



Review

Estrogens, Estrogen Receptors and Tumor Microenvironment in Ovarian Cancer

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Abstract: Ovarian cancer is one of the most common cancers in women and the most concerning issues in gynecological oncology in recent years. It is postulated that many factors may contribute to the development of ovarian cancer, including hormonal imbalance. Estrogens are a group of hormones that have an important role both in physiological and pathological processes. In ovarian cancer, they may regulate proliferation, invasiveness and epithelial to mesenchymal transition. Estrogen signaling also takes part in the regulation of the biology of the tumor microenvironment. This review summarizes the information connected with estrogen receptors, estrogens and their association with a tumor microenvironment. Moreover, this review also includes information about the changes in estrogen receptor expression upon exposition to various environmental chemicals.

Keywords: estrogens; estrogen receptors; ovarian cancer; proliferation; tumor microenvironment



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1. Introduction

Ovarian cancer (OC) is one of the most common cancers in women in developing countries. In recent years, it has been postulated that OC is one of the most concerning issues in gynecological oncology, causing more than 200,000 deaths worldwide in 2020 [1]. Due to a late diagnosis, treatment is challenging and the 5-year survival rate is still under 50%. As in every cancer, when OC is diagnosed early, most women may be treated effectively using standard therapeutic approaches. Unfortunately, once OC spreads to the pelvic and abdominal organs or/and beyond the peritoneal cavity, treatment becomes much more difficult. OCs are a molecularly truly diverse group of cancers. They may be classified based on their origin into epithelial, stromal and germ cells. Most ovarian neoplasms are of an epithelial origin. Another classification, based on histology and carcinogenesis, includes high-grade and low-grade ovarian neoplasms. The stage of ovarian cancer can also be presented on the FIGO scale, where stage I is the least advanced disease and stage IV is the most advanced disease [2]. Treatment of ovarian cancer mostly depends on surgery and chemotherapy treatment. Recently, it has been shown that usage of both 4-Hydroxy-Tamoxifen (4-OHT) and Gatifotuzumab has better outcomes, and may increase the efficacy of treatment [3]. This discovery seems to be important since the authors noticed that there is an association between TA-MUC1 and estrogen receptors (ERs) [3]. Although many research studies have been carried out on ovarian cancer, a detailed molecular mechanism has not been revealed yet. As far as we know, the expression of GPER1 in ovarian cancer tissues is not clear. To achieve a better understanding, and then better treatment, it is necessary to have a better knowledge about OC, especially in the case of both classical and non-classical ERs and tumor microenvironment (TME). Therefore, the purpose of this review is to summarize recent literature concerning ERs, the involvement of TME regarding them and the progression of OC.

2. Estrogens, ER Expression and Signaling Pathways

It is known that estrogens modulate both physiological and pathophysiological processes [4]. Nevertheless, they also have a vital role in the functioning of the cardiovascular and immune systems [5,6]. The ovaries are the central reproductive organs of women that produce hormones such as testosterone, progesterone and estrogen [7]. Hormones produced by the ovaries play a vital role not only in the ovaries but also in the endometrium, where they stimulate changes in the endometrium during periodic changes [8]. Therefore, proper functioning of the ovaries is necessary for, among others, maintaining pregnancy. Among estrogens, we can distinguish estrone (E1), 17 β -estradiol (E2) and estriol (E3), of which E2 is the most dominant estrogen [4]. It was observed that estrogens may stimulate the growth of ovarian cancer cells in vitro [9]. For example, Matsumura et al. showed that E2 stimulates the proliferation of Caov-3 and OVCAR3 cells via Akt/ERK cascades [10]. Interestingly, it was noticed that hormone replacement therapy (HRT) in postmenopausal women is associated with increased incidence and mortality of OC [11]. Therefore, the question was whether physicians should treat women after resection of ovaries with HRT. Fortunately, Eeles et al. stated that women after surgical treatment of OC can safely take HRT, to help them overcome inconvenience associated with the resection of the ovaries and lack of production of hormones [12]. It was also presented that HRT administration is related to the reduced risk of colorectal cancer (CRC) incidence [13]. In turn, it was observed that HRT increases the recurrence of breast cancer (BC) with positive hormone receptors [14]. Side effects of HRT were also linked with cardiovascular diseases [15]. Nevertheless, the statement regarding HRT and coronary heart disease has changed in recent years [16]. Although the role of HRT and its negative impact on women's health have been discussed for several decades, no clear conclusions have yet been drawn. Recent results are hopeful, suggesting that HRT does not have as many negatives as initially thought, and significantly increases the quality of life. Cellular signaling of estrogens is mediated via ERs. ERs are a family of transcription factors that control the biological function of estrogens via regulating gene transcription through estrogen response elements (EREs). Estrogens may modulate the biology of cancer cells by affecting processes like cell proliferation, invasion, apoptosis, cell cycle and inflammation (Figure 1).

Among ERs, we can distinguish both classical (ER α and ER β) and non-classical (G-protein-coupled estrogen receptor 1 (GPER1)) receptors. Research on the expression of classical ERs seems consistent and we can come to conclusions. The ratio between estrogen receptors (ER α and ER β) has a significant role in ovarian cancer development [17]. Together with the progression of ovarian cancer, the expression of ER α increases, while the expression of ER β decreases. Nevertheless, in the case of GPER1, it is difficult to draw clear conclusions. On the one hand, some studies reported that GPER1 has a potentially tumor-promoting role in OC and may predict lower patient survival [18,19]. Its expression was also observed to be correlated with the histological grade of the OC. On the other hand, others have not observed any correlation between GPER1 expression and clinical stage and/or patient survival or even GPER1 expression being correlated with higher survival [20–22]. Because studies are not consistent, it seems that more studies should be performed to obtain confirmation. ER α and ER β are encoded by *ESR1* and *ESR2* genes, respectively. *ESR1* is located on chromosome 6 (6q25.1), while *ESR2* is on chromosome 14 (14q23.2) [23]. Both estrogen receptors were found to be located in the cell membrane, but they were also found in cytoplasmic organelles, including mitochondria and the endoplasmic reticulum [24]. ERs consist of six domains: A/B, C, D and E/F starting from amino and ending at carboxyl terminals (Figure 2). The wild type of ER α and ER β protein has a mass of 66 and 59 kDa, respectively. The A/B domain is responsible for the specificity of the receptor, the C domain is the DNA-binding site and the D domain connects the C domain with the E domain and stabilizes the binding to the DNA in the C domain. The E/F domains are responsible for binding to the ligand. AF1 and AF2 are mandatory for the activation of both ER α and ER β . GPER1, also known as GPR30, is a non-classical estrogen receptor encoded by the *GPER* gene that is located on chromosome 7 (7p22.3). Normally, GPER1 is present

in the endoplasmic reticulum; however, its presence was also observed in the plasma membrane [25]. It was proved that GPER1 binds with high affinity with estradiol. After binding, GPER1 is responsible for the rapid activation of numerous signaling pathways. It promotes the production of cAMP and activation of the epidermal growth factor receptor (EGFR) and it affects signaling pathways like PI3K/Akt and ERK/MAPK [25,26].

