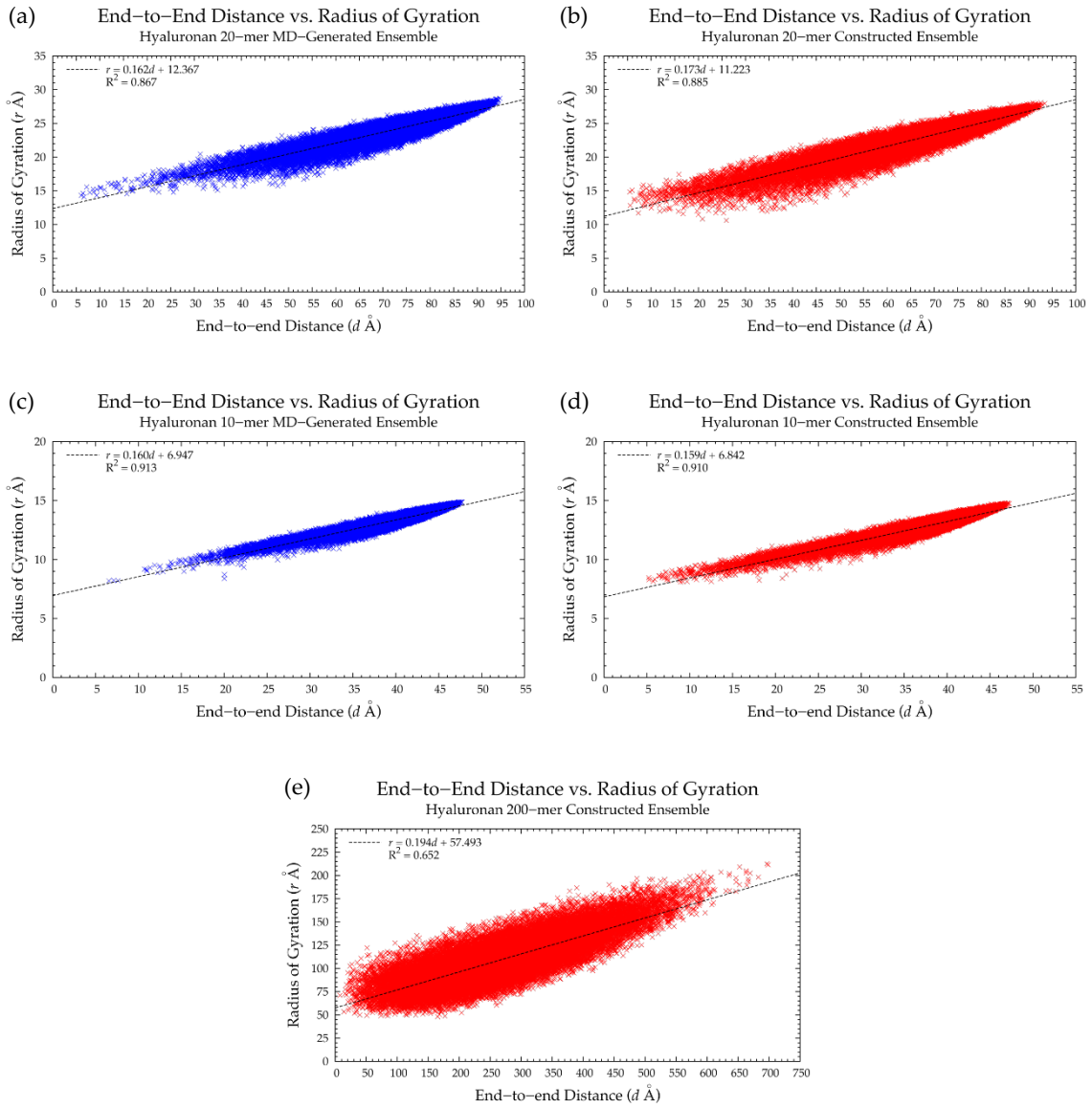


Figure S2. Scatterplots of radius of gyration as a function of end-to-end distance in MD-generated and constructed ensembles of hyaluronan (a,b) 20-mer and (c,d) 10-mer, respectively, and (e) constructed ensemble of hyaluronan 200-mer; each plot has 40,000 samples and shows linear regression and R^2 .

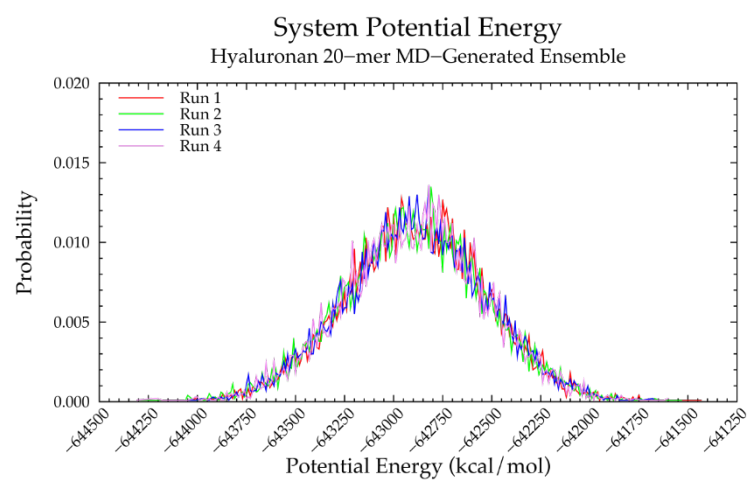
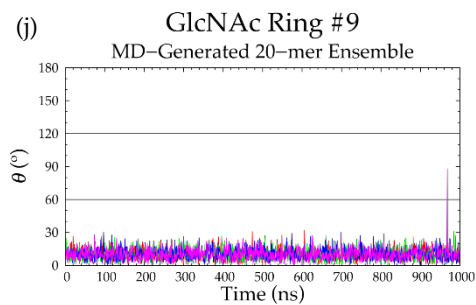
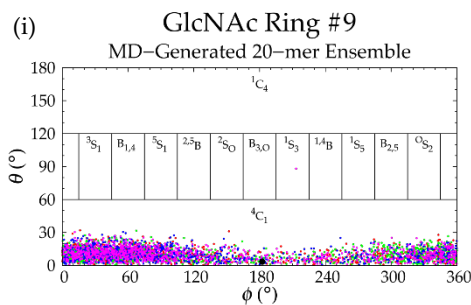
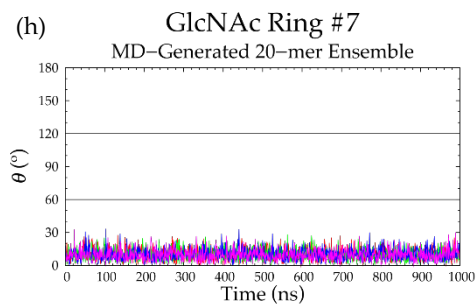
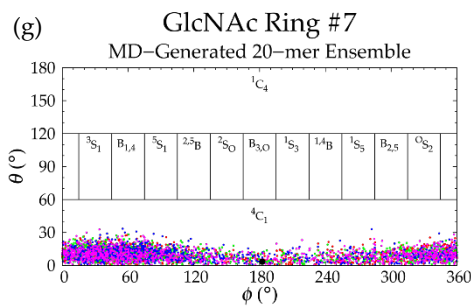
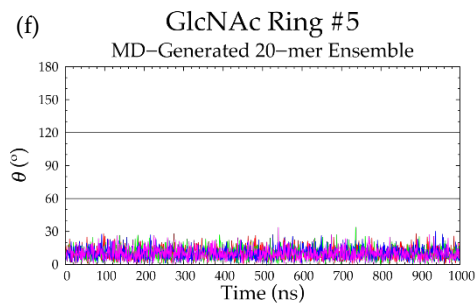
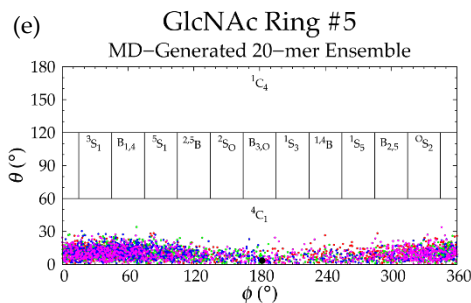
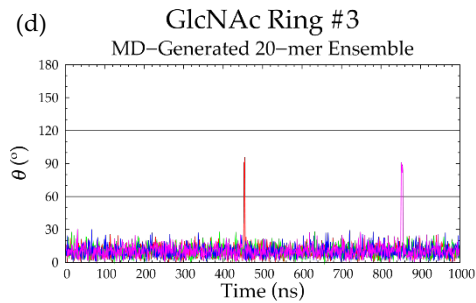
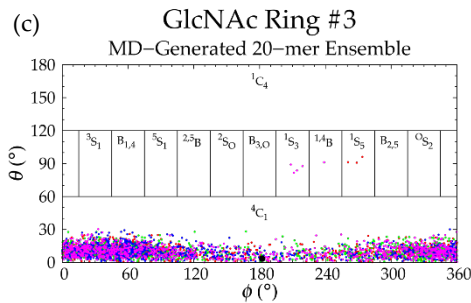
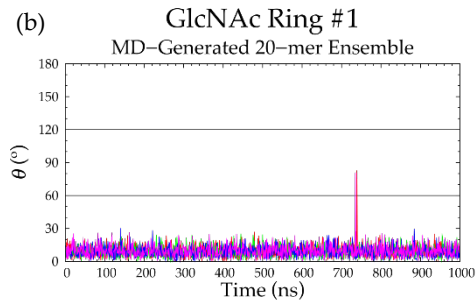
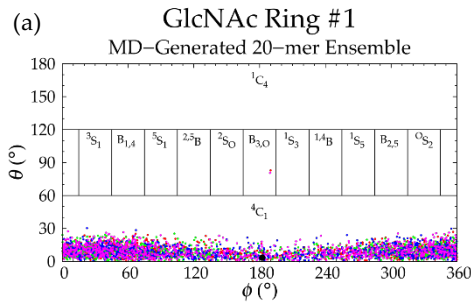


Figure S3. System potential energy probability distribution of the MD-generated hyaluronan 20-mer ensemble; each MD run is represented by a different color.



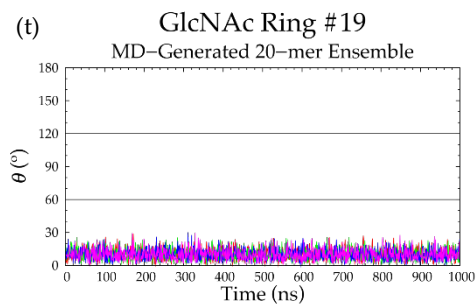
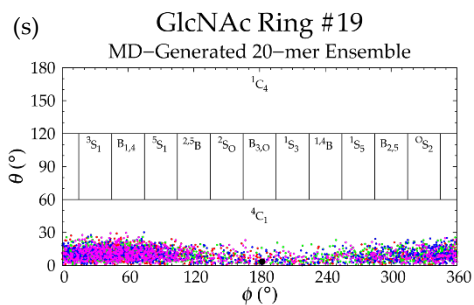
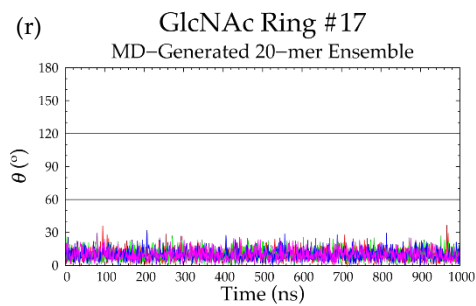
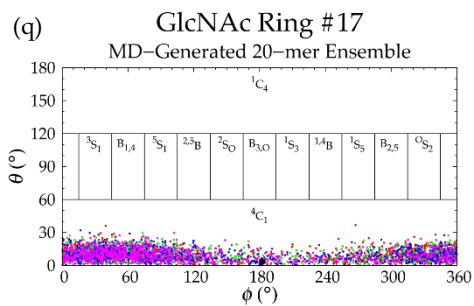
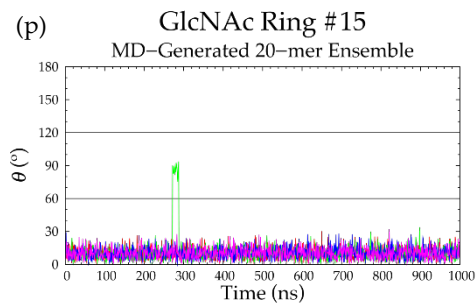
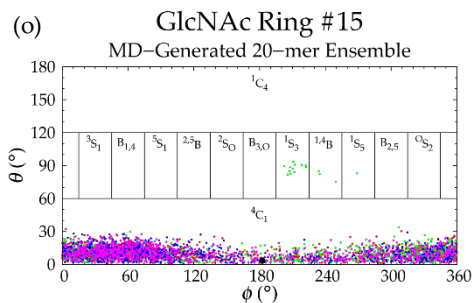
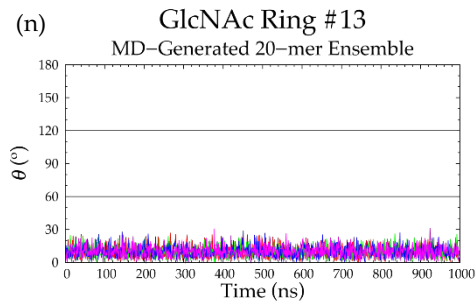
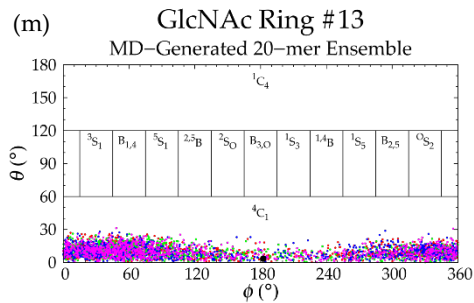
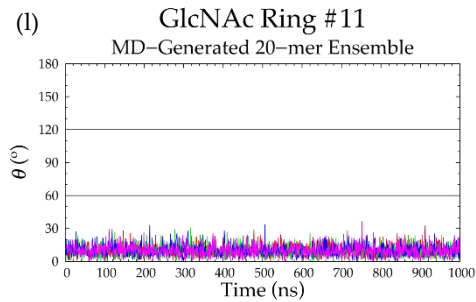
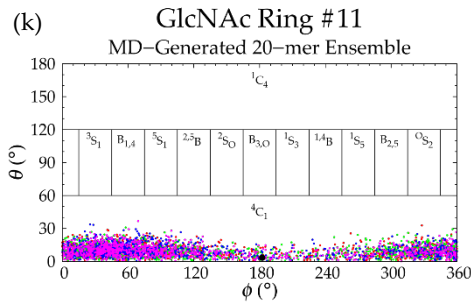
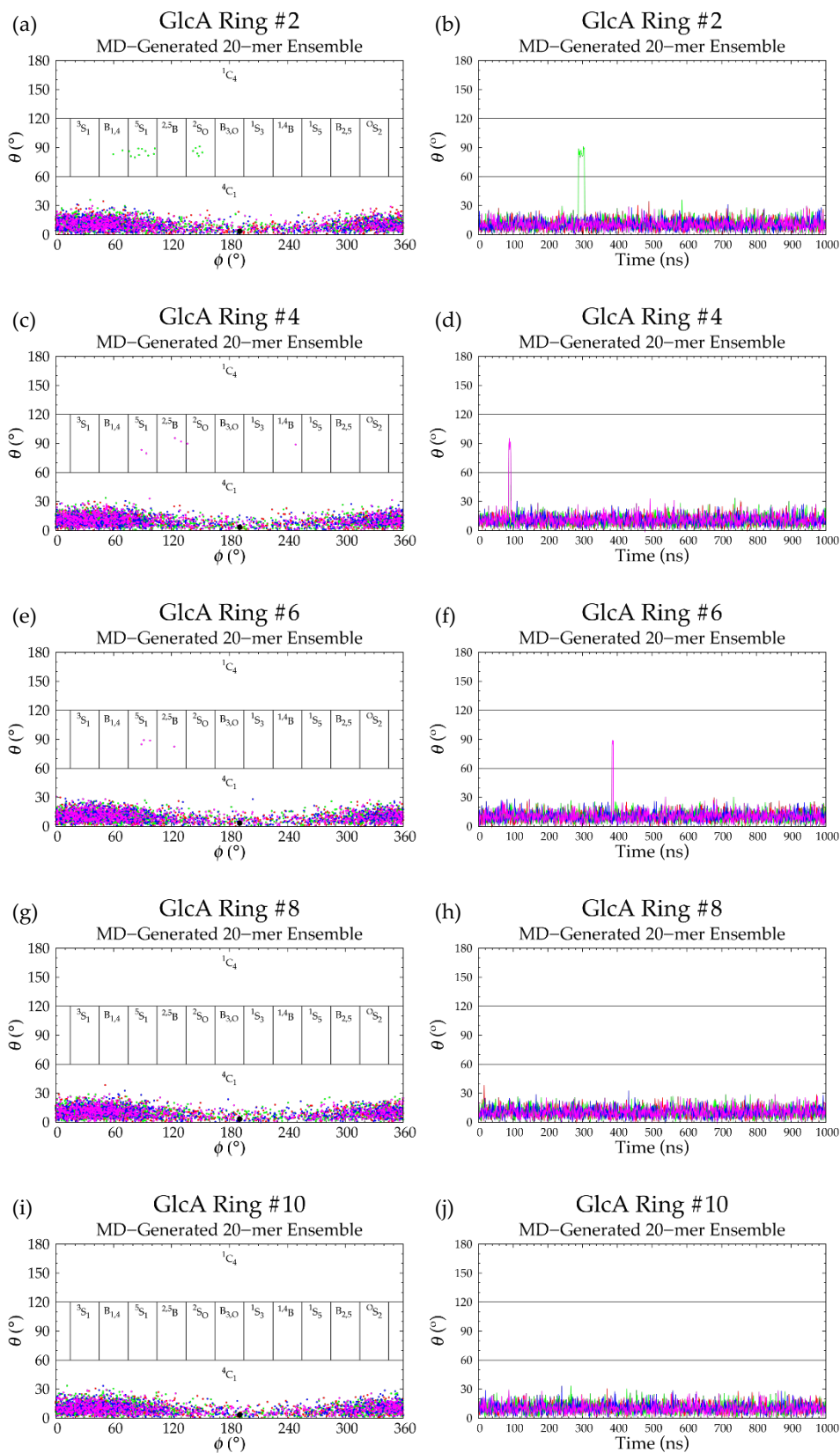


Figure S4. (a,c,e,g,i,k,m,o,q,s) Cremer-Pople plots and (b,d,f,h,j,l,n,p,r,t) Cremer-Pople parameter θ timeseries for each GlcNAc monosaccharide ring in the MD-generated hyaluronan 20-mer ensemble; monosaccharides are numbered from reducing to non-reducing end; each of the 4 runs is represented by different color.



