



Supplemental Tables and Figures

Table S1: Deletion strains sensitive (n=135) to polygodial ranked as 0 and 1. In bold are hypersensitive mutants (n=66). Gene functions and cellular locations were taken from the SGD database.

Biological process	Genes
Mitochondrial genome maintenance and normal morphology	<i>GET1, PCP1, YME2</i>
Mitochondria electron transport chain	<i>QCR10</i>
Mitochondria ATPase	<i>ATP2</i>
Mitochondrial protein maturation	<i>ISA1</i>
Mitochondrial import, folding, maturation, degradation	<i>YME1</i>
Mitochondrial TCA cycle	<i>KGD2</i>
Mitochondrial NADP biosynthesis	<i>POS5</i>
Mitochondrial riboflavin biosynthesis	<i>RIB4</i>
Peroxisome protein deubiquitination	<i>UBP15</i>
Vacuolar acidification, V-ATPase	<i>VMA1, VMA3, VMA4, VMA7, VMA8, VMA13, VMA16, VPH2</i>
Vacuolar degradation autophagy	<i>ATG8, ATG12, ATG10, ATG13</i>
Vesicular trafficking to vacuole, vacuolar protein sorting	<i>VAM6, VAM3, GYP1, VAM7, VPS16, VPS54, VPS61, VPS63, VPS65</i>
Golgi P-type ATPase	<i>PMR1</i>
ER Golgi endosome trafficking	<i>CHS5, RGP1, YPT6, VPS35, VPS52, VPS54, SEC22, SEC28, YSP1</i>
ER protein targeting to membrane, translocation	<i>SEC66, SGT2, GET1, GET4</i>
ER Golgi protein N-linked glycosylation	<i>OST4, OCH1</i>
ER ergosterol biosynthesis	<i>ERG3, ERG2, ERG28</i>
ER sterol transport	<i>SIP3</i>
ER lipid droplet formation and triacylglyceride homeostasis	<i>VPS66/LOA1</i>
ER glycosylphosphatidylinositol biosynthesis	<i>LAS21</i>
ER-associated degradation (ERAD)	<i>SHP1</i>
Nuclear envelope phospholipid biosynthesis, lipid droplets	<i>NEM1</i>
Chromatin organization, structure, remodelling	<i>EAF1, ULS1, RTF1, ARP5</i>
Plasma membrane transport	<i>FUR4, TRK1</i>
Cell wall synthesis	<i>SMI1, FKS1</i>
Bud site selection & axial budding	<i>BUD25</i>
Cytoskeleton organization and cellular morphogenesis	<i>BEM2</i>
Actin stabilization	<i>TPM1</i>
Transcription regulation	<i>SWI3, TAF14, SSN3, GAL11, CSE2, SIP3, BUR2, GON7, ROX3, SRB2, SNF6, SNF12, SRB5, RTF1, ADA2, UME6, RPN4, MED2, TUP1, RPA14</i>

Mitosis, meiosis	<i>DCC1, CTF4, CNM67, VPS54, HTL1, CIK1, MRE11</i>
mRNA export from nucleus	<i>SAC3, NPL3</i>
Nuclear protein import	<i>MOG1</i>
DNA repair	<i>FYV6</i>
Amino acid synthesis	<i>TYR1, CPA1, ADE3, PRO1, PRO2, SER1, SER2, TRP1, TRP2, TRP5, CPR2, ARG2</i>
Ribosomal protein; translation and protein synthesis	<i>RPL13B, MRPL23, TIF3, CPA1 <i>u</i>ORF</i>
Ribosome biogenesis	<i>SAC3, BMT5</i>
Signal transduction	<i>TPK1, IRA2, PHO85, CLA4, NBP2, STE11</i>
Glycolysis	<i>PFK2, ADH1</i>
Glutathione biosynthesis, binding	<i>GSH1, URE2</i>
Cyanamide metabolism	<i>DDI3</i>
Trehalose synthesis	<i>TPS2</i>
Nucleotide metabolism	<i>ADE3, ADK1</i>
Intracellular copper transport	<i>CCS1</i>
Mating	<i>SCP160</i>
Unknown function	<i>ECM33, BOP2, JJJ2, YGL024W, YML012C-A, YER091C-A, YDR433W, YDR157W, YMR102C, YOR331C, YNL296W</i>

Table S2: Mutants resistant to polygodial ranked as 5 and 6 (n=233). In bold are hyper-resistant mutants (n=47). Gene functions and cellular locations were taken from the SGD database.