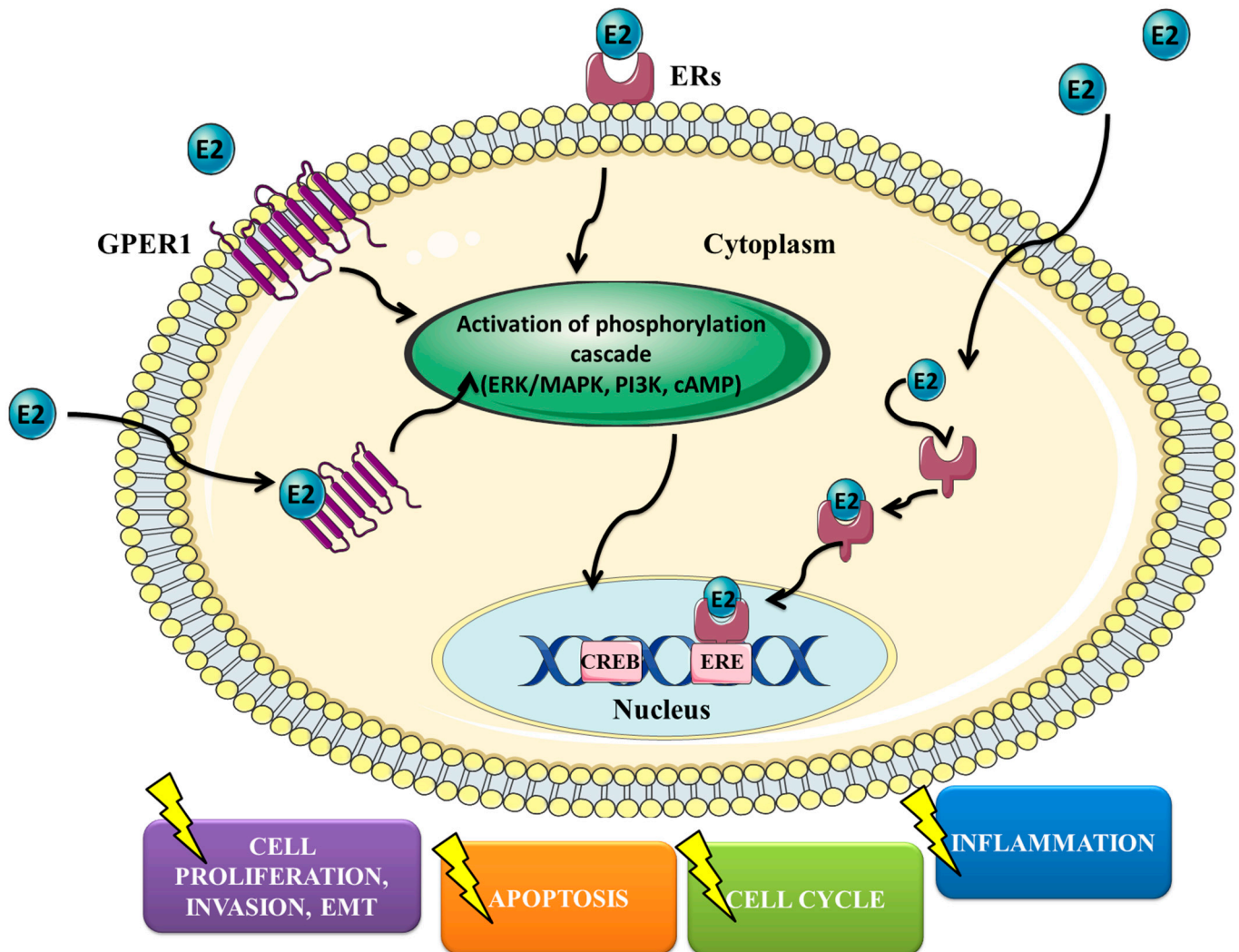


Figure 1. Main mechanisms of estrogen-induced change in the biology of ovarian cancer cells. E2—estradiol, ERs—estrogen receptors, EMT—epithelial to mesenchymal transition, GPER1—G-protein-coupled estrogen receptor 1, ERE—estrogen response element, CREB—cAMP-response-element-binding protein.

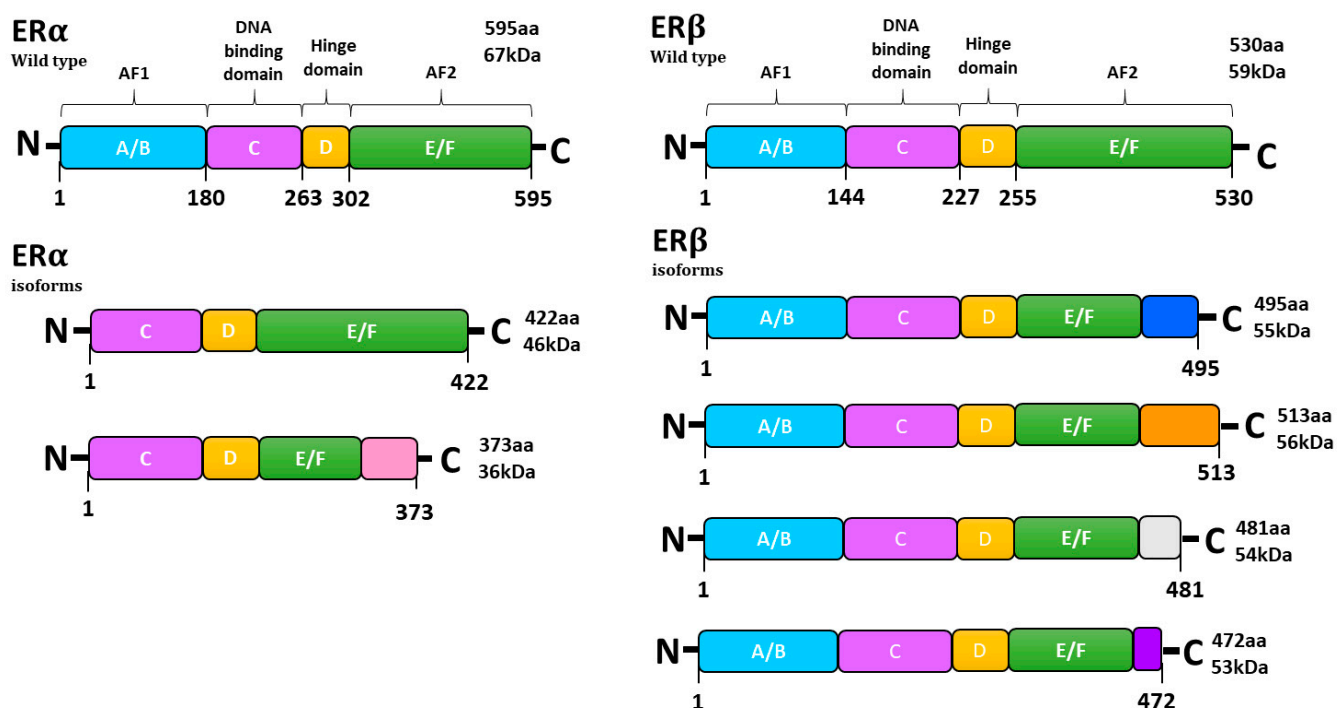


Figure 2. Structure of estrogen receptor α and estrogen receptor β and their isoforms. ER α —estrogen receptor α , ER β —estrogen receptor β , AF1—activation function 1, AF2—activation function 1.

3. Ovarian Cancer Proliferation, EMT and Cell Invasiveness

ER α was reported to modulate the expression of genes associated with cell proliferation and tumor growth in epithelial ovarian cancer [27]. In turn, Bossard et al. showed that mice with increased expression of ER β have reduced tumor growth [28]. Moreover, increased expression of ER β in BG-1 cells significantly decreased cell proliferation stimulated with estradiol [28]. Similar results were obtained in other cell lines, like breast and prostate cancer, where transfection of ER β decreased motility and invasion of cells [27]. Therefore, it is believed that ER α has pro-cancerous abilities, while ER β is anti-cancerous, which underlines that this disproportion may have a crucial role in ovarian carcinogenesis [28–30]. It is well known that estrogens stimulate proliferation, while anti-estrogen drugs abolish the proliferation of ovarian cancer both in vitro and in vivo [27]. Their ability to enhance the proliferation of neoplastic cells along molecular pathways may occur in a receptor-dependent and receptor-independent manner [31]. The first way is strongly associated with the ER α receptor. After binding estrogen with ER α , a signal cascade is triggered, which causes increased transcription of genes associated with cancer progression such as c-fos, c-myc, growth factors and cyclins that regulate cell cycle progression [31]. Migration of ovarian cancer cells and epithelial to mesenchymal transition (EMT) may be also stimulated with estrogens acting via ER α by decreasing the expression of E-cadherin and increasing EMT-related transcription factors: Snail and Slug [32]. It was also postulated that estrogens may affect adhesion to extracellular matrix proteins via ER α [30]. Increased expression of ER β resulted in a decreased number of cells in the S phase and an increase in G2/M in a BG-1 cell line. Regulation of cell cycle progression was also seen through cyclin D1 and A2 modulation. Moreover, ER β was found to modulate total retinoblastoma (Rb), its phosphorylated form, pAkt, cyclin D1 and A2 [28]. Also, ER β was proposed to modulate the expression and activity of ER α ; therefore, its clinical utilization may be worth it [28]. In another study, in a different cell line, the antitumoral effect of ER β , however, was independent of ER α and estradiol [28]. The discrepancies between the cell lines may be due, among others, to the different mutations that occur in them. In OVCAR3 and OAW-42 cells, the usage of four different ER β agonists resulted in decreased proliferation [33]. According

to the previous results, the knockdown of ER β stimulated the growth of OAW-42 cells [33]. Schöler-Toprak et al. also showed that the acting of the ER β agonists is related to β -catenin (CTNNB1) and amyloid β precursor protein (APP) in OAW-42 cells. In SKOV3 cells, increased expression of ER β resulted in decreased growth and migration [34]. The authors observed that these effects were associated with the modulation of cyclin-dependent kinase inhibitor p21 (WAF1), cyclin A2 transcripts and fibulin 1c [34]. Banerjee et al. showed that activation of ER β with a newly developed agonist (OSU-ERb-12) abolishes the ability to grow and migrate and invasiveness of ovarian cancer cells [35]. Because EMT is strongly associated with the migration and invasion of cancer cells, an observation considering the role of ER α and ER β has also been made. It was shown that increased expression of ER β results in increased E-cadherin (E-cad) and decreased Snail expression [35]. At the same time, it was proved that ER β agonists may downregulate stemness markers like SOX2, Oct4 and Nanog. Non-genomic effects that may stimulate cell proliferation rely on binding to GPER and thus induce extracellular-signal-regulated kinase (ERK), phosphoinositide 3-kinase (PI3K) and epidermal growth factor (EGFR) signaling [31]. Modulation of GPER1 has been shown to affect ovarian cancer cell growth. Yan et al. showed that the selective GPER-1 agonist (G-1) in a dose of 10 nM increases migration and invasiveness of an ER α -negative cell line via promotive production and activation of MMP-9 [36]. Knockdown of GPER-1 agreed with previous results—it resulted in a reduction in migration and invasion [36]. Changes in invasion, proliferation and migration also affected another cell line, SKOV3, where GPER1 knockdown resulted in their decrease and was also associated with changes in the expression and activity of MMP-2 and MMP-9 [37]. Additionally, Yan et al. presented that GPER1 may modulate the expression of ER α and ER β [37]. In turn, it was also shown that usage of G-1 may decrease proliferation and induce G2/M cell cycle arrest [22]. It was presented that G-1 promotes the activation of mitotic-promoting factor (MPF) and phosphorylation of nuclear mitotic apparatus protein 1 (NuMA) [38]. Treatment with G-1 also causes a decrease in the number of cells in the G1 phase, an increase in prophase and a decline in metaphase, anaphase, telophase and cytokinesis. Wang et al. showed that G1 inhibits the proliferation of SKOV3 and IGROV-1 cell lines in a dose-dependent manner and that it disrupts the morphology of these cells [38]. The differences between these works may be associated with different doses used for experiments.

