

## **Supplementary materials**

### **Mother–fetus immune cross-talk coordinates “extrinsic”/”intrinsic” embryo gene expression noise and growth stability**

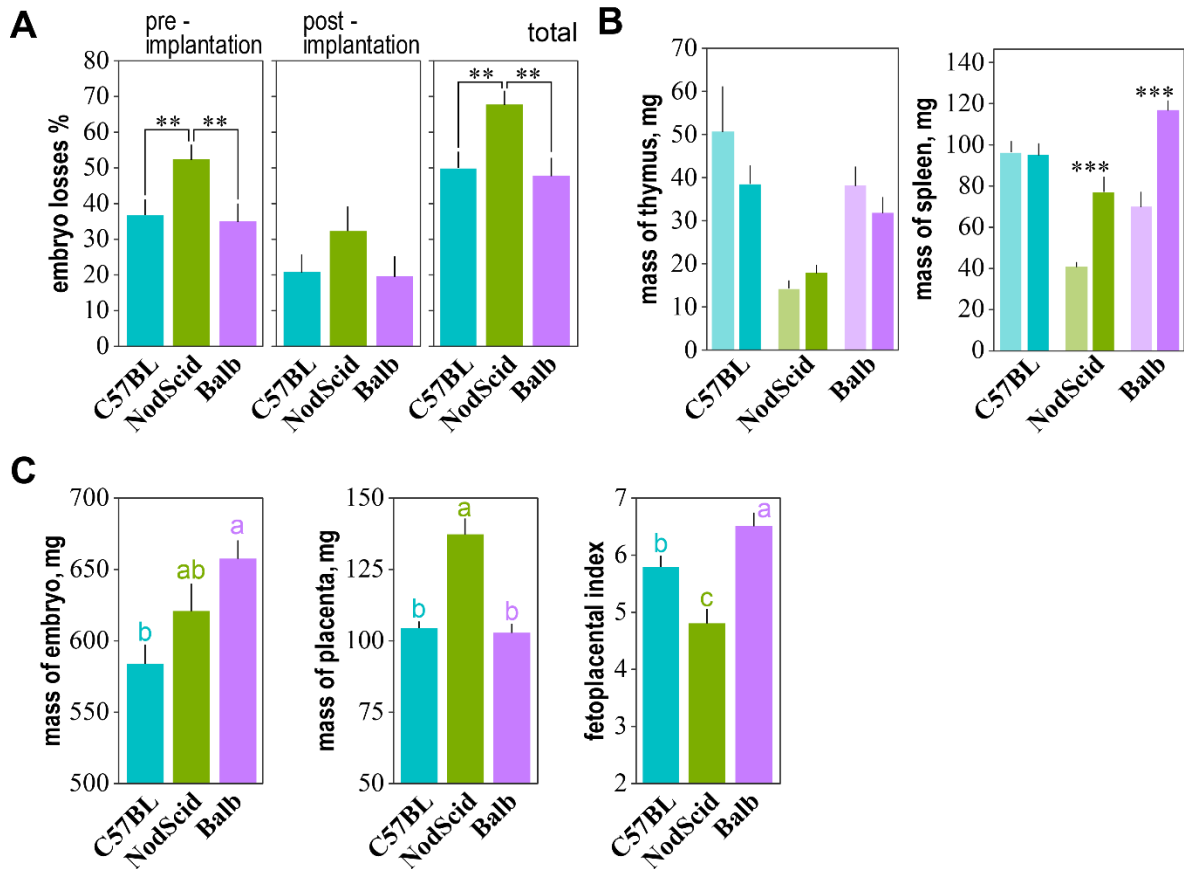
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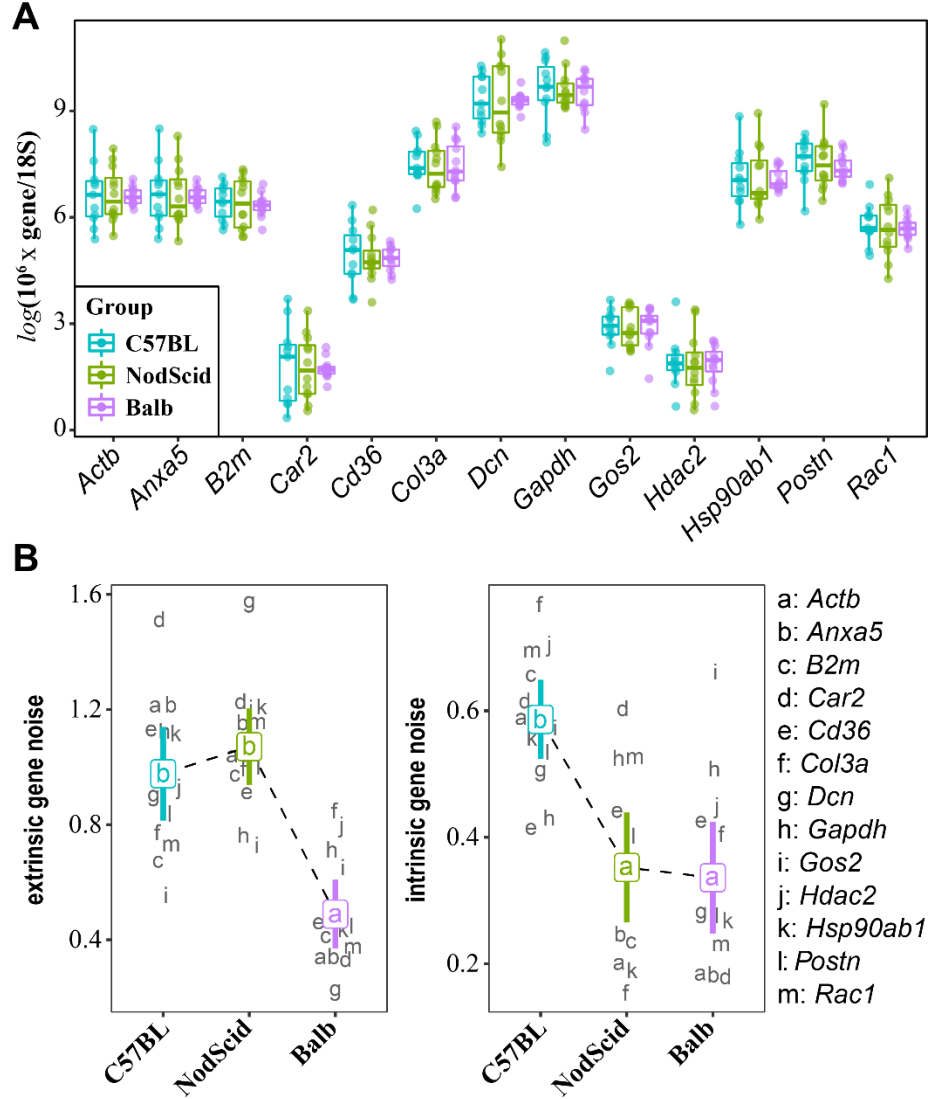