Biological process	Genes
Mitochondrial transcription and translation	<i>PET494, MSK1, MEF1, MEF2, MTF2, MRM2, MRPL38, MRPL35, SLS1, TUF1, MSE1, RSM27, MRX5</i>
Mitochondrial genome maintenance, fusion, fission and normal morphology	<i>MCP2, CCE1, RIM1, ACO1, MGM1, DNM1, UGO1</i>
Mitochondria ATPase	<i>ATP15, ATP17, AFG3</i>
Mitochondrial inner membrane electron transport chain	<i>QCR9</i>
Mitochondrial protein folding	<i>CPR3, MDJ1</i>
Mitochondrial TCA cycle	<i>FUM1, PDX1, CIT1, SDH5, ACO1, ACO2</i>
Mitochondrial amino acid biosynthesis	<i>GDH3, ALT1, THI4, AAT1</i>
Mitochondrial transporter activity	<i>MDL1, HOT13, FLX1, MRS2</i>
Mitochondrial lipid homeostasis	<i>UPS2, MCP2</i>
Mitochondrial unknown function	<i>MRP8, FMP33, FMP48, FMP49</i>
Peroxisome biogenesis	<i>PEX30</i>
Vacuolar protein sorting, MVB, ESCRT, CORVET	<i>SNA3, MIL1, CPS1, YCK3, VPS8, VPS9, VPS21, VPS36, VPS60, VPS25, VPS27, MVB12, SSH4, LST4, ENV7</i>
Vacuolar transport	<i>AVT3, BPT1, CCC1</i>
Vacuolar carboxypeptidases	<i>ATG42, CPS1</i>
Golgi glycosylation	<i>MNN4</i>
ER Golgi transport	<i>COY1</i>

ER enzymes and transport	<i>ICE2, ERV2, YBR137W</i>
ER cargo exit	<i>TED1</i>
ER lipid homeostasis	<i>OSH2, APQ12</i>
ER inheritance	<i>SHE3</i>
Actin cytoskeleton reorganization, microtubules and endocytosis	<i>RVS167, RVS161, SLA1, END3, ACF2, SRV2, GVP36, TPM2, PAC11, CIN1</i>
Plasma membrane transport	<i>QDR1, AQR1, HXT10, ALR2, FRE7, BOR1, AMF1,</i>
Cell wall	<i>FIT3, KDX1, PRY3, TOS1</i>
Bud site selection & axial budding	<i>BUD2, BUD20, BUD21, BUD26, BUD28, AXL2</i>
Mating	<i>FUS3, KDX1, PRY3</i>
Mitosis and Meiosis	<i>HOP1, CLG1, IRC8, RMR1, HOS4, REC8, SMA2, GMC1</i>
Nuclear protein, protein quality control	<i>CMR1</i>
DNA repair and excision	<i>RAD7, RAD16, EAF7, MMS4, REV7, UBC13, YNK1</i>
Transcription regulators	<i>MSS11, TEA1, RFX1, MRK1, HAP4, CSI1, URC2, SPT7, HAA1, EAF7, HAP2, NOT3, HAP5, YAP7, TYE7, ACM1, HOS2, LAG2</i>
RNA processing	<i>NNF2, MUD1, CAF40, PES4</i>
Ribosomal protein; translation and protein synthesis	<i>RPP2A, RPL24B, RPL37A, RPS28B, RPL29, RPL24A, RPL23A, RPL23B, RPL16A, RPL41B, RPL13A, RPS16B, RPS17B, RPS16A, RPL27B, RPL19A, RPS25A, RPL1B, RPS30A, RPS1B</i>
Ribosome biogenesis	<i>SYO1, NOP12, GVP36</i>
Signal transduction, GSE/EGO complex	<i>MDS3, GTR1, GTR2, LTV1, PIB2, CKA2, RCN1, CNB1, SIP5, GIS4</i>
Ubiquitination & proteasome complex activity	<i>UBI4, UBR2, UBC13, PRE9, MMS2, MUB1, NPL4, UBR1, HUL5, ELA1</i>
Nucleotide biosynthesis	<i>YNK1, URA4, AMD1</i>
Cytoplasmic L-methionine salvage	<i>UTR4</i>
Glycogen biosynthesis, glycolysis and gluconeogenesis	<i>TDH3, GSY2</i>
Glycerol metabolism	<i>GPD2</i>
NADH-dependent aldehyde reductase	<i>YNL134C</i>
NAD biosynthesis	<i>YEF1</i>
Undefined/ unclear/unknown	<i>IRC19, OCA4, OCA5, GDS1, FHN1, STB6, NQM1, PBP4, FMP33, FMP48, BSC4, CMR3, CMG1, SDD2, YPR078C, YOR333C, YOL079W, YOR309C, YNR005C, YML084W, YMR141C, YML083C, YKL147W, YLR407W, YLR169W, YLR125W, YLR184W, YLL047W, YKL075C, YJR056C, YIL161W, YJL028W, YIL141W, YGR219W, YGR137W, YGR122C-A, YFL032W, YFL015C, YER038W-A, YDR442W, YBR013C, YDL094C, YDL062W, YBR287W, YML095C-A, YBR144C, YBR138C, YJL017W, YJL206C-A, YBR096W, YGR153W</i>