4. Interaction of Environmental Chemicals, Estrogen Receptors and Ovarian Cancer Proliferation

Some substances present in the environment as pollution, food/cosmetic additives and many others may have an impact on the hormone-dependent tissues, due to the structural and functional similarity to naturally occurring estrogens. Substances like these are called xenoestrogens and may directly and/or indirectly bind with ERs and thus change the biology of cells. Some of these compounds may be of a natural origin, others synthetic. Genistein belongs to a subgroup of isoflavones (the flavonoid family), at physiological concentrations, activates the nuclear estrogen receptors ER α and ER β and affects TGF β signaling pathways [39]. Genistein is also the most common natural substance with estrogen activity. Chan et al. showed that genistein and daidzein (another member of the isoflavone family) suppress the proliferation, motility and invasiveness of ovarian cancer cells via modulation of the expression of ER β [40]. Moreover, they also observed that changes in the behavior of cells were associated with increased expression of p21 and E-cad, and with decreased expression of vimentin (VIM). The modulation of PI3K/AKT signaling was also described [40]. A similar observation has been made for resveratrol, a naturally occurring phytoestrogen, with downregulation expression of ER α IGF-1R, p-IRS-1, p-Akt1/2/3 and cyclin D1 [41]. Sang et al. showed that bisphenol A (BPA) induces the proliferation of ovarian cancer cells via the regulation of matrix metalloproteinase-2 (MMP-2), matrix metalloproteinase-9 (MMP-9) and intercellular cell adhesion molecule-1 (IMAC-1), but the addition of an ER α inhibitor abolished this effect, suggesting that BPA promotes ovarian cancer cells via the ER α signaling pathway [42].

This statement seems to be confirmed by Sang et al. The authors showed that proliferation, migration, invasion and adhesion stimulated with BPA in OVCAR3 cells depend on the activity of ER α [42]. Hwang et al. showed that genistein can reduce BPA-stimulated proliferation in BG-1 cells [43]. Moreover, Hwang et al. also presented that the mechanism is associated with the regulation of cell cycle progression [41]. Liu et al. observed that histamine in a dose of 50 ng/mL induces the proliferation of OVCAR3 cells after 48 h by regulating the expression of both ER α and ER β [44]. The same team also observed that apigenin, a natural flavonoid found in many plant species, inhibits histamine-induced proliferation via the PI3K/AKT/mTOR pathway [44]. Similar results were obtained in other cell lines like cervical cancer or breast cancer [44,45]. The PI3K/AKT/mTOR pathway was also involved in the regulation of apoptosis and autophagy with Tanshinone I (Tan-I), an extract from the Chinese medicinal herb *Salvia miltiorrhiza* Bunge [46]. In our study, we observed that estrogenic mycotoxin (Alternariol; AOH) stimulates apoptosis in ovarian cancer cells via ER α and also modulates migration, proliferation and invasion [47]. Nevertheless, the effect on the expression of ERs in both studies has not been shown. Aconitine, a substance produced by plants, has also been described in the context of ovarian cancer. Wang et al. presented that aconitine decreased cell viability, colony formation and migration of A2780 cells. The same team also showed that treatment with aconite induces DNA damage and apoptosis in cells via regulation of the expression of ER β and also other factors connected with estrogen signaling like vascular endothelial growth factor (VEGF) and hypoxia-inducible factor 1 α (HIF1 α) [48]. Ataei et al. showed that cadmium chloride induces the proliferation of ovarian cancer cells by affecting the expression of ERs and then activation of the ERK1/2/MAPK pathway and c-jun, c-fos and foxo3a transcription factors. It was also shown that the cadmium acting via ERs may affect the expression of progesterone receptors [49]. Some compounds were also investigated taking into consideration non-classical estrogenic pathways. Hoffmann et al. showed that tetrabromobisphenol A (TBBPA) stimulates the proliferation of OVCAR3 and KGN cells via the GPR30 pathway [50]. The addition of a GPER1 antagonist reversed this effect [50]. Summary information regarding the regulation of ERs expression is presented in Table 1.

Table 1. Summarized information about compounds and their influence on ER expression in ovarian cancer cells. ERs—estrogen receptors, ER α —Estrogen receptor α , ER β —Estrogen receptor β , \uparrow —upregulation, \downarrow —downregulation.

Substance	Concentration	Time of Exposition (h)	Cell Line	ERs	Literature
Histamine	50 ng/mL	48	OVCAR3	\uparrow ER α \downarrow ER β	[44]
Aconitine	100, 200 and 400 μ g/mL	24	A2780	\uparrow ER β	[49]
Genistein	10 and 50 μ M	24	SKOV3, OVCAR3, A2780CP	\uparrow ER β	[40]
Daidzein	10 and 50 μ M	24	SKOV3, OVCAR3, A2780CP	\uparrow ER β	[40]
Bisphenol A	10 μ M	6 and 24	BG-1	\uparrow ER α	[41,43]
Cadmium Chloride	0.01 μ M	24	OVCAR3	\uparrow ER α \uparrow ER β	[51]

5. Tumor Microenvironment (TME) in the Progression of Ovarian Cancer

In recent years, non-cancerous cells constituting TME are believed to be critical mediators of tumor progression. Moreover, the importance of the interplay between tumor cells, stromal cells, immune cells and extracellular molecules in TME is emphasized as it has a profound effect on antitumor immunity and immunotherapeutic response [52]. The importance of TME is emphasized with the fact that it is chosen as a therapeutic target in cancer treatment and enjoys great interest in both research and clinical trials [53]. Nevertheless, one of the main difficulties in targeting the TME in cancer therapy is that host cells or non-cellular components of the TME may have different associations with tumor cells [53]. Therefore, further research on the TME in cancers is necessary because there is

a lack of in-depth knowledge about these cells and their interactions that results in failed therapy. As it was proved, each cancer is different in terms of cells that belong to TME; however, we can distinguish cell types that are always included in TME: cancer-associated fibroblasts (CAFs), tumor-associated macrophages (TAMs), myeloid-derived suppressor cells (MDSCs), lymphocytes T and B, natural killer cells (NK cells) and endothelial cells [52]. CAFs, TAMs and MDSCs have a crucial role in cancer cell proliferation. Estrogen signaling is also known to play an important role in the regulation of the immunological response [6]. Their role is also visible in the TME. Both ERs and aromatase, which is a key enzyme in the production of estrogens, are expressed in cells that belong to the TME [54]. For example, expression of ER α and ER β was observed in CAFs and TAMs in local TME of ovarian cancer [54]. The interaction of the cells in the organisms is mostly tissue-specific and depends on many factors. However, based on the literature survey, many of the effects stimulated by compartments of TME are mediated via PI3K/MAPK/Akt pathways, which are described as well-known ER-mediated pathways. Epithelial ovarian cancer (EOC) is in some kind unique among other solid tumors in the context of TME since tumor cells migrate from the primary tumor to create malignant ascites in the peritoneal cavity [55]. Malignant ascites also have TME and ascite-derived tumor cells occur as single floating cells or also as spheroids [55]. It is generally known that microRNAs (miRNA) are short, non-coding RNAs that regulate gene expression [56,57]. Their dysregulation has been observed in most types of cancer including ovarian cancer. Recently, it has been proposed that they also may have an influence on TME [56,57].

