

Molecular Diversity of the Genus *Plagiorchis* Lühe, 1899 in Snail Hosts of Central Europe with Evidence of New Lineages

Petra Kunderd^{1,2}, Camila Pantoja³, Kristýna Janovcová¹ and Miroslava Soldánová^{2,*}

¹ Faculty of Science, University of South Bohemia in České Budějovice, Branišovská 1645/31a, 370 05 České Budějovice, Czech Republic; petra.kunderd@paru.cas.cz (P.K.); janovk04@prf.jcu.cz (K.J.)

² Institute of Parasitology, Biology Centre, Czech Academy of Sciences, Branišovská 1160/31, 370 05 České Budějovice, Czech Republic

³ Laboratório de Biologia de Trematoda, Instituto de Ciências Biológicas, Universidade Federal de Minas Gerais, Belo Horizonte 31270-901, MG, Brazil; camilaspantoja@yahoo.com.br

* Correspondence: soldanova@paru.cas.cz; Tel.: +420-387-775-484

Supplementary Table S1. *Plagiorchis* species reported from European freshwater snails. Records from Russia include only the European part. The reference numbering does not correspond to that in the main text.

<i>Plagiorchis</i> species	Snail host	Country	Reference
<i>Plagiorchis elegans</i> (Rudolphi, 1802)	<i>Galba truncatula</i>	Spain	[1]
	<i>Lymnaea stagnalis</i>	Austria, Belarus, Czech Republic, Denmark, Finland, Germany, Ireland, Poland, Russia, Slovakia, Ukraine	[2], [3], [4–9,10 ¹], [11 ²], [12–14, 15 ¹], [16–20], [15 ¹], [2, 21–24], [25], [2, 10 ¹], [26,27]
	<i>Radix auricularia</i>	Czech Republic, Germany, Ukraine	[4], [19,28], [29]
	<i>Radix peregra</i> ³	Czech Republic, Finland, Germany	[4], [12,13], [28]
	<i>Radix</i> spp.	Belarus	[3]
	<i>Stagnicola corvus</i>	Ukraine	[27]
	<i>Stagnicola fuscus</i>	Ireland	[15 ¹]
	<i>Stagnicola palustris</i>	Belarus, Czech Republic, Denmark, Germany, Ukraine	[3], [4], [11 ²], [17,28], [27]
<i>Plagiorchis koreanus</i> Ogata, 1938	<i>Radix auricularia</i>	Czech Republic	[10 ¹]
<i>Plagiorchis laricola</i> Skrjabin, 1924	<i>Lymnaea stagnalis</i>	Czech Republic	[30–32]
<i>Plagiorchis maculosus</i> (Rudolphi, 1802)	<i>Lymnaea stagnalis</i>	Bulgaria, Czech Republic, Denmark, Finland, Ireland, Poland	[33], [4,5,10 ¹ ,32,34], [11 ²], [15 ¹], [15 ¹], [24]
	<i>Radix auricularia</i>	Czech Republic	[4]
	<i>Radix balthica</i> ³	Denmark	[11 ²]
<i>Plagiorchis megalorchis</i> Rees, 1952	<i>Lymnaea stagnalis</i>	Great Britain	[35]
<i>Plagiorchis muelleri</i> Tkach and Sharpilo, 1990	<i>Radix balthica</i> ³	Ireland	[15 ¹]
<i>Plagiorchis multiglandularis</i> Semenov, 1927	<i>Lymnaea stagnalis</i>	Belarus, Russia, Ukraine	[3], [36], [27]
	<i>Myxas glutinosa</i>	Russia	[36]
	<i>Radix ovata</i> ³	Russia	[36]
	<i>Radix</i> spp.	Belarus	[3]
	<i>Stagnicola palustris</i>	Russia	[36]
<i>Plagiorchis mutationis</i> Panova, 1927	<i>Lymnaea stagnalis</i>	Ukraine	[37,38]
<i>Plagiorchis nanus</i> (Rudolphi, 1802)	<i>Lymnaea stagnalis</i>	Belarus	[3]
	<i>Myxas glutinosa</i>	Russia	[36]
	<i>Radix ovata</i> ³	Russia	[36]
<i>Plagiorchis neomidis</i> Brendow, 1970	<i>Ampullaceana balthica</i>	Germany	[20]
	<i>Lymnaea stagnalis</i>	Slovakia	[10 ¹]
	<i>Radix ampla</i> ³	Germany	[18]
	<i>Radix auricularia</i>	Czech Republic	[4,32]
	<i>Radix ovata</i> ³	Czech Republic, Germany	[4], [18]
	<i>Radix peregra</i> ³	Czech Republic, Germany	[4,39], [18,40]
	<i>Radix</i> spp.	Belarus	[3]
<i>Plagiorchis vespertilionis</i> (Müller, 1780)	<i>Lymnaea stagnalis</i>	Belarus	[3]
<i>Plagiorchis</i> sp. 1 <i>sensu</i> Soldánová et al. [41]	<i>Radix balthica</i> ³	Iceland, Norway	[15 ¹], [41 ²]
<i>Plagiorchis</i> sp. 2 <i>sensu</i> Soldánová et al. [41]	<i>Radix balthica</i> ³	Finland, Iceland, Norway	[15 ¹], [15 ¹], [41 ²]
<i>Plagiorchis</i> sp. 3 <i>sensu</i> Soldánová et al. [41]	<i>Radix balthica</i> ³	Iceland, Ireland, Norway	[15 ¹], [15 ¹], [41 ²]
<i>Plagiorchis</i> sp. 4 <i>sensu</i> Soldánová et al. [41]	<i>Radix balthica</i> ³	Norway	[41 ²]
<i>Plagiorchis</i> sp. 5 <i>sensu</i> Soldánová et al. [41]	<i>Radix balthica</i> ³	Finland, Ireland, Norway	[15 ¹], [15 ¹], [41 ²]

