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Abstract: The discovery of a large variety of functions of vitamin D_3 and its metabolites has led to the design and synthesis of a vast amount of vitamin D_3 analogues in order to increase the potency and reduce toxicity. The introduction of highly electronegative fluorine atom(s) into vitamin D_3 skeletons alters their physical and chemical properties. To date, many fluorinated vitamin D_3 analogues have been designed and synthesized. This review summarizes the molecular structures of fluoro-containing vitamin D_3 analogues and their synthetic methodologies.

Keywords: synthesis; fluorine; vitamin D₃; metabolite; A-ring; CD-ring; side-chain

1. Introduction

Fluorine is one of the halogens, known as a small and the most electronegative element. Fluorine substitution offers a variety of advantages, such as changing the pKa and dipole moment of the molecule, improving the chemical or metabolic stability, and enhancing the binding affinity to the target protein. Furthermore, it has a small atomic radius, similar to that of a hydrogen atom. Due to their unique properties, fluorine atoms have been incorporated into many drugs, drug candidates, and agricultural chemicals [1–12]. The contribution of fluorine to drug development and medicinal chemistry, and the life science as well as material science fields, is widely recognized around the world [13–17]. Many scientists have been engaged in the practical synthesis of organofluorine compounds, including fluorinated vitamin D_3 analogues. Vitamin D_3 (VD₃) is a fat-soluble vitamin whose biological function depends on metabolic activation by CYP enzymes [18]. Bioactivation of VD_3 requires sequential oxidation steps at C-25 and C-1 catalyzed by vitamin D 25-hydroxylase (CYP2R1) and 25-hydroxyvitamin D_3 -1 α -hydroxylase (CYP27B1), respectively. The resulting B-secosterol, 1α , 25-dihydroxyvitamin D₃ [1α , 25(OH)₂D₃ (1)], is the fully active and hormonal form of VD₃. Moreover, 1α ,25(OH)₂D₃ (1) and 25-hydroxyvitamin D₃ [25(OH)D₃ (2)] are degraded via hydroxylation at C23 or C24 catalyzed by $1\alpha_2$ -dihydroxyvitamin D₃-24-hydroxylase (CYP24A1) [19]. C23 hydroxylation and subsequent three-step oxidation lead to vitamin D_3 -26,23-lactone (3). On the other hand, C24 hydroxylation and subsequent five-step oxidation lead to calcitroic acid (4) (Scheme 1) [19,20].

For slowing or preventing the biological degradation of the VD₃ side chain, replacing C-H with C-F bond(s) at appropriate positions should prolong their half-life in vivo, since a C-F bond is stronger than a C-H bond chemically. The introduction of fluorine atoms into VD₃ analogues can also alter electron distribution, which can confer lower pKa at the hydroxy group(s), change the dipole moment, and influence the conformation because of their marked electron-withdrawing properties. Because of these unique properties, both academic institutions and industries have designed and synthesized numerous fluorinated VD₃ analogues, similarly to other bioactive compounds with fluorine atoms. Most of them show fluorination at the main metabolic site(s) and/or neighboring hydroxy group of the VD₃ molecule, namely either an A-ring or a side-chain moiety or both. This review introduces fluorinated VD₃ analogues and their synthetic methodologies, including some basic biological activities.



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Scheme 1. Deactivation metabolic pathways of $25(OH)D_3$ (2) and 1α , $25(OH)_2D_3$ (1).

2. A-Ring Fluorinated VD₃ Analogues

On the VD₃ A-ring, 1α -hydroxylation is the final and essential step to produce the hormonal form of VD₃ from 25(OH)D₃ (**2**), and 1α ,25(OH)₂D₃ (**1**) exerts a range of physiological activities by binding to the vitamin D receptor (VDR). The A-ring moiety is anchored in the VDR ligand-binding pocket through four hydrogen-bonding interactions, i.e., the 1α -hydroxy group to Ser233 and Arg270, and the 3β-hydroxy group to Tyr143 and Ser274 [21]. As expected, based on the importance of the A-ring moiety, many A-ring fluorinated VD₃ analogues have been synthesized and evaluated regarding their biological activities.

2.1. 1-Fluorinated VD₃ Analogues

DeLuca and coworkers described the first synthesis of 1-fluoro-VD₃ for the purpose of studying the possibility of using it as a kind of VD₃ antagonist against 1 α -hydroxylase in 1979 [22]. They prepared 1 α -hydroxyvitamin D₃-3-acetate by the selective acetylation of 1 α -hydroxyvitamin D₃ [1 α (OH)D₃ (5)] and used it as a starting material. The fluorination step was achieved by *N*,*N*-(diethylamino)sulfur trifluoride (DAST) to afford 1-fluorovitamin D₃ (6) (Scheme 2). The authors did not assign its C1 configuration in the report. The biological evaluation revealed that 6 demonstrated a relative preference for stimulating bone calcium mobilization with respect to intestinal calcium transport after metabolism in vivo, and 6 was a weak agonist that could not be used as an anti-vitamin D agent.