5.1. Cancer-Associated Fibroblasts (CAFs)

CAFs are the most predominant stromal cells that create TME. Zhang et al. observed that an increased amount of CAFs was in EOC than in benign tumors [52]. Moreover, CAFs isolated from EOC lesions were able to increase the invasion and migration of ovarian cancer cells [58,59]. Interestingly, Jin et al. showed that collapsin response mediator protein-2 (CRMP2) participates in these modulations through activation of the hypoxia-inducible factor (HIF)-1 α –glycolysis signaling pathway [60]. CAFs relieve substances like chemokines and growth factors, which then stimulate pathways associated with tumor growth and progression [61]. Thongchot et al. showed that interleukin-8 (IL-8) released from CAFs increases the migration of ovarian cancer cells [62]. Similarly, fibroblast growth factor-1 (FGF-1) released from CAFs increased the growth of SKOV3 cells via the FGFR4/MAPK/ERK pathway [63]. In turn, Zhang et al. showed that CAFs induce EMT in OC cells via the Wnt/ β -catenin pathway. Interestingly, it was also shown that progranulin (PGRN) stimulates the proliferation and invasion of ovarian cancer cells, indirectly via CAFs [64]. A similar observation has been made for TGF- β [65]. Wu et al. observed that collagen type XI alpha 1 (COL11A1) is upregulated in CAFs [66]. Moreover, they also noticed that its modulation may have an important role in the biology of OC, resulting in decreased invasiveness and tumor formation, giving hope that COL11A1 may have a key role in the future in the treatment of OC with elevated levels of TGF- β 3 [66]. Yue et al. noticed that CAFs increase the metastasis character of ovarian cancer cells via the PI3K/Akt pathway [67]. Additionally, increased expression of E2—the responsive gene—was observed in CAFs compared to normal fibroblasts [68,69]. Kim et al. presented that expression of GLIS1 (Glis Family Zinc Finger 1) is increased both at gene and protein levels in CAFs. Moreover, the knockdown of GLIS1 decreased migration and invasion of ovarian cancer cells, suggesting that this factor may have an important role in the progression of OC [70]. It was observed that miRNAs in CAFs may affect their reprogramming. In the ovarian cancer microenvironment, and more specifically in the context of CAFs, miR-214 and miR-155 have been described at low and high expression, respectively [71]. It was further demonstrated that by disrupting their expression, it was possible to reduce the growth and metastasis of ovarian cancer through loss of the CAF-like phenotype, which seems to be very important in ovarian cancer treatment [71].

5.2. Tumor-Associated Macrophages (TAMs)

It is generally known that macrophages belong to phagocytic cells and that they regulate immunological response. Nevertheless, it has also been shown that they may regulate the invasion and metastasis of cancer cells, mainly because they can create an inflammatory environment, which may stimulate mutations, growth, proliferation, metastases and many others [72]. In primary OC as well as in ascites, the main population of immune cells consists of macrophages. They may originate from embryonic yolk sacs and bone-marrow-derived monocytes. When macrophages (M0) are recruited to TME, dependent on stimuli, they differentiate into the M1 or M2 subtype [73]. M1 macrophages inhibit the progression of cancer via the secretion of cytokines like IL-12, TNF α or IFN γ [73]. In turn, M2 macrophages stimulate the proliferation of cancer cells via the secretion of matrix metalloproteinases, IL-4, IL-5, IL-6 and other factors like VEGF [73]. In the OC microenvironment, TAMs mainly occur as the M2 phenotype, highly expressing scavenger receptor class B (CD163) and the mannose receptor (MR, CD204) [55]. Despite chemokines and cytokines that are well known to modulate M0 macrophage polarization, in recent years, increasing evidence indicates that microRNAs may also play an important role. Ying et al. showed that miR-222-3p secreted by OC cells stimulates the polarization of M0 cells into M2 via the SOCS3/STAT3 pathway [74]. MiRNA-940, miRNA-200b, miRNA-181c-5p and miRNA-141-3p have also been found to promote the M2-like phenotype [75–78]. In turn, Jiang et al. showed that miR-217 inhibits polarization into M2 cells by affecting IL-6 and the JAK3/STAT3 signaling pathway [79]. Both cancer cells and cells belonging to the TME secrete substances that cause their mutual interaction. Earlier, we indicated substances secreted by cancer cells that affect the polarization of macrophages. Nevertheless, polarized macrophages also secrete substances that can further affect cancer cells. It has been shown that M2 macrophages increase the proliferation and progression of OC [80]. Steitz et al. showed that TAMs derived from ascites promote invasion of HGSC via transforming growth factor beta-induced (TGFB1) protein and tenascin C (TNC) [80]. Zeng et al. presented that M2-like macrophages secrete epidermal growth factor (EGF) and thus affect EGFR-ERK signaling, leading to the progression of OC [81]. Ke et al. showed that TAMs increase the invasion of OC cells via the TLR signaling pathway and its downstream nuclear factor NF κ B and microtubule-associated proteins' (MAPs) kinases [82]. A co-culture of SKOV3 cells with macrophages resulted in increased migration and invasion [83,84]. It also increased the expression of NF κ B, CXCL16 and CXCR6 and also affected the PI3K/Akt signaling pathway [83].

5.3. Myeloid-Derived Suppressor Cells (MDSCs)

MDSCs are described as a heterogeneous population of cells of a myeloid origin in various stages of differentiation without mature myeloid markers [85]. It was observed that MDSCs are present in the peripheral blood derived from women with EOC [86]. In normal conditions, immature myeloid cells (IMCs) differentiate into mature forms like granulocytes, macrophages or dendritic cells [85]. Pathological conditions result in an expansion of IMCs due to the blocking of the differentiation and making them MDSCs. Interestingly, it was shown that obesity also has an impact on MDSCs and ovarian cancer [87]. Yang et al. showed that in obese mice, the proportion of MDSCs in peripheral blood was higher than in healthy mice [87]. MDSCs present higher expression of immune suppressive factors like *arginase 1* (ARG1), and inducible nitric oxide synthase (iNOS). A characteristic is also increased expression of CD33+ on their surface and increased production of nitric oxide (NO) and reactive oxygen species (ROS) [85,88]. MDSCs have immune suppressive functions by directly affecting T and NK cells and also stimulating angiogenesis, proliferation, invasion and metastases of the tumor [89–91]. Cui et al. showed that MDSCs increased cancer stemness via inhibition of T cell activation and affection of the expression of miRNA101 and the corepressor gene C-terminal binding protein-2 (CtBP2) [91]. Li et al. also observed that MDSCs enhance the stemness of EOC cells [86]. The authors also verified genes that were modulated during a co-culture of MDSCs and SKOV3 cells and they observed that

colony-stimulating factor 2 (CSF2), intercellular adhesion molecule 1 (ICAM1), baculoviral IAP repeat-containing 3 (BIRC3), TNF α -induced protein 3 (TNFAIP3) and interleukin-32 (IL-32) increased significantly [86]. Zheng et al. showed that upregulation of miRNA-211 decreased MDSC proliferation and affected pathways like NF- κ B, PI3K/Akt and STAT3 [92]. Taki et al. presented that knockdown of Snail in mouse OC cells resulted in an inhibited growth and decreased amount of MDSC cells. Snail was also described as a factor that recruits MDSC in the OC environment via regulation of the expression of the interleukin-8 receptor, beta (CXCR2) ligands [93]. Vascular endothelial growth factor (VEGF) has also been proposed as a factor that recruits MDSCs [94]. Nevertheless, usage of anti-VEGF therapy increases granulocyte–monocyte colony-stimulating factor (GM-CSF) expression and thus recruits MDSCs; therefore, Horikawa et al. suggested that targeting GM-CSF may help overcome resistance to anti-VEGF therapy [95]. Summarized effects of TAMs, CAFs and MDSCs are presented in Figure 3.

