



Review

# Activated Hepatic Stellate Cells in Hepatocellular Carcinoma: Their Role as a Potential Target for Future Therapies

Esraa Ali <sup>1,\*</sup>, Andriy Trailin <sup>1,\*</sup>, Filip Ambrozkiwicz <sup>1</sup>, Václav Liška <sup>2,3</sup> and Kari Hemminki <sup>1,4</sup>

<sup>1</sup> Laboratory of Translational Cancer Genomics, Biomedical Center, Faculty of Medicine in Pilsen, Charles University, Alej Svobody 1665/76, 32300 Pilsen, Czech Republic

<sup>2</sup> Laboratory of Cancer Treatment and Tissue Regeneration, Biomedical Center, Faculty of Medicine in Pilsen, Charles University, Alej Svobody 1665/76, 32300 Pilsen, Czech Republic

<sup>3</sup> Department of Surgery University Hospital and Faculty of Medicine in Pilsen, Charles University, Alej Svobody 80, 32300 Pilsen, Czech Republic

<sup>4</sup> Department of Cancer Epidemiology, German Cancer Research Center, Im Neuenheimer Feld 280, 69120 Heidelberg, Germany

\* Correspondence: andriy.trailin@lfp.cuni.cz; Tel.: +420-377-593-862

**Abstract:** Hepatocellular carcinoma (HCC) is a global healthcare challenge, which affects more than 815,000 new cases every year. Activated hepatic stellate cells (aHSCs) remain the principal cells that drive HCC onset and growth. aHSCs suppress the anti-tumor immune response through interaction with different immune cells. They also increase the deposition of the extracellular matrix proteins, challenging the reversion of fibrosis and increasing HCC growth and metastasis. Therapy for HCC was reported to activate HSCs, which could explain the low efficacy of current treatments. Conversely, recent studies aimed at the deactivation of HSCs show that they have been able to inhibit HCC growth. In this review article, we discuss the role of aHSCs in HCC pathophysiology and therapy. Finally, we provide suggestions for the experimental implementation of HSCs in HCC therapies.

**Keywords:** hepatocellular carcinoma; hepatic stellate cells; fibrosis regression; therapeutic studies



**Citation:** Ali, E.; Trailin, A.; Ambrozkiwicz, F.; Liška, V.; Hemminki, K. Activated Hepatic Stellate Cells in Hepatocellular Carcinoma: Their Role as a Potential Target for Future Therapies. *Int. J. Mol. Sci.* **2022**, *23*, 15292. <https://doi.org/10.3390/ijms232315292>

Academic Editor: Nam Deuk Kim

Received: 19 October 2022

Accepted: 2 December 2022

Published: 4 December 2022

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



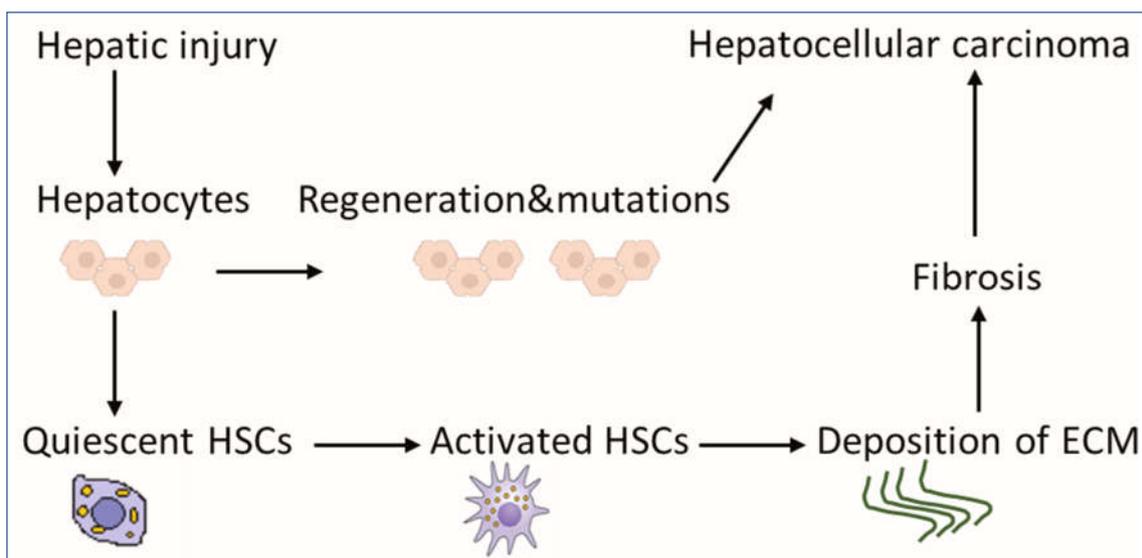
**Copyright:** © 2022 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

Liver cancer is a global health problem, with an estimated increase of 32% by 2040 [1]. Representing 90% of liver cancers, hepatocellular carcinoma (HCC) causes 700,000 deaths annually.

Although HCC pathogenesis is complex and varies depending on underlying etiology, the usual background setting for HCC is liver injury, chronic inflammation, irreversible fibrosis, and cirrhosis [2]. In fact, 80–90% of HCC develops in the fibrotic or cirrhotic liver [3]. Hepatic stellate cells (HSCs) play a key role in this sequence of events, contributing mainly to liver fibrosis and cirrhosis. They are liver-specific mesenchymal cells, which are located in the perisinusoidal space in contact with different cell types [4]. In a healthy liver, HSCs exist in a quiescent non-proliferative state as an important source of paracrine, autocrine, and chemoattractant factors to maintain hepatic homeostasis [5]. Quiescent HSCs are very sensitive to extracellular pro-fibrotic signals [6] and contain numerous vitamin A lipid droplets, which are essential for the proper function of the immune system [7]. When toxins or viruses injure the liver, damaged hepatocytes and immune cells secrete signals, which could activate HSCs into myofibroblast-like cells [8]. Activated HSCs produce an extracellular matrix (ECM) at the site of injury as a temporary protective scar to prevent further damage, initiating the first steps of fibrosis [8,9]. Long-acting agents maintain the activation of HSCs, increasing their capabilities for proliferation and migration [10]. Activated HSCs produce more ECM, leading to chronic fibrosis and cirrhosis and eventually to HCC (Figure 1) [6]. Despite significant advances in the treatment of HCC, drug-resistance is a critical obstacle [11], and the 5-year survival rate is low (5–14%) [12], but survival rates of greater than 20% have been reached in some regions [13]. Therefore, there is a

potential to increase survival rates. Given the fact that available therapies can activate HSCs, synchronous targeting aHSCs may be beneficial for patients [14]. In this review, we emphasize the role of aHSCs in HCC. We clarify how aHSCs suppress the immune response in the tumor microenvironment. We provide insights into the contribution of HSCs to slow down fibrosis regression and to increase deposition of ECM proteins, which may favor HCC growth and metastasis. In addition, we focus on the ability of conventional therapies to activate HSCs, whereas studies of aHSC deactivation might be an important strategy to improve HCC treatment. Finally, we suggest how to reinforce experiments that target aHSCs.