Figure S1: An example of a read-out from the chemogenomic screen. The same mutant strains from a 96-well plate were spotted on these three agar plates. The top agar plate contains DMSO (diluent of polygodial) and is a control plate. The middle agar plate contains 2 $\mu\text{g/mL}$ polygodial (to detect hypersensitive strains) while the bottom plate contains 6 $\mu\text{g/mL}$ polygodial (to detect resistant strains). Ranking was based on the growth of the mutants relative to the WT BY4742. Lower ranks (0-2) represent poor growth, 3 represents WT and more growth than WT ranks from 4-6.

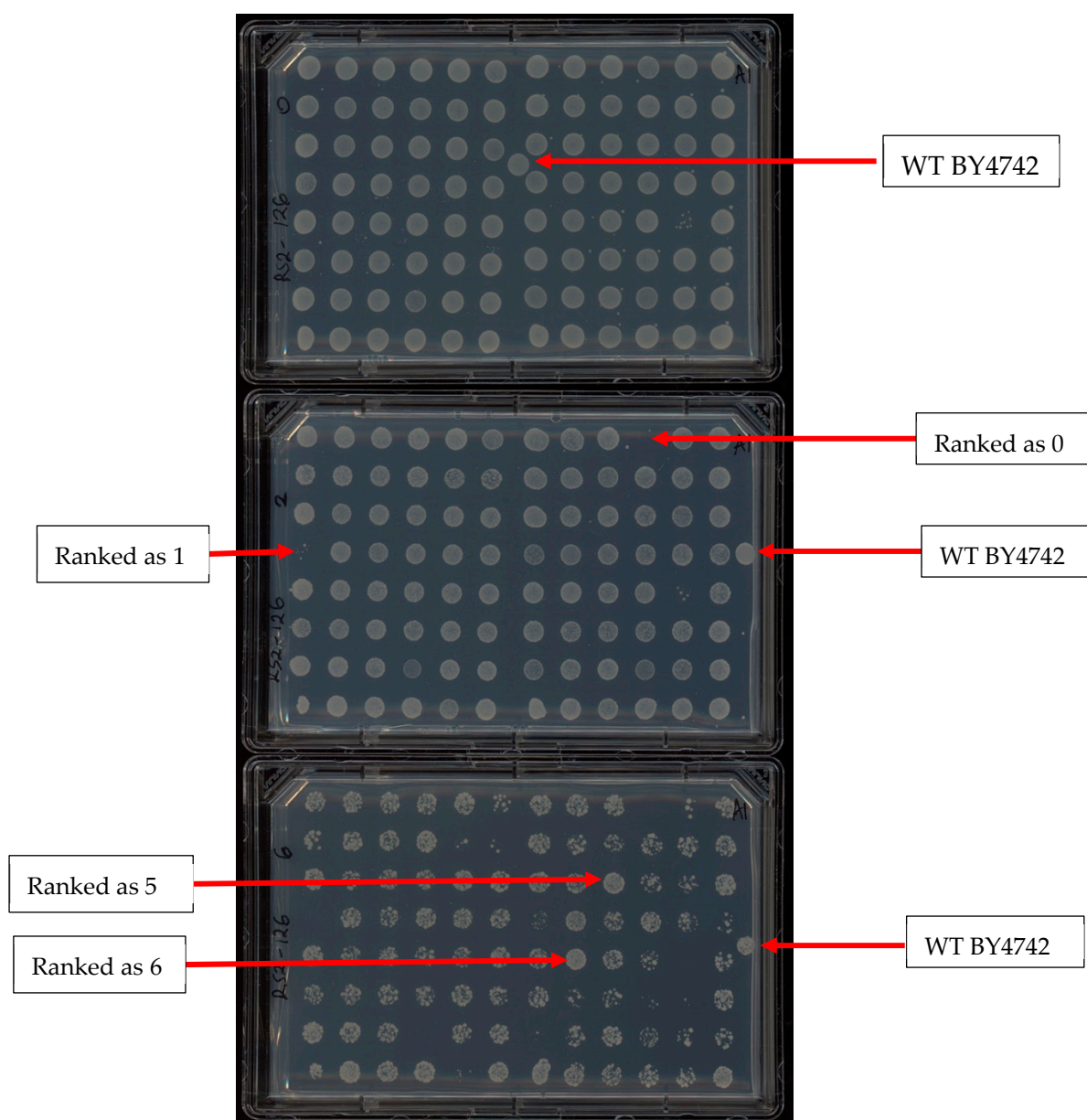


Figure S2: A clustering visualization of all polygodial sensitive strains using the STRING database. The colored circular nodes represent proteins, while the edges joining the nodes represent interactions (protein-protein), either established known interactions or predicted interactions.