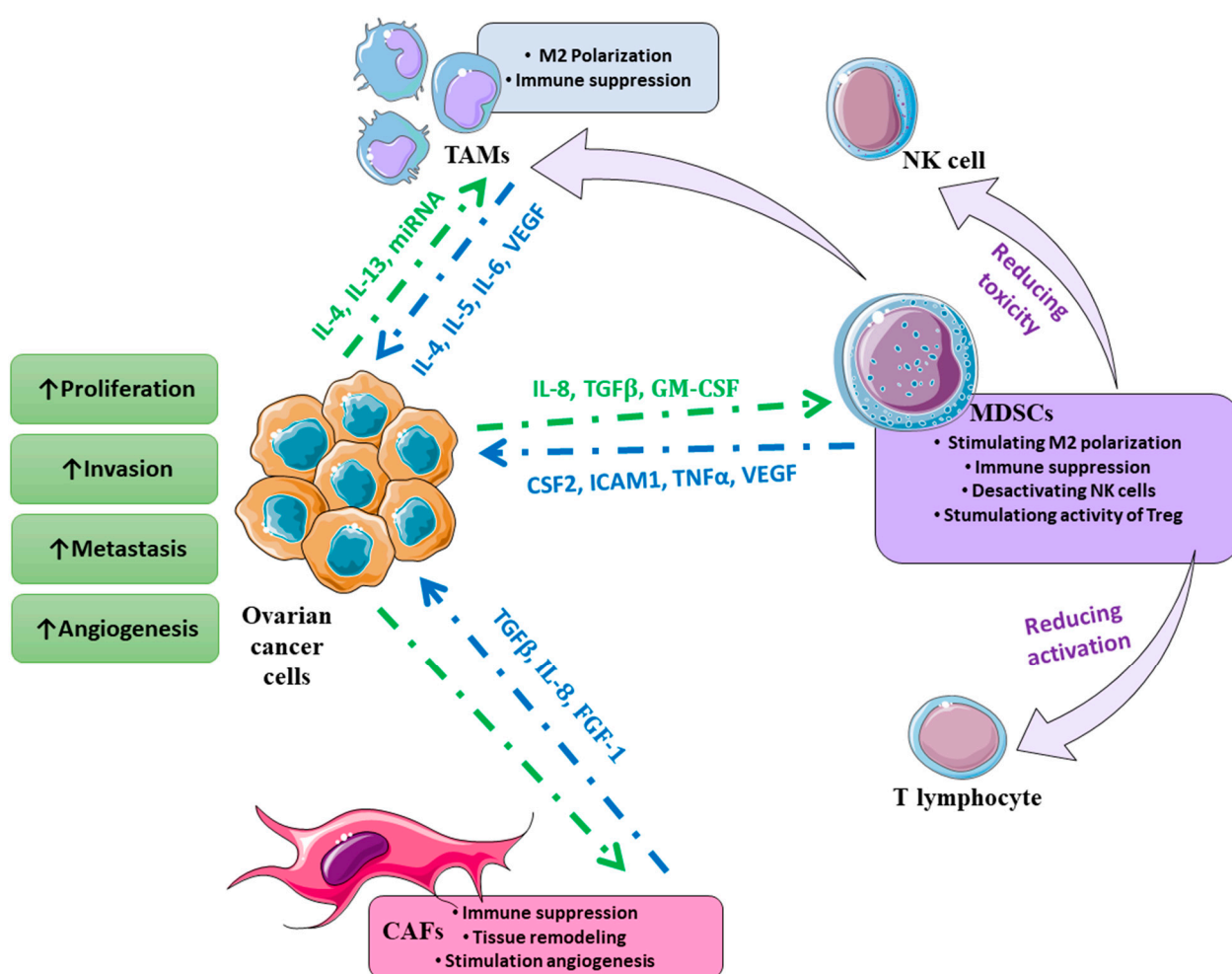


Figure 3. Schema presenting the interaction of the compartments of TME that stimulate ovarian cancer. TAMs—Tumor-Associated Macrophages, MDSCs—Myeloid-Derived Suppressor Cells, CAFs—Cancer-Associated Fibroblasts. The graphical illustration was prepared by using images from Servier Medical Art by Servier. Minor modifications were made (e.g., the color of the stock images) (https://smart.servier.com/smart_image/, accessed on 21 August 2023).

6. Conclusions

In recent years, there is more and more evidence that estrogens play a key role in the formation of all kinds of hormone-dependent cancers, such as breast, prostate or ovarian cancer. From this review of the literature, it is clear that estrogens or estrogen-like compounds, acting through the ERs, regulate various cellular processes such as proliferation,

EMT, invasiveness, differentiation and inflammation in ovarian cancer cells. It seems that all receptors play a very important role in the process of ovarian carcinogenesis, and keeping their proportions in a physiological state is necessary to stay healthy. Moreover, it seems that estrogens are also necessary for the regulation of the TME because most of the estrogen-related pathways are disrupted and thus contribute to the pro-tumor function of the TME. In conclusion, estrogens and estrogen-like compounds play an important role in ovarian carcinogenesis through direct or indirect involvement in molecular mechanisms stimulating the growth and proliferation of cancer cells. Further studies are needed to elucidate all molecular mechanisms of estrogen signaling in ovarian cancer cells and the components of TME, due to their crucial role in cancer progression and therapy. Enriched knowledge about estrogens, estrogen receptors and the tumor microenvironment may encourage us to look for new therapeutic options for patients.

7. Limitations of the Review

During the review associated with tumor microenvironment (TME), we focused on cells that are associated with ovarian cancer proliferation; nevertheless, it should be emphasized that more types of cells are included in TME and they may also be worth reviewing.

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References

1. World Cancer Research Fund International. Ovarian Cancer Statistics. Available online: <https://www.wcrf.org/cancer-trends/ovarian-cancer-statistics/> (accessed on 22 June 2023).
2. Nowotwór Jajnika Stadia Zaawansowania. Krajowy Rejestr Nowotworów. Available online: <https://onkologia.org.pl/pl/nowotwor-jajnika-stadia-zaawansowania#page-main-image> (accessed on 8 August 2023).
3. Heublein, S.; Page, S.; Mayr, D.; Schmoekel, E.; Trillsch, F.; Marmé, F.; Mahner, S.; Jeschke, U.; Vattai, A. Potential interplay of the gatipotuzumab epitope TA-MUC1 and estrogen receptors in ovarian cancer. *Int. J. Mol. Sci.* **2019**, *20*, 295. [CrossRef] [PubMed]
4. Boon, W.C.; Chow, J.D.Y.; Simpson, E.R. The multiple roles of estrogens and the enzyme aromatase. *Prog. Brain Res.* **2010**, *181*, 209–232. [CrossRef] [PubMed]
5. Knowlton, A.A.; Lee, A.R. Estrogen and the cardiovascular system. *Pharmacol. Ther.* **2012**, *135*, 54–70. [CrossRef]
6. Chakraborty, B.; Byemerwa, J.; Krebs, T.; Lim, F.; Chang, C.Y.; McDonnell, D.P. Estrogen Receptor Signaling in the Immune System. *Endocr. Rev.* **2023**, *44*, 117–141. [CrossRef] [PubMed]
7. Richards, J.S.; Pangas, S.A. The ovary: Basic biology and clinical implications. *J. Clin. Investig.* **2010**, *120*, 963–972. [CrossRef]
8. Xu, X.L.; Huang, Z.Y.; Yu, K.; Li, J.; Fu, X.W.; Deng, S.L. Estrogen Biosynthesis and Signal Transduction in Ovarian Disease. *Front. Endocrinol.* **2022**, *13*, 827032. [CrossRef] [PubMed]
9. Cuna, S.; Hoffmann, P.; Pujol, P. Estrogens and epithelial ovarian cancer. *Gynecol. Oncol.* **2004**, *94*, 25–32. [CrossRef] [PubMed]
10. Matsumura, S.; Ohta, T.; Yamanouchi, K.; Liu, Z.; Sudo, T.; Kojimahara, T.; Seino, M.; Narumi, M.; Tsutsumi, S.; Takahashi, T.; et al. Activation of estrogen receptor α by estradiol and cisplatin induces platinum-resistance in ovarian cancer cells. *Cancer Biol. Ther.* **2017**, *18*, 730–739. [CrossRef]
11. Group, C.; Cancer, O. Menopausal hormone use and ovarian cancer risk: Individual participant meta-analysis of 52 epidemiological studies. *Lancet* **2015**, *385*, 1835–1842. [CrossRef]
12. Eeles, R.A.; Morden, J.P.; Gore, M.; Mansi, J.; Glees, J.; Wenzl, M.; Williams, C.; Kitchener, H.; Osborne, R.; Guthrie, D.; et al. Adjuvant hormone therapy may improve survival in epithelial ovarian cancer: Results of the AHT randomized trial. *J. Clin. Oncol.* **2015**, *33*, 4138–4144. [CrossRef]
13. Symer, M.M.; Wong, N.Z.; Abelson, J.S.; Milsom, J.W.; Yeo, H.L. Hormone Replacement Therapy and Colorectal Cancer Incidence and Mortality in the Prostate, Lung, Colorectal, and Ovarian Cancer Screening Trial. *Clin. Color. Cancer* **2018**, *17*, e281–e288. [CrossRef] [PubMed]