**Figure 1.** Scheme for the contribution of HSCs to liver pathology. HSCs exist in a quiescent state, containing numerous vitamin A lipid droplets. When the liver is injured, damaged hepatocytes mediate HSC activation, which could produce a large amount of ECM, leading to fibrosis as an indirect mechanism of HCC. Mutations during the regeneration of hepatocytes may lead directly to the development of HCC. Abbreviations: HSCs: hepatic stellate cells, HCC: hepatocellular carcinoma and ECM: extracellular matrix.

## 2. The Role of aHSCs in HCC

### 2.1. The Suppression of the Antitumor Immune Response by aHSCs

In HCC, aHSCs receive signals from individual immune cells, and, in turn, they produce soluble mediators, acting on surrounding immune cells [15]. HSC mediators could orchestrate both innate and adaptive immunity, resulting in an immunosuppressive tumor microenvironment (Table 1) [7]. Reduction of antitumor responses was shown in immunocompetent mice after co-transplantation of HSCs and HCC cells [16]. Such cotransplantation of HSCs inhibited systemically lymphocyte infiltration, which promoted tumor cell proliferation and, therefore, HCC growth; the size of the tumors was HSC dose-dependent [16]. Previous experiments addressing HCC–HSC interactions were performed on immunodeficient mice, and, therefore, the effect of HSC on the immune system was not investigated. Using immunocompetent mice, the authors were able to define HSC-immune interactions in HCC [16].

The antigen-directed cytotoxicity of T lymphocytes (TLs) boosts the immune response against cancer [17]. Activation and proliferation of TL in tumor tissue, predominantly CD8+ and CD4+ T lymphocytes, can control HCC progression [18]. HSCs can exert their immunomodulatory activities by downregulating the number and function of CD4+ and CD8+ TLs [19]. Contrary to quiescent HSCs, aHSCs in mice and humans expressed programmed death-ligand 1 (PD-L1) to inhibit TL responses [7]. PD-L1 expressed by HSCs can induce TL apoptosis, attenuate TL infiltration, and suppress TL-mediated cytotoxicity,

therefore inhibiting TL responses and enabling tumor cells to escape the host immune response [20]. In addition, HSCs may prevent the local stimulation of naive TLs [21]. In Hepa1–6 cells, activated HSCs induced the death of activated TLs and reduced the cytotoxicity of cancer-specific TLs, which resulted in the increased proliferation and migration of cancer cells [22]. More investigation of the role of aHSCs in the apoptosis of TLs in HCC patients is needed.

aHSCs also induce expansion of two suppressive immune cell populations; myeloid-derived suppressor cells (MDSCs) [14] and T helper 17 (Th17) cells, a subset of CD4+ effector T cells [23]. MDSCs play a pivotal negative role in the immune response through the inhibition of cytotoxic T cells and recruitment of regulatory T cells, which results in tumor progression [24]. HSCs induce MDSC accumulation in the tumor tissue by the stimulation of the COX2–PGE2–EP4 pathway [25]. Inhibition of this pathway in murine orthotopic HCC models downregulated MDSCs and HCC growth [25]. Immunosuppressive functions of Th17 cells may contribute to HCC progression [26]. IL-17A produced by Th17 could increase cancer cell motility via the activation of the nuclear factor- $\kappa$ B (NF- $\kappa$ B) transcript factor, increasing HCC metastasis [27]. Culturing CD4+ cells with HSCs (extracted from hepatitis B virus-related fibrotic liver tissue) increased the percentages of Th17 cells [23]. HSCs may secrete high levels of interleukin-6 as a critical initiator of Th17 expansion and tumor necrosis factor- $\alpha$  as a key regulator of Th17 differentiation [28]. Interestingly, previous data indicated suppression of Th17 differentiation by mouse HSCs [29]. Critical evaluation of Th17-HSCs interactions could be addressed in appropriate mouse models of HCC.

Macrophages polarize in the liver with strong plasticity into pro-inflammatory M1 or anti-inflammatory M2 in response to local signals from the tumor microenvironment [30,31]. M1 macrophages are thought to be tumoricidal, while M2 macrophages are usually believed to promote tumorigenesis and tumor progression [32]. M2 macrophages in HCC promote the invasion and migration of tumor cells [33]. M2 macrophage-derived CCL22 was proven to enhance tumor migration through the activation of epithelial–mesenchymal transition [34]. aHSCs recruited CCL2/CCR2 pathway in HCC cell lines to stimulate M2 phenotypic transformation [35]. M2 macrophage polarization could lead to the progression of HCC [36].

Natural killer (NK) cells defend the body against tumors by engaging death-inducing receptors to stimulate cancer cell apoptosis. HCC patients with low intratumoral NK cells infiltration have shorter disease-free survival [37]. In animal models of fibrosis, transforming growth factor- $\beta$  secreted by HSC could inhibit NK cell function [38]. On the other hand, NK cells could induce apoptosis of aHSCs in hepatitis C virus-infected patients [39] and mouse models of fibrosis [40]. Studies on the interaction between aHSCs and NK cells in HCC models should be investigated. Dendritic cells (DCs) can activate antitumor immunity by priming TL against cancer-progression-associated antigens. HSCs induce the expression of dendritic-cell-derived immunoglobulin receptor 2 (DIgR2), which inhibits DC-induced antigen-specific TL responses [41]. DIgR2 was shown to bind to the receptor in TLs, suppressing TL proliferation, cytokine production, and cytotoxic TL activity [41]. Co-culturing of tumor-HSCs (isolated from the tumor) to DCs induced the expression of DIgR2, in contrast to quiescent HSCs, which had no significant effect on DIgR2 expression. Considering quiescent HSCs in such studies boosts the role of activated HSCs in HCC [41].

Although the role of immune system in liver cancer is complex [42], the overall role of aHSCs in immune regulation is pro-oncogenic [16]. Exploring the interaction between different immune cells and HSCs in established HCC models could highlight the main targets to improve immune surveillance against HCC.

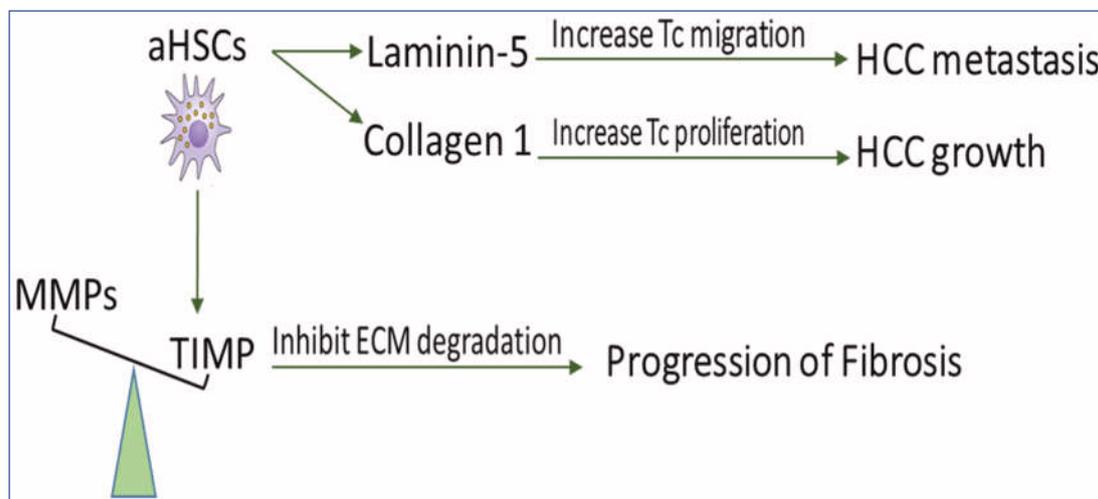
**Table 1.** Immunosuppressive functions of HSCs.