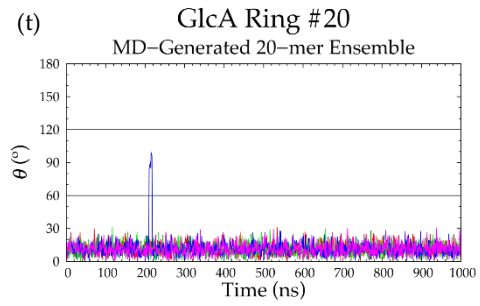
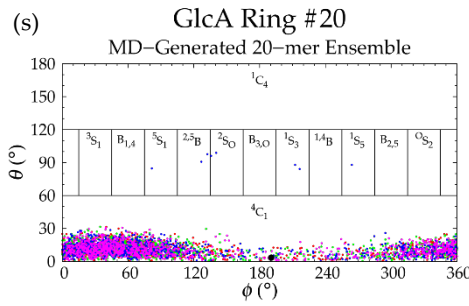
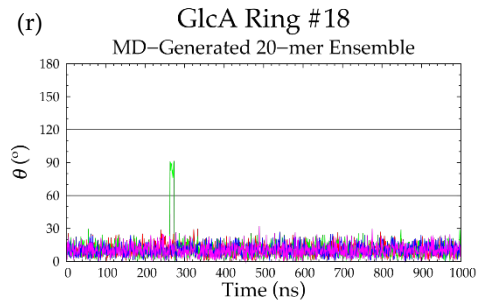
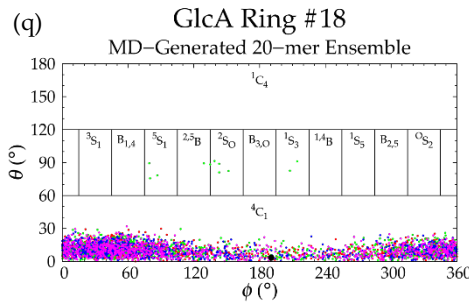
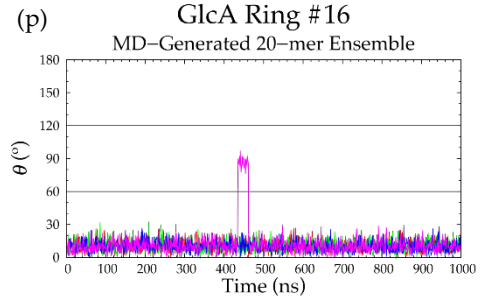
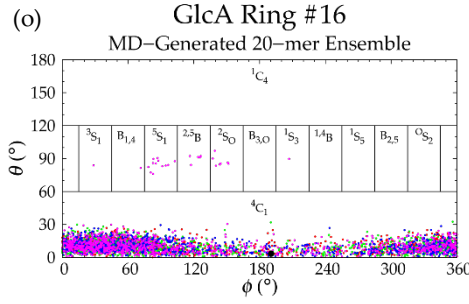
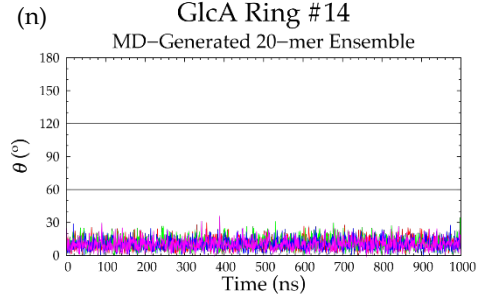
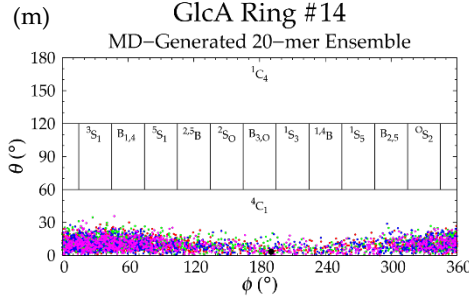
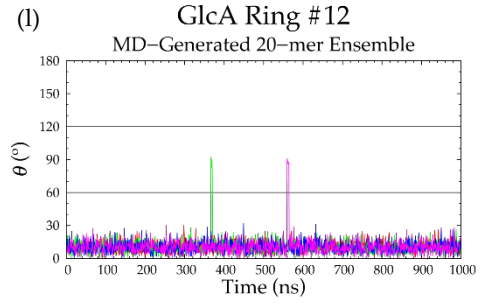
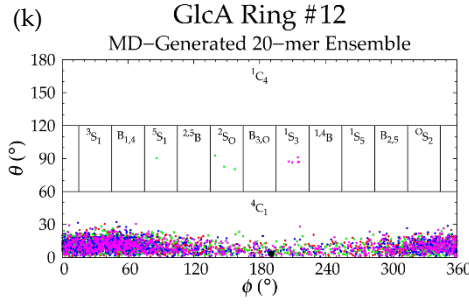


Figure S5. (a,c,e,g,i,k,m,o,q,s) Cremer-Pople plots and (b,d,f,h,j,l,n,p,r,t) Cremer-Pople parameter θ timeseries for each GlcA monosaccharide ring in the MD-generated hyaluronan 20-mer ensemble; monosaccharides are numbered from reducing to non-reducing end; each of the 4 runs is represented by different color.

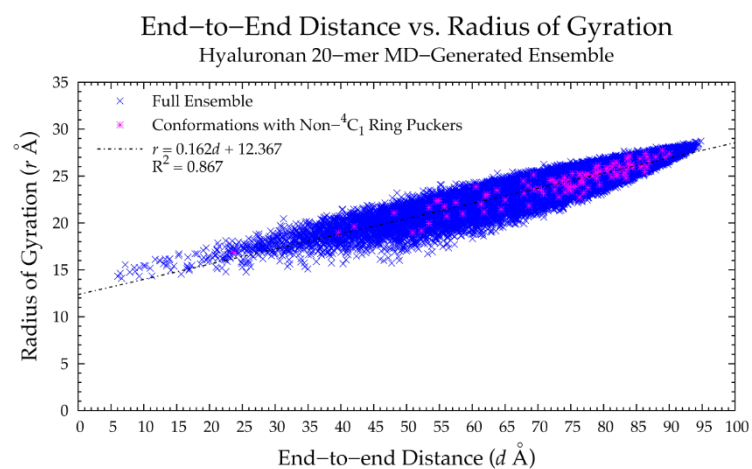
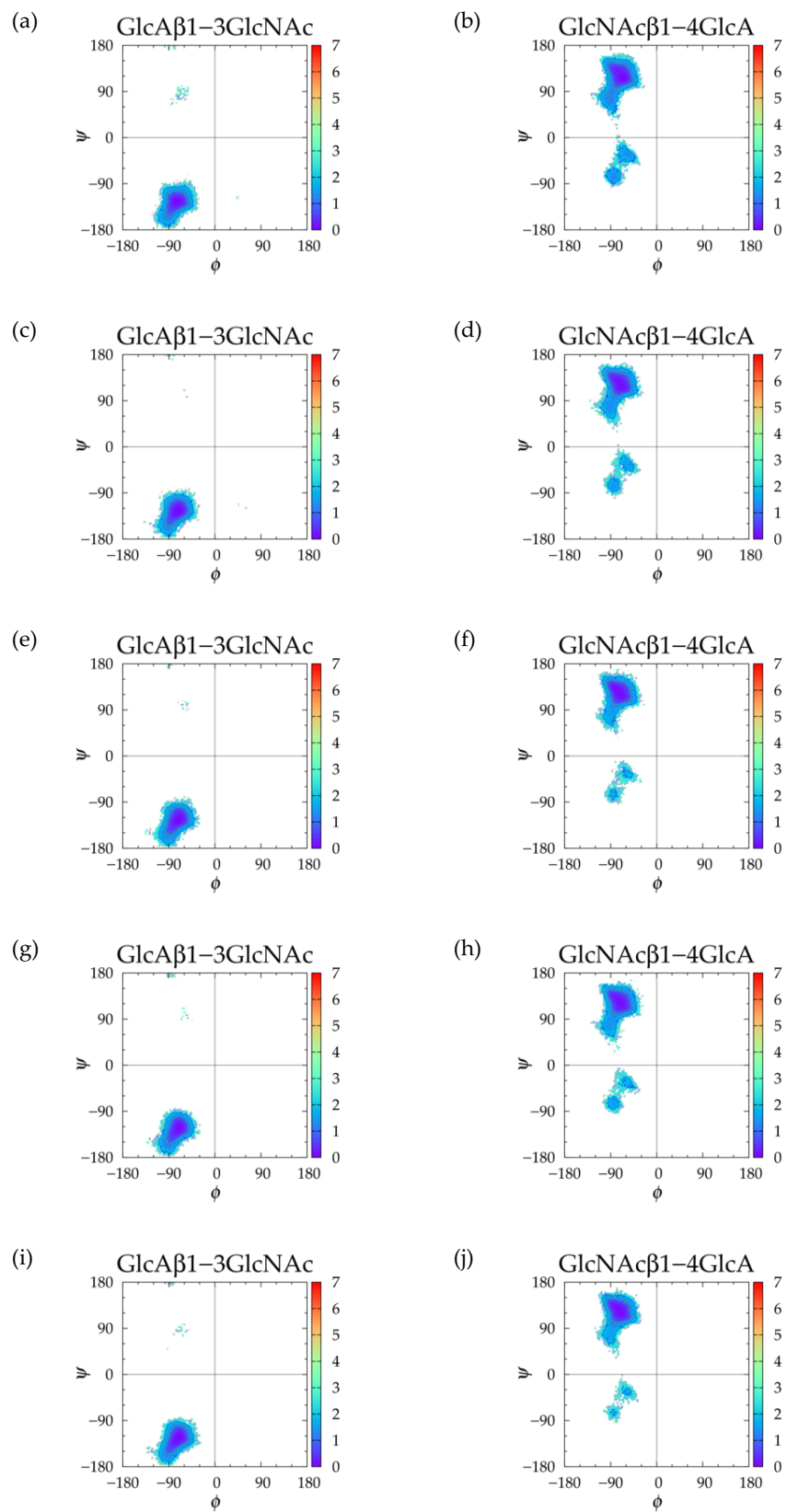


Figure S6. Scatterplot of radius of gyration as a function of end-to-end distance in MD-generated hyaluronan 20-mer conformations with non- 4C_1 ring puckers (pink) overlaid on data for full MD-generated hyaluronan 20-mer ensemble (blue) and corresponding linear regression (*Figure S1a*).



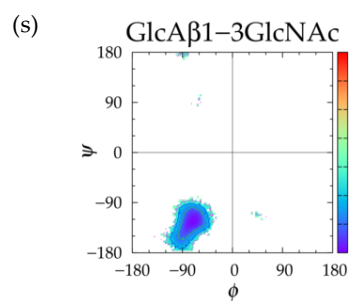
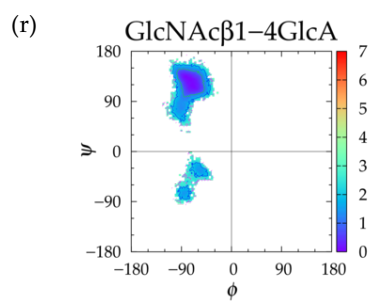
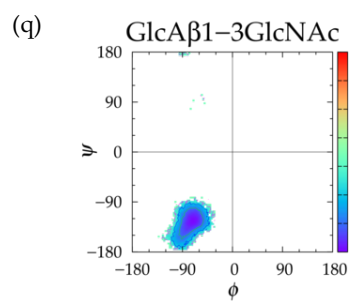
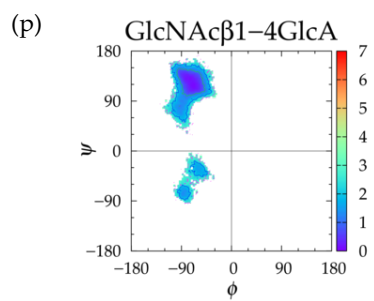
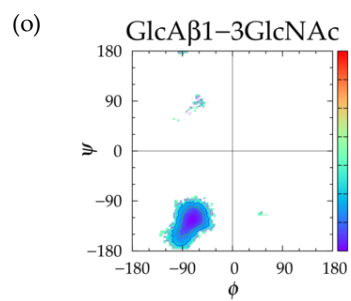
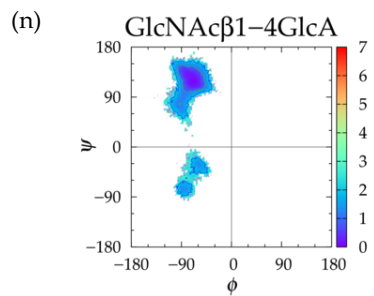
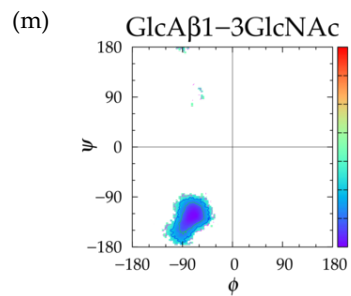
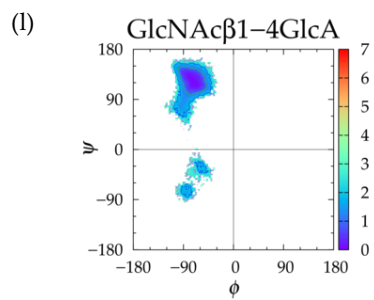
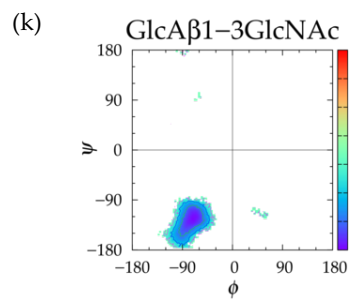


Figure S7. $\Delta G(\phi, \psi)$ plots for each glycosidic linkage in the MD-generated hyaluronan 20-mer ensemble; (a) GlcA2→GlcNAc1, (b) GlcNAc3→GlcA2, (c) GlcA4→GlcNAc3, (d) GlcNAc5→GlcA4, (e) GlcA6→GlcNAc5, (f) GlcNAc7→GlcA6, (g) GlcA8→GlcNAc7, (h) GlcNAc9→GlcA8, (i) GlcA10→GlcNAc9, (j) GlcNAc11→GlcA10, (k) GlcA12→GlcNAc11, (l) GlcNAc13→GlcA12, (m) GlcA14→GlcNAc13, (n) GlcNAc15→GlcA14, (o) GlcA16→GlcNAc15, (p) GlcNAc17→GlcA16, (q) GlcA18→GlcNAc17, (r) GlcNAc19→GlcA18, and (s) GlcA20→GlcNAc19; monosaccharides are numbered from reducing to non-reducing end; ϕ, ψ separated into 2.5° bins.