**Figure S1. Characteristics of C57BL/6J fetuses and surrogate mothers' immunocompetent organs response.**

**A)** Pre-, post-implantation, and total embryo losses in C57BL/6J, NOD-SCID, and BALB/c surrogate mothers. \*\* - Student's t-test,  $p < 0.01$ .

**B)** Females' thymus and spleen mass in non-pregnant females (light-colored bars) and in pregnant (day 16.5 of surrogate pregnancy) females (dark-colored bars). \*\*\* -  $p < 0.001$ , t-test; the number of non-pregnant/pregnant females: C57BL/6J ( $m = 3/13$ ), NOD-SCID ( $m = 5/16$ ) and BALB/c ( $m = 5/13$ ).

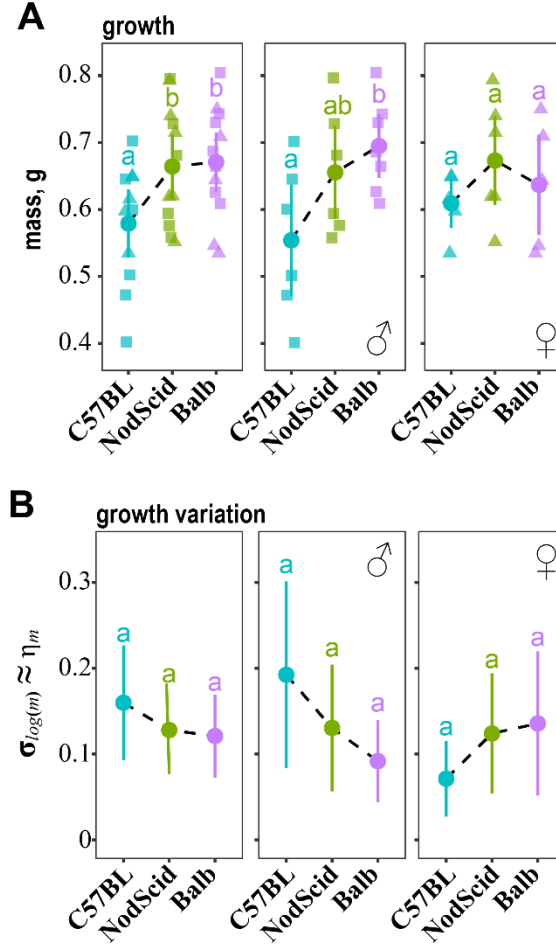
**C)** C57BL/6J embryo weight, placenta weight, and fetoplacental index on day 16.5 of surrogate pregnancy. Letters indicate significant statistical differences at  $p < 0.05$ , LSD-test; the number of embryos by surrogate mother group: C57BL/6J ( $n = 61$ ), NOD-SCID ( $n = 46$ ) and BALB/c ( $n = 49$ ).



**Figure S2. The impact of mother-fetus MHC-mediated interactions on gene-wise “extrinsic”/“intrinsic” fluctuations in left/right forelimbs of C57BL/6J embryos.**

**A)** Left/right mean RT-qPCR  $\Delta Ct = \log(10^6 g/18S)$  values for each gene measured on day 16.5 of gestation in forelimbs of C57BL/6J embryos, developing in C57BL/6J, NOD-SCID, and BALB/c surrogate mothers. ANOVA and pair-wise comparisons revealed no significant differences between surrogate mother groups'  $\Delta Ct$ s of selected genes.

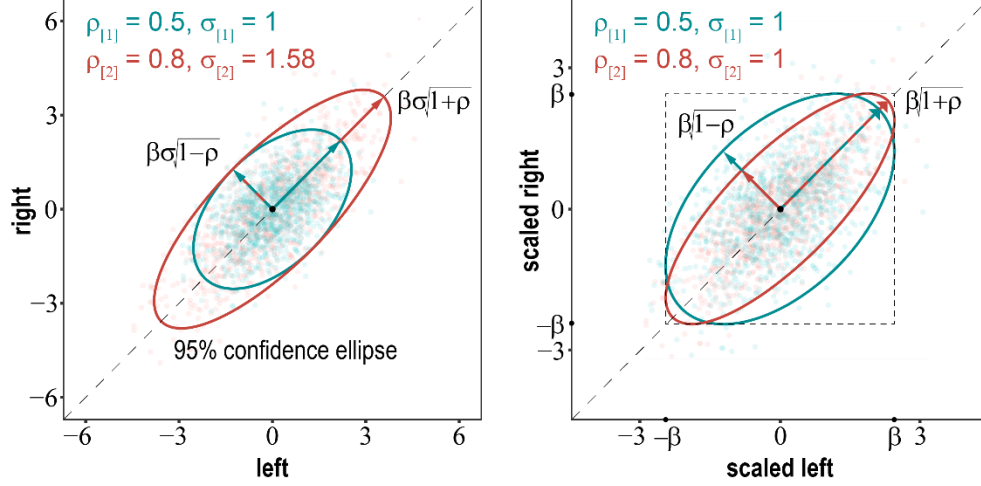
**B)** Estimations of “extrinsic” ( $\eta_{ext.} \approx \sqrt{2\lambda_1}$ ) and “intrinsic” ( $\eta_{int.} \approx \sqrt{2\lambda_2}$ ) gene noise for the selected genes following PCA decomposition of left/right gene expression ( $\Delta Ct$ ) matrices for each gene (eq. (9)). Colored boxes indicate means, whiskers - 95% confidence intervals, letters within mean boxes - multiple t-test comparisons with significant differences at  $FDR < 0.05$ .