Mediator	Immune cell	Response	Possible Effect
PD-L1	T lymphocytes	TL apoptosis, attenuation of TL infiltration, and suppression of TL-mediated cytotoxicity [20]	
Dendritic-cell-derived immunoglobulin receptor 2	Dendritic cells	Inhibition of DC-induced antigen-specific TL responses [41]	HCC growth
COX2-PGE2-EP4	MDSCs	MDSC accumulation [25]	
Transforming growth factor- $\beta$	NK cells	Inhibition of NK cell function [38]	
Interleukin-6 and tumor necrosis factor- $\alpha$	Th17	Th17 expansion and Th17 differentiation [28]	HCC metastasis
CCL2/CCR2	Macrophages	Stimulation of M2 macrophages phenotypic transformation [35]	

HSCs produce soluble mediators acting on surrounding immune cells, resulting in a negative immune response. The immune regulatory role of HSCs may lead to HCC growth and metastasis. HSCs: hepatic stellate cells, PD-L1: programmed death-ligand 1, MDSCs: myeloid-derived suppressor cells, NK: natural killer, Th17: T helper 17, HCC: hepatocellular carcinoma, and TL: T lymphocyte.

## 2.2. HSCs Upregulate the Deposition of ECM for the Development of Fibrosis and HCC

Under normal conditions, the rate of ECM production in the liver equals that of its degradation, resulting in no net accumulation of the matrix. Fibrogenesis occurs when there is an imbalance between ECM production and degradation [43], resulting in the impairment of liver functions, which may eventually lead to cirrhosis and HCC [44]. Fibrosis and cirrhosis are reported clinically as reversible processes [45]. Reversibility of liver fibrosis depends mainly on the degradation of ECM [46]. aHSCs are able to increase matrix protein synthesis, which might lead to the irreversibility of fibrosis and favor progress and metastasis of HCC (Figure 2).



**Figure 2.** The scheme demonstrates the role of HSCs in disturbing ECM balance for the development of fibrosis and HCC. aHSCs express TIMP glycoproteins which inhibit MMP-mediated degradation of ECM and, therefore, prevent fibrosis regression. They also secrete collagen I, which increases tumor cell proliferation and produces Ln-5, which promotes tumor cell migration. MMPs: matrix metalloproteinases, TIMPs: tissue inhibitors of MMP, aHSCs: activated hepatic stellate cells, HCC: hepatocellular carcinoma, ECM: extracellular matrix, and TC: tumor cell.

The reversibility of fibrosis and cirrhosis is dependent on the activity of matrix metalloproteinases (MMPs) [47]. MMPs are a group of enzymes involved in the degradation of

ECM-proteins, which are blocked by tissue inhibitors of MMP (TIMPs). It has been reported that prolonged expression of TIMPs, even after withdrawal of fibrogenic factors, slows the regression of liver fibrosis [47]. In a rat model of regressed liver fibrosis, the reversibility of fibrosis was increased in parallel with a marked decrease in TIMP expression [48]. Fully activated HSCs release and upregulate expression of TIMP-1 and TIMP-2, which inactivate MMPs through proteolytic cleavage [43,45,48]. Targeting activated HSCs in vivo decreased the expression of TIMP-1 and TIMP-2 and resulted in attenuated liver fibrosis [49]. Impairment of HSCs activation in mice downregulated TIMP-1 and diminished alcohol-induced steatohepatitis [50]. Interestingly, the addition of activated MMP-2 to aHSCs in culture enhanced the apoptosis of HSCs [51].

Increased production of ECM proteins, such as collagen I and laminin-5 (Ln-5), is associated with the growth and metastasis of HCC. Collagen I promotes HCC cell proliferation by regulating the integrin  $\beta$ 1/FAK signaling pathway [52]. HSCs produce collagen I [53], which has been associated with the increased aggressiveness of HCC [54], where silencing its expression in HSCs may treat liver fibrosis [55].

Upregulation of Ln-5 in HCC patients promotes the migration of tumor cells, which is directly related to poor prognosis and tumor metastasis [56]. HCC grows in a microenvironment enriched with Ln-5 produced by HSCs [57]. In human HCC tissues, Ln-5 was distributed mainly along aHSCs, stimulating tumor cell migration [58]. Blocking antibodies against Ln-5 in HCC cell lines in the presence of HSCs inhibited tumor metastasis [58], while the presence of HSCs or Ln-5 in HCC cell lines increased resistance to sorafenib [57].

Normalization of ECM may represent an important therapeutic strategy for HCC [59]. Analysis of HSC-secreted proteins that control components of ECM could identify possible targets for HCC treatment.

### 3. Activation and Deactivation of HSCs as a Result of Therapy

#### 3.1. Activation of HSCs by Conventional Therapy

Current conventional therapy can activate HSCs, which could explain its limited success in curing HCC (Table 2). Chemotherapy can cause activation of HSCs through stroma-derived factor 1 and hypoxia-inducible factor 1  $\alpha$ . These mechanisms often regulate, unite, or intersect with other pathways to activate HSCs [14]. Transarterial chemoembolization (TACE) is clinically recommended for patients with advanced-stage HCC. However, the long-term results of TACE in HCC might be compromised by TACE-induced hepatic hypoxia and subsequent HSC activation [60]. In HCC animal models, TACE activated HSCs and induced prominent hepatic fibrogenesis [61]. In contrast, latent HSCs countered cancer growth by increasing the cytotoxicity of chemotherapeutics such as doxorubicin [62]. Treating rat hepatoma cells with quiescent HSCs and doxorubicin enhanced the efficacy of doxorubicin and led to faster tumor cell death [62].

Sorafenib, as an anti-tumor molecular inhibitor, can impede HCC cell proliferation but can also activate HSCs through the mitogen-activated protein kinase (MAPK) signaling pathway [63]. In mice, combined delivery of sorafenib with an inhibitor for MAPK could prevent the activation of HSCs, resulting in anti-fibrotic properties [63]. Coculturing of HSC-LX2 in Huh7 cell lines induced sorafenib resistance [64]. The interaction between sorafenib and aHSC might affect the success rate of this molecular inhibitor.

The main challenge to classic radiotherapy is its side effects on the surrounding tissues [65]. Upon radiotherapy, HSCs were activated and accumulated in the patient's liver [66]. In a hepatoma cell line, radiotherapy activated HSCs through the toll-like receptor 4 pathway and increased the potential of HCC metastasis [67]. The activation of HSCs by radiation is a key process underlying hepatic fibrosis, which could promote radioresistance and tumor recurrence [14].

Although radiofrequency ablation (RFA) is increasingly incorporated into HCC treatment, available data indicate its ability to stimulate residual tumor growth and cause tumor recurrence [68]. It was reported that RFA could activate HSC through inflammatory

cytokine-mediated pathways. Elevated levels of interleukin 6 and a massive accumulation and migration of activated HSCs were recorded in mice after RFA [69].

Defining and analyzing the interconnected factors between HSC activation and conventional therapies could help to enhance the efficacy of current treatment modalities.