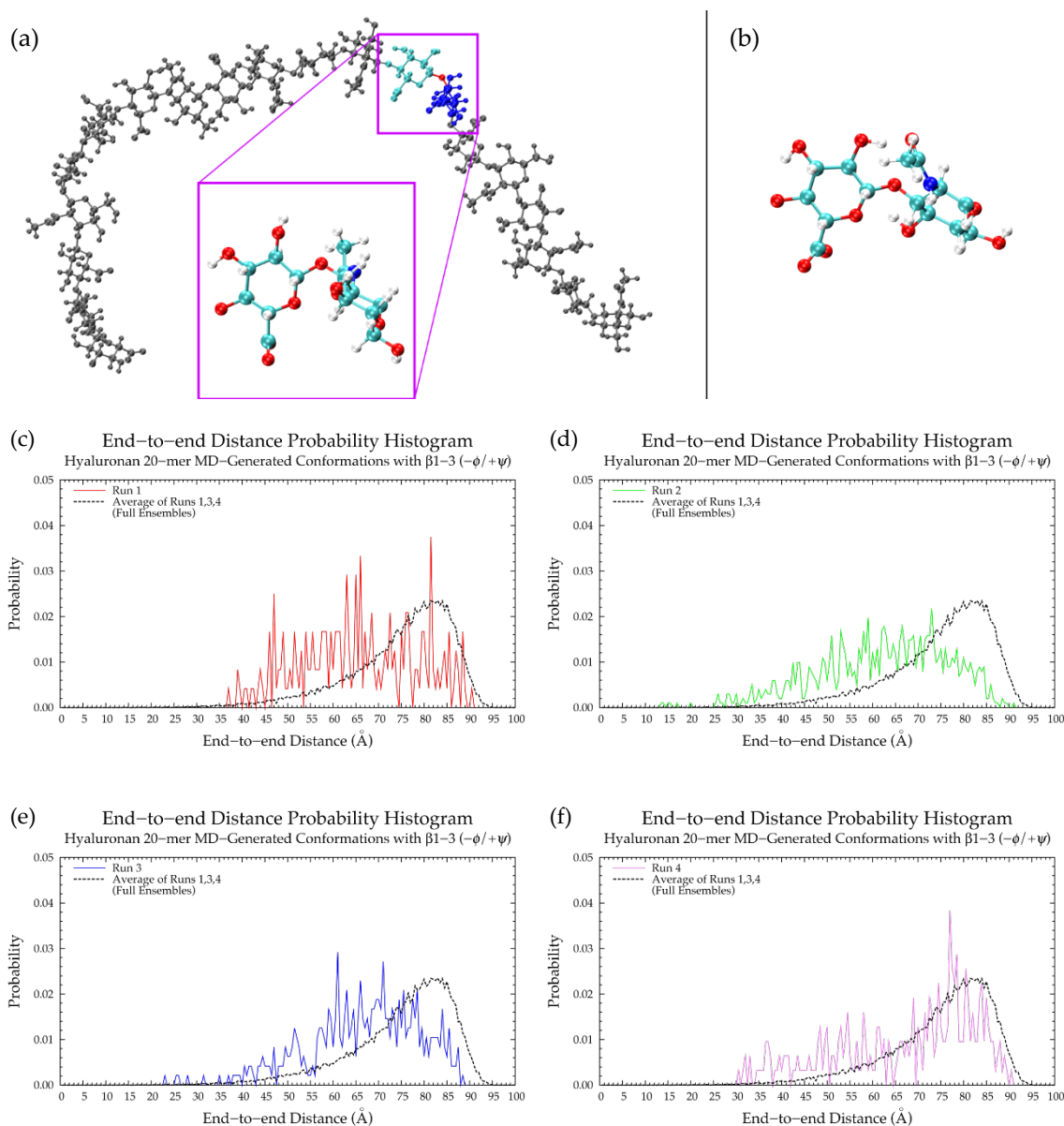


Figure S8. Snapshots from hyaluronan 20-mer MD: (a) 20-mer conformation with GlcA 8 (cyan) β 1-3 GlcNAc 7 (blue) linkage dihedrals near $\Delta G(\phi, \psi)$ min II ($\phi = -52.1^\circ$ and $\psi = +91.0^\circ$), which causes a kink (linker oxygen is red), and closeup of this disaccharide unit, (b) closeup of the same disaccharide unit with linkage dihedrals near $\Delta G(\phi, \psi)$ min I ($\phi = -70.9^\circ$ and $\psi = -119.1^\circ$). (c-f) End-to-end distance probability distributions of MD-generated hyaluronan 20-mer conformations with β 1-3 linkages with $-\phi, +\psi$ dihedrals in each of the four runs; as the end-to-end distance distribution from run 2 appeared to be an outlier, these data were compared to the average end-to-end distance distribution of all snapshots in MD runs 1, 3, and 4 (black dashed line).

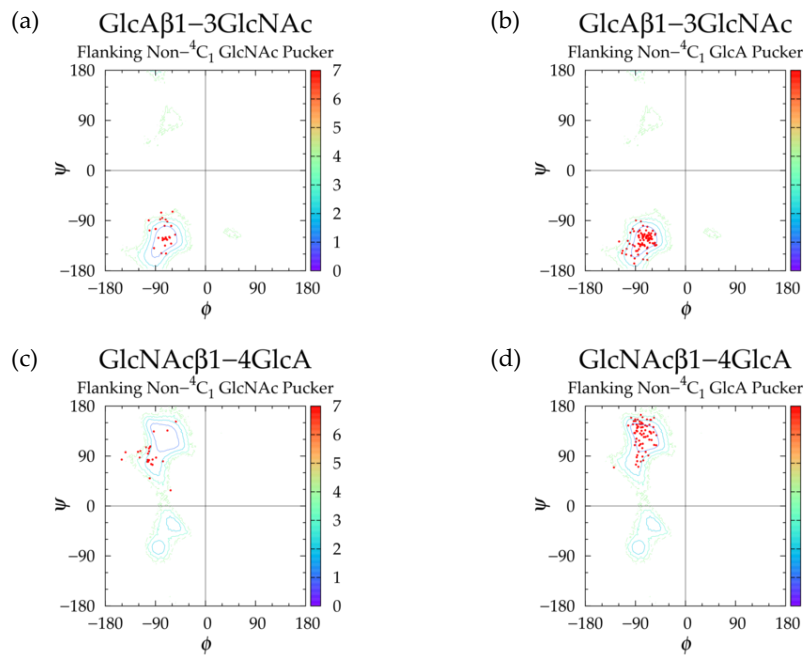


Figure S9. Scatterplots for dihedrals ϕ and ψ of glycosidic linkages flanking non-⁴C₁ ring puckers in the MD-generated hyaluronan 20-mer ensemble: (a) GlcA β 1-3GlcNAc flanking GlcNAc, (b) GlcA β 1-3GlcNAc flanking GlcA, (c) GlcNAc β 1-4GlcA flanking GlcNAc, (d) GlcNAc β 1-4GlcA flanking GlcA; contour lines come from corresponding aggregated MD-generated $\Delta G(\phi, \psi)$ data.

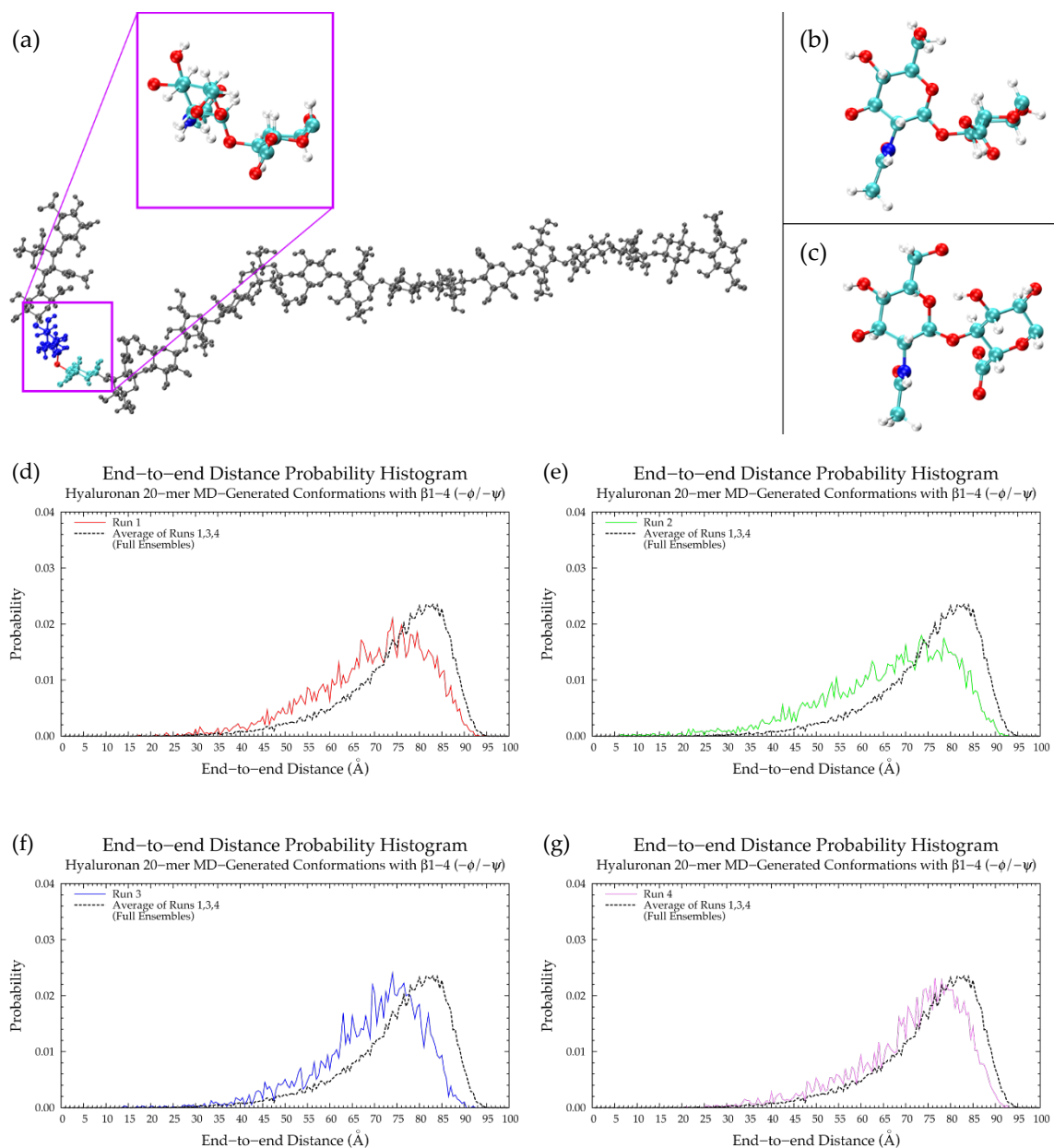


Figure S10. Snapshots from hyaluronan 20-mer MD: (a) 20-mer conformation with GlcNAc 17 (blue) β 1-4 GlcA 16 (cyan) linkage dihedrals near $\Delta G(\phi, \psi)$ min II ($\phi = -86.8^\circ$ and $\psi = -78.6^\circ$), which causes a kink (linker oxygen is red), and closeup of this disaccharide unit, (b) closeup of this disaccharide unit with linkage dihedrals near $\Delta G(\phi, \psi)$ min II' ($\phi = -55.7^\circ$ and $\psi = -39.7^\circ$), (c) closeup of the same disaccharide unit with linkage dihedrals near $\Delta G(\phi, \psi)$ min I ($\phi = -68.3^\circ$ and $\psi = +115.2^\circ$). (d-g) End-to-end distance probability distributions of MD-generated hyaluronan 20-mer conformations with β 1-4 linkages with $-\phi, -\psi$ dihedrals in each of the four runs; as the end-to-end distance distribution from run 2 appeared to be an outlier, these data were compared to the average end-to-end distance distribution of all snapshots in MD runs 1, 3, and 4 (black dashed line).

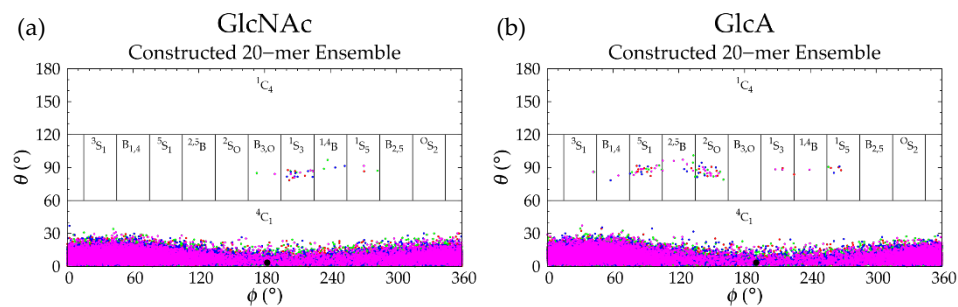


Figure S11. Cremer–Pople data for (a) GlcNAc and (b) GlcA in the constructed hyaluronan 20-mer ensemble; each of the 4 runs is represented by different color and the force-field geometry is represented by a single large black dot; each run contains 10,000 parameter sets.

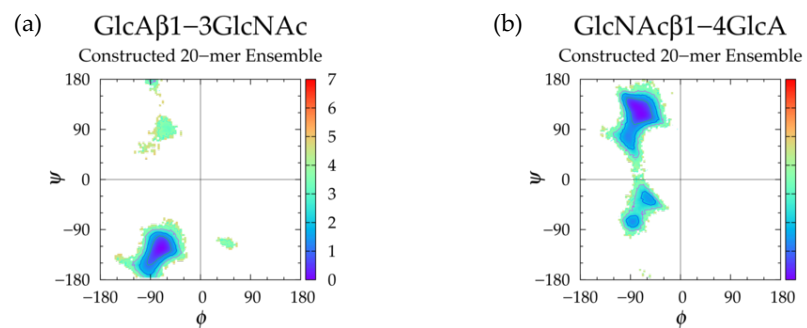


Figure S12. $\Delta G(\phi, \psi)$ in the constructed hyaluronan 20-mer ensemble for aggregated (a) GlcA β 1-3GlcNAc and (b) GlcNAc β 1-4GlcA glycosidic linkage data; contour lines every 1 kcal/mol.

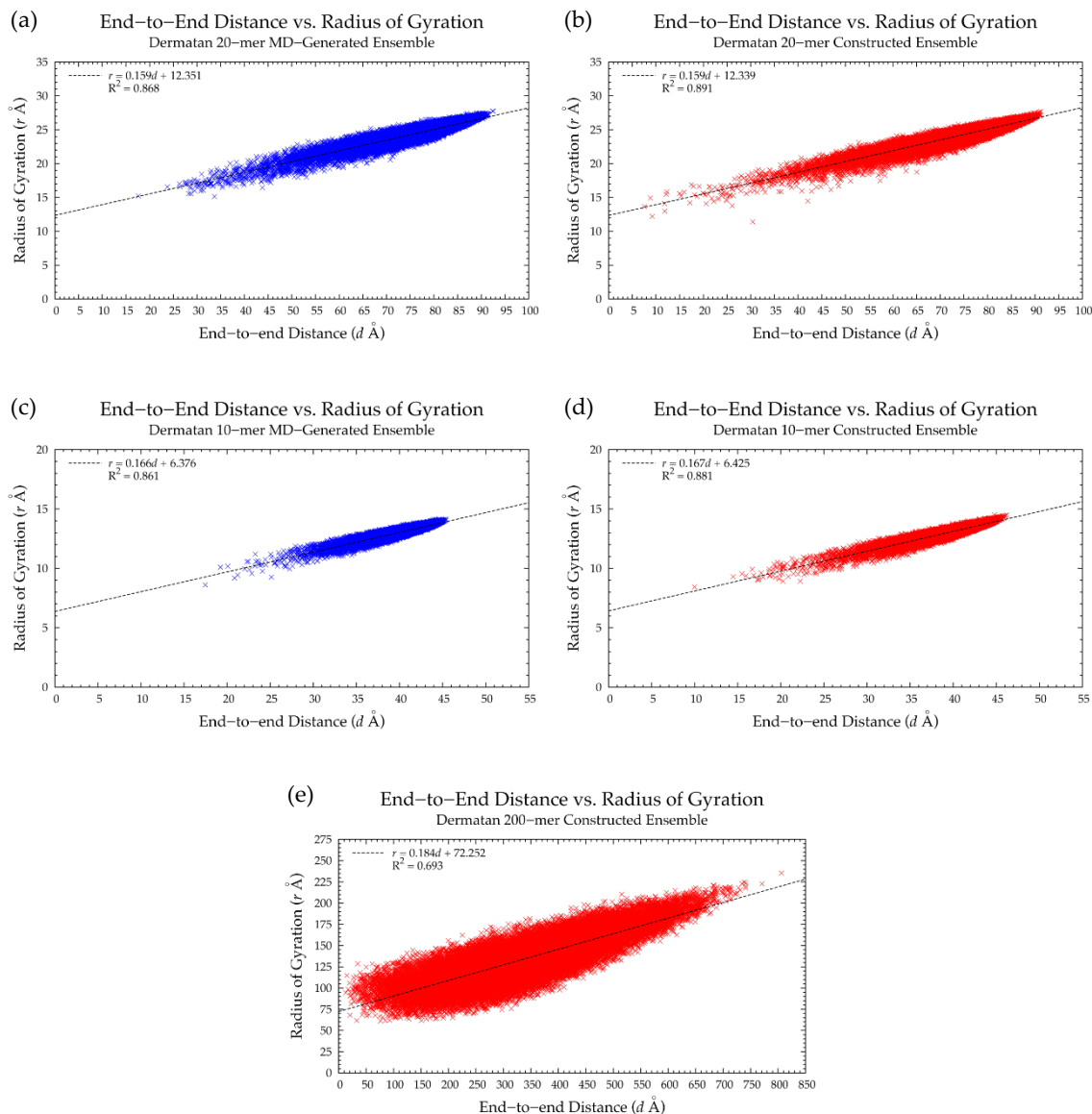


Figure S13. Scatterplots of radius of gyration as a function of end-to-end distance in MD-generated and constructed ensembles of non-sulfated dermatan (a,b) 20-mer and (c,d) 10-mer, respectively, and (e) constructed ensemble of non-sulfated dermatan 200-mer; each plot has 40,000 samples and shows linear regression and R^2 .