**Figure S3. The effect of surrogate mother MHC haplotype on C57BL/6J embryos mass and its fluctuation.**

**A)** Non-adjusted mass of C57BL/6J embryos developed in C57BL/6J, NOD-SCID, and BALB/c surrogate mothers; see Figure 3C.

**B)** GAMLSS estimations of  $\hat{\sigma}_{\log(m)} \approx \hat{\eta}_m$  for non-adjusted embryo mass; see Figure 3D.



**Figure S4. Correlation is not a proper measure of the FA/”intrinsic” variability.**

To show that the correlation could be a misleading measure of the FA, let’s assume a trait  $x$  under two conditions [1] (green) and [2] (red). Under [1], the correlation between left and right  $\rho_{l,r[1]} = 0.5$ , and  $\sigma_{x[1]} = 1.0$ ; under [2],  $\rho_{l,r[2]} = 0.8$  and  $\sigma_{x[2]} = \frac{\sigma_{x[1]}\sqrt{1-\rho_{l,r[1]}}}{\sqrt{1-\rho_{l,r[2]}}} = 1.58$ .

Left panel, from the eq. (2), the “extrinsic” component of the DI of  $x_{[1]}$  is greater than that of  $x_{[2]}$ :

$$\text{Var}[E(x_{[1]}|\xi)] \approx \frac{1}{2}\sigma_{x[1]}^2(1 + \rho_{l,r[1]}) = 0.75, \text{ and } \text{Var}[E(x_{[2]}|\xi)] \approx 2.25.$$

From the eq. (3), the “intrinsic” component of the DI (FA) is equal for  $x_{[1]}$  and  $x_{[2]}$ :

$$E[\text{Var}(x_{[1]}|\xi_{[1]})] \approx \frac{1}{4}FA_{x[1]} \approx \frac{1}{2}\sigma_{x[1]}^2(1 - \rho_{l,r[1]}) = 0.25, \text{ and } E[\text{Var}(x_{[2]}|\xi_{[2]})] \approx 0.25.$$

Ellipses correspond to 95% confidence ellipses,  $\beta = \sqrt{\chi_{2,\alpha=0.05}^2} \approx 2.45$ .

Right panel, estimation of the FA using only the correlation leads to a conclusion that  $FA_{x[1]} > FA_{x[2]}$ . The such discrepancy stems from that that correlation is estimated as the covariance of standardized variables. In other words, information on the fluctuation of  $x$  ( $\sigma_x^2$ ) is dropped off. Rewriting the FA for the standardized trait  $x$ :  $FA_x \approx 2(1 - \rho_{l,r})$  (eq. (2)), it is easy to see that omitting the changes in  $\sigma_x^2$  could result in a misleading conclusion about changes in the  $FA_x$ .