Next, DeLuca et al. designed and synthesized 1α ,25-difluorovitamin D₃ (7) in 1981 [23]. As shown in Scheme 3, it was synthesized from either 1α ,25(OH)₂D₃ 3-acetate (8) or 1α ,25(OH)₂-3,5-cyclovitamin D₃ (9) using DAST as a fluorination reagent. The authors explained that the stereochemistry at C1 was 1α in the report, but they revised it to 1β in 1984 [24].



Scheme 2. DeLuca's direct fluorination approach to 1-fluorovitamin D_3 (6).



Scheme 3. Direct C1 and C25 fluorination approach to $1\alpha_{2}$ -difluorovitamin D₃ (7).

The authors evaluated the biological properties of 1α ,25-difluorovitamin D₃ (7) and demonstrated that it had essentially no vitamin D activity. These results strongly supported the idea that C1 and C25 hydroxylation are essential aspects of vitamin D function. Initially, since both the C1 and C25 positions of VD₃ were blocked with fluorine atoms, it might have shown the anti-vitamin D activity of 25-hydroxylation of VD₃ in vivo. However, 7 did not exhibit the expected inhibitory activity against the 25-hydroxylation of VD₃.

In 1984, 1 α -fluorovitamin D₃ (**10**) and 1 α -fluoro-25(OH)D₃ (**11**) were synthesized utilizing a steroid A-ring epoxide as the starting material in order to confirm the stereochemistry at C1 (Scheme 4) [24,25].



Scheme 4. Synthesis of 1α -fluoro-VD₃ (10) and 1α -fluoro-25(OH)D₃ (11) from sterols.

A comparison of the spectral data of the report [24] and previous ones [22,23] revealed that the reported 1α -fluorovitamin D₃ in 1979 [22] and 1α ,25-difluorovitamin D₃ in 1981 [23] were, in fact, 1 β -fluorovitamin D₃ and 1 β ,25-difluorovitamin D₃, respectively.

On the other hand, although 1α -fluoro-25(OH)D₃ (11) showed no stimulation of intestinal calcium transport or bone calcium mobilization activities at a dosage level of 1.3 µg, its binding affinity to chick intestine VDR was 30 times greater than that of 25(OH)D₃.

A convergent synthetic route to 1-fluoro-VD₃ analogues was described by Uskoković and coworkers using an A-ring key fragment, a 1 α -fluorinated A-ring precursor (**12**), which was prepared from (*S*)-(+)-carvone (**13**), in 1990 (17 steps in 4%) (Scheme 5) [26], as well as an A ring from VD₃ in 1991 (12 steps) (Scheme 6) [27]. A lithium anion of the A-ring phosphine oxide (**12**) underwent a Wittig–Horner coupling reaction with the 8-keto-CD-ring (**14**) to afford the desired 1 α -fluoro-25(OH)D₃ (**11**).



Scheme 5. Uskoković's approach to 1α-fluoro-25(OH)D₃ (11) using the Wittig–Horner reaction.



Scheme 6. Alternative convergent synthetic approach to 1α -fluoro-25(OH)D₃ (11) using the A-ring moiety (12), available from the A-ring of VD₃.

Later, Uskoković's group utilized the 1α -fluoro-A-ring (12) to synthesize six 1α -fluoro-VD₃ analogues with two different side chains at C20 (Gemini analogues). They prepared six 8-keto-CD-rings (16–21) starting from the methyl ester (15) and coupled it with 12 under basic conditions (Scheme 7) [28]. The anticancer activity of these compounds was tested, but 1α -fluorination did not effectively promote the activity.



Scheme 7. Various 1α -fluoro-VD₃ analogues with six different double side chains were synthesized using the Wittig–Horner olefination at the coupling stage with the 1α -fluoro-A-ring part (**12**).

In 2019, Uesugi and colleagues published the first synthesis of 1-fluoro-19-norVD₃ analogues [29]. They constructed 1-hydroxy-19-norvitamin D₃ structures with a modified Julia olefination method [30], and the direct deoxyfluorination of C1 yielded the corresponding 1-fluorinated analogues (**22,23**). In this reaction sequence, shown in Scheme 8, they also obtained 3-fluoro-19-norVD₃ analogues (**24,25**). Some of these analogues were poor VDR binders but showed potent sterol regulatory element-binding protein (SREBP) inhibitory activity via inducing SREBP cleavage-activating protein (SCAP) degradation [29].



Scheme 8. Synthesis of 1- and 3-fluoro-19-norvitamin D₃ analogues (22–25) using direct deoxyfluorination reactions.

2.2. 2-Fluorinated VD₃ Analogues

Several 2-fluorinated VD₃ analogues have been reported to date, because fluorine substitution at this position can change the A-ring conformation and pKa value of the neighboring 1 α - and 3 β -hydroxy groups. The first synthesis of 2 β -fluoro-VD₃ was reported by Ikekawa and coworkers in 1980, in which nucleophilic fluorination using KHF₂ to 1,2 α -epoxycholesterol gave 2 β -fluoro-1 α (OH)D₃ (26), (Scheme 9) [31]. It was noted that the biological activity of 26 was increased [32].