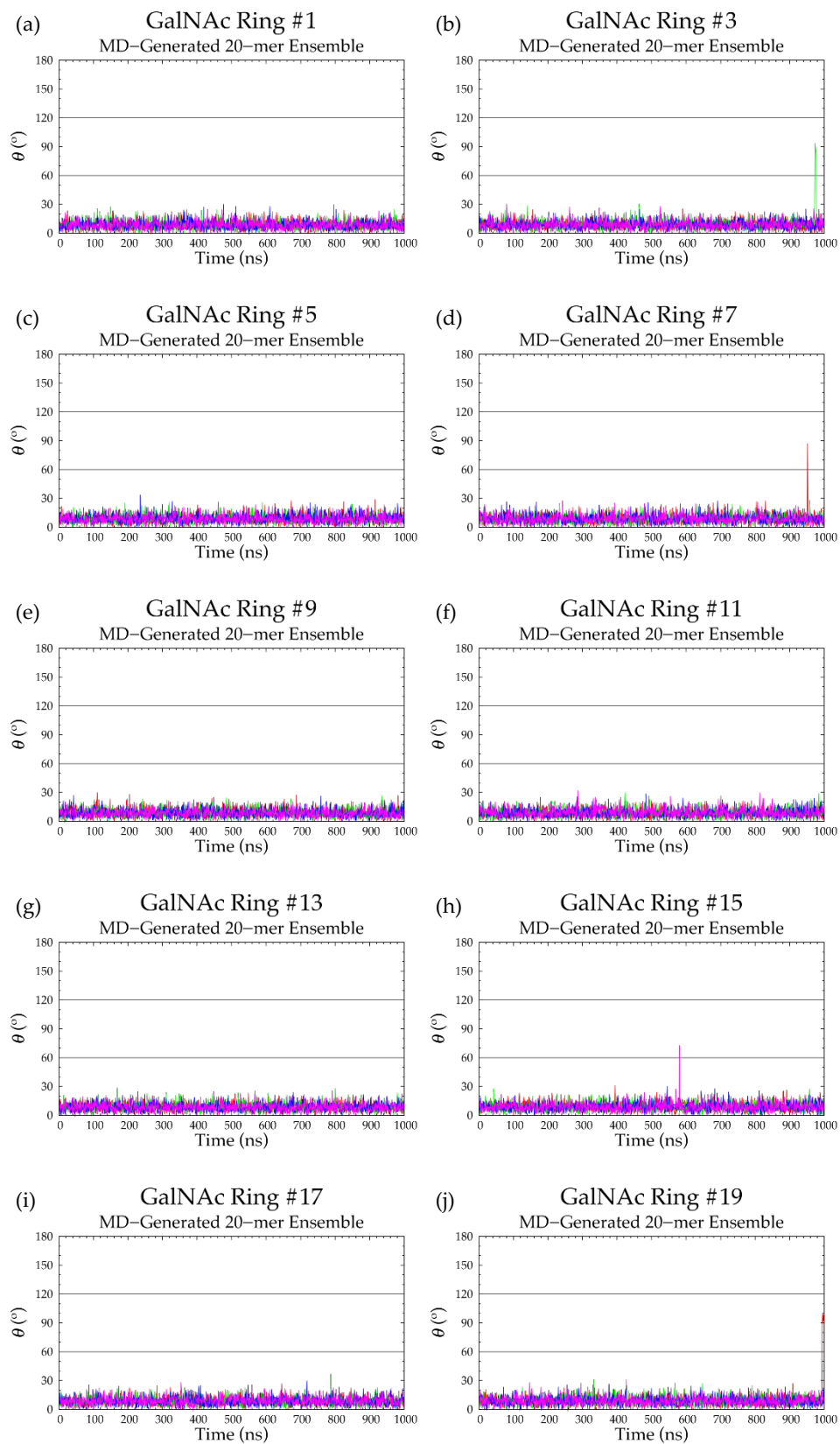


Figure S14. (a-j) Cremer-Pople parameter θ timeseries for each GalNAc monosaccharide ring in the MD-generated non-sulfated dermatan 20-mer ensemble; monosaccharides are numbered from reducing to non-reducing end; each of the 4 runs is represented by different color.

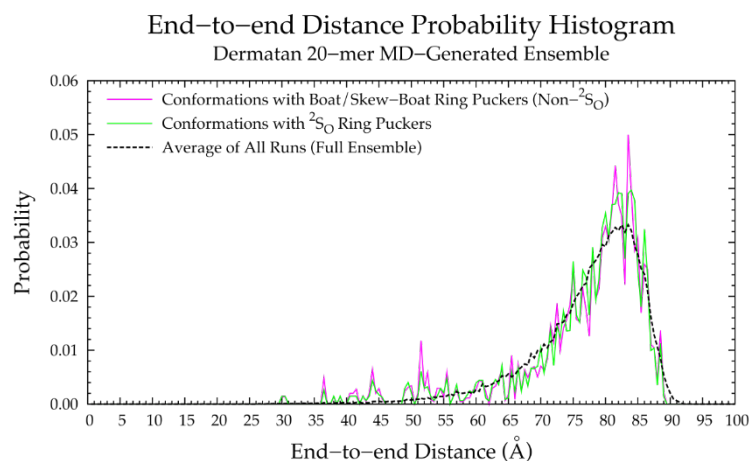
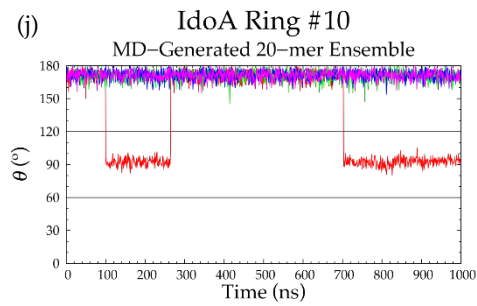
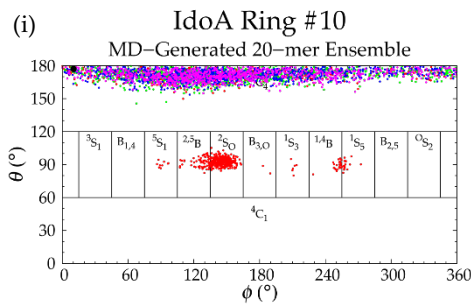
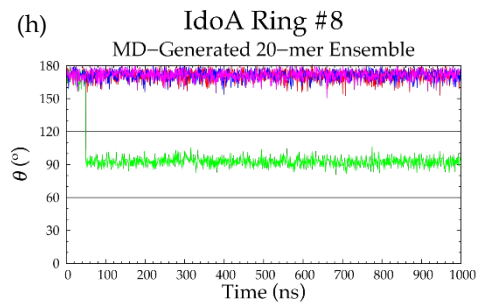
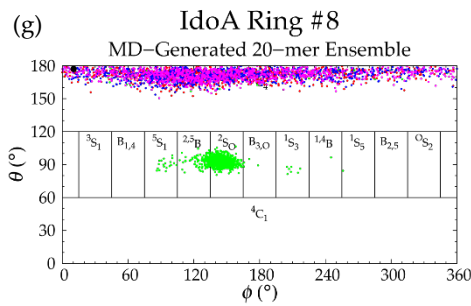
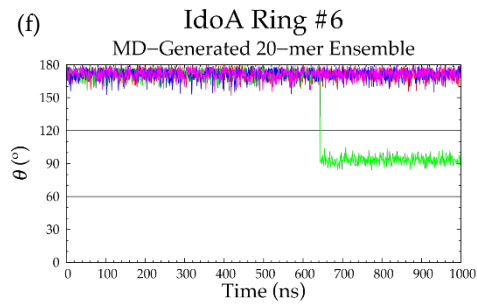
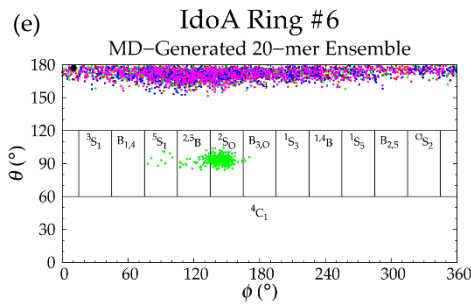
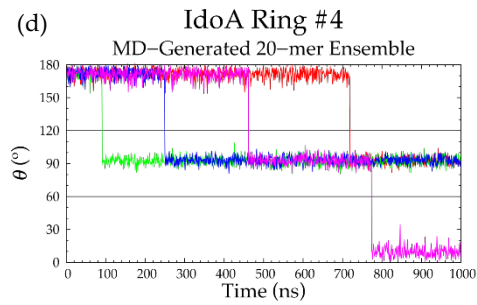
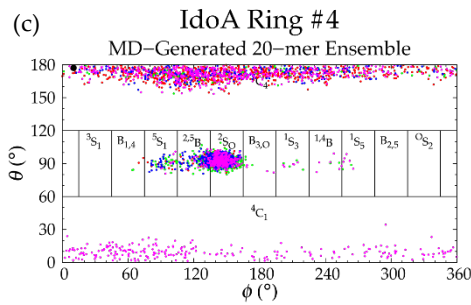
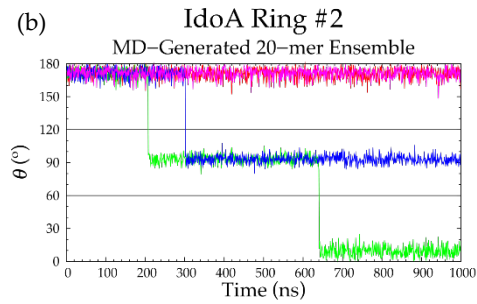
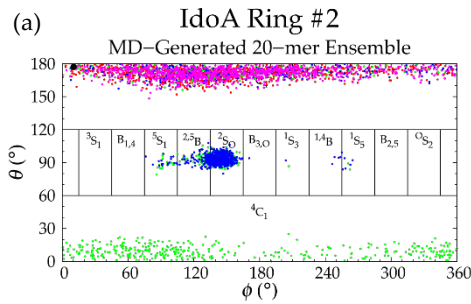


Figure S15. End-to-end distance distributions of MD-generated non-sulfated dermatan 20-mer conformations with boat/skew-boat ring puckers that cause a kink in the polymer chain, i.e. non- 2S_0 (pink solid line; most probable end-to-end distance is 83.5 Å) and 2S_0 conformations (green solid line; most probable end-to-end distance is 84.0 Å) and the average of all four runs in the full MD-generated ensemble (black dashed line; most probable end-to-end distance is 83.5 Å); probabilities were calculated for end-to-end distances sorted into 0.5 Å bins.



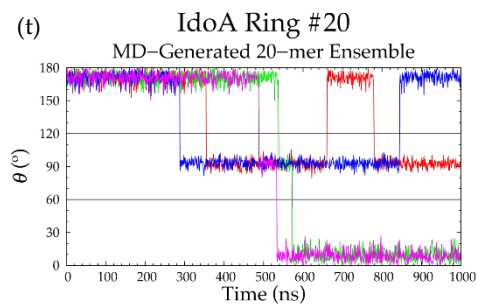
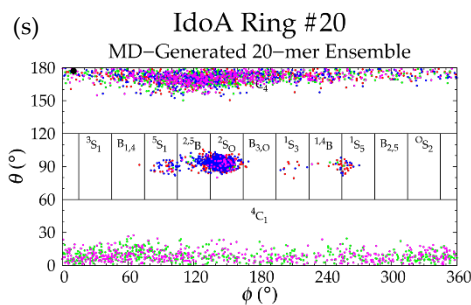
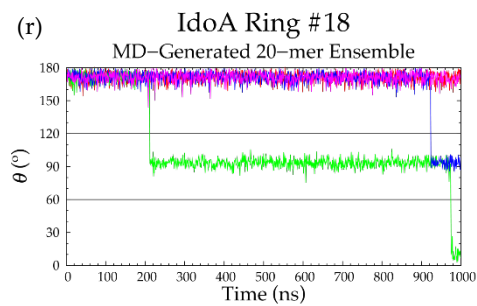
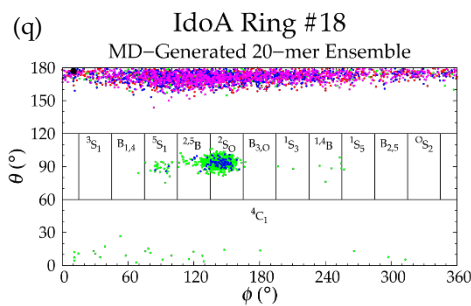
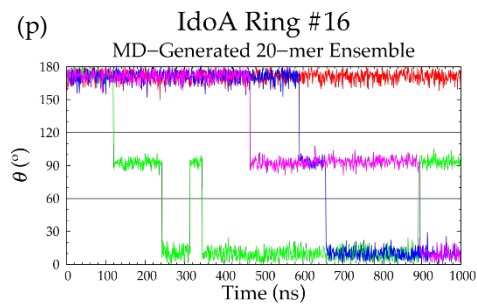
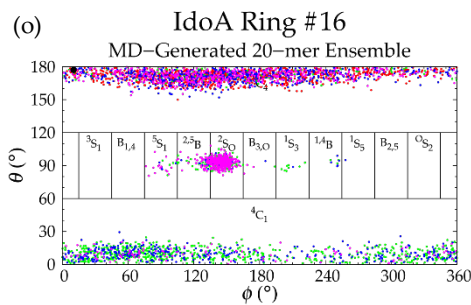
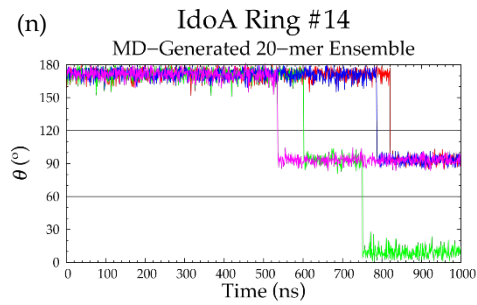
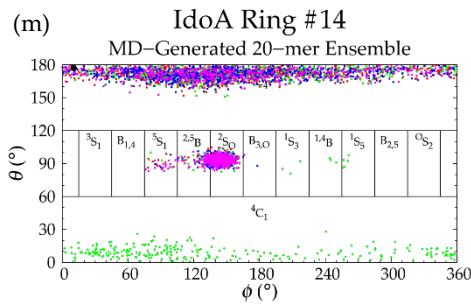
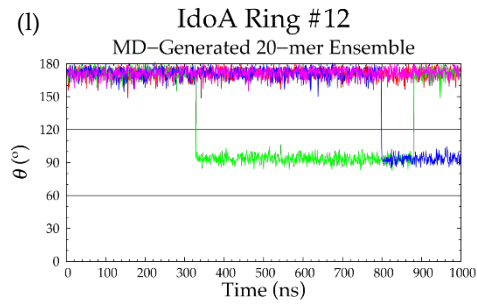
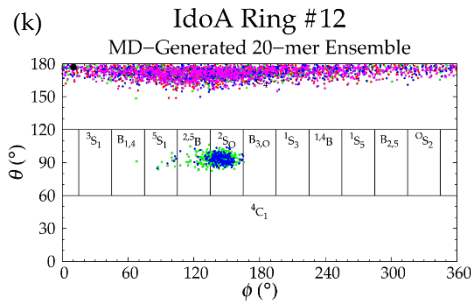


Figure S16. (a,c,e,g,i,k,m,o,q,s) Cremer-Pople plots and (b,d,f,h,j,l,n,p,r,t) Cremer-Pople parameter θ timeseries for each IdoA monosaccharide ring in the MD-generated non-sulfated dermatan 20-mer ensemble; monosaccharides are numbered from reducing to non-reducing end; each of the 4 runs is represented by different color.