**Table S1. Primers used for RT-qPCR**

<b>Gene</b>	<b>Description</b>	<b>Ensembl</b>	<b>Forward</b>	<b>Reverse</b>
<i>Actb</i>	Actin, beta	ENSMUSG00000029580	GGCTGTATTCCCCTCCATCG	CCAGTTGGTAACAATGCCATGT
<i>Anxa5</i>	Annexin A5	ENSMUSG00000027712	ATCCTGAACCTGTTGACATCCC	AGTCGTGAGGGCTTCATCATA
<i>B2m</i>	Beta-2 microglobulin	ENSMUSG00000060802	TTCTGGTGCTTGTCTCACTGA	CAGTATGTTTCGGCTTCCCATTCT
<i>Car2</i>	Carbonic anhydrase 2	ENSMUSG00000027562	CGGATGGATTGGCTGTTTTGG	GCACGCTTCCCCTTTGTTTTTA
<i>Cd36</i>	CD36 antigen	ENSMUSG00000002944	ATGGGCTGTGATCGGAACTG	GTCTTCCCAATAAGCATGTCTCC
<i>Col3a1</i>	Collagen, type III, alpha 1	ENSMUSG00000026043	CTGTAACATGGAACTGGGGAAA	CCATAGCTGAACTGAAAACCACC
<i>Dcn</i>	Decorin	ENSMUSG00000019929	TCTTGGGCTGGACCATTGAA	CATCGGTAGGGGCACATAGA
<i>Gapdh</i>	Glyceraldehyde-3-phosphate dehydrogenase	ENSMUSG00000057666	AGGTCGGTGTGAACGGATTTG	TGTAGACCATGTAGTTGAGGTCA
<i>G0s2</i>	G0/G1 switch gene 2	ENSMUSG00000009633	AAGGAGATGATGGCGCAGAAG	GCTGCACACCGTCTCAACTA
<i>Hdac2</i>	Histone deacetylase 2	ENSMUSG00000019777	TGTGCTTGCCATCCTCGAATT	AGCTTCCTCAACACCATCACC
<i>Hsp90ab1</i>	Heat shock protein 90 alpha, class B member 1	ENSMUSG00000023944	TCAAACAAGGAGATTTTCCTCCG	GCTGTCCAACCTAGAAGGGTC
<i>Postn</i>	Periostin, osteoblast specific factor	ENSMUSG00000027750	GAGGTCTCCAAGGTCACAAAGT	TGTGTCTCCCTGAAGCAGTCT
<i>Rac1</i>	RAS-related C3 botulinum substrate 1	ENSMUSG00000001847	GAGACGGAGCTGTTGGTAAAA	ATAGGCCCAAGATTCACTGGTT
<i>Rn18s</i>	18S ribosomal RNA	ENSMUSG00000119584	CTCAACACGGGAAACCTCAC	CGCTCCACCAACTAAGAACG

**Table S2. The number of implanted and live C57BL/6J embryos in experimental surrogate mother groups (mean  $\pm$  SE)**

<b>Surrogate mother group</b>	<b>n</b>	<b>implanted embryo</b>	<b>live embryos</b>
C57BL/6J	13	5.92 $\pm$ 1.05	4.69 $\pm$ 1.01
NOD-SCID	16	4.25 $\pm$ 0.98	2.88 $\pm$ 0.75
BALB/c	13	4.69 $\pm$ 1.31	3,77 $\pm$ 1.06

**Table S3. GAMLSS models of the relationships between the distribution parameters (mean -  $\mu$ , dispersion -  $\sigma$ ) of logarithm-transformed embryo mass ( $\log(m)$ ) and individual estimates of pulled “intrinsic” gene noise ( $\eta_{int.}$ ) for C57BL/6J embryos**

**A) Models for  $\log(m)$**

group	GAMLSS model	estimate ( $\hat{\alpha}, \hat{\beta}$ ) $\pm$ s.e.	p
all <sup>(a)</sup>	(1) $\mu_{\log(m)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.264<math>\pm</math>0.028</b>	<b>&lt; 0.001</b>
	(2) $\sigma_{\log(m)} \sim \beta_0 + \beta \log(\eta_{int.})^{(b)}$	<b>0.777<math>\pm</math>0.276</b>	<b>0.008</b>
C57BL/6J	(1) $\mu_{\log(m)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.208<math>\pm</math>0.024</b>	<b>&lt; 0.001</b>
	(2) $\sigma_{\log(m)} \sim \beta_0 + \beta \log(\eta_{int.})$	<b>3.524<math>\pm</math>1.168</b>	<b>0.019</b>
NOD-SCID	(1) $\mu_{\log(m)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.255<math>\pm</math>0.054</b>	<b>0.002</b>
	(2) $\sigma_{\log(m)} \sim \beta_0 + \beta \log(\eta_{int.})$	0.715 $\pm$ 0.594	0.263
BALB/c	(1) $\mu_{\log(m)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.241<math>\pm</math>0.046</b>	<b>0.001</b>
	(2) $\sigma_{\log(m)} \sim \beta_0 + \beta \log(\eta_{int.})'$	0.877 $\pm$ 0.534	0.139