Scheme 9. Ikekawa's approach to 2β -fluoro- $1\alpha(OH)D_3$ (**26**) via 2β -fluorination of A-ring epoxide using a nucleophilic fluorination reaction.

Next, 2α -fluorovitamin D₃ (27) was also synthesized, and its biological activity was tested in 1986 [32]. Electrophilic 2α -fluorination of 3,6 β -diacetoxycholest-2-ene (28) using CsSO₄F gave 2α -fluoroketone (29). To construct the B-secosteroidal structure, conventional photochemical conversion and subsequent thermal isomerization were applied (Scheme 10) [32]. The biological effects of 27 on intestinal calcium transport and bone calcium mobilization as well as the serum calcium concentration at a dosage level of 500 ng for rats were measured, and the activities were found to be essentially equivalent to those of VD₃ itself.



Scheme 10. Introduction of the 2α -fluoro group to VD₃ using an electrophilic fluorination reaction.

The synthesis and biological activities of 2β -fluoro- 1α ,25-dihydroxyvitamin D₃ (**30**) were reported by Scheddin et al. in 1998 [33]. The synthetic route was similar to Ikekawa's [31], as shown in Scheme 11, and the biological evaluation revealed that the synthetic 2β -fluoro- 1α ,25-(OH)₂D₃ (**30**) exhibited greater potency in vitro, for example, six-times higher affinity for VDR, nearly identical affinity for the vitamin D-binding protein (DBP), and 90-times higher antiproliferative activity toward C3H10T1/2 cells under serum-containing conditions, as well as five-times greater adipogenesis inhibitory activity than the natural hormone 1α ,25(OH)₂D₃ (**1**).



Scheme 11. Synthesis of 2β -fluoro- 1α , 25-(OH)₂D₃ (30).

The catalytic asymmetric stereoselective synthesis of the A-ring precursor of the 19-nor type 2α -fluorovitamin D₃ analogue (**31**) and its synthesis were reported by Mikami et al. [**34**,**35**]. The regio- and stereo-selective 2α -fluorination was achieved via a ring opening reaction of chiral epoxide (**32**) mediated by HfF₄/Bu₄NH₂F₃, the asymmetric catalytic carbonyl-ene cyclization was used to construct the 6-membered A-ring precursor, and the subsequent coupling reaction with the CD ring afforded 2α -fluoro-19-normaxacalcitol (**31**) (Scheme 12). This 2α -fluoro-22-oxa-19-nor analogue (**31**) had very low DBP-binding affinity but four-times stronger VDR-binding potency than its 22-oxa-19-nor counterpart and also showed significant transactivation activity [**34**]. It was also shown that **31** was highly effective in inhibiting metastatic tumor growth in vivo without toxicity in terms of hypercalcemia and weight loss [**35**].



Scheme 12. Mikami's group synthesized 2α -fluoro-19-normaxacalcitol (**31**) using a convergent coupling method between the 2α -fluoro-A-ring part, prepared from regio- and stereoselective fluorination to **32**, followed by asymmetric catalytic carbonyl-ene cyclization, and the appropriate 8-keto-CD-ring.

Posner's group showed the first example of synthesis of the 2,2-difluorovitamin D_3 analogue in 2002 [36]. They synthesized the racemic 2,2-difluoro substituted A-ring phosphine oxide (33) from trifluoroethanol (7% in 13 steps). A coupling reaction of 33 with the 8-keto-CD-ring and subsequent deprotection yielded the 2,2-difluoro-1,25-dihydroxyvitamin D_3 analogues in the ratio of 5:1 (1 α ,3 β (34):1 β ,3 α) (Scheme 13). The diastereomers were

separated with reversed-phase HPLC to afford the target 2,2-difluoro- 1α ,25(OH)₂D₃ (**3**). Biological evaluation revealed that **34** exhibited antiproliferative activity similar to that of 1α ,25(OH)₂D₃ (**1**) and was 2-3 times more transcriptionally active than **1** in rat osteosarcoma cells, even though the human VDR-binding affinity was 9.6% relative to that of **1**. Compound **34** showed strong calcemic activity in vivo.



Scheme 13. Posner's synthetic route to 2,2-difluoro- 1α ,25-dihydroxyvitamin D₃ (**34**) from 2,2-difluoro-A-ring (**33**) via an inverse-electron-demand Diels–Alder reaction between a pyrone diene and difluorovinyl ether.

2.3. 3-Fluorinated VD₃ Analogues

The C3 position of VD₃ has a β -hydroxy group, and substitution of the hydroxy with a fluorine atom is of fundamental interest. There are several reports of replacing the C3-hydroxy group with a fluorine atom in order to demonstrate the expected positive effects on biological activity.