<i>Plagiorchis</i> sp. 6 <i>sensu</i> Soldánová et al. [41]	<i>Radix balthica</i> ³	Ireland, Norway	[15 ¹], [41 ²]
<i>Plagiorchis</i> sp. 7 <i>sensu</i> Soldánová et al. [41]	<i>Myxas glutinosa</i>	Finland	[15 ¹]
	<i>Radix balthica</i> ³	Iceland, Ireland, Norway	[15 ¹], [15 ¹], [41 ²]
<i>Plagiorchis</i> sp. 8 <i>sensu</i> Kudlai et al. [15]	<i>Radix balthica</i> ³	Finland, Iceland, Ireland	[15 ¹]
<i>Plagiorchis</i> sp. 9 <i>sensu</i> Kudlai et al. [15]	<i>Stagnicola fuscus</i>	Ireland	[15 ¹]
<i>Plagiorchis</i> sp. CR <i>sensu</i> Zikmundová et al. [10]	<i>Lymnaea stagnalis</i>	Czech Republic	[10 ¹]
<i>Plagiorchis</i> sp. <i>sensu</i> Duan et al. [11]	<i>Lymnaea stagnalis</i>	Denmark	[11 ²]
	<i>Radix balthica</i> ³	Denmark	[11 ²]
	<i>Stagnicola palustris</i>	Denmark	[11 ²]
<i>Plagiorchis</i> sp. <i>sensu</i> Huguenin et al. [42]	<i>Lymnaea stagnalis</i>	France	[42 ²]
<i>Plagiorchis</i> sp. 1	<i>Lymnaea stagnalis</i>	Slovakia, Germany, Serbia	[2], [2,43], [43,44]
<i>Plagiorchis</i> sp. 2	<i>Lymnaea stagnalis</i>	Slovakia	[2]
<i>Plagiorchis</i> sp. II	<i>Radix ovata</i> ³	Russia	[36]
<i>Plagiorchis</i> sp.	<i>Galba truncatula</i>	Bulgaria	[45]
	<i>Lymnaea stagnalis</i>	Germany	[18]
	<i>Radix auricularia</i>	Germany, Switzerland	[46], [47]
	<i>Radix euphratica</i>	Georgia	[48]
	<i>Stagnicola corvus</i>	Ukraine	[38]