14. Poggio, F.; Del Mastro, L.; Bruzzone, M.; Ceppi, M.; Razeti, M.G.; Fregatti, P.; Ruelle, T.; Pronzato, P.; Massarotti, C.; Franzoi, M.A.; et al. Safety of systemic hormone replacement therapy in breast cancer survivors: A systematic review and meta-analysis. *Breast Cancer Res. Treat.* **2022**, *191*, 269–275. [[CrossRef](#)] [[PubMed](#)]
15. Johansson, T.; Fowler, P.; Ek, W.E.; Skalkidou, A.; Karlsson, T.; Johansson, A. Oral Contraceptives, Hormone Re-placement Therapy, and Stroke Risk. *Stroke* **2022**, *53*, 3107–3115. [[CrossRef](#)] [[PubMed](#)]
16. Lobo, R.A. Hormone-replacement therapy: Current thinking. *Nat. Rev. Endocrinol.* **2017**, *13*, 220–231. [[CrossRef](#)]
17. Pujol, P.; Rey, J.-M.; Nirde, P.; Roger, P.; Gastaldi, M.; Laffargue, F.; Rochefort, H.; Maudelonde, T. Differential Expression of Estrogen Receptor- α and - β Messenger RNAs as a Potential Marker of Ovarian Carcinogenesis. *Cancer Res.* **1998**, *58*, 5367–5373.
18. Fujiwara, S.; Terai, Y.; Kawaguchi, H.; Takai, M.; Yoo, S.; Tanaka, Y.; Tanaka, T.; Tsunetoh, S.; Sasaki, H.; Kanemura, M.; et al. GPR30 regulates the EGFR-Akt cascade and predicts lower survival in patients with ovarian cancer. *J. Ovarian Res.* **2012**, *5*, 35. [[CrossRef](#)]
19. Smith, H.O.; Arias-Pulido, H.; Kuo, D.Y.; Howard, T.; Qualls, C.R.; Lee, S.J.; Verschraegen, C.F.; Hathaway, H.J.; Joste, N.E.; Prossnitz, E.R. GPR30 predicts poor survival for ovarian cancer. *Gynecol. Oncol.* **2009**, *114*, 465–471. [[CrossRef](#)]
20. Kolkova, Z.; Casslén, V.; Henic, E.; Ahmadi, S.; Ehinger, A.; Jirstrom, K.; Casslén, B. The G protein-coupled estrogen receptor 1 (GPER/GPR30) does not predict survival in patients with ovarian cancer. *J. Ovarian Res.* **2012**, *5*, 9. [[CrossRef](#)]
21. Fraungruber, P.; Kaltfofen, T.; Heublein, S.; Kuhn, C.; Mayr, D.; Burges, A.; Mahner, S.; Rathert, P.; Jeschke, U.; Trillsch, F. G Protein-Coupled Estrogen Receptor Correlates with Dkk2 Expression and Has Prognostic Impact in Ovarian Cancer Patients. *Front. Endocrinol.* **2021**, *12*, 564002. [[CrossRef](#)]
22. Ignatov, T.; Modl, S.; Thulig, M.; Weißenborn, C.; Treeck, O.; Ortmann, O.; Zenclussen, A.C.; Costa, S.D.; Kalinski, T.; Ignatov, A. GPER-1 acts as a tumor suppressor in ovarian cancer. *J. Ovarian Res.* **2013**, *6*, 51. [[CrossRef](#)]
23. Chen, P.; Li, B.; Ou-Yang, L. Role of estrogen receptors in health and disease. *Front. Endocrinol.* **2022**, *13*, 839005. [[CrossRef](#)]
24. Levin, E.R. Plasma Membrane Estrogen Receptors. *Trends Endocrinol. Metab.* **2009**, *20*, 477–482. [[CrossRef](#)]
25. Revankar, C.M.; Cimino, D.F.; Sklar, L.A.; Arterburn, J.B.; Prossnitz, E.R. A transmembrane intracellular estrogen receptor mediates rapid cell signaling. *Science* **2005**, *307*, 1625–1630. [[CrossRef](#)]
26. Xu, E.; Xia, X.; Jiang, C.; Li, Z.; Yang, Z.; Zheng, C.; Wang, X.; Du, S.; Miao, J.; Wang, F.; et al. GPER1 Silencing Sup-presses the Proliferation, Migration, and Invasion of Gastric Cancer Cells by Inhibiting PI3K/AKT-Mediated EMT. *Front. Cell Dev. Biol.* **2020**, *8*, 591239. [[CrossRef](#)]
27. Simpkins, F.; Garcia-Soto, A.; Slingerland, J. New insights on the role of hormonal therapy in ovarian cancer. *Steroids* **2013**, *78*, 530–537. [[CrossRef](#)]
28. Bossard, C.; Busson, M.; Vindrieux, D.; Gaudin, F.; Machelon, V.; Brigitte, M.; Jacquard, C.; Pilon, A.; Balaguer, P.; Balabanian, K.; et al. Potential Role of Estrogen Receptor Beta as a Tumor Suppressor of Epithelial Ovarian Cancer. *PLoS ONE* **2012**, *7*, e44787. [[CrossRef](#)]
29. Thomas, C.; Gustafsson, J.Å. The different roles of ER subtypes in cancer biology and therapy. *Nat. Rev. Cancer* **2011**, *11*, 597–608. [[CrossRef](#)]
30. Langdon, S.P.; Herrington, C.S.; Hollis, R.L.; Gourley, C. Estrogen signaling and its potential as a target for therapy in ovarian cancer. *Cancers* **2020**, *12*, 1647. [[CrossRef](#)] [[PubMed](#)]
31. Mungenast, F.; Thalhammer, T. Estrogen biosynthesis and action in ovarian cancer. *Front. Endocrinol.* **2014**, *5*, 192. [[CrossRef](#)] [[PubMed](#)]
32. Park, S.H.; Cheung, L.W.T.; Wong, A.S.T.; Leung, P.C.K. Estrogen regulates snail and slug in the down-regulation of E-cadherin and induces metastatic potential of ovarian cancer cells through estrogen receptor α . *Mol. Endocrinol.* **2008**, *22*, 2085–2098. [[CrossRef](#)] [[PubMed](#)]
33. Schüler-Toprak, S.; Moehle, C.; Skrzypczak, M.; Ortmann, O.; Treeck, O. Effect of estrogen receptor β agonists on proliferation and gene expression of ovarian cancer cells. *BMC Cancer* **2017**, *17*, 319. [[CrossRef](#)] [[PubMed](#)]
34. Treeck, O.; Pfeiler, G.; Mitter, D.; Latrich, C.; Piendl, G.; Ortmann, O. Estrogen receptor β 1 exerts antitumoral effects on SK-OV-3 ovarian cancer cells. *J. Endocrinol.* **2007**, *193*, 421–433. [[CrossRef](#)] [[PubMed](#)]
35. Banerjee, A.; Cai, S.; Xie, G.; Li, N.; Bai, X.; Lavudi, K.; Wang, K.; Zhang, X.; Zhang, J.; Patnaik, S.; et al. A Novel Estrogen Receptor β Agonist Diminishes Ovarian Cancer Stem Cells via Suppressing the Epithelial-To-Mesenchymal Transition. *Cancers* **2022**, *14*, 2311. [[CrossRef](#)] [[PubMed](#)]
36. Yan, Y.; Liu, H.; Wen, H.; Jiang, X.; Cao, X.; Zhang, G.; Liu, G. The novel estrogen receptor GPER regulates the migration and invasion of ovarian cancer cells. *Mol. Cell. Biochem.* **2013**, *378*, 1–7. [[CrossRef](#)]
37. Yan, Y.; Jiang, X.; Zhao, Y.; Wen, H.; Liu, G. Role of GPER on proliferation, migration and invasion in ligand-independent manner in human ovarian cancer cell line SKOV3. *Cell Biochem. Funct.* **2015**, *33*, 552–559. [[CrossRef](#)]
38. Wang, C.; Lv, X.; He, C.; Hua, G.; Tsai, M.Y.; Davis, J.S. The G-protein-coupled estrogen receptor agonist G-1 sup-presses proliferation of ovarian cancer cells by blocking tubulin polymerization. *Cell Death Dis.* **2013**, *4*, e869. [[CrossRef](#)]
39. Goh, Y.X.; Jalil, J.; Lam, K.W.; Husain, K.; Premakumar, C.M. Genistein: A Review on its Anti-Inflammatory Properties. *Front. Pharmacol.* **2022**, *13*, 820969. [[CrossRef](#)]
40. Chan, K.K.L.; Siu, M.K.Y.; Jiang, Y.X.; Wang, J.J.; Leung, T.H.Y.; Ngan, H.Y.S. Estrogen receptor modulators genistein, daidzein and ERB-041 inhibit cell migration, invasion, proliferation and sphere formation via modulation of FAK and PI3K/AKT signaling in ovarian cancer. *Cancer Cell Int.* **2018**, *18*, 65. [[CrossRef](#)]