The synthesis of 3β -fluoro-3-deoxyvitamin D₃ (**35**) from 3β -fluorocholesta-5,7-diene (**36**) via photochemical transformation followed by thermal isomerization was described by Segal et al. in 1976 [37]. It was found that 3β -fluoro-3-deoxyvitamin D₃ (**35**) had an antirachitic effect analogous to VD₃. On the other hand, in 1978, Mazur and coworkers designed and synthesized 3β -fluoro-3-deoxy-1 α -hydroxyvitamin D₃ (**37**) to elicit VD₃ activity. The 3β -fluoro group was constructed from (6*R*)-hydroxy-3,5-cyclovitamin D₃ (**38**) by treatment with HF (Scheme 14) [38]. The biological activities of the newly synthesized analogue were tested, and the fluoro-analogue (**37**) could actively induce the formation of a calcium-binding protein and stimulate intestinal calcium absorption in rachitic chicks.



Scheme 14. Segal's and Mazur's 3β-fluoro-3-deoxyvitamin D₃ syntheses.

Later, in 1985, Kumar and coworkers reported the synthetic route to 3β -fluoro-3deoxyvitamin D₃ (**35**) starting from 7-dehydrocholesterol (**39**) via 3-deoxy-3-fluoro-7dehydrocholesterol (**36**) [39]. The B-ring protected PTAD (4-phenyl-1,2,4-triazoline-3,5dione) adduct (**40**) was reacted with DAST to yield a fluorinated product (**41**), and subsequent deprotection of **41** gave 3-deoxy-3-fluoro-7-dehydrocholesterol (**36**) (Scheme 15). To investigate the influence of the replacement of the C3-hydroxy group with fluorine, they compared the biological activities of VD₃, 3-deoxyvitamin D₃, and 3β -fluoro-3deoxyvitamin D₃ (**35**) and revealed that **35** was less active than VD₃ and more active than 3-deoxyvitamin D₃ in terms of intestinal calcium transport and bone calcium mobilization in vivo.



Scheme 15. Synthesis of 3β -fluoro-3-deoxyvitamin D₃ (35) starting from 7-dehydrocholesterol (39).

As mentioned in Section 2.1, Uesugi and colleagues reported the first synthesis of 3-fluoro-19-norVD₃ analogues and evaluated their SREBP inhibitory activity vs. VDR activity (see Scheme 8) [29].

2.4. 4-Fluorinated VD₃ Analogues

Yamada and coworkers reported 4,4-difluorovitamin D_3 (42) and 4,4-difluoro-1 α ,25(OH)₂ D_3 (43) in an A-ring conformational study in 1999, and these analogues were synthesized starting with enones (44,45), which were constructed from ergosterol, respectively [40]. The difluorination step was achieved by electrophilic fluorination under thermodynamic conditions (Scheme 16). The binding affinity of 4,4-difluoro-1 α ,25(OH)₂ D_3 (43) for VDR was only ca. 1% of that of the natural hormone, 1 α ,25(OH)₂ D_3 (1).



Scheme 16. Yamada's approach to 4,4-difluorovitamin D₃ (42,43) using a thermodynamic fluorination reaction.

3. Introduction of a Fluorine Atom into the Triene Part of VD₃

3.1. 19-Fluorinated VD₃ Analogues

In 1980, the first attempt to synthesize 19,19-difluorovitamin D_3 (46) starting from 19-oxocholesteryl acetate (47) via photoirradiation and thermal isomerization reactions using diene (48) failed, because the final thermal isomerization step by [1,7]-sigmatropic rearrangement did not proceed in the presence of the difluoromethyl group at the C10 position (Scheme 17) [41].



Scheme 17. Mazur's trial to synthesize 19,19-difluorovitamin D₃ (46).

Later, in 1996, Yamada and coworkers showed a novel synthetic route to the first (10*E*)and (10*Z*)-19-fluorovitamin D₃ (**50**,**51**) [42]. They prepared VD₃-SO₂ adducts (**49**) from VD₃, and fluorination at the C19 position was achieved by electrophilic fluorination using (PhSO₂)₂NF in the presence of LiHMDS (Scheme 18A). In 2000, the same group showed the regioselective introduction of a fluorine atom to the C19 position and synthesized both (10*E*)- and (10*Z*)-19-fluoro-1 α ,25(OH)₂D₃ (**52**,**53**) [43]. The binding affinity of (10*Z*)-19-fluoro-1 α ,25(OH)₂D₃ for VDR was ca. 10% compared with that of 1α ,25(OH)₂D₃ (**1**). The alternative synthetic route to **52** and **53** from a 10-oxo-19-norVD₃ derivative was also established by the same group in 2001 (Scheme 18B) [44].



Scheme 18. Yamada's approach to (10*E*)- and (10*Z*)-19-fluorovitamin D₃ (**50**,**51**) in (**A**) and (10*E*)- and (10*Z*)-19-fluoro- 1α ,25(OH)₂D₃ (**52**,**53**) analogues in (**A**,**B**).

3.2. 6- and 7-Fluorinated VD₃ Analogues

The synthesis of 6-fluorovitamin D_3 (54) was described by Dauben et al. in 1985, and the synthetic route started from 6-oxo-cholestanyl acetate (55) [45]. The key intermediate, 6-fluoro-7-dehydrocholesteryl acetate (56), was synthesized by allowing 55 to react with piperidinosulfur trifluoride in the presence of sulfuric acid. To construct the triene system, 6-fluoro-7-dehydrocholesterol was irradiated, followed by thermal [1,7]-sigmatropic hydrogen rearrangement, to give the desired 6-fluorovitamin D_3 (54) (Scheme 19). The obtained 6-fluorovitamin D_3 (54) was air-sensitive, and decomposition proceeded. The biological profile of the analogue was evaluated in vivo, revealing that 54 had no biological effect on either intestinal calcium absorption or bone calcium mobilization. However, it significantly inhibited both VD₃- and 1α ,25(OH)₂D₃-mediated intestinal calcium absorption through a direct interaction with VDR [46].