¹ Molecular (DNA sequences available) and morphological identification; ² Molecular identification only;

³ *Peregriana peregra* (former synonym *Radix peregra* and *R. labiata*); *Ampullaceana ampla* (former synonym *Radix ampla*), *A. balthica* (former synonym *Radix balthica* and *R. ovata*) according to Aksenova et al. [49] and Horsák et al. [50]

Supplementary Table S2. The composition of birds in Lakes Medard and Most based on literature data and online databases [51–61]. Definitive bird hosts of *Plagiorchis* spp. (see footnote) are provided based on records in the Host–parasite Database of the Natural History Museum, London [62] and additional literature data [63–69]. The reference numbering does not correspond to that in the main text.

Family	Lake Medard	Lake Most
	Species	
Accipitridae	<i>Buteo buteo</i> , <i>Circus aeruginosus</i> ¹ , <i>Milvus milvus</i>	<i>Accipiter nisus</i> ^{1,3} , <i>A. gentilis</i> ^{1,2} , <i>B. buteo</i> , <i>B. lagopus</i> ⁴ , <i>C. aeruginosus</i> ¹ , <i>C. cyaneus</i> ^{1,3} , <i>C. pygargus</i> , <i>Haliaeetus albicilla</i> ³ , <i>M. milvus</i> , <i>Milvus migrans</i> , <i>Pernis apivorus</i>
Acrocephalidae	<i>Acrocephalus scirpaceus</i> ¹ , <i>A. arundinaceus</i> ¹	<i>A. scirpaceus</i> ¹ , <i>A. schoenobaenus</i> ^{1,2} , <i>A. arundinaceus</i> ¹ , <i>A. palustris</i> , <i>Hippolais icterina</i> ¹
Aegithalidae	<i>Aegithalos caudatus</i>	<i>A. caudatus</i>
Alaudidae	<i>Alauda arvensis</i> ^{1,2}	<i>A. arvensis</i> ^{1,2} , <i>Lullula arborea</i>
Alcedinidae	<i>Alcedo atthis</i>	<i>A. atthis</i>
Anatidae	<i>Alopochen aegyptiacus</i> , <i>Anas clypeata</i> ^{1,3} , <i>A. crecca</i> ² , <i>A. penelope</i> ⁴ , <i>A. platyrhynchos</i> ^{1,4} , <i>A. strepera</i> , <i>Anser albifrons</i> ^{1,4} , <i>An. anser</i> , <i>An. fabalis</i> ³ , <i>Aythya ferina</i> ^{1,2} , <i>Ay. fuligula</i> ^{1,2} , <i>Ay. marila</i> ⁴ , <i>Bucephala clangula</i> ³ , <i>Clangula hyemalis</i> ^{1,4} , <i>Cygnus olor</i> , <i>Melanitta fusca</i> ³ , <i>M. nigra</i> ⁴ , <i>Mergellus albellus</i> ³ , <i>Mergus merganser</i> ^{1,3} , <i>Netta rufina</i>	<i>Al. aegyptiacus</i> , <i>A. acuta</i> ^{1,3} , <i>A. clypeata</i> ^{1,3} , <i>A. crecca</i> ² , <i>A. penelope</i> ⁴ , <i>A. platyrhynchos</i> ^{1,4} , <i>A. querquedula</i> ² , <i>A. strepera</i> , <i>An. albifrons</i> ^{1,4} , <i>An. anser</i> , <i>An. fabalis</i> ³ , <i>An. indicus</i> , <i>Ay. ferina</i> ^{1,2} , <i>Ay. fuligula</i> ^{1,2} , <i>Ay. marila</i> ⁴ , <i>Branta leucopsis</i> ⁴ , <i>B. clangula</i> ³ , <i>C. hyemalis</i> ^{1,4} , <i>Cy. olor</i> , <i>M. fusca</i> ³ , <i>M. nigra</i> ⁴ , <i>Me. albellus</i> ³ , <i>Me. merganser</i> ^{1,3} , <i>Mergus serrator</i> ³ , <i>Tadorna tadorna</i> , <i>T. ferruginea</i> , <i>N. rufina</i>