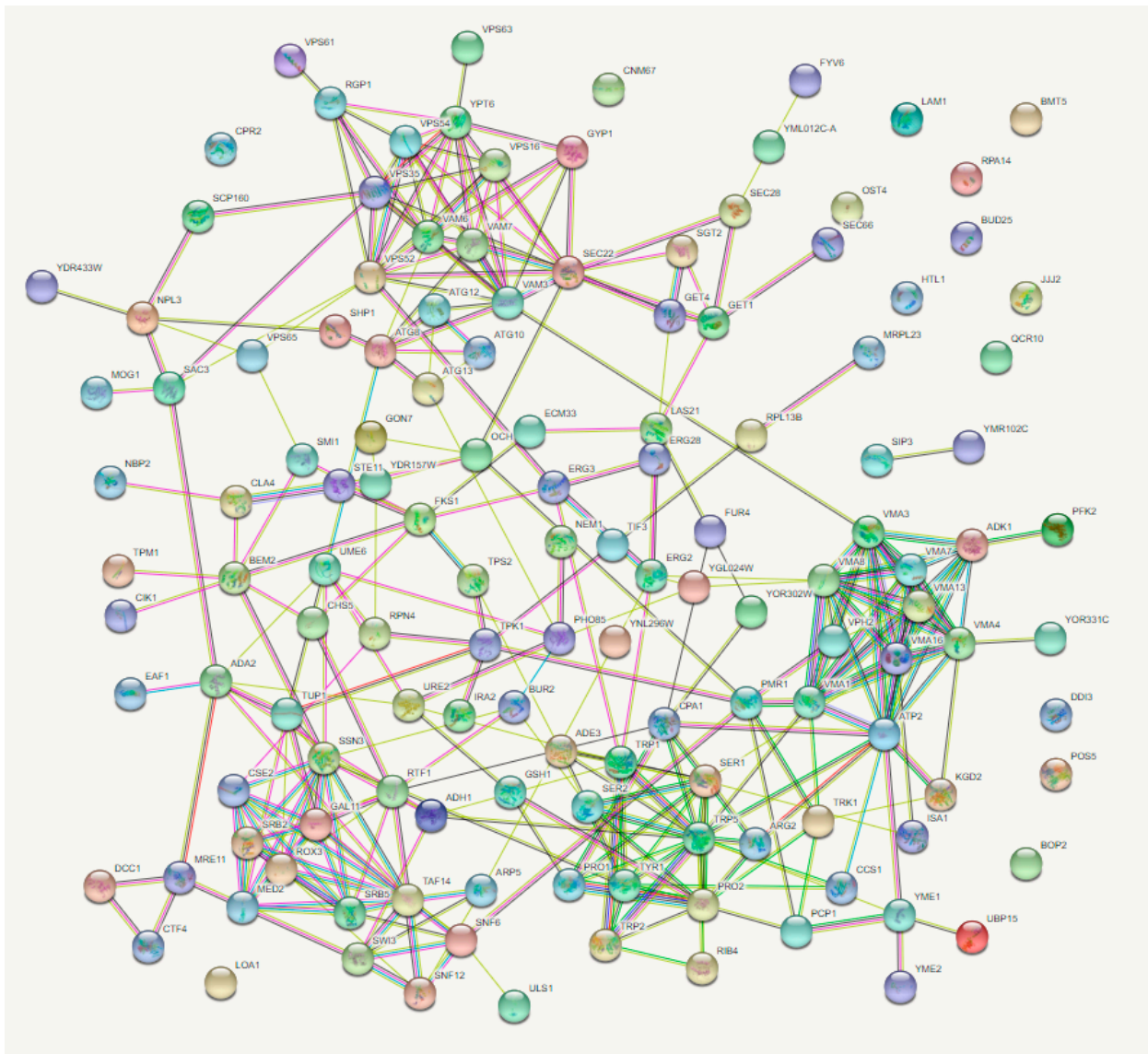


Figure S3: A clustering visualization of all polygodial resistant strains using the STRING database. The colored circular nodes represent proteins, while the edges joining the nodes represent interactions (protein-protein), either established known interactions or predicted interactions.

