Table S3. Bacterial strains, plasmids and primers used in this study.

Name	Description	Origin
Strain		
Pectobacterium atrosepticum SCRI1043	Wild type	[110]
Pectobacterium atrosepticum SCRI1043 Δ*rpoS*	Mutant strain of SCRI1043 containing the kanamycin cassette in the chromosome; *Km^R*	This study
Pectobacterium atrosepticum SCRI1043 Δ*rpoS* complementation mutant	Mutant strain of SCRI1043 containing the kanamycin cassette in the chromosome and the *rpoS* complementation construct on pGEM:*rpoS*; *Km^R Amp^R*	This study
Escherichia coli cc118	Host for suicidal vector pKNG101; *Δ(ara, leu) araD ΔlacX 74 galE galK PhoA20 thi-1 rpsE rpoB argE (am) recA1, Sm^R*	[111]
Escherichia coli HH26/pNJ5000	Mobilizing strain for conjugative transfer of the suicide vector pKNG101 into *P. atrosepticum* cells; *tra+; Tet^R*	[112]
Escherichia coli NovaBlue	*endA 1hsdR17 (rK12–mK12+) supE44 thi-1 recA1 gyrA96 relA1 lacF' [proA+ B+ lacIqZ ΔM15::Tn10 (Tet^R)]*	Novagen
Plasmids		
pGEM-T Easy	Linearized vector for cloning; *f1 ori Amp^R lacZ*	Promega
pGEM:*rpoS*	*f1 ori Amp^R lacZ rpoS*	This study
pGEM:*ΔrpoS*;Km^R	*f1 ori Amp^R lacZ Km^R*	This study
pKD4	Matrix for PCR amplification of kanamycin resistance cassette; *oriRγ rgnB bla Km^R*	[113]
pKNG101	Suicide mobilized vector for inactivation of target genes; *pir-ori R6K mobRK2 sacB Sm^R*	[97]
pKNG101:*ΔrpoS*;Km^R	Suicide plasmid carrying mutant locus *ΔrpoS*;Km^R; *Km^R Sm^R sacB*	This study
pGEM:*rpoS*; complementation construct	*f1 ori Amp^R lacZ rpoS*	This study
Primers		
Primer name	**Primer sequence 5'—3'**	
Primers for mutagenesis		
up *rpoS* F	TGGCAACAACGCTTCTGGTG	
dn *rpoS* R	GAACTGAATGAAATCAAGGCGGAG	

dn *rpoS* KmF	CCCATGTCAGCCGTTAAGACGCTGGAAGATGTGGGTCG
up *rpoS* KmR	CCAGCCTACACAATCGATGGGACATCGCCACGC
Km *rpoS* F	GCGATGTCCCATCGATTGTGTAGGCTGGAGCTGCTTC
Km *rpoS* R	ATCTTCCAGCGTCTTAACGGCTGACATGGGAATTAGC
Check *rpoS* F	CGTATAGCCTGAATGTCGGGCAATC
Check *rpoS* R	GCCATCAACTGGTTGCGTTGC
T7F	TAATACGACTCACTATAGGG
Sp6R	TAAATCCACTGTGATATCTTA
Primers for complementation construct	
Comp *rpoS* F	CGATACGATGCTAGTCCGTGAGC
Comp *rpoS* R	CTGGCGAATATCATACGCTAATCG
Primers for qRT-PCR	
*rpoS*_F	CTATTTGGGAGAGATTGGCTATTCGC
*rpoS*_R	CCAGACCACGATTGTTGTAACGGC
*rpoD*_F	CTAATGCGGAAGAAGACATTGCCCC
*rpoD*_R	CTTCTCACGCGCCAGTTCAGGATCG
*Ffh*_F	GTGGCATCATCCCTTTCATACCGC
*Ffh*_R	CGATTCAGGAACGTGCTAAACCAGAG
*recA*_F	GTGGATATTGATGCCTTCGCCG
*recA*_R	GTTTTATGCCTCTGTTCGTTTGGATATTC
*ECA0603_cfa6*_F	CGTGGTCGGGCATTCTGTAG
*ECA0603_cfa6*_R	CGGCGTAACATCTTCAACGG
*ECA0609_cfl*_F	CACAGACGAAATCGCATCTATATTTTAC
*ECA0609_cfl*_R	CTGGCTTATCACCATCTGATTTGTG
*ECA4106_pck*_F	CTCGGCAAACGTCGGCG
*ECA4106_pck*_R	CTTGATGGTTTTGGCGTAGCAG
*ECA3517_trxC*_F	CTTGGTGCGGTCCCTGCG
*ECA3517_trxC*_R	CGGTTCGGCTTCGGTATTGAC
*ECA1700_flgM*_F	CAATGTCCTGCGTGCCTGG
*ECA1700_flgM*_R	GCACACAGTCACTGAAGCCG
*ECA3238_iscR*_F	CTCATCCAGCGTGATGTTATTCAGG
*ECA3238_iscR*_R	CTGTTGGTATGGTTATTTCTGCCGTC
*ECA3882_dnaK*_F	CAGCACCAATGGCAACGG
*ECA3882_dnaK*_R	GAAACTGGAATCACTGGTAGAAGAGC
*ECA3014_treC*_F	GGTCTGTGGATTTTGGGCGG
*ECA3014_treC*_R	GGGCTGAAATACATCCTGACTGAAC
*ECA2112_hrpW*_F	CTGAGCACTGCTGTTAGTGATTTC
*ECA2112_hrpW*_R	CTGAAAAACGTGGTGTTTGG
*ECA2767_dps*_F	CTGGAACATGCGTGGCG
*ECA2767_dps*_R	CAGCGACGTTTGTTTACCCAC
*ECA2180_uvrD*_F	GCTTCAAGAGGAGATCTCTACATACAT
*ECA2180_uvrD*_R	GGAAGAGCGGCGTAAGTTTT
*ECA3430*_F	CTGGGCAAGACATACAGTGAAGGTAAG
*ECA3430*_R	CGCTCGGATGTAGCCTGC
*ECA3441*_F	GAAAGCGGCTAAAGCTGAGATTGCC
*ECA3441*_R	CATGAAGGGCAGCAAAATCGTC
*ECA4067_pelA*_F	CTGCTCGCAGCCCAACCG
*ECA4067_pelA*_R	GACGACTTTACCGCTTGAATC
*ECA4068_pelB*_F	CACGCCAGACAATGACACGAC
*ECA4068_pelB*_R	GCATGAACCAGACCACCACGC