Scheme 19. Synthesis of 6-fluorovitamin D₃ (54) using piperidinosulfur trifluoride as a fluorination reagent.

Furthermore, 6-fluoro-1 α ,25-dihydroxy-19-norvitamin D₃ (57) and 7-fluoro-1 α ,25-dihydroxy-19-norvitamin D₃ (58) were synthesized by the Teijin research group in 2004 [47]. Takenouchi et al. prepared a C6-fluorinated A-ring (59) and C7-fluorinated CD-ring (60), respectively. A Ni-catalyzed cross-coupling reaction with each CD-ring- or A-ring-activated alkene counterpart was used to construct the diene structures of 57 and 58 (Scheme 20). These compounds possessed 10–70% VDR binding affinity of that of 1α ,25(OH)₂D₃ and potential activity to induce HL-60 cell differentiation.



Scheme 20. Efficient introduction of the C6- and C7-fluorovinyl unit to the A-ring or CD-ring followed by construction of the diene structures using a Ni-catalyzed cross-coupling reaction.

4. CD-Ring Fluorinated VD₃ Analogues: 11-Fluorinated VD₃ Analogues

To our knowledge, only one report has been published on the synthesis of CD-ring fluoro-VD₃ analogues. In 1994, De Clercq and coworkers designed and synthesized 11 α - and 11 β -fluorovitamin D₃ analogues (**61–64**) with the aim of inducing a conformational change from *s-trans* to *s-cis* at the C6-C7 single bond via expected hydrogen bond formation between the C11-F and C1 α -OH groups [48]. Enone (**65**), readily available from Grundmann's ketone, was used as a starting material. Epoxidation of **65**, followed by reductive opening with lithium dimethylcuprate, gave the C11 α -OH group was treated with *N*-(2-chloro-1,1,2-trifluoroethyl)diethylamine (FAR) to give C11 α -F and C11 β -F isomers as well as an elimination product at a ratio of 3:1:1 (Scheme 21). NMR spectra analyses of the 11-fluoro-1 α (OH)D₃ showed that all analogues had a large *J*₆₋₇ coupling constant, reflecting an almost exclusive *s-trans* extended geometry without intramolecular hydrogen bonding between C11-F and 1 α -OH.



Scheme 21. Stereoselective hydroxylation, coupling with the A-ring precursor, and subsequent fluorination at the C11 position.

5. Side-Chain Fluorinated VD₃ Analogues

The CYP24A1 pathway is well-known as the deactivation pathway of both $25(OH)D_3$ (2) and 1α , $25(OH)_2D_3$ (1) [19]. Varieties of side-chain-fluorinated VD₃ analogues have been actively synthesized because of the expected slower catabolism resulting from the presence of fluorine atoms at the oxidation site or adjacent area.

5.1. 22-Fluorinated VD₃ Analogues

In 1986, Kumar and coworkers described the synthesis of 22-fluorovitamin D_3 (67) starting from (22*S*)-cholest-5-ene-3 β ,22-diol (68) [49]. Selective protection of the C3 hydroxy group as an acetate, followed by fluorination at the C22 position by DAST, gave 22-fluorocholest-5-en-3 β -acetate (69). After forming the C5-7 diene unit in the B ring, photolysis and thermal isomerization yielded the target 22-fluorovitamin D_3 (67) without assigning C22 stereochemistry (Scheme 22). They tested the biological activities of the analogue in vitro and in vivo, referring to the potency of intestinal calcium transport, serum calcium level, calcium-binding protein induction, plasma vitamin D-binding protein (DBP), and VDR affinities, and concluded that the introduction of a fluorine atom to C22 resulted in the compound, with weak biological activities and poor binding to DBP compared with VD₃ itself.



Scheme 22. Synthesis of 22-fluorovitamin D₃ (67) from (22S)-22-hydroxycholesterol (68).

5.2. 23-Fluorinated VD₃ Analogues

As mentioned in the Introduction, the C23 position of VD₃ is one of the essential metabolic sites of CYP24A1; therefore, C23-fluorinated VD₃ analogues have been designed and synthesized based on the idea of blocking the oxidative position. Ikekawa and colleagues achieved the first synthesis of a C23-fluoro-VD₃ analogue, 23,23-difluoro-25(OH)D₃ (70), in 1984 [50]. For the synthesis of 70, the triene structure was constructed by applying the well-established route through 5,7-diene steroids, and the difluoro unit was introduced using DAST into a reactive α -ketoester (71) (Scheme 23).