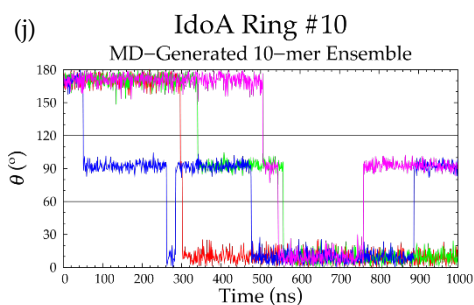
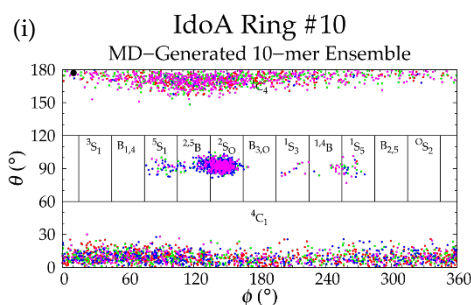
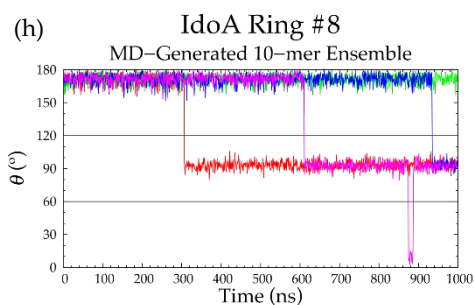
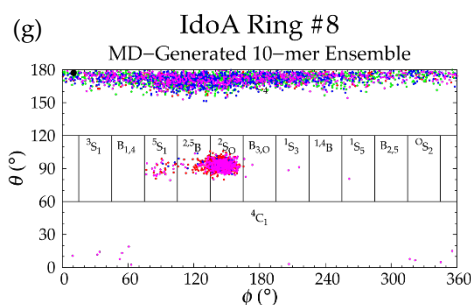
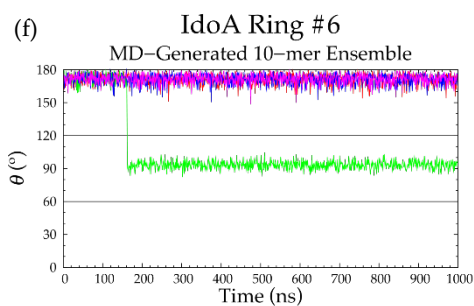
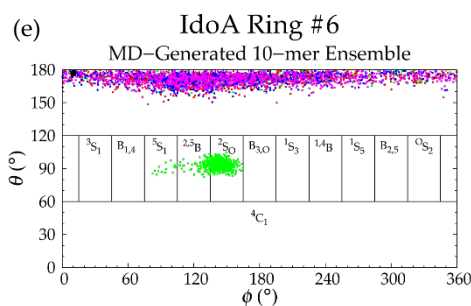
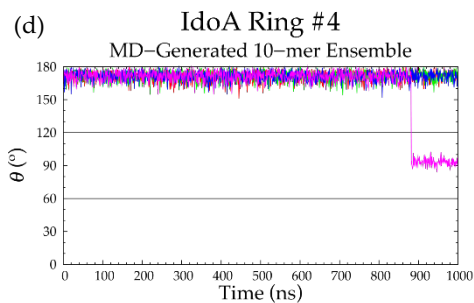
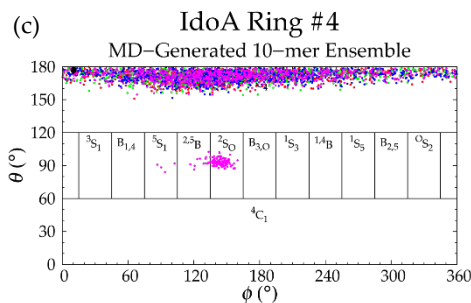
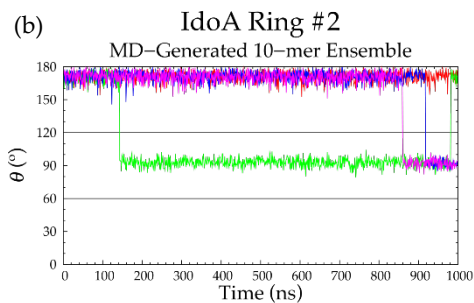
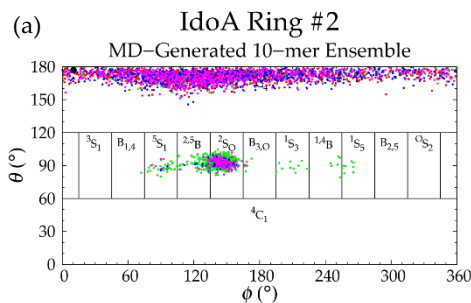


Figure S17. (a,c,e,g,i) Cremer-Pople plots and (b,d,f,h,j) Cremer-Pople parameter θ timeseries for each IdoA monosaccharide ring in the MD-generated non-sulfated dermatan 10-mer ensemble; monosaccharides are numbered from reducing to non-reducing end; each of the 4 runs is represented by different color.

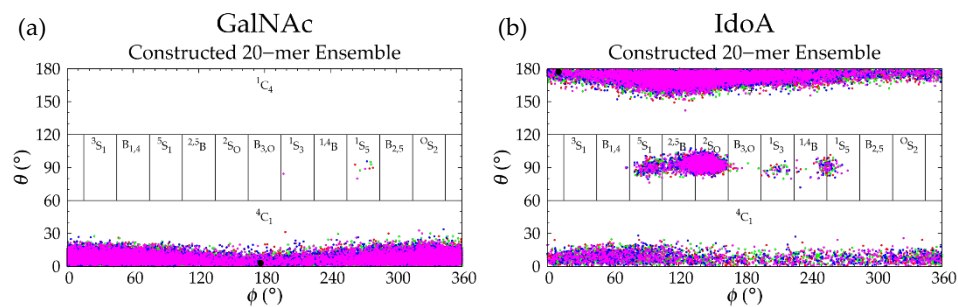


Figure S18. Cremer–Pople data for (a) GalNAc and (b) IdoA in the constructed non-sulfated dermatan 20-mer ensemble; each of the 4 runs is represented by different color and the force-field geometry is represented by a single large black dot; each run contains 10,000 parameter sets.

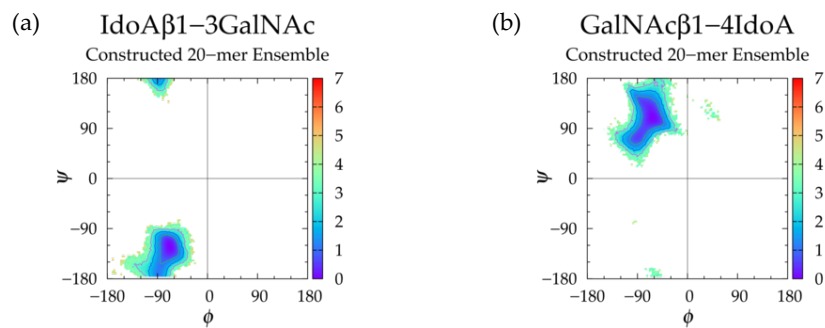


Figure S19. $\Delta G(\phi, \psi)$ in the constructed non-sulfated dermatan 20-mer ensemble for aggregated (a) IdoA β 1-3GalNAc and (b) GalNAc β 1-4IdoA glycosidic linkage data; contour lines every 1 kcal/mol.

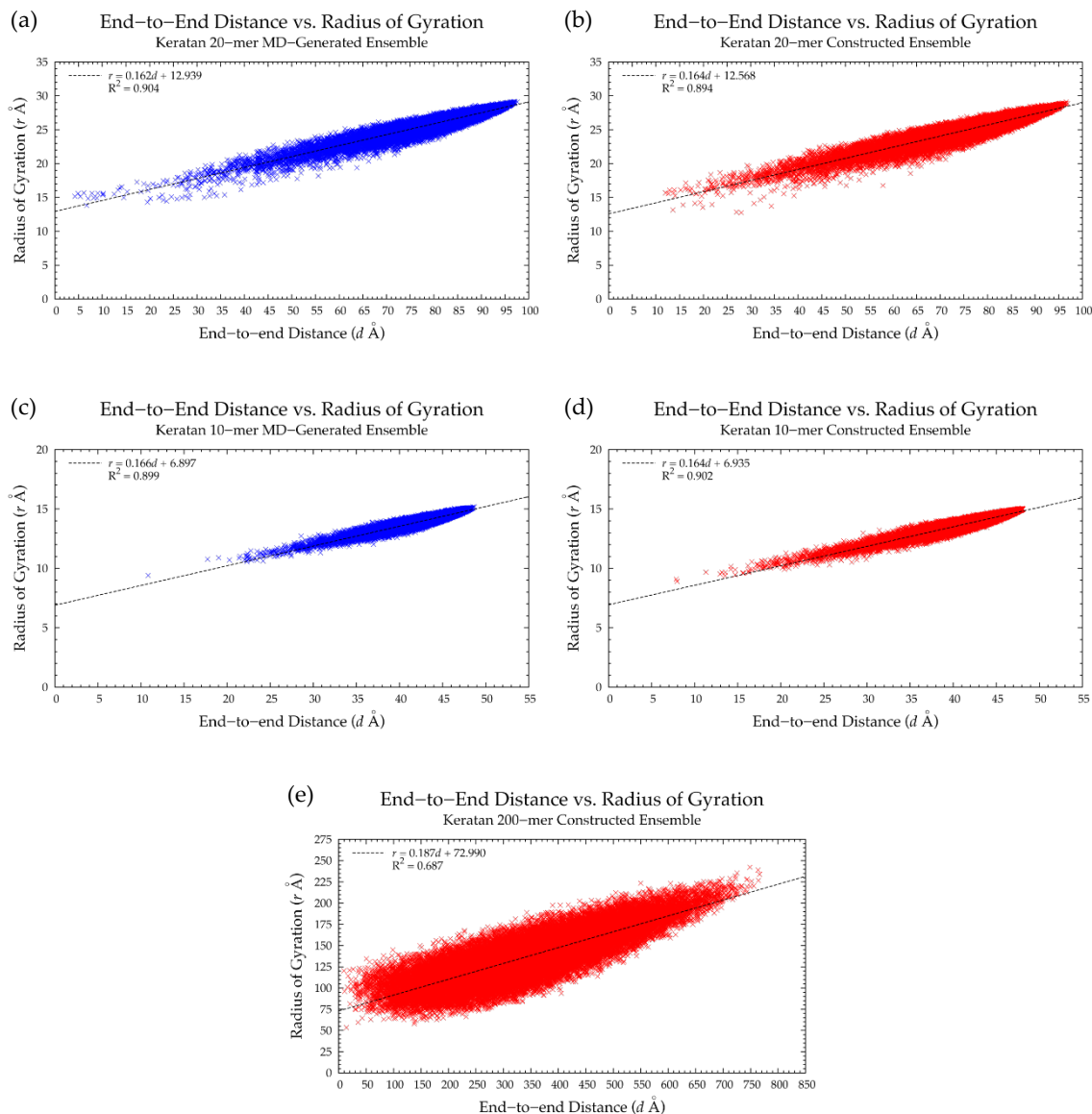
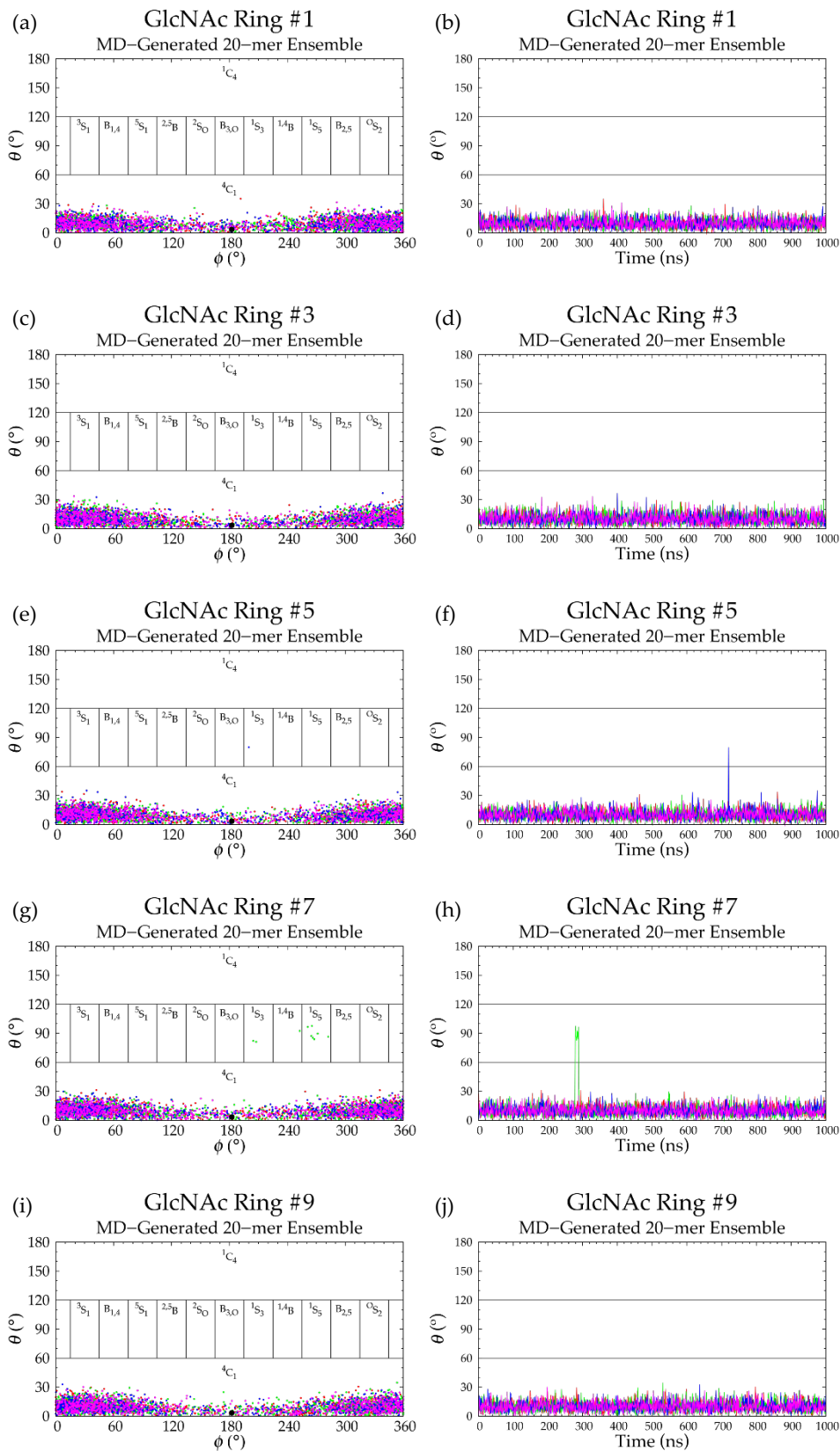


Figure S20. Scatterplots of radius of gyration as a function of end-to-end distance in MD-generated and constructed ensembles of non-sulfated keratan (a,b) 20-mer and (c,d) 10-mer, respectively, and (e) constructed ensemble of non-sulfated keratan 200-mer; each plot has 40,000 samples and shows linear regression and R^2 .