Apodidae	<i>Apus apus</i> ^{1,2}	<i>A. apus</i> ^{1,2}
Ardeidae	<i>Ardea cinerea</i> , <i>Botaurus stellaris</i> , <i>Egretta alba</i>	<i>A. cinerea</i> , <i>A. purpurea</i> , <i>Ardeola ralloides</i> , <i>Botaurus stellaris</i> , <i>Casmerodius albus</i> , <i>E. alba</i> , <i>Ixobrychus minutus</i>
Bombycillidae	<i>Bombycilla garrulus</i> ³	
Calcariidae	<i>Plectrophenax nivalis</i> ⁴	<i>P. nivalis</i> ⁴
Charadriidae		<i>Charadrius dubius</i> ¹ , <i>Vanellus vanellus</i> ^{1,2}
Ciconiidae		<i>Ciconia ciconia</i> , <i>Ciconia nigra</i>
Columbidae	<i>Columba livia</i> f. <i>domestica</i> , <i>Columba palumbus</i>	<i>Columba oenas</i> , <i>C. livia</i> f. <i>domestica</i> , <i>C. palumbus</i> , <i>Streptopelia turtur</i> , <i>Streptopelia decaocto</i>
Corvidae	<i>Corvus corax</i> ^{1,2} , <i>Garrulus glandarius</i> ¹ , <i>Pica pica</i> ^{1,2}	<i>C. corax</i> ^{1,2} , <i>C. corone</i> ^{1,2} , <i>C. cornix</i> ^{1,2} , <i>C. monedula</i> , <i>G. glandarius</i> ¹ , <i>P. pica</i> ^{1,2}
Cuculidae		<i>Cuculus canorus</i> ^{1,2}
Emberizidae	<i>Emberiza schoeniclus</i> ^{1,2} , <i>Emberiza citrinella</i> ^{1,2}	<i>E. schoeniclus</i> ^{1,2} , <i>E. hortulana</i> ¹ , <i>E. citrinella</i> ^{1,2} , <i>Miliaria calandra</i>
Falconidae	<i>Falco tinnunculus</i> ¹	<i>Falco peregrinus</i> ³ , <i>F. tinnunculus</i> ¹ , <i>F. subuteo</i> ^{1,2}
Fringillidae	<i>Carduelis carduelis</i> ¹ , <i>C. chloris</i> ² , <i>C. flammea flammea</i> ² , <i>C. spinus</i> ²	<i>C. cannabina</i> , <i>C. carduelis</i> ¹ , <i>C. chloris</i> ² , <i>C. flammea flammea</i> ² , <i>C. spinus</i> ² , <i>Coccothraustes coccothraustes</i> , <i>Fringilla coelebs</i> ¹
Gaviidae	<i>Gavia arctica</i> ³ , <i>Gavia immer</i> ³	<i>G. arctica</i> ³ , <i>G. stellata</i> ⁴
Gruidae		<i>Grus grus</i> ³
Hirundinidae	<i>Delichon urbicum</i> ^{1,2} , <i>Hirundo rustica</i> ^{1,2} , <i>Picus viridis</i> , <i>P. canus</i>	<i>D. urbicum</i> ^{1,2} , <i>Dendrocopos minor</i> , <i>Hirundo daurica</i> ¹ , <i>H. rustica</i> ^{1,2} , <i>Jynx torquilla</i> , <i>P. viridis</i> , <i>Riparia riparia</i> ^{1,2}
Laniidae	<i>Lanius excubitor</i> ^{1,2}	<i>Lanius collurio</i> ^{1,2} , <i>L. excubitor</i> ^{1,2}
Laridae	<i>Chlidonias niger</i> ¹ , <i>Larus argentatus</i> ^{1,4} ,	<i>C. niger</i> ¹ , <i>Chroicocephalus ridibundus</i> ² , <i>L.</i>