### 3.2. Pharmacological Approaches to Deactivate HSCs

Pharmacological trials that target activated HSCs suggest a new paradigm that “hitting one target leads to a domino effect”. This is easy to understand since targeting aHSCs will affect subsequently HSC-induced immunosuppression, drug resistance, and tumor metastasis (Table 2).

The trials are based mainly on targeting molecular pathways of HSCs activation. TGF- $\beta$  signaling is considered the key pathway that drives HSC activation [70]. Targeting TGF- $\beta$  emerges as an effective therapeutic option to revert the activation of HSCs and stop the progress of HCC. Galunisertib is a small-molecule selective inhibitor of TGF- $\beta$  receptor type I. The combination of galunisertib and sorafenib in patients with advanced HCC showed acceptable safety and prolonged overall survival [71]. Imatinib simultaneously and rather selectively inhibits TGF- $\beta$  signaling. In mice, El-Mezayen et al. targeted HSCs using imatinib–nanomedicine therapy resulting in outstanding anti-fibrotic effects with reduced cytotoxicity [72].

Another pathway for HSC activation implies the involvement of peripheral nerves. Peripheral nerves secrete substance P (SP), which transmits the information via a neurokinin-1 receptor (NK-1R)—expressed on HSCs. SP activates HSCs through the SP/NK-1R signal pathway [73]. In vivo, the combination of doxorubicin with capsaicin, as a blocker to “SP-HSCs”, could effectively inhibit drug resistance and HCC metastasis [73].

PD-L1 is required for HSC activation by stabilizing TGF- $\beta$  receptors [74]. Targeting HSC PD-L1 in mice suppressed HSC activation and growth of intrahepatic cholangiocarcinoma [74]. Nivolumab, a blocker of PD-L1, was applied in the treatment of advanced HCC patients who were resistant to sorafenib [75]. New therapeutic approaches that combine targeting HSCs with traditional treatment could become a gold standard to cure HCC.

**Table 2.** Illustration of the effect of current treatments and new pharmacological studies on HSCs.

Approach	Effect on HSCs	Possible Result	Ref.
TACE	Activate HSCs	Induce prominent hepatic fibrogenesis	[61]
Sorafenib	Activate HSCs	Resistance to sorafenib	[64]
Sorafenib and MAPK inhibitor	Prevent HSC activation	Anti-fibrotic effect	[63]
Radiotherapy	Activate HSCs	Increase HCC metastasis	[67]
Radiofrequency ablation	Activate HSCs	Tumor recurrence	[69]
Galunisertib and sorafenib	Deactivate HSCs	Prolonged overall survival in HCC patients	[71]
Imatinib–nanomedicine	Deactivate HSCs	Outstanding anti-fibrotic effects	[72]
Doxorubicin and capsaicin	Deactivate HSCs	Inhibit drug resistance and HCC metastasis	[73]
Nivolumab	Deactivate HSCs	Treat advanced HCC	[75]

Conventional therapies are able to activate HSCs, which could improve tumor growth. In contrast, the deactivation of HSCs by new pharmacological products could inhibit HCC progress and arise as an effective therapeutic strategy. HSCs: hepatic stellate cells, TACE: transarterial chemoembolization, MAPK: mitogen-activated protein kinase, and HCC: hepatocellular carcinoma.

## 4. Suggestions for the Application of HSCs in the Treatment of HCC

Growing evidence of the role of HSCs in HCC suggests that HSC-related therapies might revolutionize the therapy for HCC. Yet, several gaps still limit the application of HSC deactivation in the treatment of HCC.

The first obstacle is the obscure nature of molecular mechanisms of HSC activation and deactivation [76]. In particular, quiescent HSCs shift to the activated state in a dynamic

complex scenario with different subpopulations, which could orchestrate the course of HCC [77]. Filliol et al. showed how quiescent or least-activated HSCs express hepatic growth factor, which limits HCC growth, but in advanced disease stages, fully activated HSCs express collagen I, promoting tumor proliferation [77]. The signals, which modulate these HSC subpopulations, have not been investigated. In addition, during fibrosis regression, activated HSCs can undergo apoptosis or revert to an inactivate phenotype, which is distinct from quiescent HSCs [78]. We may address those challenges by analyzing gene expression signatures for quiescent, activated, and inactivated HSCs in parallel with transcriptional analysis of their cellular neighborhood during different stages of HCC. Administering HSCs to experimental animals in a different experimental setting (fibrosis, HCC) and tracking their interactions could give us a better understanding of the role of HSCs in different liver pathological conditions.

The second obstacle is that HSCs account only for 5% to 8% of total liver cells, and targeting HSCs may be blocked by the condensed nature of the perisinusoidal space [79]. Targeting HSCs with free drugs could cause severe systemic toxicity, and the benefits are restricted due to their poor solubility, short half-life, and low bioavailability [73]. The application of drug carrier systems, which deliver drugs to specific tissues [80], may solve this problem [79]. Delivering drugs to aHSCs would be expected to boost potency with decreased side effects [72]. When nanomicelles were modified to target aHSCs in vivo, they suppressed the activation of HSCs and inhibited fibrosis development safely and efficiently [81].

Controlled studies to define the main factors of HSC activation and the proper delivery system to target it should promote the potential of HSCs in the context of HCC therapies. The final step is to define the inclusion and exclusion criteria for such therapies. The inclusion criteria need to consider HCC staging and hepatic microenvironment, including immune cells and ECM materials.

## 5. Conclusions

HSCs play an exemplary role in the tumor regulating different cytokines and growth factors for the progress of HCC. They interact with different immune cells to suppress HCC immunosurveillance. Targeting HSC-related immune suppression would improve immune response to HCC. aHSCs disturb the balance of ECM proteins, which leads to the progression of fibrosis and HCC. Exploring how HSCs destabilize ECM could identify possible targets for HCC treatment. As current therapies activate HSCs, deactivation of HSCs has become a critical therapeutic strategy. Targeting aHSCs can be developed by studying their molecular activation mechanisms and selecting proper targeting methods.

**Author Contributions:** Conceptualization, E.A.; writing—original draft, E.A.; writing—review and editing, F.A., K.H., V.L. and A.T.; supervision, K.H.; funding acquisition, K.H. and V.L. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research has received funding from the European Union's Horizon 2020 research and innovation programme under grant agreement N°856620, grants of Ministry of Health of the Czech Republic AZV NU21-03-00506 and AZV NU21-03-00145, Cooperatio Program, research area SURG, and by the project National Institute for Cancer Research—NICR (Programme EXCELES, ID Project No. LX22NPO5102), funded by the European Union—Next Generation EU.