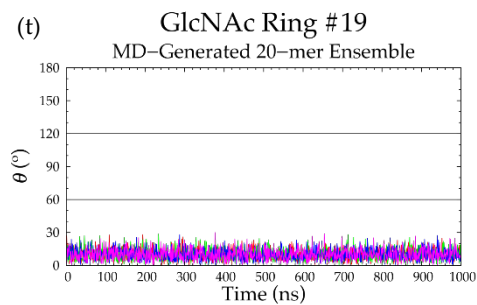
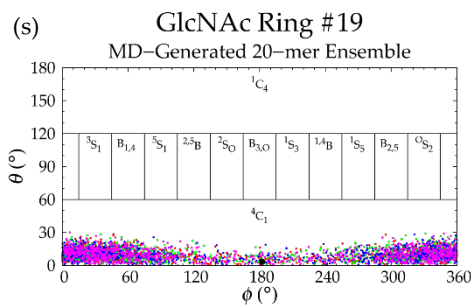
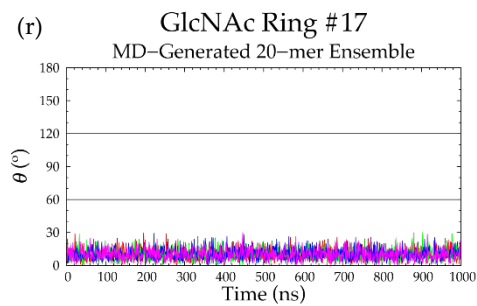
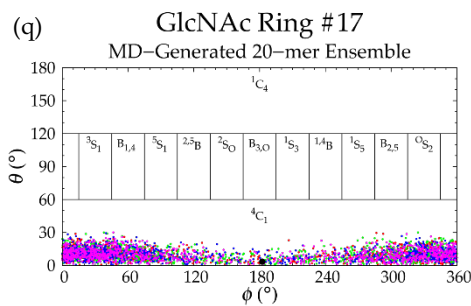
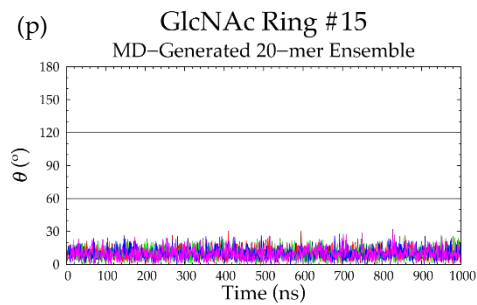
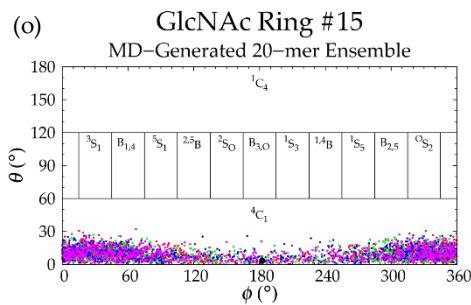
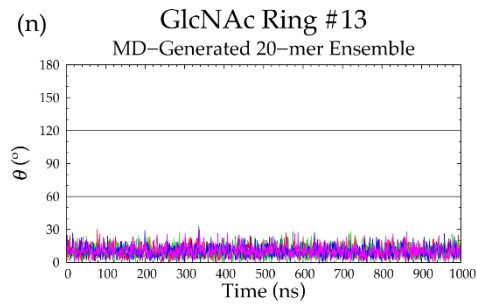
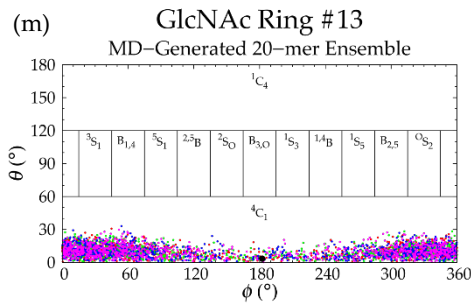
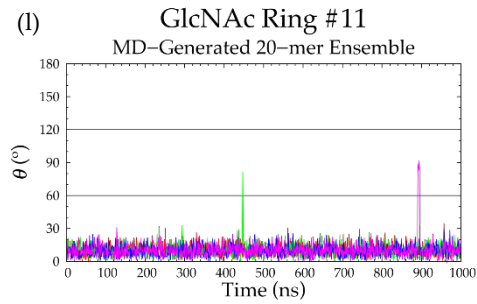
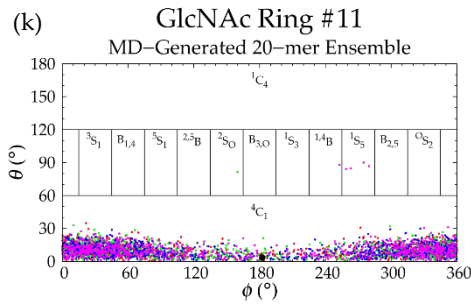


Figure S21. (a,c,e,g,i,k,m,o,q,s) Cremer-Pople plots and (b,d,f,h,j,l,n,p,r,t) Cremer-Pople parameter θ timeseries for each GlcNAc monosaccharide ring in the MD-generated non-sulfated keratan 20-mer ensemble; monosaccharides are numbered from reducing to non-reducing end; each of the 4 runs is represented by different color.

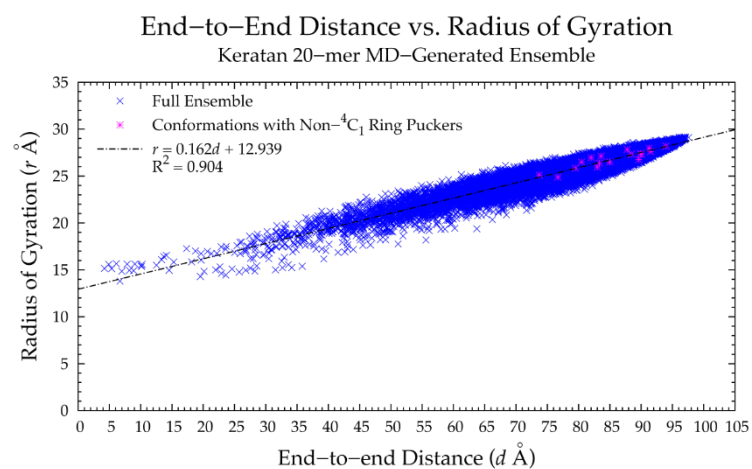


Figure S22. Scatterplot of radius of gyration as a function of end-to-end distance in MD-generated non-sulfated keratan 20-mer conformations with non- 4C_1 ring puckers overlaid on data for full MD-generated non-sulfated keratan 20-mer ensemble and corresponding linear regression (*Figure S34a*).

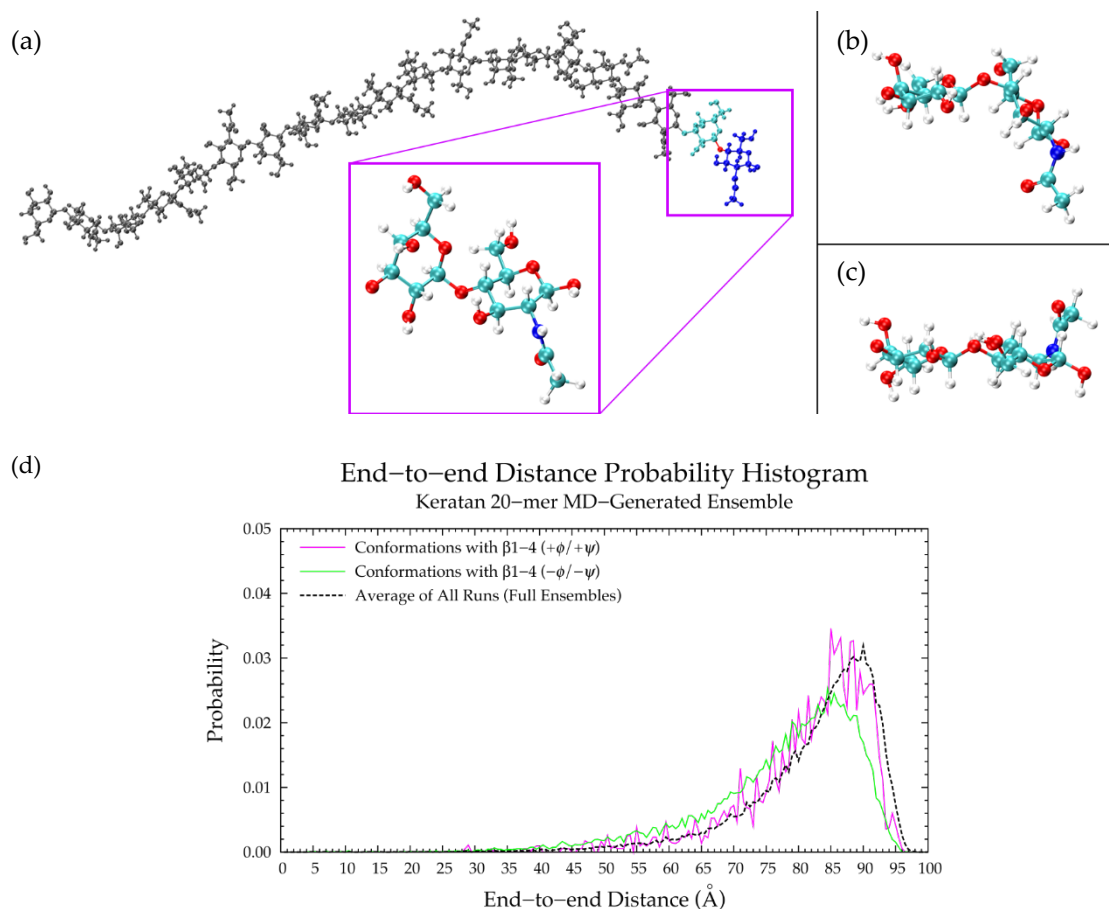


Figure S23. Snapshots from non-sulfated keratan 20-mer MD: (a) 20-mer conformation with Gal 2 (cyan) $\beta 1-4$ GlcNAc 1 (blue) linkage dihedrals in tertiary basin ($\phi = +13.8^\circ$ and $\psi = +131.1^\circ$), which causes a slight bend (linker oxygen is red), and closeup of this disaccharide unit, (b) closeup of this disaccharide unit with linkage dihedrals in secondary basin, i.e. near $\Delta G(\phi, \psi)$ min II ($\phi = -98.8^\circ$ and $\psi = -71.0^\circ$), (c) closeup of the same disaccharide unit with linkage dihedrals in primary basin near $\Delta G(\phi, \psi)$ min I ($\phi = -86.2^\circ$ and $\psi = +100.5^\circ$). (d) End-to-end distance probability distributions of MD-generated non-sulfated keratan 20-mer conformations with $\beta 1-4$ linkages with $+\phi, +\psi$ (pink solid line; most probable end-to-end distance is 85.0 Å) and $-\phi, -\psi$ (green solid line; most probable end-to-end distance is 84.5 Å) dihedrals in all four runs; these data were compared to the average end-to-end distance distribution of all snapshots in all four MD runs (black dashed line; most probable end-to-end distance is 90.0 Å).

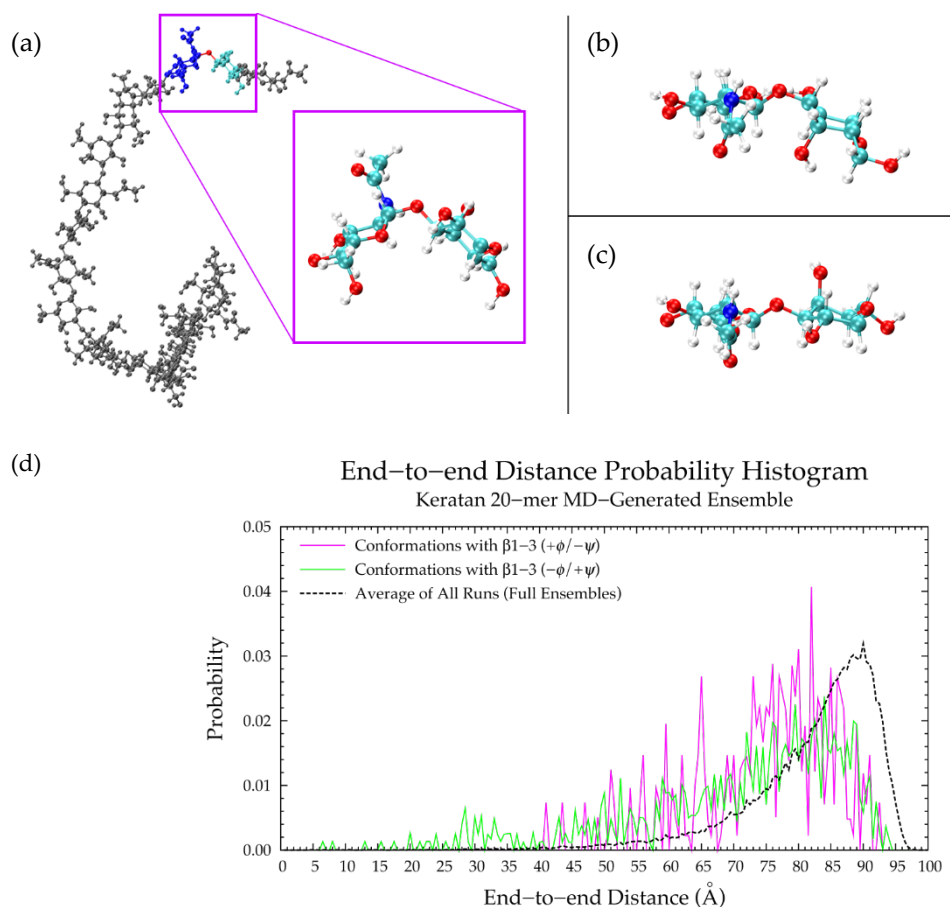


Figure S24. Snapshots from non-sulfated keratan 20-mer MD: (a) 20-mer conformation with GlcNAc 3 (blue) $\beta 1-3$ Gal 2 (cyan) linkage dihedrals in tertiary basin ($\phi = +38.8^\circ$ and $\psi = -127.3^\circ$), which causes a kink (linker oxygen is red), and closeup of this disaccharide unit, (b) closeup of GlcNAc 5 (blue) $\beta 1-3$ Gal 4 (cyan) disaccharide unit with linkage dihedrals in secondary basin ($\phi = -95.3^\circ$ and $\psi = +39.9^\circ$), (c) closeup of the same disaccharide unit with linkage dihedrals in primary basin ($\phi = -80.7^\circ$ and $\psi = -138.2^\circ$). (d) End-to-end distance probability distributions of MD-generated non-sulfated keratan 20-mer conformations with $\beta 1-3$ linkages with $+\phi, -\psi$ (pink solid line; most probable end-to-end distance is 82.0 Å) and $-\phi, +\psi$ (green solid line; most probable end-to-end distance is 84.0 Å) dihedrals in all four runs; these data were compared to the average end-to-end distance distribution of all snapshots in all four MD runs (black dashed line; most probable end-to-end distance is 90.0 Å).

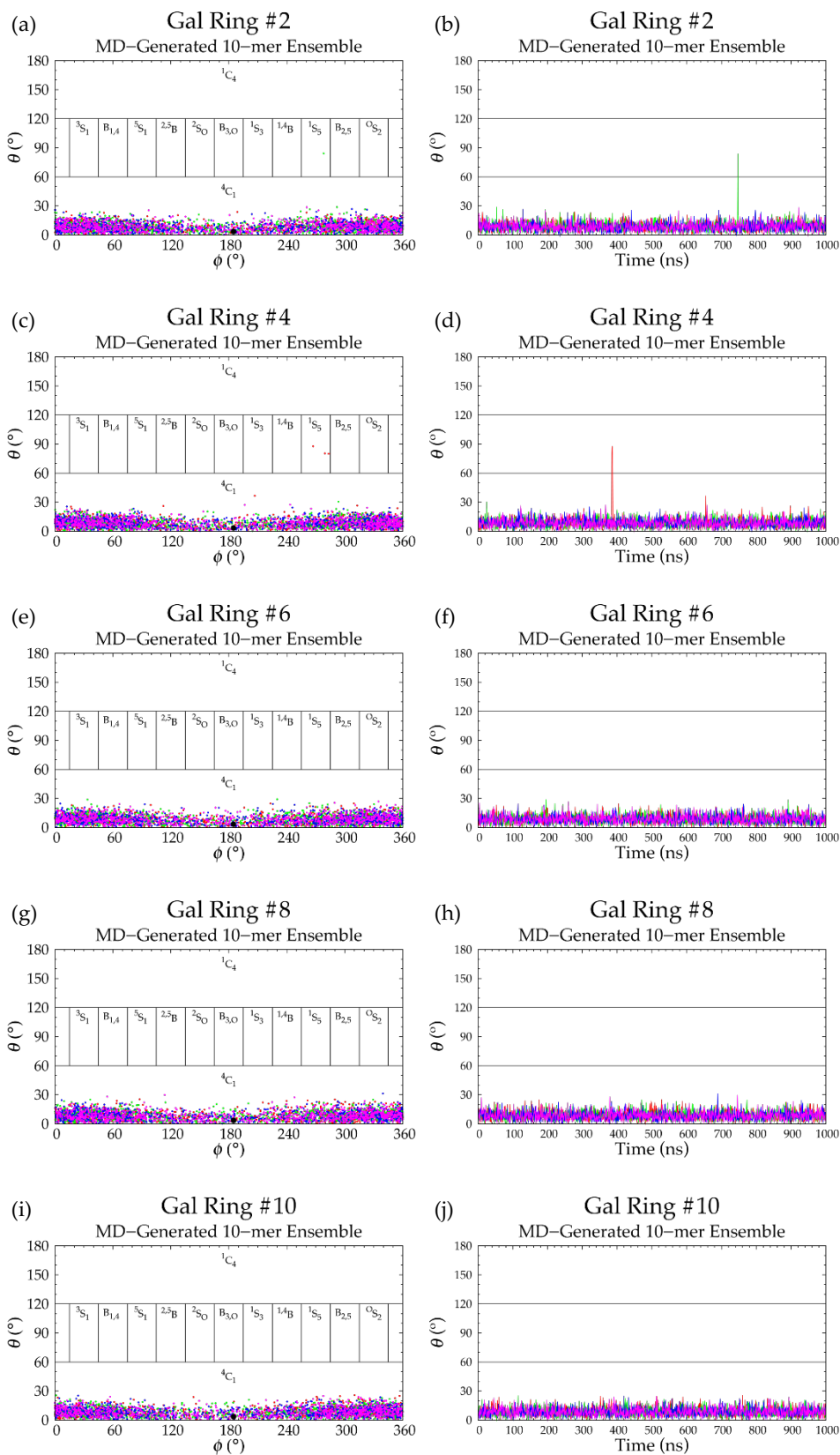


Figure S25. (a,c,e,g,i) Cremer-Pople plots and (b,d,f,h,j) Cremer-Pople parameter θ timeseries for each Gal monosaccharide ring in the MD-generated non-sulfated keratan 10-mer ensemble; monosaccharides are numbered from reducing to non-reducing end; each of the 4 runs is represented by different color.