	<i>L. cachinnans</i> , <i>L. canus</i> ^{1,3} , <i>L. fuscus</i> ^{1,4} , <i>L. ridibundus</i> ^{1,3} , <i>L. minutus</i> ¹ , <i>Sterna</i> <i>sandvicensis</i>	<i>argentatus</i> ^{1,4} , <i>L. cachinnans</i> , <i>L. canus</i> ^{1,3} , <i>L.</i> <i>fuscus</i> ^{1,4} , <i>L. ridibundus</i> ^{1,3} , <i>L. michahellis</i> , <i>L.</i> <i>melanocephalus</i> ¹
Locustellidae		<i>Locustella naevia</i>
Meropidae		<i>Merops apiaster</i> ¹
Motacillidae	<i>Anthus trivialis</i> ^{1,2} , <i>A. pratensis</i> ^{1,3} , <i>A.</i> <i>spinoletta</i> , <i>Motacilla alba</i> ^{1,2}	<i>A. campestris</i> , <i>A. trivialis</i> ^{1,2} , <i>A. pratensis</i> ^{1,3} , <i>A.</i> <i>spinoletta</i> , <i>M. alba</i> ^{1,2} , <i>M. flava</i> ^{1,2}
Muscicapidae	<i>Erithacus rubecula</i> ¹ , <i>Oenanthe</i> <i>oenanthe</i> ^{1,2} , <i>Saxicola torquatus</i> ^{1,3} , <i>S.</i> <i>rubetra</i> ^{1,2}	<i>E. rubecula</i> ¹ , <i>Luscinia svecica cyanecula</i> ³ , <i>L.</i> <i>megarhynchos</i> , <i>O. oenanthe</i> ^{1,2} , <i>Phoenicurus</i> <i>ochruros</i> , <i>S. torquatus</i> ^{1,3} , <i>S. rubetra</i> ^{1,2} , <i>Muscicapa</i> <i>striata</i> ^{1,2}
Oriolidae		<i>Oriolus oriolus</i> ¹
Pandionidae		<i>Pandion haliaetus</i> ³
Panuridae		<i>Panurus biarmicus</i>
Paridae	<i>Parus major</i> ^{1,2} , <i>P. caeruleus</i>	<i>P. major</i> ^{1,2} , <i>P. caeruleus</i>
Passeridae	<i>Passer domesticus</i> ^{1,2} , <i>P. montanus</i> ^{1,2}	<i>P. domesticus</i> ^{1,2} , <i>P. montanus</i> ^{1,2}
Phalacrocoracidae	<i>Phalacrocorax carbo</i>	<i>P. carbo</i>
Phasianidae		<i>Phasianus colchicus</i> , <i>Coturnix coturnix</i> ¹
Phylloscopidae	<i>Phylloscopus collybita</i> ¹ , <i>P. trochilus</i> ^{1,2}	<i>P. collybita</i> ¹ , <i>P. trochilus</i> ^{1,2}
Picidae	<i>Dendrocopos major</i> ^{1,2} , <i>Picus viridis</i> ¹ , <i>P.</i> <i>canus</i>	<i>D. major</i> ^{1,2} , <i>D. minor</i> ^{1,2} , <i>Jynx torquilla</i> ² , <i>P. viridis</i> ¹
Podicipedidae	<i>Podiceps auritus</i> , <i>P. cristatus</i> ¹ , <i>P.</i> <i>grisegena</i> ¹ , <i>P. nigricollis</i> ¹ , <i>Tachybaptus</i> <i>ruficollis</i> ¹	<i>P. auritus</i> , <i>P. cristatus</i> ¹ , <i>P. grisegena</i> ¹ , <i>P.</i> <i>nigricollis</i> ¹ , <i>T. ruficollis</i> ¹
Prunellidae		<i>Prunella modularis</i> ^{1,2}
Rallidae	<i>Fulica atra</i> , <i>Gallinula chloropus</i>	<i>F. atra</i> , <i>G. chloropus</i> , <i>Porzana porzana</i> ² , <i>Rallus</i> <i>aquaticus</i>
Remizidae	<i>Remiz pendulinus</i>	<i>R. pendulinus</i>
Scolopacidae	<i>Actitis hypoleucos</i> ^{1,2} , <i>Numenius</i> <i>arquata</i> ^{1,2} , <i>Tringa nebularia</i> ³	<i>A. hypoleucos</i> ^{1,2} , <i>Calidris alpina</i> ^{1,4} , <i>Gallinago</i> <i>gallinago</i> ² , <i>N. arquata</i> ^{1,2} , <i>Philomachus pugnax</i> ^{1,4} , <i>Tringa glareola</i> ^{1,2} , <i>T. nebularia</i> ³ , <i>T. ochropus</i> ^{1,3} , <i>T.</i> <i>totanus</i> ^{1,2} , <i>T. stagnatilis</i> ¹ , <i>T. erythropus</i> ⁴
Sittidae		<i>Sitta europaea</i> ^{1,2}
Strigidae		<i>Bubo bubo</i> ²
Sturnidae	<i>Sturnus vulgaris</i> ^{1,2}	<i>S. vulgaris</i> ^{1,2}
Sylviidae	<i>Sylvia atricapilla</i> ¹ , <i>S. communis</i>	<i>S. atricapilla</i> ¹ , <i>S. borin</i> ^{1,2} , <i>S. communis</i> , <i>S.</i> <i>curruca</i> ² , <i>S. nisoria</i> ¹
Troglodytidae	<i>Troglodytes troglodytes</i>	<i>T. troglodytes</i>
Turdidae	<i>Turdus pilaris</i> ^{1,2} , <i>T. merula</i> ¹	<i>T. pilaris</i> ¹ , <i>T. philomelos</i> ^{1,2} , <i>T. merula</i> ¹
Upupidae		<i>Upupa epops</i> ¹