**B) Models for litter-size adjusted  $\log(m)$**

group	GAMLSS model	estimate ( $\hat{\alpha}, \hat{\beta}$ ) $\pm$ s.e.	p
all	(1) $\mu_{\log(m)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.196<math>\pm</math>0.021</b>	<b>&lt; 0.001</b>
	(2) $\sigma_{\log(m)} \sim \beta_0 + \beta \log(\eta_{int.})'$	<b>0.979<math>\pm</math>0.247</b>	<b>&lt; 0.001</b>
C57BL/6J	(1) $\mu_{\log(m)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.144<math>\pm</math>0.017</b>	<b>&lt; 0.001</b>
	(2) $\sigma_{\log(m)} \sim \beta_0 + \beta \log(\eta_{int.})'$	<b>3.031<math>\pm</math>0.667</b>	<b>0.003</b>
NOD-SCID	(1) $\mu_{\log(m)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.24<math>\pm</math>0.039</b>	<b>&lt; 0.001</b>
	(2) $\sigma_{\log(m)} \sim \beta_0 + \beta \log(\eta_{int.})'$	<b>1.309<math>\pm</math>0.449</b>	<b>0.015</b>
BALB/c	(1) $\mu_{\log(m)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.103<math>\pm</math>0.021</b>	<b>0.001</b>
	(2) $\sigma_{\log(m)} \sim \beta_0 + \beta \log(\eta_{int.})'$	<b>1.801<math>\pm</math>0.506</b>	<b>0.007</b>

<sup>(a)</sup> - all embryos.

<sup>(b)</sup> – Note that from the Taylor expansion for the moments  $\sigma_{\log(m)} \approx \frac{\mu_m}{\sigma_m} = \eta_m$ . Assuming a normal distribution of  $\log(m)$ :  $\sigma_{\log(m)} = \sqrt{\log(1 + \eta_m^2)}$ , and for  $\eta_m^2 < 1$ ,  $\sigma_{\log(m)} \approx \eta_m$ .



**Table S4. GAMLSS models of the relationships between the distribution parameters ( $\mu$ ,  $\sigma$ ) of  $\log(PLGF)$  (PLGF concentrations, ng/l) and individual estimates of pulled “intrinsic” gene noise ( $\eta_{int.}$ ) for C57BL/6J embryos**

group	GAMLSS model	estimate ( $\hat{\alpha}, \hat{\beta}$ ) $\pm$ s.e.	p
all	(1) $\mu_{\log(PLGF)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.551<math>\pm</math>0.081</b>	<b>&lt; 0.001</b>
	(2) $\sigma_{\log(PLGF)} \sim \beta_0 + \beta \log(\eta_{int.})$	<b>1.424<math>\pm</math>0.314</b>	<b>&lt; 0.001</b>
C57BL/6J	(1) $\mu_{\log(PLGF)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.555<math>\pm</math>0.092</b>	<b>0.001</b>
	(2) $\sigma_{\log(PLGF)} \sim \beta_0 + \beta \log(\eta_{int.})$	1.823 $\pm$ 0.896	0.081
NOD-SCID	(1) $\mu_{\log(PLGF)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.524<math>\pm</math>0.105</b>	<b>0.001</b>
	(2) $\sigma_{\log(PLGF)} \sim \beta_0 + \beta \log(\eta_{int.})$	<b>2.338<math>\pm</math>0.555</b>	<b>0.003</b>
BALB/c	(1) $\mu_{\log(PLGF)} \sim \alpha_0 + \alpha \log(\eta_{int.})$	<b>-0.353<math>\pm</math>0.089</b>	<b>0.004</b>
	(2) $\sigma_{\log(PLGF)} \sim \beta_0 + \beta \log(\eta_{int.})$	0.372 $\pm$ 0.53	0.502