Supplementary material

Tuning G-Quadruplex Structures with Lipids. Towards Designing Hybrid Scaffolds for Oligonucleotide Delivery

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(9*H*-fluoren-9-yl)methyl-(1-(((2*R*,3*R*)-1,3-dihydroxybutan-2-yl)amino)-6-octanamido-1-oxohexan-2-yl)carbamate



(9*H*-fluoren-9-yl)methyl-(1-(((2*R*,3*R*)-1,3-dihydroxybutan-2-yl)amino)-6-octanamido-1-oxo-6-tetradecanamidohexan-2-yl)carbamate



(9*H*-fluoren-9-yl)methyl-(1-(((2*R*,3*R*)-3-hydroxy-1-(trityloxy)butan-2-yl)amino)-6-octanamido-1-oxohexan-2-yl)carbamate



(9*H*-fluoren-9-yl)methyl-(1-(((2*R*,3*R*)-3-hydroxy-1-(trityloxy)butan-2-yl)amino)-6-tetradecanamidohexan-2-yl)carbamate



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Figure S1. Native 12% polyacrylamide gel electrophoresis (PAGE) analysis of antisense Gquadruplex constructs containing A. 3'-lipid-threoninol based modifications (**15-20** and **24**) and B. 5'-lipid moieties (**19-26**). Conditions: 1X PBS supplemented with 100 mM KCI. The gel was stained with SYBR-green. The gels have been reversed to visualize the singled-stranded oligonucleotide on the top (fast runners) and the spots corresponding to quadruplex at the bottom (slow runners).



В



26 25 24 23 22 21 20 19 Luc

Figure S2. ThT assay using a fixed concentration of G-quadruplex (69 nM). ThT dye was added at increasing concentrations and recording its fluorescence intensity after binding to G-constructs



Name	Sequence	Backbone	Modification (mod)	Ка	∑r²	∑Fexp ²	Σr ² /ΣFexp ²
9	TGGGGT	PO	unmod.	1.60E+6	64.37	69886.6	0.09%
10	TGGGGGGT	PO	unmod.	1.51E+6	53.78	51076	0.11%
11	TGGGGT_mod	PO	3'_Thr_C8 ^a	1.41E+6	55.2	39085.3	0.14%
13	TGGGGGGT_ <i>mod</i>	РО	3'_Thr_C8 ^a	1.40E+6	92.7	69169	0.13%
14	TGGGGGGT_ <i>mod</i>	РО	3'_Thr_C14 ^a	1.30E+6	72.6	49417.29	0.15%
15	<i>mod _Luc-</i> TGGGGT	PS/PO	5'_ <i>Luc</i> _Thr_C8 ^b	1.42E+6	262	1.31E+06	0.02%
16	<i>mod _Luc-</i> TGGGGT	PS/PO	5'_ <i>Luc</i> _Thr_C14 ^b	4.11E+6	318.7	1211540.5	0.03%
19	Luc-TGGGGT	PS/PO	unmod.	1.44E+6	985.1	6643506.3	0.01%

Table S1. Affinity constants of lipid oligonucleotide and LOCs by ThT fluorescence spectrometry post G-quadruplex formation

Luc sequence: d(5'-<u>CGTTTCCTTTGTTCTGGA</u>-3'); unmod: unmodified; PO: phosphodiester; PS: phosphorothioate (underlined); ^aOligonucleotide conjugates containing hydrophobic threoninol-based derivatives with distinct length of saturated alkyl chains (C8 or C14); ^bLOC containing the *Luc* sequence at the 5'-end of the G-rich sequence.

Figure S3. Cytotoxicity analysis on HEK293 cells of antisense G-quadruplex conjugates containing four G-tetrads and six G-tetrads (see Table 1). Four concentrations of antisense G-quadruplex conjugates ranging from 60 nM to 600 nM were used. All antisense conjugates were incubated up to 24 hours at 37 °C. Data were means ±SD of three independent experiments



Figure S4. Flow cytometry analysis involving the effect of G4_25 and conjugate 26 when transfecting HeLa cells at 60 nM. (A) First row: non-fluorescent labelled cell populations (Blank) (left), selected region R2 (center) and histogram of non-fluorescence cells (right); second row: forward scatter dot plot of G4_25 (left); forward scatter dot plot of conjugate 26 (center) and histogram (right) in the presence of HeLa cells at 60 nM. The Flowing Software 2.5.1 was used to measure the relationship between untreated cell and positive cell populations.



Figure S5. Flow cytometry analysis involving the effect of G4_25 and conjugate 26 when transfecting HeLa cells at 300 nM. (A) First row: non-fluorescent labelled cell populations (Blank) (left), selected region R2 (center) and histogram of non-fluorescence cells (right); second row: forward scatter dot plot of 26 (left); forward scatter dot plot of conjugate G4_25 (center) and histogram (right) in the presence of HeLa cells at 300 nM. The Flowing Software 2.5.1 was used to measure the relationship between untreated cell and positive cell populations.