41. Kang, N.H.; Hwang, K.A.; Lee, H.R.; Choi, D.W.; Choi, K.C. Resveratrol regulates the cell viability promoted by 17 β -estradiol or bisphenol A via down-regulation of the cross-talk between estrogen receptor α and insulin growth factor-1 receptor in BG-1 ovarian cancer cells. *Food Chem. Toxicol.* **2013**, *59*, 373–379. [\[CrossRef\]](#)
42. Sang, C.; Song, Y.; Jin, T.W.; Zhang, S.; Fu, L.; Zhao, Y.; Zou, X.; Wang, Z.; Gao, H.; Liu, S. Bisphenol A induces ovarian cancer cell proliferation and metastasis through estrogen receptor- α pathways. *Environ. Sci. Pollut. Res. Int.* **2021**, *28*, 36060–36068. [\[CrossRef\]](#)
43. Hwang, K.A.; Park, M.A.; Kang, N.H.; Yi, B.R.; Hyun, S.H.; Jeung, E.B.; Choi, K.C. Anticancer effect of genistein on BG-1 ovarian cancer growth induced by 17 β -estradiol or bisphenol A via the suppression of the crosstalk between estrogen receptor alpha and insulin-like growth factor-1 receptor signaling pathways. *Toxicol. Appl. Pharmacol.* **2013**, *272*, 637–646. [\[CrossRef\]](#) [\[PubMed\]](#)
44. Liu, M.; Zhang, Y.; Xu, Q.; Liu, G.; Sun, N.; Che, H.; He, T. Apigenin Inhibits the Histamine-Induced Proliferation of Ovarian Cancer Cells by Downregulating ER α /ER β Expression. *Front. Oncol.* **2021**, *11*, 682917. [\[CrossRef\]](#) [\[PubMed\]](#)
45. Pham, T.H.; Page, Y.L.; Percevault, F.; Ferrière, F.; Flouriot, G.; Pakdel, F. Apigenin, a partial antagonist of the estrogen receptor (Er), inhibits er-positive breast cancer cell proliferation through Akt/foxm1 signaling. *Int. J. Mol. Sci.* **2021**, *22*, 470. [\[CrossRef\]](#)
46. Zhou, J.; Jiang, Y.Y.; Chen, H.; Wu, Y.C.; Zhang, L. Tanshinone I Attenuates the Malignant Biological Properties of Ovarian Cancer by Inducing Apoptosis and Autophagy via the Inactivation of PI3K/AKT/MTOR Pathway. *Cell Prolif.* **2020**, *53*, e12739. [\[CrossRef\]](#)
47. Koziół, M.J.; Habrowska-Górczyńska, D.E.; Urbanek, K.A.; Domińska, K.; Piastowska-Ciesielska, A.W.; Kowalska, K. Estrogen Receptor α Mediates Alternariol-Induced Apoptosis and Modulation of the Invasiveness of Ovarian Cancer Cells. *Toxicol. Lett.* **2023**, *386*, 9–19. [\[CrossRef\]](#) [\[PubMed\]](#)
48. Wang, X.; Lin, Y.; Zheng, Y. Antitumor effects of aconitine in A2780 cells via estrogen receptor β -mediated apoptosis, DNA damage and migration. *Mol. Med. Rep.* **2020**, *22*, 2318–2328. [\[CrossRef\]](#)
49. Ataei, N.; Aghaei, M.; Panjehpour, M. Cadmium induces progesterone receptor gene expression via activation of estrogen receptor in human ovarian cancer cells. *Res. Pharm. Sci.* **2018**, *13*, 500–508. [\[CrossRef\]](#)
50. Hoffmann, M.; Gogola, J.; Kotula-Balak, M.; Ptak, A. Stimulation of ovarian cell proliferation by tetrabromobisphenol A but not tetrachlorobisphenol A through G protein-coupled receptor 30. *Toxicol. Vitro.* **2017**, *45*, 54–59. [\[CrossRef\]](#)
51. Ataei, N.; Aghaei, M.; Panjehpour, M. Evidences for involvement of estrogen receptor induced ERK1/2 activation in ovarian cancer cell proliferation by Cadmium Chloride. *Toxicol. Vitro.* **2019**, *56*, 184–193. [\[CrossRef\]](#)
52. Quail, D.F.; Joyce, J.A. Microenvironmental regulation of tumor progression and metastasis. *Nat. Med.* **2013**, *19*, 1423–1437. [\[CrossRef\]](#)
53. Xiao, Y.; Yu, D. Tumor Microenvironment as a Therapeutic Target in Cancer. *Pharmacol. Ther.* **2021**, *221*, 107753. [\[CrossRef\]](#) [\[PubMed\]](#)
54. Rothenberger, N.J.; Somasundaram, A.; Stabile, L.P. The Role of the Estrogen Pathway in the Tumor Microenvironment. *Int. J. Mol. Sci.* **2018**, *19*, 611. [\[CrossRef\]](#) [\[PubMed\]](#)
55. Nowak, M.; Klink, M. The Role of Tumor-Associated Macrophages in the Progression and Chemoresistance of Ovarian Cancer. *Cells* **2020**, *9*, 1299. [\[CrossRef\]](#) [\[PubMed\]](#)
56. Pan, Z.; Niu, G.; Cao, C.; Tian, Y. Role of MicroRNAs in Remodeling the Tumor Microenvironment (Review). *Int. J. Oncol.* **2020**, *56*, 407–416. [\[CrossRef\]](#)
57. Soon, P.; Kiaris, H. MicroRNAs in the Tumour Microenvironment: Big Role for Small Players. *Endocr. Relat. Cancer* **2013**, *20*, R257–R267. [\[CrossRef\]](#)
58. Zhang, Y.; Tang, H.; Cai, J.; Zhang, T.; Guo, J.; Feng, D.; Wang, Z. Ovarian cancer-associated fibroblasts contribute to epithelial ovarian carcinoma metastasis by promoting angiogenesis, lymphangiogenesis and tumor cell invasion. *Cancer Lett.* **2011**, *303*, 47–55. [\[CrossRef\]](#)
59. Dai, J.M.; Sun, K.; Li, C.; Cheng, M.; Guan, J.H.; Yang, L.N.; Zhang, L. Cancer-associated fibroblasts contribute to cancer metastasis and apoptosis resistance in human ovarian cancer via paracrine SDF-1 α . *Clin. Transl. Oncol.* **2023**, *25*, 1606–1616. [\[CrossRef\]](#)
60. Jin, Y.; Bian, S.; Wang, H.; Mo, J.; Fei, H.; Li, L.; Chen, T.; Jiang, H. CRMP2 derived from cancer associated fibroblasts facilitates progression of ovarian cancer via HIF-1 α -glycolysis signaling pathway. *Cell Death Dis.* **2022**, *13*, 675. [\[CrossRef\]](#)
61. Xing, F.; Saidou, J.; Watabe, K. Cancer associated fibroblasts (CAFs) in tumor microenvironment. *Front. Biosci.* **2010**, *15*, 166–179. [\[CrossRef\]](#)
62. Thongchot, S.; Jamjuntra, P.; Therasakvichya, S.; Warnnissorn, M.; Ferraresi, A.; Thuwajit, P.; Isidoro, C.; Thuwajit, C. Interleukin-8 released by cancer-associated fibroblasts attenuates the autophagy and promotes the migration of ovarian cancer cells. *Int. J. Oncol.* **2021**, *58*, 14. [\[CrossRef\]](#)
63. Sun, Y.; Fan, X.; Zhang, Q.; Shi, X.; Xu, G.; Zou, C. Cancer-associated fibroblasts secrete FGF-1 to promote ovarian proliferation, migration, and invasion through the activation of FGF-1/FGFR4 signaling. *Tumor Biol.* **2017**, *39*, 1010428317712592. [\[CrossRef\]](#) [\[PubMed\]](#)
64. Dong, T.; Yang, D.; Li, R.; Zhang, L.; Zhao, H.; Shen, Y.; Zhang, X.; Kong, B.; Wang, L. PGRN promotes migration and invasion of epithelial ovarian cancer cells through an epithelial mesenchymal transition program and the activation of cancer associated fibroblasts. *Exp. Mol. Pathol.* **2016**, *100*, 17–25. [\[CrossRef\]](#)