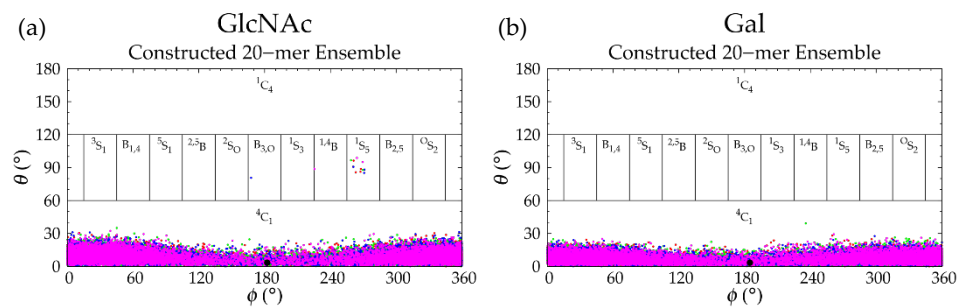


Figure S26. Cremer–Pople data for (a) GlcNAc and (b) Gal in the constructed non-sulfated keratan 20-mer ensemble; each of the 4 runs is represented by different color and the force-field geometry is represented by a single large black dot; each run contains 10,000 parameter sets.

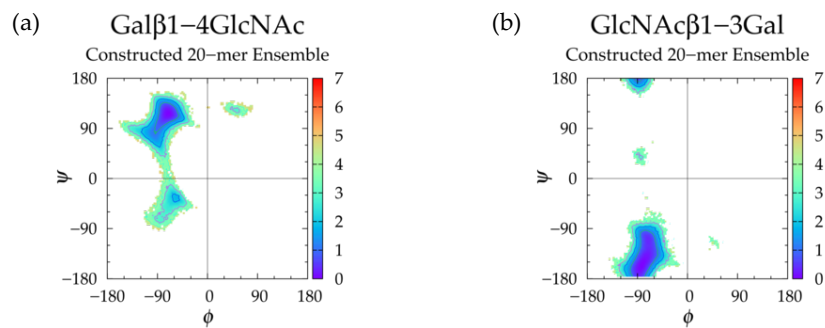


Figure S27. $\Delta G(\phi, \psi)$ in the constructed non-sulfated keratan 20-mer ensemble for aggregated (a) Galβ1-4GlcNAc and (b) GlcNAcβ1-3Gal glycosidic linkage data; contour lines every 1 kcal/mol.

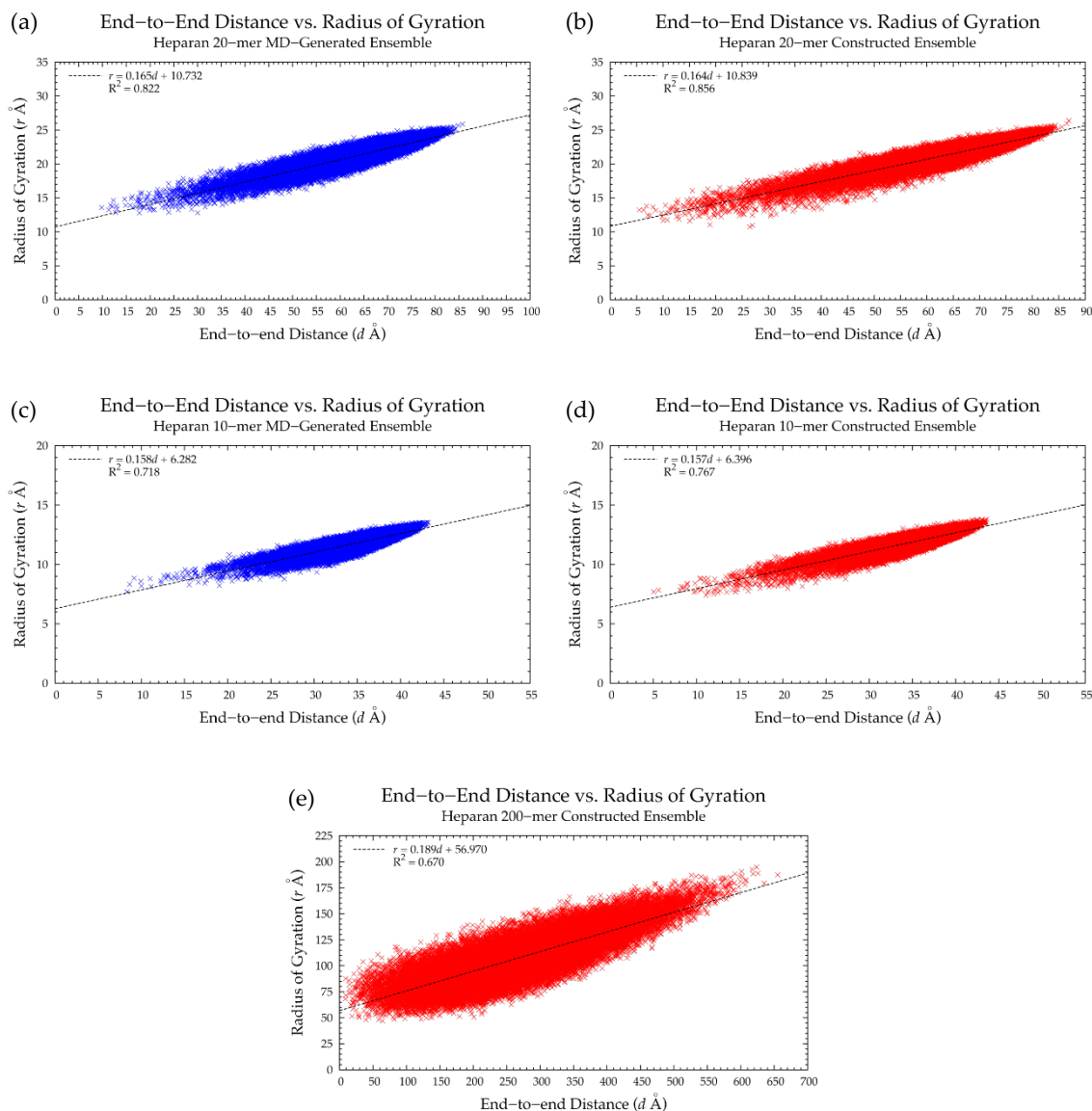


Figure S28. Scatterplots of radius of gyration as a function of end-to-end distance in MD-generated and constructed ensembles of non-sulfated heparan (a,b) 20-mer and (c,d) 10-mer, respectively, and (e) constructed ensemble of non-sulfated heparan 200-mer; each plot has 40,000 samples and shows linear regression and R^2 .

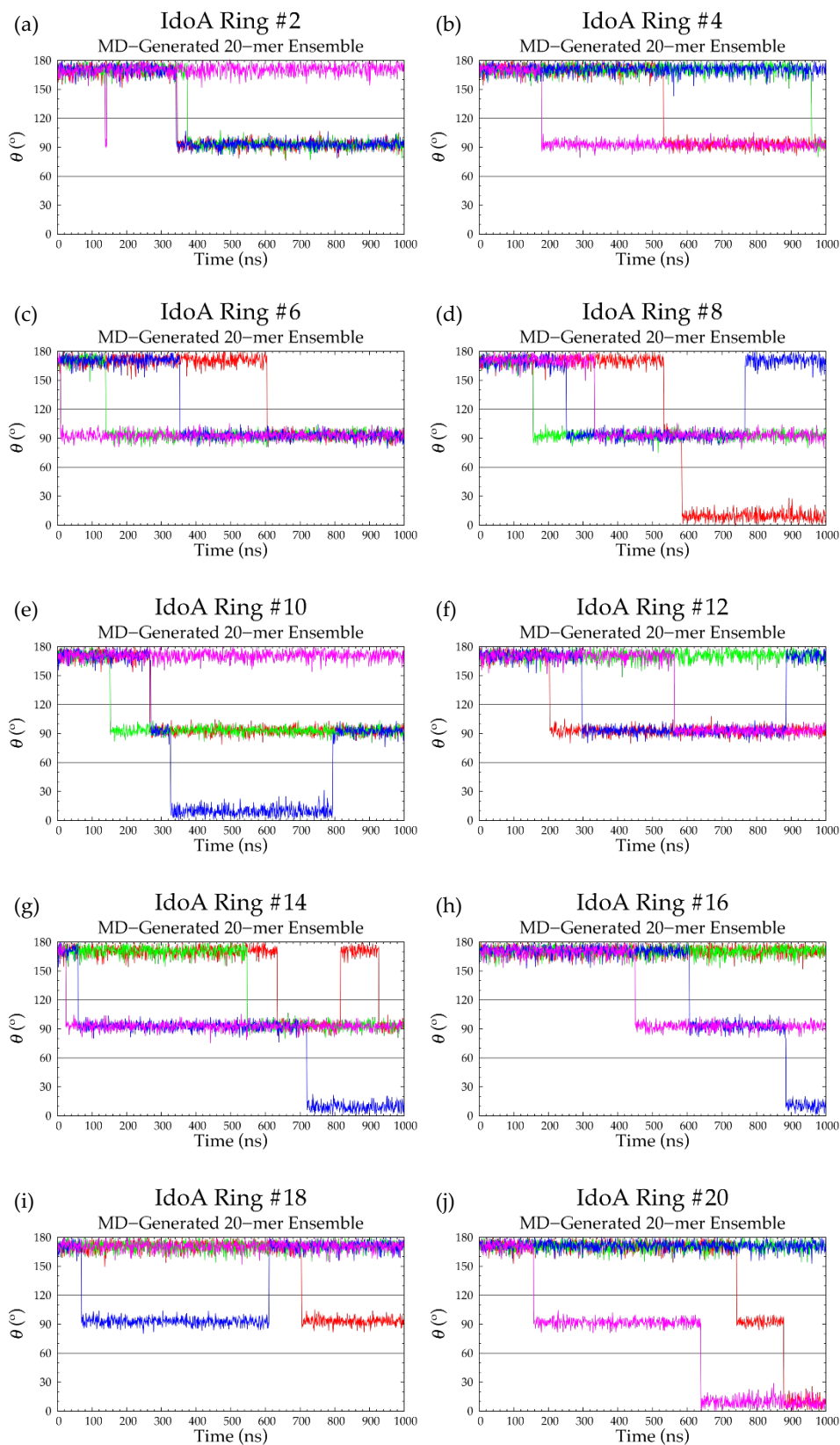


Figure S29. (a-j) Cremer-Pople parameter θ timeseries for each IdoA monosaccharide ring in the MD-generated non-sulfated heparan 20-mer ensemble; monosaccharides are numbered from reducing to non-reducing end; each of the 4 runs is represented by different color.

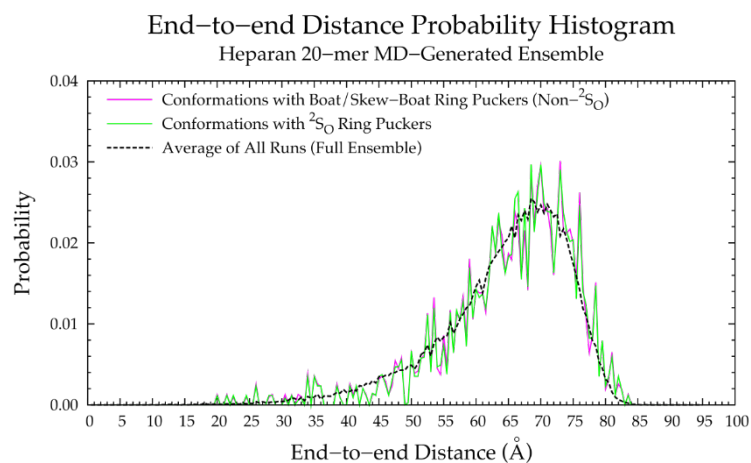


Figure S30. End-to-end distance distributions of MD-generated non-sulfated heparan 20-mer conformations with boat/skew-boat ring puckers that cause a kink in the polymer chain, i.e. non- 2S_0 (pink solid line; most probable end-to-end distance is 73.0 Å) and 2S_0 conformations (green solid line; most probable end-to-end distance is 68.5 Å) and the average of all four runs in the full MD-generated ensemble (black dashed line; most probable end-to-end distance is 68.5 Å); probabilities were calculated for end-to-end distances sorted into 0.5 Å bins.

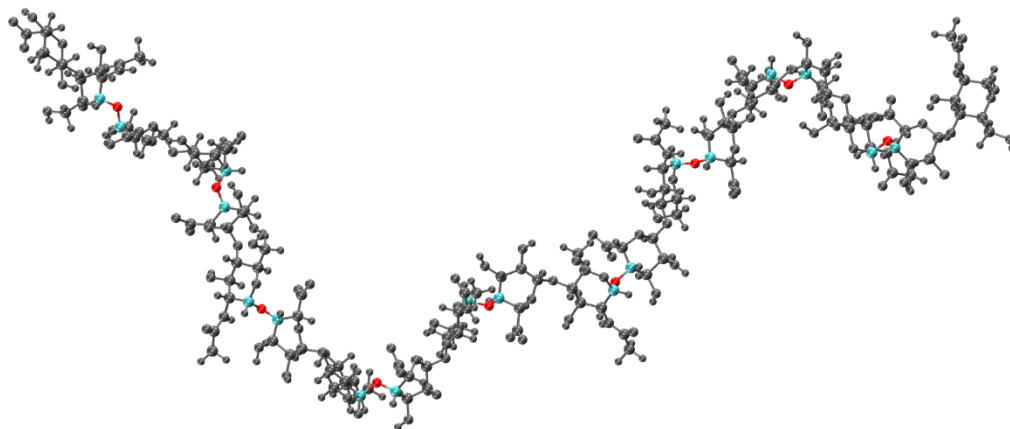


Figure S31. Snapshot of non-sulfated heparan 20-mer from MD simulation; GlcNAc α 1-4IdoA linkages are highlighted; linker oxygen atoms in red and carbon atoms (GlcNAc C₁ and IdoA C₄) in cyan.

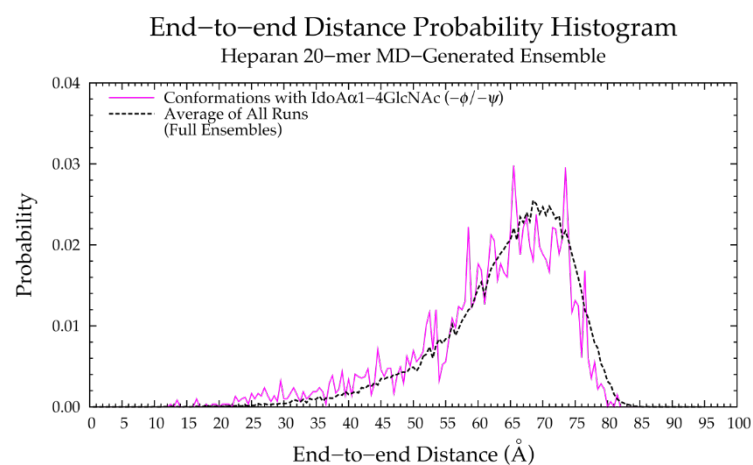


Figure S32. End-to-end distance probability distribution of MD-generated non-sulfated heparan 20-mer conformations with IdoA α 1-4GlcNAc linkages with $-\phi$, $-\psi$ dihedrals aggregated across all four MD runs (pink solid line; top two most probable end-to-end distances are 65.5 Å and 73.5 Å); these data were compared to the average end-to-end distance distribution of all snapshots in all four MD runs (black dashed line; most probable end-to-end distance is 68.5 Å).