**Acknowledgments:** We would like to thank Marie Rajtmajerová for help in manuscript proofreading.

**Conflicts of Interest:** The authors declare no conflict of interest.

## Abbreviations

HCC	hepatocellular carcinoma
aHSCs	activated hepatic stellate cells
ECM	extracellular matrix
TLs	T lymphocytes

PD-L1	programmed death-ligand 1
MDSCs	myeloid-derived suppressor cells
Th17	T helper 17 cells
NF- $\kappa$ B	nuclear factor- $\kappa$ B
IL6	interleukin-6
NK	natural killer
IL-10	interleukin-10
TGF $\beta$	transforming growth factor- $\beta$
DCs	dendritic cells
MMPs	metallo-proteinases
TIMPs	tissue inhibitors of MMP
Ln-5	laminin-5
TACE	transarterial chemoembolization
MAPK	mitogen-activated protein kinase
RFA	radiofrequency ablation
NK-1R	neurokinin-1 receptor
SP	substance P
DIgR2	dendritic-cell-derived immunoglobulin receptor 2

## References

- Ramani, A.; Tapper, E.B.; Griffin, C.; Shankar, N.; Parikh, N.D.; Asrani, S.K. Hepatocellular Carcinoma-Related Mortality in the USA, 1999–2018. *Am. J. Dig. Dis.* **2022**, *67*, 4100–4111. [[CrossRef](#)] [[PubMed](#)]
- Chidambaranathan-Reghupaty, S.; Fisher, P.B.; Sarkar, D. Hepatocellular carcinoma (HCC): Epidemiology, etiology and molecular classification. 2021, 149, 1–61. *Adv. Cancer Res.* [[CrossRef](#)]
- Affo, S.; Yu, L.-X.; Schwabe, R.F. The Role of Cancer-Associated Fibroblasts and Fibrosis in Liver Cancer. *Annu. Rev. Pathol. Mech. Dis.* **2017**, *12*, 153–186. [[CrossRef](#)] [[PubMed](#)]
- Zhang, C.-Y.; Yuan, W.-G.; He, P.; Lei, J.-H.; Wang, C.-X. Liver fibrosis and hepatic stellate cells: Etiology, pathological hallmarks and therapeutic targets. *World J. Gastroenterol.* **2016**, *22*, 10512–10522. [[CrossRef](#)] [[PubMed](#)]
- Kanel, G.C.; Korula, J. General Aspects of the Liver and Liver Diseases. *Atlas Liver Pathol.* **2011**, 3–15. [[CrossRef](#)]
- Barry, A.; Baldeosingh, R.; Lamm, R.; Patel, K.; Zhang, K.; Dominguez, D.A.; Kirton, K.J.; Shah, A.P.; Dang, H. Hepatic Stellate Cells and Hepatocarcinogenesis. *Front. Cell Dev. Biol.* **2020**, *8*, 709. [[CrossRef](#)]
- Weiskirchen, R.; Tacke, F. Cellular and molecular functions of hepatic stellate cells in inflammatory responses and liver immunology. *Hepatobiliary Surg. Nutr.* **2014**, *3*, 344–363. [[CrossRef](#)]
- Yin, C.; Evason, K.J.; Asahina, K.; Stainier, D.Y. Hepatic stellate cells in liver development, regeneration, and cancer Find the latest version: Review series Hepatic stellate cells in liver development, regeneration, and cancer. *J. Clin. Invest.* **2013**, *123*, 1902–1910. [[CrossRef](#)]
- Krizhanovsky, V.; Yon, M.; Dickins, R.A.; Hearn, S.; Simon, J.; Miething, C.; Yee, H.; Zender, L.; Lowe, S.W. Senescence of Activated Stellate Cells Limits Liver Fibrosis. *Cell* **2008**, *134*, 657–667. [[CrossRef](#)]
- Carloni, V.; Luong, T.V.; Rombouts, K. Hepatic stellate cells and extracellular matrix in hepatocellular carcinoma: More complicated than ever. *Liver Int.* **2014**, *34*, 834–843. [[CrossRef](#)]
- Mossenta, M.; Busato, D.; Dal Bo, M.; Macor, M.; Toffoli, G. Novel Nanotechnology Approaches to Overcome Drug Resistance in the Treatment of Hepatocellular Carcinoma: Glypican 3 as a Useful Target for Innovative Therapies. *Int. J. Mol. Sci.* **2022**, *23*, 10038. [[CrossRef](#)]
- Sarveazad, A.; Agah, S.; Babahajian, A.; Amini, N.; Bahardoust, M. Predictors of 5 year survival rate in hepatocellular carcinoma patients. *J. Res. Med Sci. Off. J. Isfahan Univ. Med. Sci.* **2019**, *24*, 86. [[CrossRef](#)]
- Hemminki, K.; Försti, A.; Hemminki, O.; Liska, V.; Hemminki, A. Long-term survival trends for primary liver and pancreatic cancers in the Nordic countries. *JHEP Reports* **2022**, *4*, 100602. [[CrossRef](#)] [[PubMed](#)]
- Ruan, Q.; Wang, H.; Burke, L.J.; Bridle, K.R.; Li, X.; Zhao, C.-X.; Crawford, D.H.G.; Roberts, M.; Liang, X. Therapeutic modulators of hepatic stellate cells for hepatocellular carcinoma. *Int. J. Cancer* **2020**, *147*, 1519–1527. [[CrossRef](#)] [[PubMed](#)]
- Wu, M.; Miao, H.; Fu, R.; Zhang, J.; Zheng, J. Hepatic Stellate Cell: A Potential Target for Hepatocellular Carcinoma. *Curr. Mol. Pharmacol.* **2020**, *13*, 261–272. [[CrossRef](#)] [[PubMed](#)]
- Zhao, W.; Zhang, L.; Yin, Z.; Su, W.; Ren, G.; Zhou, C.; You, J.; Fan, J.; Wang, X. Activated hepatic stellate cells promote hepatocellular carcinoma development in immunocompetent mice. *Int. J. Cancer* **2011**, *129*, 2651–2661. [[CrossRef](#)] [[PubMed](#)]
- Waldman, A.D.; Fritz, J.M.; Lenardo, M.J. A guide to cancer immunotherapy: From T cell basic science to clinical practice. *Nat. Rev. Immunol.* **2020**, *20*, 651–668. [[CrossRef](#)]
- Zheng, X.; Jin, W.; Wang, S.; Ding, H. Progression on the Roles and Mechanisms of Tumor-Infiltrating T Lymphocytes in Patients With Hepatocellular Carcinoma. *Front. Immunol.* **2021**, *12*, 1480. [[CrossRef](#)]