Scheme 23. Synthesis of 23,23-difluoro-25(OH)D₃ (**70**) using α -ketoester (**71**) as a key intermediate, and the structure of 23,23-difluoro-1 α ,25(OH)₂D₃ (**72**) is also shown.

The same group continued to report 23,23-difluoro- 1α ,25(OH)₂D₃ (**72**) by the enzymatic 1 α -hydroxylation of **70** in 1985 [51]. Their biological activities were evaluated, and 23,23-difluoro-25(OH)D₃ (**70**) was 5-10 times less active than 25(OH)D₃ in stimulating intestinal calcium transport, bone calcium mobilization, mineralization of rachitic bone, etc., and 23,23-difluoro- 1α ,25(OH)₂D₃ (**72**) was one-seventh as active as 1α ,25(OH)₂D₃ in binding to VDR.

Ikeda and coworkers of the Sumitomo research group reported the synthesis of (23R)-23,26,26,26,27,27,27-heptafluoro-1 α ,25(OH)₂D₃ (73) and its 23*S* isomer (74) in 2000 [52]. The starting methyl ketone was available from VD₂, and a subsequent hexafluoroacetone (HFA) aldol reaction gave a 23-oxo derivative, which was reduced to 23*R*- and 23*S*-secondary alcohols that could be separated and subjected to deoxyfluorination using DAST to afford 73 and 74 (Scheme 24). Both analogues showed higher VDR-binding affinity and HL-60 cell differentiation activity than falecalcitriol.



Scheme 24. Synthesis of 23,26,26,26,27,27,27-heptafluoro- 1α ,25(OH)₂D₃ using DAST for the seventh fluorine introduction.

Later, in 2019, our group synthesized (23*R*)-23-fluoro-25(OH)D₃ (**75**) and its 23*S*-isomer (**76**) starting from the Inhoffen–Lythgoe diol via the key intermediate 23-hydroxy-CD- rings (**77,78**) (Scheme 25) [53]. The preliminary biological evaluation revealed that the 23*S*-isomer (**76**) showed higher resistance to CYP24A1 metabolism than its 23*R*-isomer (**75**).



Scheme 25. Stereoselective C23-fluorination of 23-hydroxy-CD-rings using PyFluor.

5.3. 24-Fluorinated VD₃ Analogues

Since oxidation of the hydroxy function at the C24 position catalyzed by CYP24A1 is one of the important pathways to deactivate both $25(OH)D_3$ and 1α , $25(OH)_2D_3$, developing practical methods to construct the C24-fluoro unit on the VD₃ skeleton has been pursued since 1979. The first synthesis of the 24,24-difluorovitamin D₃ analogue was reported independently by Takayama's group [54] and Kobayashi-Ikekawa's group [55]. Both synthetic routes involved the key intermediate (**79**), and the two groups synthesized the same analogue, 24,24-difluoro-25(OH)D₃ (**80**). For the construction of the 24,24-difluoro unit, Takayama and coworkers utilized the reaction of α -ketoester (**81**), which was derived from lithocholic acid, with DAST. On the other hand, Kobayashi et al. used the reaction of steroidal enol ether, derived from cholic acid, with difluorocarbene (Scheme 26).



Scheme 26. Takayama's and Kobayashi-Ikekawa's synthetic routes to 24,24-difluoro-25(OH)D₃ (80).

In 1980, Kobayashi-DeLuca's group subsequently demonstrated that kidney homogenates from the chicken converted 24,24-difluoro-25(OH)D₃ (80) to 24,24-difluoro- 1α ,25(OH)₂D₃ (82) [56]. The results of the biological evaluation revealed that 82 and its nonfluorinated counterpart 1α ,25(OH)₂D₃ (1) equipotently stimulate intestinal calcium transport and bone calcium mobilization in vivo, while in another in vitro system, 82 was found to be four times more potent than 1α ,25(OH)₂D₃ (1).

In 1990, Kumar's group synthesized 24,24-difluoro-25-hydroxy-26,27-dihomovitamin D₃ (83) and its 1 α -hydroxy analogue (84) from 3 β -hydroxy-22,23-dinorcholenic acid using a Reformatsky reaction in their synthetic route (Scheme 27) [57]. Both analogues showed similar biological activities, such as intestinal calcium transport and bone calcium mobilization in vivo, to those of the respective nonfluorinated counterparts and also 25(OH)D₃ or 1 α ,25(OH)₂D₃.



Scheme 27. Kumar's synthetic approach to 24,24-difluorovitamin D homologues (83,84) using the Reformatsky reaction.

In 1992, two alternative linear synthetic routes to 24,24-difluoro- 1α ,25(OH)₂D₃ (82) were reported by Takayama et al. using the Reformatsky reaction with ethyl bromodifluoroacetate or Horner–Emmons reaction as the key step, respectively (Scheme 28) [58,59]. The starting material of the former was 1α -hydroxydehydroepiandrosterone, with a 3.8% overall yield of 82 [58], and that of the latter was vitamin D₂, with a 9.3% overall yield of 82 [59].



Scheme 28. Takayama's two improved synthetic approaches to 24,24-difluoro- $1\alpha,25$ (OH)₂D₃ (82).