¹ Possible definitive host of *Plagiorchis* spp.

² Native or breeding birds in Central Europe

³ Migratory birds (occur in Central Europe during a relatively short period of the year on migration between breeding and non-breeding areas)

⁴ Native or breeding birds in the sub-Arctic

Supplementary Table S3. Nucleotide comparison of *cox1* mtDNA sequences of *Plagiorchis* spp. (compatible with data from Europe) based on 324 nt long alignment. The percentage of *p*-distance is presented below the diagonal, while the count of variable nucleotides is displayed above the diagonal. Sequences generated in this study are indicated in bold. Provided as a separate file in the Excel format.

Supplementary Table S4. Nucleotide comparison of *cox1* mtDNA sequences of *Plagiorchis* spp. (compatible with data from North America) based on 371 nt long alignment. The percentage of *p*-distance is presented below the diagonal, while the count of variable nucleotides is displayed above the diagonal. Sequences generated in this study are indicated in bold. Provided as a separate file in the Excel format.

Supplementary Table S5. Nucleotide comparison of the partial 28S rDNA sequences of *Plagiorchis* spp. based on 1119 nt long alignment. The percentage of *p*-distance is presented below the diagonal, while the count of variable nucleotides is displayed above the diagonal. Sequences generated in this study are indicated in bold. Provided as a separate file in the Excel format.

Supplementary Table S6. Overview of newly generated sequences of snail hosts infected with *Plagiorchis* spp.