19. Xu, Y.; Huang, Y.; Xu, W.; Zheng, X.; Yi, X.; Huang, L.; Wang, Y.; Wu, K. Activated Hepatic Stellate Cells (HSCs) Exert Immunosuppressive Effects in Hepatocellular Carcinoma by Producing Complement C3. *OncoTargets Ther.* **2020**, *ume 13*, 1497–1505. [[CrossRef](#)]
20. Charles, R.; Chou, H.-S.; Wang, L.; Fung, J.; Lu, L.; Qian, S. Human Hepatic Stellate Cells Inhibit T-Cell Response Through B7-H1 Pathway. *Transplantation* **2013**, *96*, 17–24. [[CrossRef](#)]
21. Schildberg, F.A.; Wojtalla, A.; Siegmund, S.V.; Endl, E.; Diehl, L.; Abdullah, Z.; Kurts, C.; Knolle, P.A. Murine hepatic stellate cells veto CD8 T cell activation by a CD54-dependent mechanism. *Hepatology* **2011**, *54*, 262–272. [[CrossRef](#)] [[PubMed](#)]
22. Zhao, W.; Su, W.; Kuang, P.; Zhang, L.; Liu, J.; Yin, Z.; Wang, X. The role of hepatic stellate cells in the regulation of T-cell function and the promotion of hepatocellular carcinoma. *Int. J. Oncol.* **2012**, *41*, 457–464. [[CrossRef](#)] [[PubMed](#)]
23. Li, X.; Su, Y.; Hua, X.; Xie, C.; Liu, J.; Huang, Y.; Zhou, L.; Zhang, M.; Li, X.; Gao, Z. Levels of hepatic Th17 cells and regulatory T cells upregulated by hepatic stellate cells in advanced HBV-related liver fibrosis. *J. Transl. Med.* **2017**, *15*, 1–11. [[CrossRef](#)] [[PubMed](#)]
24. Cheng, J.-N.; Yuan, Y.-X.; Zhu, B.; Jia, Q. Myeloid-Derived Suppressor Cells: A Multifaceted Accomplice in Tumor Progression. *Front. Cell Dev. Biol.* **2021**, *9*. [[CrossRef](#)]
25. Xu, Y. Activated hepatic stellate cells promote liver cancer by induction of myeloid-derived suppressor cells through cyclooxygenase-2. *Oncotarget* **2016**, *7*, 8866–8878. [[CrossRef](#)]
26. Lee, H.L.; Jang, J.W.; Lee, S.W.; Yoo, S.H.; Kwon, J.H.; Nam, S.W.; Bae, S.H.; Choi, J.Y.; Han, N.I.; Yoon, S.K. Inflammatory cytokines and change of Th1/Th2 balance as prognostic indicators for hepatocellular carcinoma in patients treated with transarterial chemoembolization. *Sci. Rep.* **2019**, *9*, 3260. [[CrossRef](#)]
27. Li, J.; Lau, G.K.-K.; Chen, P.L.; Dong, S.-S.; Lan, H.Y.; Huang, X.-R.; Li, Y.; Luk, J.; Yuan, Y.; Guan, X.-Y. Interleukin 17A Promotes Hepatocellular Carcinoma Metastasis via NF- $\kappa$ B Induced Matrix Metalloproteinases 2 and 9 Expression. *PLoS ONE* **2011**, *6*, e21816. [[CrossRef](#)]
28. Liao, R.; Sun, J.; Wu, H.; Yi, Y.; Wang, J.-X.; He, H.-W.; Cai, X.-Y.; Zhou, J.; Cheng, Y.-F.; Fan, J.; et al. High expression of IL-17 and IL-17RE associate with poor prognosis of hepatocellular carcinoma. *J. Exp. Clin. Cancer Res.* **2013**, *32*, 3–11. [[CrossRef](#)]
29. Ichikawa, S.; Mucida, D.; Tyznik, A.J.; Kronenberg, M.; Cheroutre, H. Hepatic Stellate Cells Function as Regulatory Bystanders. *J. Immunol.* **2011**, *186*, 5549–5555. [[CrossRef](#)]
30. Ricketts, T.D.; Prieto-Dominguez, N.; Gowda, P.S.; Ubil, E. Mechanisms of Macrophage Plasticity in the Tumor Environment: Manipulating Activation State to Improve Outcomes. *Front. Immunol.* **2021**, *12*. [[CrossRef](#)]
31. Braga, T.T.; Agudelo, J.S.H.; Camara, N.O.S. Macrophages During the Fibrotic Process: M2 as Friend and Foe. *Front. Immunol.* **2015**, *6*, 602. [[CrossRef](#)]
32. Wanderley, C.W.; Colón, D.F.; Luiz, J.P.M.; Oliveira, F.F.; Viacava, P.R.; Leite, C.A.; Pereira, J.A.; Silva, C.M.; Silva, C.R.; Silva, R.L.; et al. Paclitaxel reduces tumor growth by reprogramming tumor-associated macrophages to an M1- profile in a TLR4-dependent manner. *Cancer Res.* **2018**, *78*, 5891–5900. [[CrossRef](#)] [[PubMed](#)]
33. Liu, G.; Yin, L.; Ouyang, X.; Zeng, K.; Xiao, Y.; Li, Y. M2 Macrophages Promote HCC Cells Invasion and Migration via miR-149-5p/MMP9 Signaling. *J. Cancer* **2020**, *11*, 1277–1287. [[CrossRef](#)] [[PubMed](#)]
34. Yeung, O.W.; Lo, C.-M.; Ling, C.-C.; Qi, X.; Geng, W.; Li, C.-X.; Ng, K.T.; Forbes, S.J.; Guan, X.-Y.; Poon, R.T.; et al. Alternatively activated (M2) macrophages promote tumour growth and invasiveness in hepatocellular carcinoma. *J. Hepatol.* **2015**, *62*, 607–616. [[CrossRef](#)]
35. Xi, S.; Zheng, X.; Li, X.; Jiang, Y.; Wu, Y.; Gong, J.; Jie, Y.; Li, Z.; Cao, J.; Sha, L.; et al. Activated Hepatic Stellate Cells Induce Infiltration and Formation of CD163+ Macrophages via CCL2/CCR2 Pathway. *Front. Med.* **2021**, *8*, 627927. [[CrossRef](#)]
36. Wang, C.; Ma, C.; Gong, L.; Guo, Y.; Fu, K.; Zhang, Y.; Zhou, H.; Li, Y. Macrophage Polarization and Its Role in Liver Disease. *Front. Immunol.* **2021**, *12*, 5381. [[CrossRef](#)] [[PubMed](#)]
37. Yu, M.; Li, Z. Natural killer cells in hepatocellular carcinoma: Current status and perspectives for future immunotherapeutic approaches. *Front. Med.* **2017**, *11*, 509–521. [[CrossRef](#)]
38. Mossanen, J.C.; Tacke, F. Role of lymphocytes in liver cancer. *OncoImmunology* **2013**, *2*, e26468. [[CrossRef](#)]
39. Glässner, A.; Eisenhardt, M.; Krämer, B.; Körner, C.; Coenen, M.; Sauerbruch, T.; Spengler, U.; Nattermann, J. NK cells from HCV-infected patients effectively induce apoptosis of activated primary human hepatic stellate cells in a TRAIL-, FasL- and NKG2D-dependent manner. *Lab. Investig.* **2012**, *92*, 967–977. [[CrossRef](#)]
40. Radaeva, S.; Sun, R.; Jaruga, B.; Nguyen, V.T.; Tian, Z.; Gao, B. Natural Killer Cells Ameliorate Liver Fibrosis by Killing Activated Stellate Cells in NKG2D-Dependent and Tumor Necrosis Factor-Related Apoptosis-Inducing Ligand-Dependent Manners. *Gastroenterology* **2006**, *130*, 435–452. [[CrossRef](#)]
41. Xia, T.-H. Tumor-specific hepatic stellate cells (tHSCs) induces DlgR2 expression in dendritic cells to inhibit T cells. *Oncotarget* **2017**, *8*, 55084–55093. Available online: [www.impactjournals.com/oncotarget/](http://www.impactjournals.com/oncotarget/) (accessed on 15 August 2017). [[CrossRef](#)]
42. HCC Monitor. New evidence supports a key role of the immune system in HCC, HCC monitor. *Target. Oncol.* **2016**, *2*, 3. Available online: <https://www.targetedonc.com> (accessed on 15 August 2017).
43. Zois, C.D.; Baltayiannis, G.H.; Karayiannis, P.; Tsianos, E.V. Systematic review: Hepatic fibrosis-regression with therapy. *Aliment. Pharmacol. Ther.* **2008**, *28*, 1175–1187. [[CrossRef](#)] [[PubMed](#)]