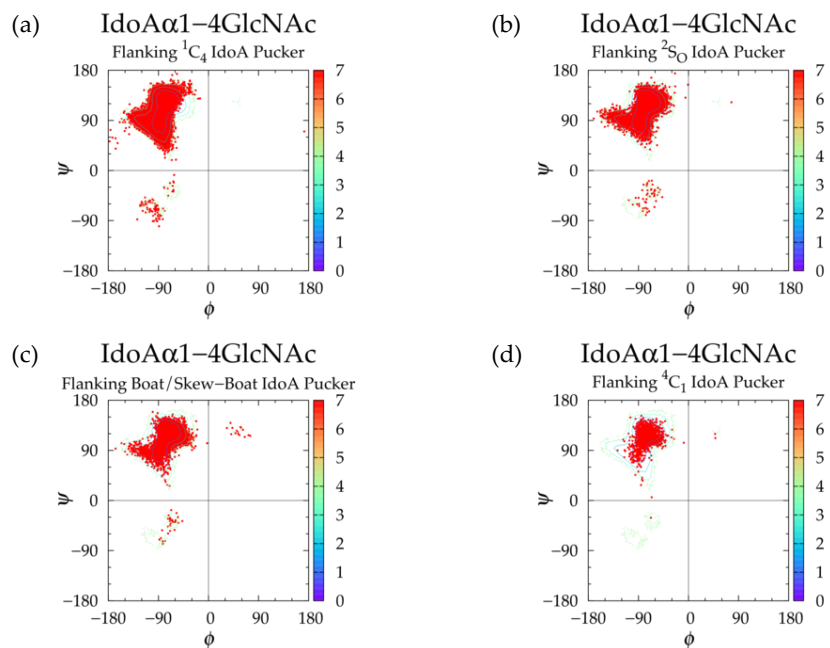


Figure S33. Scatterplots for dihedrals ϕ and ψ of IdoA α 1-4GlcNAc linkages flanking different IdoA conformations in the MD-generated non-sulfated heparan 20-mer ensemble: (a) 1C_4 , (b) 2S_0 , (c) boat/skew-boat (non- 2S_0), and (d) 4C_1 ; contour lines come from corresponding aggregated MD-generated $\Delta G(\phi, \psi)$ data.

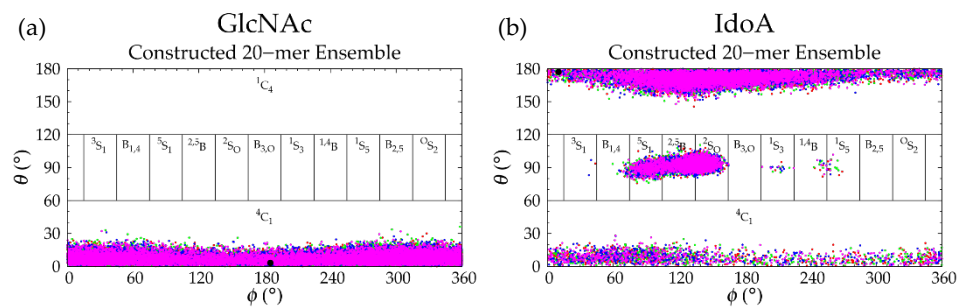


Figure S34. Cremer–Pople data for (a) GlcNAc and (b) IdoA in the constructed non-sulfated heparan 20-mer ensemble; each of the 4 runs is represented by different color and the force-field geometry is represented by a single large black dot; each run contains 10,000 parameter sets.

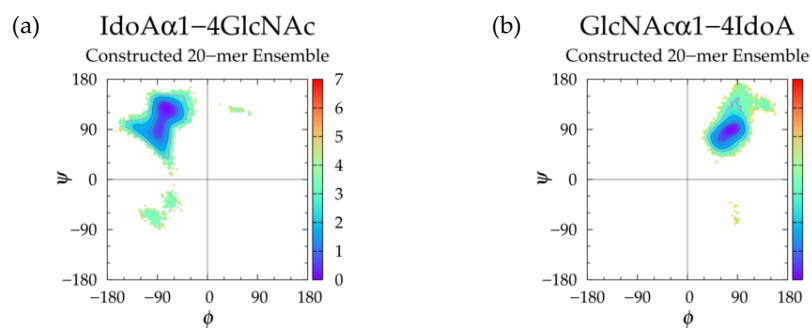


Figure S35. $\Delta G(\phi, \psi)$ in the constructed non-sulfated heparan 20-mer ensemble for aggregated (a) IdoA α 1-4GlcNAc and (b) GlcNAc α 1-4IdoA glycosidic linkage data; contour lines every 1 kcal/mol.

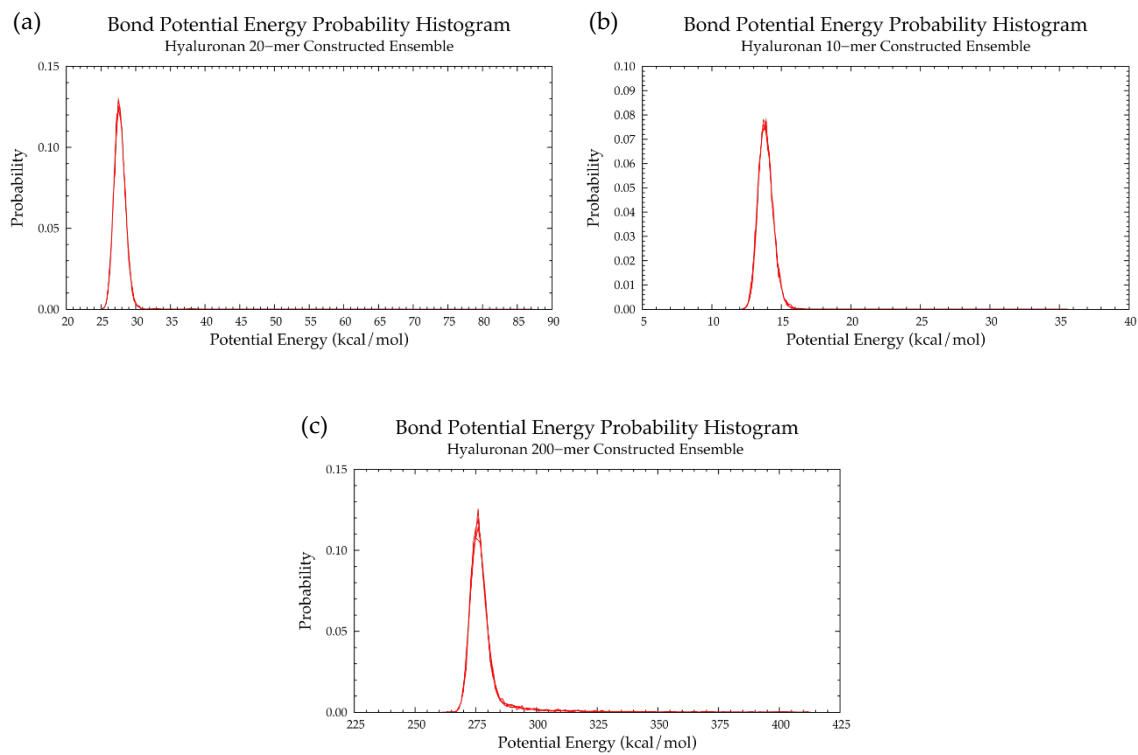


Figure S36. Bond potential energy probability distributions from constructed ensembles of hyaluronan (a) 20-mer (cutoff = 128.5 kcal/mol), (b) 10-mer (cutoff = 112.8 kcal/mol), and (c) 200-mer (cutoff = 412.2 kcal/mol).

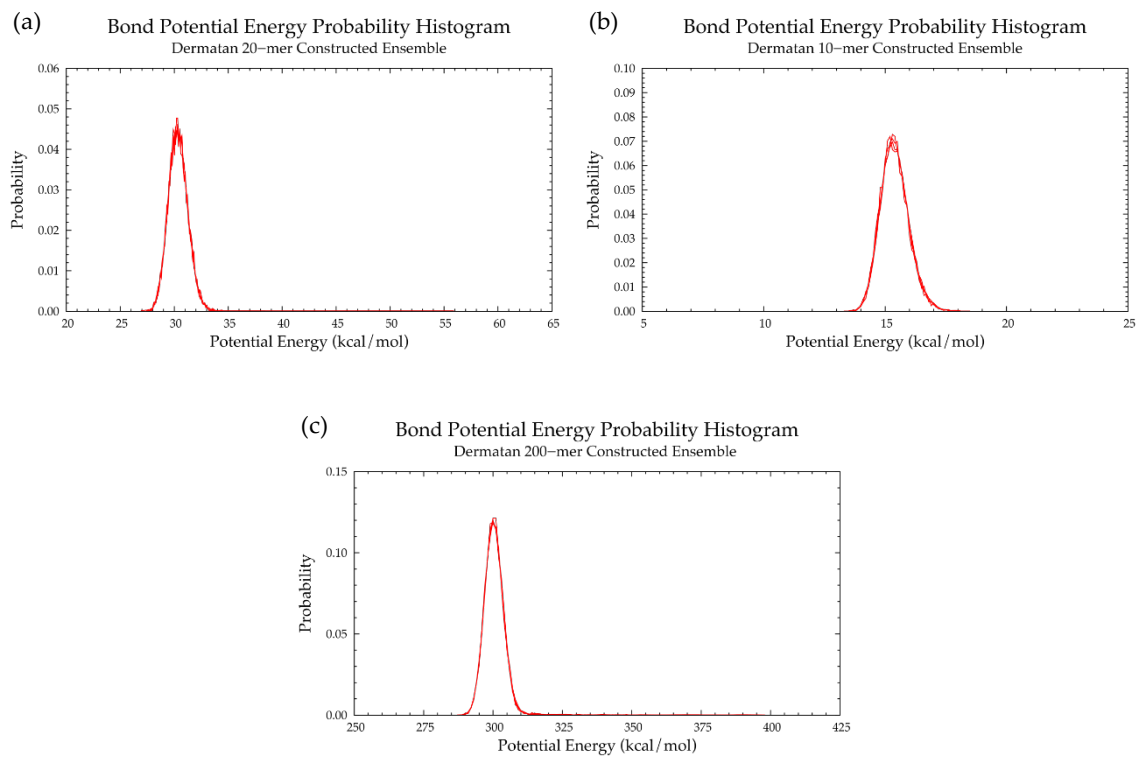


Figure S37. Bond potential energy probability distributions from constructed ensembles of non-sulfated dermatan (a) 20-mer (cutoff = 131.9 kcal/mol), (b) 10-mer (cutoff = 117.5 kcal/mol), and (c) 200-mer (cutoff = 397.7 kcal/mol).

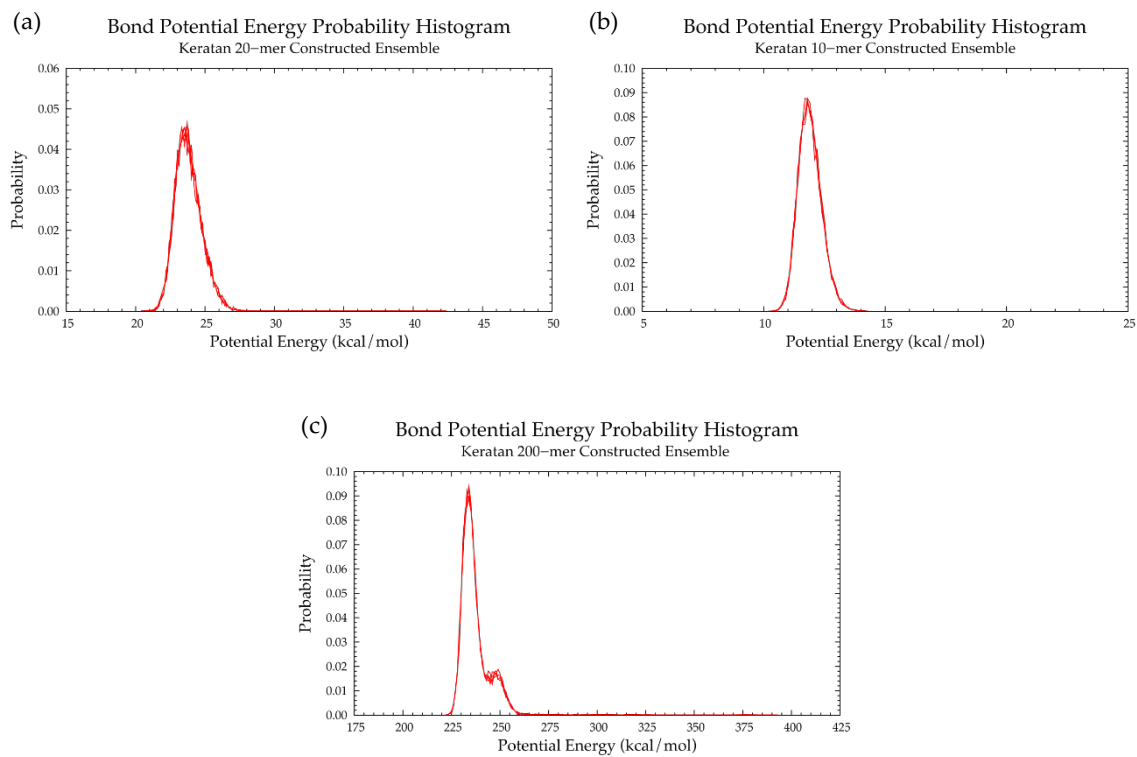


Figure S38. Bond potential energy probability distributions from constructed ensembles of non-sulfated keratan (a) 20-mer (cutoff = 126.3 kcal/mol), (b) 10-mer (cutoff = 111.5 kcal/mol), and (c) 200-mer (cutoff = 397.3 kcal/mol).

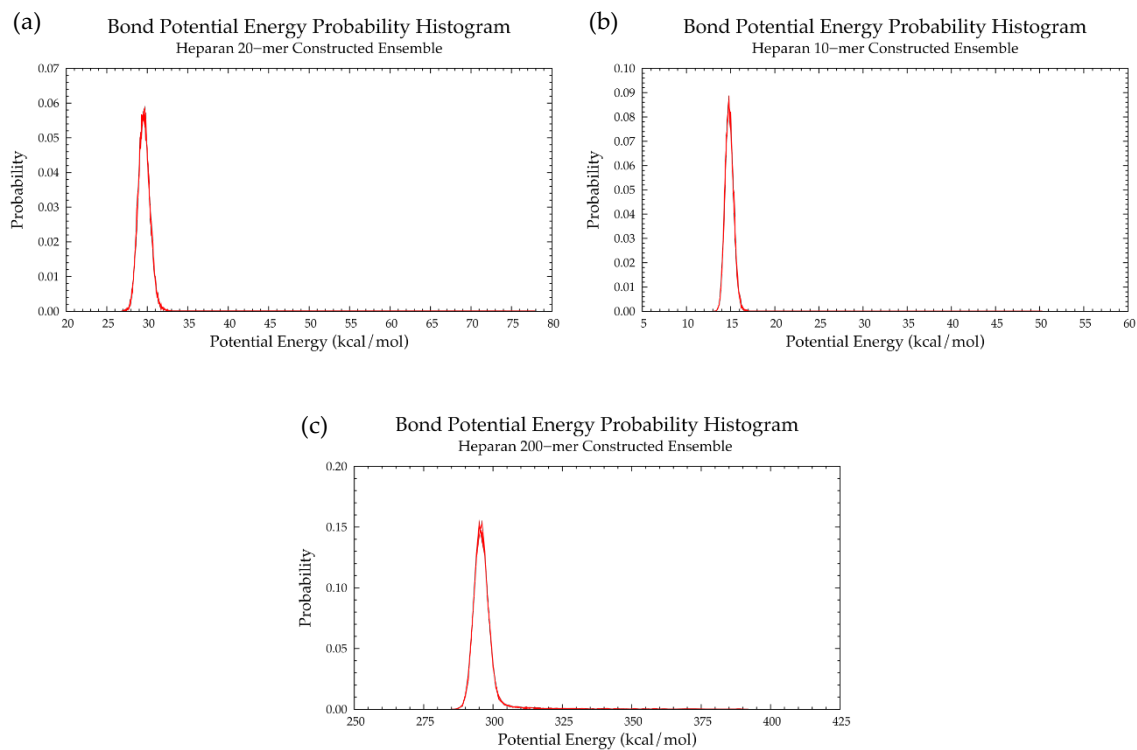


Figure S39. Bond potential energy probability distributions from constructed ensembles of non-sulfated heparan (a) 20-mer (cutoff = 130.3 kcal/mol), (b) 10-mer (cutoff = 115.8 kcal/mol), and (c) 200-mer (cutoff = 391.5 kcal/mol).