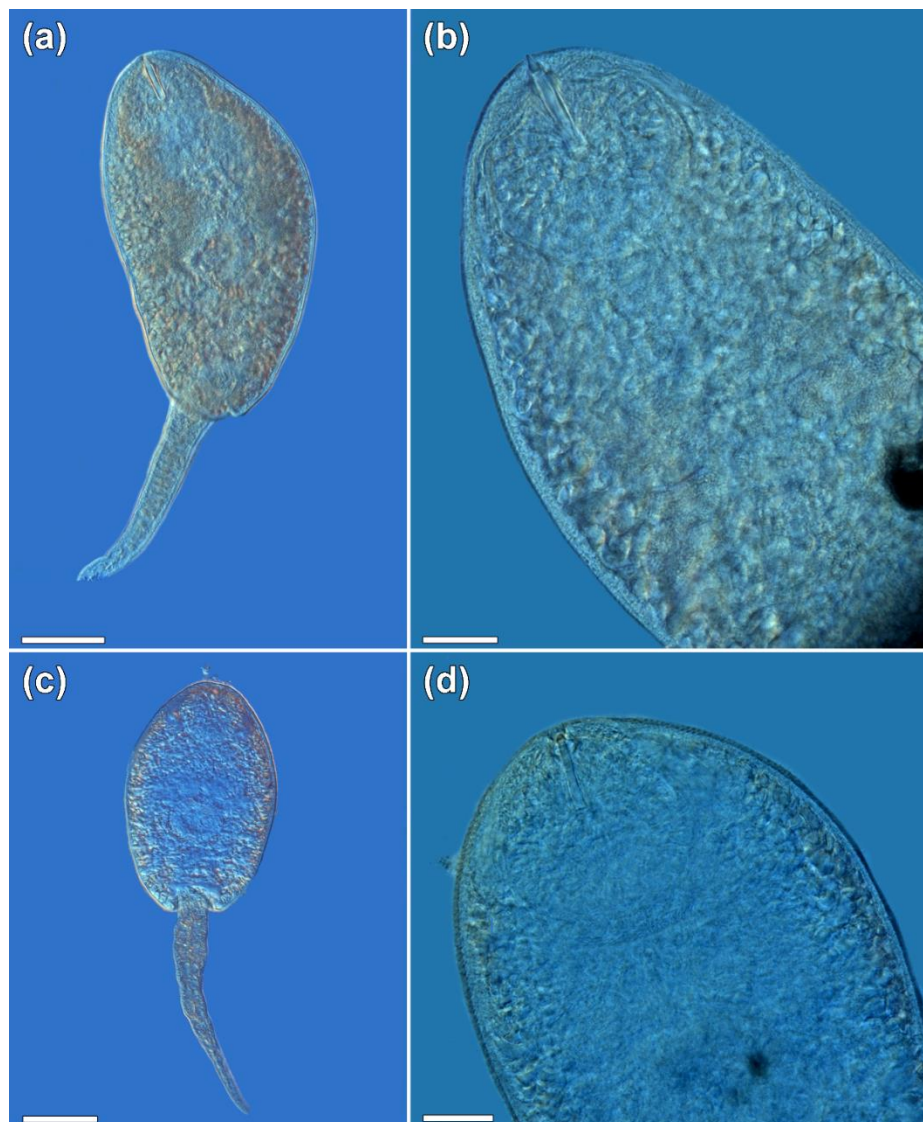
Locality	Snail host	Isolate	Genetic marker	GenBank accession numbers
Barbora	<i>Radix auricularia</i>	RA21	ITS2	PP391333
	<i>Radix auricularia</i>	RA22	ITS2	PP391334
Medard	<i>Ampullaceana balthica</i>	RA15	ITS2	PP391329
	<i>Ampullaceana balthica</i>	RA16	ITS2	PP391330
Milada	<i>Ampullaceana lagotis</i>	MIL1	ITS2	PP391341
	<i>Ampullaceana lagotis</i>	MIL2	ITS2	PP391342
Most	<i>Ampullaceana lagotis</i>	RA17	ITS2	PP391340
	<i>Ampullaceana lagotis</i>	RA18	ITS2	PP391339
Otakar	<i>Radix auricularia</i>	RA23	ITS2	PP391335
	<i>Radix auricularia</i>	RA24	ITS2	PP391336
Písník Dubina	<i>Radix auricularia</i>	RA19	ITS2	PP391331
	<i>Radix auricularia</i>	RA20	ITS2	PP391332
Spůle	<i>Radix auricularia</i>	RA27	ITS2	PP391337
	<i>Radix auricularia</i>	RA28	ITS2	PP391338