44. Arriazu, E.; de Galarreta, M.R.; Cubero, F.J.; Varela-Rey, M.; de Obanos, M.P.P.; Leung, T.M.; Lopategi, A.; Benedicto, A.; Abraham-Enachescu, I.; Nieto, N. Extracellular Matrix and Liver Disease. *Antioxidants Redox Signal.* **2014**, *21*, 1078–1097. [[CrossRef](#)] [[PubMed](#)]
45. Jung, Y.K.; Yim, H.J. Reversal of liver cirrhosis: Current evidence and expectations. *Korean J. Intern. Med.* **2017**, *32*, 213–228. [[CrossRef](#)]
46. Sun, M.; Kisseleva, Y. Reversibility of liver fibrosis. In *Clinics and Research in Hepatology and Gastroenterology*; Elsevier Masson SAS: Amsterdam, The Netherlands, 2015; pp. S60–S63. [[CrossRef](#)]
47. Kisseleva, T.; Brenner, D. Hepatic stellate cells and the reversal of fibrosis. *J. Gastroenterol. Hepatol.* **2006**, *21*, S84–S87. [[CrossRef](#)]
48. Arthur, M.J.; Mann, D.A.; Iredale, J.P. Tissue inhibitors of metalloproteinases, hepatic stellate cells and liver fibrosis. *J. Gastroenterol. Hepatol.* **1998**, *13*, S33–S38. [[CrossRef](#)]
49. Zhang, Y.; Li, Y.; Mu, T.; Tong, N.; Cheng, P. Hepatic stellate cells specific liposomes with the Toll-like receptor 4 shRNA attenuates liver fibrosis. *J. Cell. Mol. Med.* **2021**, *25*, 1299–1313. [[CrossRef](#)]
50. Arab, J.P.; Cabrera, D.; Sehrawat, T.S.; Jalan-Sakrikar, N.; Verma, V.K.; Simonetto, D.; Cao, S.; Yaqoob, U.; Leon, J.; Freire, M.; et al. Hepatic stellate cell activation promotes alcohol-induced steatohepatitis through Igfbp3 and SerpinA12. *J. Hepatol.* **2020**, *73*, 149–160. [[CrossRef](#)]
51. Hartland, S.N.; Murphy, F.; Aucott, R.L.; Abergel, A.; Zhou, X.; Waung, J.; Patel, N.; Bradshaw, C.; Collins, J.; Mann, D.; et al. Active matrix metalloproteinase-2 promotes apoptosis of hepatic stellate cells via the cleavage of cellular N-cadherin. *Liver Int.* **2009**, *29*, 966–978. [[CrossRef](#)]
52. Zheng, X.; Liu, W.; Xiang, J.; Liu, P.; Ke, M.; Wang, B.; Lv, Y. Collagen I promotes hepatocellular carcinoma cell proliferation by regulating integrin  $\beta$ 1/FAK signaling pathway in nonalcoholic fatty liver. *Oncotarget* **2017**, *8*, 95586–95595. Available online: [www.impactjournals.com/oncotarget](http://www.impactjournals.com/oncotarget) (accessed on 10 November 2017).
53. Zhang, R.; Ma, M.; Lin, X.-H.; Liu, H.-H.; Chen, J.; Chen, J.; Gao, D.-M.; Cui, J.-F.; Ren, Z.-G.; Chen, R.-X. Extracellular matrix collagen I promotes the tumor progression of residual hepatocellular carcinoma after heat treatment. *BMC Cancer* **2018**, *18*, 901. [[CrossRef](#)] [[PubMed](#)]
54. Ma, H.-P.; Chang, H.-L.; Bamodu, O.A.; Yadav, V.K.; Huang, T.-Y.; Wu, A.T.H.; Yeh, C.-T.; Tsai, S.-H.; Lee, W.-H. Collagen 1A1 (COL1A1) Is a Reliable Biomarker and Putative Therapeutic Target for Hepatocellular Carcinogenesis and Metastasis. *Cancers* **2019**, *11*, 786. [[CrossRef](#)]
55. Yu, F.; Lin, Z.; Zheng, J.; Gao, S.; Lu, Z.; Dong, P. Suppression of collagen synthesis by Dicer gene silencing in hepatic stellate cells. *Mol. Med. Rep.* **2014**, *9*, 707–714. [[CrossRef](#)] [[PubMed](#)]
56. Giannelli, G.; Azzariti, A.; Fransvea, E.; Porcelli, L.; Antonaci, S.; Paradiso, A. Laminin-5 offsets the efficacy of gefitinib ('Iressa') in hepatocellular carcinoma cells. *Br. J. Cancer* **2004**, *91*, 1964–1969. [[CrossRef](#)] [[PubMed](#)]
57. Azzariti, A.; Mancarella, S.; Porcelli, L.; Quatrone, A.E.; Caligiuri, A.; Lupu, L.; Giannelli, G. Hepatic Stellate Cells Induce Hepatocellular Carcinoma Cell Resistance to Sorafenib Through the Laminin-332/ $\alpha$ 3 Integrin Axis Recovery of Focal Adhesion Kinase Ubiquitination. *Hepatology* **2016**, *64*, 2103–2117. [[CrossRef](#)]
58. Santamato, A.; Fransvea, E.; Dituri, F.; Caligiuri, A.; Quaranta, M.; Niimi, T.; Pinzani, M.; Antonaci, S.; Giannelli, G. Hepatic stellate cells stimulate HCC cell migration via laminin-5 production. *Clin. Sci.* **2011**, *121*, 159–168. [[CrossRef](#)]
59. Wu, X.Z.; Chen, D.; Xie, G.R. Extracellular matrix remodeling in hepatocellular carcinoma: Effects of soil on seed? *Med. Hypotheses* **2006**, *66*, 1115–1120. [[CrossRef](#)]
60. Qu, K.; Yan, Z.; Wu, Y.; Chen, Y.; Qu, P.; Xu, X.; Yuan, P.; Huang, X.; Xing, J.; Zhang, H.; et al. Transarterial chemoembolization aggravated peritumoral fibrosis via hypoxia-inducible factor-1 $\alpha$  dependent pathway in hepatocellular carcinoma. *J. Gastroenterol. Hepatol.* **2015**, *30*, 925–932. [[CrossRef](#)]
61. Wang, Y.; Xiong, B.; Liang, B.; Zhao, H.; Li, H.; Qian, J.; Liang, H.-M.; Feng, G.-S.; Zheng, C.-S. Hepatic Parenchymal Changes following Transcatheter Embolization and Chemoembolization in a Rabbit Tumor Model. *PLoS ONE* **2013**, *8*, e70757. [[CrossRef](#)]
62. Das, D.; Fayazzadeh, E.; Li, X.; Koirala, N.; Wadera, A.; Lang, M.; Zernic, M.; Panick, C.; Nesbitt, P.; McLennan, G. Quiescent hepatic stellate cells induce toxicity and sensitivity to doxorubicin in cancer cells through a caspase-independent cell death pathway: Central role of apoptosis-inducing factor. *J. Cell. Physiol.* **2020**, *235*, 6167–6182. [[CrossRef](#)]
63. Sung, Y.-C.; Liu, Y.-C.; Chao, P.-H.; Chang, C.-C.; Jin, P.-R.; Lin, T.-T.; Lin, J.-A.; Cheng, H.-T.; Wang, J.; Lai, C.P.; et al. Combined delivery of sorafenib and a MEK inhibitor using CXCR4-targeted nanoparticles reduces hepatic fibrosis and prevents tumor development. *Theranostics* **2018**, *8*, 894–905. [[CrossRef](#)] [[PubMed](#)]
64. Chen, W.; Wu, J.; Shi, H.; Wang, Z.; Zhang, G.; Cao, Y.; Jiang, C.; Ding, Y. Hepatic Stellate Cell Coculture Enables Sorafenib Resistance in Huh7 Cells through HGF/c-Met/Akt and Jak2/Stat3 Pathways. *BioMed Res. Int.* **2014**, *2014*, 1–10. [[CrossRef](#)] [[PubMed](#)]
65. Barker, H.E.; Paget, J.T.E.; Khan, A.; Harrington, K. The tumour microenvironment after radiotherapy: Mechanisms of resistance and recurrence. *Nat. Rev. Cancer* **2015**, *15*, 409–425. [[CrossRef](#)]
66. Sempoux, C.; Horsmans, Y.; Geubel, A.; Fraikin, J.; Van Beers, B.E.; Gigot, J.; Rahier, J. Severe radiation-induced liver disease following localized radiation therapy for biliopancreatic carcinoma: Activation of hepatic stellate cells as an early event. *Hepatology* **1997**, *26*, 128–134. [[CrossRef](#)] [[PubMed](#)]