65. Yeung, T.L.; Leung, C.S.; Wong, K.K.; Samimi, G.; Thompson, M.S.; Liu, J.; Zaid, T.M.; Ghosh, S.; Birrer, M.J.; Mok, S.C. TGF- β Modulates ovarian cancer invasion by upregulating CAF-Derived versican in the tumor microenvironment. *Cancer Res.* **2013**, *73*, 5016–5028. [\[CrossRef\]](#)
66. Wu, Y.H.; Huang, Y.F.; Chang, T.H.; Chen, C.C.; Wu, P.Y.; Huang, S.C.; Chou, C.Y. COL11A1 activates cancer-associated fibroblasts by modulating TGF- β 3 through the NF- κ B/IGFBP2 axis in ovarian cancer cells. *Oncogene* **2021**, *40*, 4503–4519. [\[CrossRef\]](#) [\[PubMed\]](#)
67. Yue, H.; Li, W.; Chen, R.; Wang, J.; Lu, X.; Li, J. Stromal POSTN induced by TGF- β 1 facilitates the migration and invasion of ovarian cancer. *Gynecol. Oncol.* **2021**, *160*, 530–538. [\[CrossRef\]](#) [\[PubMed\]](#)
68. Knowler, K.C.; Chand, A.L.; Eriksson, N.; Takagi, K.; Miki, Y.; Sasano, H.; Visvader, J.E.; Lindeman, G.J.; Funder, J.W.; Fuller, P.J.; et al. Distinct nuclear receptor expression in stroma adjacent to breast tumors. *Breast Cancer Res. Treat.* **2013**, *142*, 211–223. [\[CrossRef\]](#) [\[PubMed\]](#)
69. Bae, W.J.; Kim, S.; Ahn, J.M.; Han, J.H.; Lee, D. Estrogen-responsive cancer-associated fibroblasts promote invasive property of gastric cancer in a paracrine manner via CD147 production. *FASEB J.* **2022**, *36*, e22597. [\[CrossRef\]](#)
70. Kim, M.J.; Jung, D.; Park, J.Y.; Lee, S.M.; An, H.J. GLIS1 in Cancer-Associated Fibroblasts Regulates the Migration and Invasion of Ovarian Cancer Cells. *Int. J. Mol. Sci.* **2022**, *23*, 2218. [\[CrossRef\]](#)
71. Mitra, A.K.; Zillhardt, M.; Hua, Y.; Tiwari, P.; Murmann, A.E.; Peter, M.E.; Lengyel, E. MicroRNAs Reprogram Normal Fibroblasts into Cancer-Associated Fibroblasts in Ovarian Cancer. *Cancer Discov.* **2012**, *2*, 1100–1108. [\[CrossRef\]](#)
72. Qian, B.Z.; Pollard, J.W. Macrophage Diversity Enhances Tumor Progression and Metastasis. *Cell* **2010**, *141*, 39–51. [\[CrossRef\]](#)
73. Dun, E.C.; Hanley, K.; Wieser, F.; Bohman, S.; Yu, J.; Taylor, R.N. Infiltration of tumor-associated macrophages is increased in the epithelial and stromal compartments of endometrial carcinomas. *Int. J. Gynecol. Pathol.* **2013**, *32*, 576–584. [\[CrossRef\]](#) [\[PubMed\]](#)
74. Ying, X.; Wu, Q.; Wu, X.; Zhu, Q.; Wang, X.; Jiang, L.; Chen, X.; Wang, X. Epithelial ovarian cancer-secreted exosomal miR-222-3p induces polarization of tumor-associated macrophages. *Oncotarget* **2016**, *7*, 43076–43087. [\[CrossRef\]](#)
75. Chen, X.; Ying, X.; Wang, X.; Wu, X.; Zhu, Q.; Wang, X. Exosomes derived from hypoxic epithelial ovarian cancer deliver microRNA-940 to induce macrophage M2 polarization. *Oncol. Rep.* **2017**, *38*, 522–528. [\[CrossRef\]](#)
76. Xiong, J.; He, X.; Xu, Y.; Zhang, W.; Fu, F. MiR-200b is upregulated in plasma-derived exosomes and functions as an oncogene by promoting macrophage M2 polarization in ovarian cancer. *J. Ovarian Res.* **2021**, *14*, 74. [\[CrossRef\]](#) [\[PubMed\]](#)
77. Yang, S.; Zhao, H.; Xiao, W.; Shao, L.; Zhao, C.; Sun, P. Extracellular vesicle-packaged miR-181c-5p from epithelial ovarian cancer cells promotes M2 polarization of tumor-associated macrophages via the KAT2B/HOXA10 axis. *J. Gene Med.* **2022**, *24*, e3446. [\[CrossRef\]](#) [\[PubMed\]](#)
78. Zhao, J.; Liu, L.; Zhao, W.; Lv, C.; Zhang, N.; Jia, X.; Zhang, Z. MiR-141-3p accelerates ovarian cancer progression and promotes M2-like macrophage polarization by targeting the Keap1-Nrf2 pathway. *Open Med.* **2023**, *18*, 20230729. [\[CrossRef\]](#)
79. Jiang, B.; Zhu, S.J.; Xiao, S.S.; Xue, M. MiR-217 Inhibits M2-Like Macrophage Polarization by Suppressing Secretion of Interleukin-6 in Ovarian Cancer. *Inflammation* **2019**, *42*, 1517–1529. [\[CrossRef\]](#)
80. Steitz, A.M.; Steffes, A.; Finkernagel, F.; Unger, A.; Sommerfeld, L.; Jansen, J.M.; Wagner, U.; Graumann, J.; Müller, R.; Reinartz, S. Tumor-associated macrophages promote ovarian cancer cell migration by secreting transforming growth factor beta induced (TGFBI) and tenascin C. *Cell Death Dis.* **2020**, *11*, 249. [\[CrossRef\]](#)
81. Zeng, X.Y.; Xie, H.; Yuan, J.; Jiang, X.Y.; Yong, J.H.; Zeng, D.; Dou, Y.Y.; Xiao, S.S. M2-like tumor-associated macrophages-secreted EGF promotes epithelial ovarian cancer metastasis via activating EGFR-ERK signaling and suppressing lncRNA LIMT expression. *Cancer Biol. Ther.* **2019**, *20*, 956–966. [\[CrossRef\]](#)
82. Ke, X.; Zhang, S.; Wu, M.; Lou, J.; Zhang, J.; Xu, T.; Huang, L.; Huang, P.; Wang, F.; Pan, S. Tumor-associated macrophages promote invasion via Toll-like receptors signaling in patients with ovarian cancer. *Int. Immunopharmacol.* **2016**, *40*, 184–195. [\[CrossRef\]](#)
83. Hong, L.; Wang, S.; Li, W.; Wu, D.; Chen, W. Tumor-associated macrophages promote the metastasis of ovarian carcinoma cells by enhancing CXCL16/CXCR6 expression. *Pathol. Res. Pract.* **2018**, *214*, 1345–1351. [\[CrossRef\]](#) [\[PubMed\]](#)
84. Feng, Y.; Xiao, M.; Cao, G.; Liu, H.; Li, Y.; Wang, S.; Zijteld, S.; Delvoux, B.; Xanthoulea, S.; Romano, A.; et al. Human monocytes differentiate into tumor-associated macrophages upon SKOV3 cells coculture and/or lysophosphatidic acid stimulation. *J. Inflamm.* **2022**, *19*, 11. [\[CrossRef\]](#) [\[PubMed\]](#)
85. Gabrilovich, D.I.; Nagaraj, S. Myeloid-derived suppressor cells as regulators of the immune system. *Nat. Rev. Immunol.* **2009**, *9*, 162–174. [\[CrossRef\]](#) [\[PubMed\]](#)
86. Li, X.; Wang, J.; Wu, W.; Gao, H.; Liu, N.; Zhan, G.; Li, L.; Han, L.; Guo, X. Myeloid-derived suppressor cells promote epithelial ovarian cancer cell stemness by inducing the CSF2/p-STAT3 signalling pathway. *FEBS J.* **2020**, *287*, 5218–5235. [\[CrossRef\]](#)
87. Yang, Q.; Yu, B.; Kang, J.; Li, A.; Sun, J. Obesity Promotes Tumor Immune Evasion in Ovarian Cancer Through Increased Production of Myeloid-Derived Suppressor Cells via IL-6. *Cancer Manag. Res.* **2021**, *13*, 7355–7363. [\[CrossRef\]](#)
88. Walankiewicz, M.; Grywalska, E.; Polak, G.; Kotarski, J.; Siwicki-Gieroba, D.J.; Rolinski, J. Myeloid-derived suppressor cells in ovarian cancer: Friend or foe? *Cent. J. Immunol.* **2017**, *42*, 383–389. [\[CrossRef\]](#)
89. Ostrand-Rosenberg, S.; Sinha, P. Myeloid-Derived Suppressor Cells: Linking Inflammation and Cancer. *J. Immunol.* **2009**, *182*, 4499–4506. [\[CrossRef\]](#)
90. Wang, Y.; Jia, A.; Bi, Y.; Wang, Y.; Yang, Q.; Cao, Y.; Li, Y.; Liu, G. Targeting Myeloid-Derived Suppressor Cells in Cancer Immunotherapy. *Cancers* **2020**, *12*, 2626. [\[CrossRef\]](#)

91. Cui, T.X.; Kryczek, I.; Zhao, L.; Zhao, E.; Kuick, R.; Roh, M.H.; Vatan, L.; Szeliga, W.; Mao, Y.; Thomas, D.G.; et al. Myeloid-Derived Suppressor Cells Enhance Stemness of Cancer Cells by Inducing MicroRNA101 and Suppressing the Corepressor CtBP2. *Immunity* **2013**, *39*, 611–621. [[CrossRef](#)]
92. Zheng, L.E.; Huang, M.; Ye, Y.; Sun, P. MicroRNA-211 Regulates Proliferation, Expansion, and Immune Inhibitory Function of Myeloid-Derived Suppressor Cells via Mediation of CHOP Expression. *Immunol. Investig.* **2023**, *52*, 616–634. [[CrossRef](#)]
93. Taki, M.; Abiko, K.; Baba, T.; Hamanishi, J.; Yamaguchi, K.; Murakami, R.; Yamanoi, K.; Horikawa, N.; Hosoe, Y.; Nakamura, E.; et al. Snail promotes ovarian cancer progression by recruiting myeloid-derived suppressor cells via CXCR2 ligand upregulation. *Nat. Commun.* **2018**, *9*, 1685. [[CrossRef](#)] [[PubMed](#)]
94. Horikawa, N.; Abiko, K.; Matsumura, N.; Hamanishi, J.; Baba, T.; Yamaguchi, K.; Yoshioka, Y.; Koshiyama, M.; Konishi, I. Expression of vascular endothelial growth factor in ovarian cancer inhibits tumor immunity through the accumulation of myeloid-derived suppressor cells. *Clin. Cancer Res.* **2017**, *23*, 587–599. [[CrossRef](#)] [[PubMed](#)]
95. Horikawa, N.; Abiko, K.; Matsumura, N.; Baba, T.; Hamanishi, J.; Yamaguchi, K.; Murakami, R.; Taki, M.; Ukita, M.; Hosoe, Y.; et al. Anti-VEGF therapy resistance in ovarian cancer is caused by GM-CSF-induced myeloid-derived suppressor cell recruitment. *Br. J. Cancer* **2020**, *122*, 778–788. [[CrossRef](#)] [[PubMed](#)]

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