As shown in Scheme 29A, the same group synthesized 24,24-difluoro- 1α (OH)D₃ (85) from a steroidal skeleton in 1996 [60]. This compound showed higher activity than 24,24-difluoro- 1α ,25(OH)₂D₃ (82) in intestinal calcium absorption. In 1998, Iwasaki-Takayama's group reported the synthesis of (25*R*)- and (25*S*)-24,24-difluoro- 1α ,25,26-trihydroxyvitamin D₃ (86,87), including the X-ray crystallographic analysis of a synthetic intermediate to determine C25-stereochemistry, and proved that the 25*S*-isomer (87) was the main CYP24-metabolite of 82 (Scheme 29B) [61].



Scheme 29. Synthetic routes to 24,24-difluoro-1α(OH)D₃ (85) (A) and its CYP24-metabolite (87) (B) by the Takayama group.

The 24,24-difluoro-19-norVD₃ analogues including 20-*epi*-versions (**88–91**) were reported by DeLuca et al. in 2015 [62]. In contrast to the previous synthetic methods starting from steroid skeletons, they demonstrated a convergent method utilizing the Wittig–Horner reaction between 24,24-difluoro-CD-rings (**92,93**) and a lithium salt of a phosphine oxide anion from A-ring precursors (Scheme 30). The 20*S*-derivatives showed marked bone-mobilizing activity in vivo; however, 2-methylene substitution was required for such elevated activity in the 20*R* series.

In 2019, our group developed the convergent synthesis of 24,24-difluoro- $25(OH)D_3$ (80) using a coupling reaction between the 24,24-difluoro-CD ring ketone derived from the Inhoffen–Lythgoe diol via 94 and an A-ring phosphine oxide (Scheme 31A) [63]. We subsequently synthesized a novel vitamin D-based VDR-silent SREBP inhibitor, KK-052, from 94 (Scheme 31B) [64].



Scheme 30. DeLuca's convergent approach to 24,24-difluoro- 1α ,25(OH)₂-19-norVD₃ (**88,89**) and the 2-exomethylene analogues (**90,91**) including their 20-*epi* versions using the Wittig–Horner reaction.



Scheme 31. Efficient construction of the 24,24-difluorinated CD-ring unit (94) and the subsequent A-ring coupling reaction (**A**). Synthesis of the novel SREBP-specific inhibitor KK-052 (**B**).



Ikekawa's group reported in 1979 the first synthesis of C24-monofluoro-25(OH)D₃ (95) from cholenic acid without assigning the C24 stereochemistry (Scheme 32) [55].

Scheme 32. Synthesis of 24-monofluoro-25(OH)D₃ (95) using nucleophilic fluorination as a key step.

After this, (24R)-24-fluoro- 1α ,25(OH)₂D₃ (**96**) was synthesized by Uskoković's group through linear (Scheme 33A) and convergent (Scheme 33B) synthetic routes in 1985 [65] and 1988 [66], respectively. In this case, (24R)-24-fluoro- 1α ,25(OH)₂D₃ (**96**) showed a longer plasma half-life and higher anti-rachitogenic activity than 1α ,25(OH)₂D₃ in vivo.



Scheme 33. (24*R*)-Fluoro-1 α ,25(OH)₂D₃ (**96**) was synthesized by Uskoković et al. through a linear synthetic route using **97** and optically active L-(-)-malic acid as a chiral synthon (**A**) and an alternative convergent route starting from a CD-ring part, **98** (**B**).

5.4. 25-Fluorinated VD₃ Analogues

The 25-hydroxylation is the initial metabolic conversion of VD₃ by CYP2R1 or CYP27A1 [18], and $25(OH)D_3$ (2) is known as the major circulating metabolite in the human body.

The first introduction of fluorine to C25, i.e., the synthesis of 25-fluoro-VD₃, was reported by DeLuca et al. in 1977 (Scheme 34A) [67]. C3-Selective *O*-acetylation of 25(OH)D₃ (2) and subsequent direct fluorination at the C25 position using DAST, followed by deacetylation, gave 25-fluorovitamin D₃ (99). In 1978, Stern and coworkers synthesized 25-fluoro- 1α (OH)D₃ (100) using the same approach (Scheme 34B) [68]. The results of the biological evaluation indicated that 25-fluoro- 1α (OH)D₃ (100) was approximately equipotent to 1α (OH)D₃ (5) in VDR binding and stimulating bone resorption in vitro, and the C25-fluoro substituent behaved similarly to hydrogen, without elevating its biological potency.



Scheme 34. Synthesis of 25-fluoro-VD₃ analogues via direct C25 fluorination from $25(OH)D_3$ (**A**) and $1\alpha_25(OH)_2D_3$ (**B**) using DAST as a fluorinating reagent.

As described in Section 2.1, DeLuca's group also synthesized 1,25-difluorovitamin D_3 in 1981, and this compound was devoid of any biological activity [21].