67. Shen, X.; Zhao, J.; Wang, Q.; Chen, P.; Hong, Y.; He, X.; Chen, D.; Liu, H.; Wang, Y.; Cai, X. The Invasive Potential of Hepatoma Cells Induced by Radiotherapy is Related to the Activation of Stellate Cells and Could be Inhibited by EGCG Through the TLR4 Signaling Pathway. *Radiat. Res.* **2022**, *197*, 365–375. [[CrossRef](#)]
68. Kang, T.W.; Lim, H.K.; Cha, D.I. Aggressive tumor recurrence after radiofrequency ablation for hepatocellular carcinoma. *Clin. Mol. Hepatol.* **2017**, *23*, 95–101. [[CrossRef](#)]
69. Rozenblum, N.; Zeira, E.; Bulvik, B.; Gourevitch, S.; Yotvat, H.; Galun, E.; Goldberg, S.N. Radiofrequency Ablation: Inflammatory Changes in the Periablative Zone Can Induce Global Organ Effects, including Liver Regeneration. *Radiology* **2015**, *276*, 416–425. [[CrossRef](#)]
70. Cheng, R.; Xu, H.; Hong, Y. miR221 Regulates TGF- $\beta$ 1-induced HSC activation through Inhibiting Autophagy by directly targeting LAMP2. *Mol Med Rep.* **2021**, *24*, 5. [[CrossRef](#)]
71. Kelley, R.K.; Gane, E.; Assenat, E.; Siebler, J.; Galle, P.R.; Merle, P.; Hourmand, I.O.; Cleverly, A.; Zhao, Y.; Gueorguieva, I.; et al. A Phase 2 Study of Galunisertib (TGF- $\beta$ 1 Receptor Type I Inhibitor) and Sorafenib in Patients With Advanced Hepatocellular Carcinoma. *Clin. Transl. Gastroenterol.* **2019**, *10*, e00056. [[CrossRef](#)]
72. El-Mezayen, N.S.; El-Hadidy, W.F.; El-Refaie, W.M.; Shalaby, T.; Khattab, M.M.; El-Khatib, A.S. Hepatic stellate cell-targeted imatinib nanomedicine versus conventional imatinib: A novel strategy with potent efficacy in experimental liver fibrosis. *J. Control. Release* **2017**, *266*, 226–237. [[CrossRef](#)]
73. Li, Z.; Wang, F.; Li, Y.; Wang, X.; Lu, Q.; Wang, D.; Qi, C.; Li, C.; Li, Z.; Lian, B.; et al. Combined anti-hepatocellular carcinoma therapy inhibit drug-resistance and metastasis via targeting “substance P-hepatic stellate cells-hepatocellular carcinoma” axis. *Biomaterials* **2021**, *276*, 121003. [[CrossRef](#)] [[PubMed](#)]
74. Sun, L.; Wang, Y.; Wang, X.; Navarro-Corcuera, A.; Ilyas, S.; Jalan-Sakrikar, N.; Gan, C.; Tu, X.; Shi, Y.; Tu, K.; et al. PD-L1 promotes myofibroblastic activation of hepatic stellate cells by distinct mechanisms selective for TGF- $\beta$  receptor I versus II. *Cell Rep.* **2022**, *38*, 110349. [[CrossRef](#)] [[PubMed](#)]
75. Yau, T.; Hsu, C.; Kim, T.-Y.; Choo, S.-P.; Kang, Y.-K.; Hou, M.-M.; Numata, K.; Yeo, W.; Chopra, A.; Ikeda, M.; et al. Nivolumab in advanced hepatocellular carcinoma: Sorafenib-experienced Asian cohort analysis. *J. Hepatol.* **2019**, *71*, 543–552. [[CrossRef](#)] [[PubMed](#)]
76. Zisser, A.; Ipsen, D.; Tveden-Nyborg, P. Hepatic Stellate Cell Activation and Inactivation in NASH-Fibrosis—Roles as Putative Treatment Targets? *Biomedicines* **2021**, *9*, 365. [[CrossRef](#)] [[PubMed](#)]
77. Filliol, A.; Saito, Y.; Nair, A.; Dapito, D.H.; Yu, L.-X.; Ravichandra, A.; Bhattacharjee, S.; Affo, S.; Fujiwara, N.; Su, H.; et al. Opposing roles of hepatic stellate cell subpopulations in hepatocarcinogenesis. *Nature* **2022**, *610*, 356–365. [[CrossRef](#)] [[PubMed](#)]
78. Kisseleva, T.; Brenner, D. Molecular and cellular mechanisms of liver fibrosis and its regression. *Nat. Rev. Gastroenterol. Hepatol.* **2021**, *18*, 151–166. [[CrossRef](#)]
79. Chen, Z.; Jain, A.; Liu, H.; Zhao, Z.; Cheng, K. Targeted Drug Delivery to Hepatic Stellate Cells for the Treatment of Liver Fibrosis. *J. Pharmacol. Exp. Ther.* **2019**, *370*, 695–702. [[CrossRef](#)]
80. Zabielska-Koczywaś, K.; Lechowski, R. The Use of Liposomes and Nanoparticles as Drug Delivery Systems to Improve Cancer Treatment in Dogs and Cats. *Molecules* **2017**, *22*, 2167. [[CrossRef](#)]
81. Luo, J.; Zhang, P.; Zhao, T.; Jia, M.; Yin, P.; Li, W.; Zhang, Z.-R.; Fu, Y.; Gong, T. Golgi Apparatus-Targeted Chondroitin-Modified Nanomicelles Suppress Hepatic Stellate Cell Activation for the Management of Liver Fibrosis. *ACS Nano* **2019**, *13*, 3910–3923. [[CrossRef](#)]