5.5. 26,27-Hexafluorinated VD₃ Analogues

Falecalcitriol (**101**) is 26,26,26,27,27,27-hexafluorinated VD₃ and has been approved for therapeutic use against secondary hyperparathyroidism in Japan [69,70]. Falecalcitriol (**101**) was ca. 10 times more active in increasing bone calcium mobilization than the natural hormone (**1**) in vivo [71]. Similarly to other fluorinated VD₃ analogues, it was metabolized more slowly than 1α ,25(OH)₂D₃ (**1**) by CYP24A1, and interestingly, its major metabolite (23*S*)-23-hydroxyfalecalcitriol (**102**) was equipotent to the parent compound **101** in its biological activity (Figure 1) [72–74]. The 23-hydroxylated **102** was specifically glucuronidated by UGT1A3 [75].



Figure 1. Structures of falecalcitriol (101) and its major metabolite (23S)-23-hydroxyfalecalcitriol (102).

The first synthesis of 26,26,26,27,27,27-hexafluoro-25(OH)D₃ (**103**) was reported by Kobayashi et al. in 1980 [76], and the same group subsequently synthesized 26,26,26,27,27,27-hexafluoro- 1α ,25(OH)₂D₃ (**101**) in 1982 [77]. In this case, 3β-Tetrahydropyranyloxychol-5-en-24-ol tosylate was used as a starting material, and hexafluoroacetone (HFA) was utilized for construction of the 26,26,26,27,27,27-hexafluoro unit (Scheme 35). The biological evaluation revealed that **101** was approximately 5-10-fold more active than 1α ,25(OH)₂D₃ (**1**) without inducing severe hypercalcemia.



Scheme 35. Synthesis of 26,26,26,27,27,27-hexafluoro-25(OH)D₃ (103) and 26,26,26,27,27,27-hexafluoro-1 α ,25(OH)₂D₃ (101) using hexafluoroacetone as a fluorine source.

Using the reaction of acetylides with excess HFA gas as the key step, syntheses of the hexafluoroalkynyl-VD₃ analogue (**104**), C-*seco*-hexafluoroalkynyl-VD₃ analogue (**105**), and the previously mentioned two-side-chain analogues **17-22** including alkene side chains (Section 2.1) were reported by Ohira et al. in 1992 [78], by Wu et al. in 2002 [79], and by Maehr et al. in 2009 [28], respectively (Scheme 36). Compound **104** had a potent inducing effect on the differentiation of cancer cells, with little calcium mobilization activity. Compound **105** showed comparable VDR-binding affinity to the natural hormone **1** and had strong antiproliferative activity against four cancer cell lines in vitro, with 1% calcemic activity compared with **1** in vivo. More recently, Sigüeiro et al. synthesized C22-diyne analogues with a C17-methyl group, including the hexafluoropropanol unit at the terminal (**106**), which showed potent VDR-binding affinity [80].



Scheme 36. Combination of the hexafluoropropanol unit derived from hexafluoroacetone (HFA) and triple bond(s), which brought conformational rigidity to the side chain.

Hayashi and colleagues described the introduction of the 26,26,26,27,27,27-hexafluoro unit utilizing an aldol reaction [52]. The C23 ketone was treated with HFA in the presence of LiHMDS to afford the HFA adduct (see Scheme 24 in Section 5.2).

As an alternative path to construct the hexafluoro unit, the nucleophilic trifluoromethylation of methyl esters with Ruppert–Prakash reagent (CF₃TMS) can be utilized. In our group, 26,26,26,27,27,27-hexafluoro-25(OH)D₃ (**103**) and 26,26,26,27,27,27-hexafluoro-CD ring (**107**) were synthesized in 2018 starting from the methyl esters (**108,110**) through trifluoromethylketones (**109,111**) as the intermediates (Scheme 37) [81].



Scheme 37. Synthesis of 26,26,26,27,27,27-hexafluoro-25(OH)D₃ (**103**) and synthetically useful 26,26,26,27,27,27-hexafluoro-CD-ring precursor (**107**) using efficient two-step trifluoromethylation method.

6. Summary

This review summarized the historical fluorinated VD_3 analogues with modification from the A-ring to the end of the side-chain, including their synthetic methods. With the aim of preventing or slowing their activation or degradation by CYPs, the A-ring and side-chain have been mainly focused on, and numerous VD_3 analogues containing the fluorine atom(s) have been synthesized. Hydroxylation at the C25 and C1 α positions of VD₃ is necessary for the activation process of the molecule by CYP2R1/CYP27A1 and CYP27B1, respectively; therefore, in general, the introduction of fluorine to these positions decreases the biological activity through VDR if compared to non-fluorinated 25(OH)D₃ or 1 α ,25(OH)₂D₃ as each parent VDR ligand. On the other hand, fluorination at the side chain C23, C24, and C26(27) of VD₃, where deactivating hydroxylation occurs based on CYP24A1 metabolism, produces strong VDR agonists that have a long half-life in vivo. Among them, falecalcitriol was successfully approved for the treatment of secondary hyperparathyroidism in Japan. Discovery of the new functions of VD₃ continues; for example, the potent SREBP-inhibitory activity of 25(OH)D₃ [82] and the new fluorinated analogues with their efficient synthetic methods may contribute to the treatment of patients with VD₃-function-related disease in the future.

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