Supplementary Table S7. Comparative metrical data for cercariae of *Plagiorchis vespertilionis* isolated from different snail hosts (grouped as (1) – *Ampullaceana balthica*, (2) – *A. lagotis*, (3) – *Radix auricularia*). Data are presented in micrometers as minimum to maximum values, mean in parenthesis, and as a mathematical difference between the two respective mean values (symbol Δ). See Materials and Methods for abbreviations of cercarial morphological parameters.

Snail host	<i>A. balthica</i> (1)	<i>A. lagotis</i> (2)	Δ (1–2)	<i>A. balthica</i> (1)	<i>A. lagotis</i> (2)	<i>R. auricularia</i> (3)	Δ (1–2)	Δ (1–3)	Δ (2–3)	<i>A. lagotis</i> (2)
<i>N</i> snails	3	6	3–6	5	2	5	5–2	5–5	2–5	2
<i>N</i> cercariae	4	7	4–7	33	56	24	33–56	33–56	56–24	29
Fixation	Live	Live	Live	Ethanol	Ethanol	Ethanol	Ethanol	Ethanol	Ethanol	Formalin
ToL	332–425 (363)	337–420 (388)	-25	298–446 (371)	233–437 (358)	251–407 (349)	13	22	9	263–387 (328)
BL	190–320 (242)	226–303 (261)	-19	160–290 (215)	153–263 (212)	155–245 (206)	3	9	6	171–217 (190)
BW	162–202 (178)	124–178 (149)	29	91–141 (118)	70–138 (111)	96–145 (121)	7	-3	-10	103–135 (119)
TL	99–142 (121)	110–156 (128)	-7	99–200 (156)	102–174 (148)	96–185 (143)	8	13	5	77–195 (138)
TW	25–57 (39)	29–45 (37)	2	21–38 (30)	23–36 (28)	19–35 (28)	2	2	0	22–48 (32)
OSL	57–80 (72)	48–74 (65)	7	39–60 (50)	40–58 (48)	41–64 (51)	2	-1	-3	39–57 (48)
OSW	62–73 (69)	50–69 (62)	7	44–67 (55)	41–62 (52)	49–62 (55)	3	0	-3	48–63 (57)
VSL	35–57 (47)	38–51 (45)	2	28–49 (38)	28–44 (35)	30–47 (38)	3	0	-3	32–43 (38)
VSW	44–57 (51)	37–52 (46)	5	36–53 (45)	34–54 (42)	36–58 (44)	3	1	-2	40–52 (45)
OSW/VSW	1.3–1.4 (1.4)	1.3–1.4 (1.4)	0	0.9–1.5 (1.2)	0.9–1.7 (1.2)	1.0–1.5 (1.2)	0	0	0	1.1–1.6 (1.3)
TL/BL (%)	33.0–74.9 (53.8)	38.2–58.8 (49.4)	4.4	47.8–95.7 (73.3)	58.7–85.5 (70.3)	56.3–86.4 (69.5)	3	3.8	0.8	41.4–110.9 (73.5)
SL	30.0–31.5 (30.9)	30.2–31.8 (31.0)	-0.1	28–33 (30)	27–32 (30)	27–31 (29)	0	1	1	–
SWantt	7.1–9.0 (7.8)	7.2–8.2 (7.8)	0	–	–	–	–	–	–	–
SWabt	4.5–5.3 (5.0)	4.2–5.0 (4.8)	-0.2	–	–	–	–	–	–	–
SWbt	4.7–5.2 (5.2)	4.2–5.6 (4.8)	-0.4	–	–	–	–	–	–	–
SWantt/SL (%)	22.8–28.7 (25.3)	23.8–26.3 (25.2)	0.1	–	–	–	–	–	–	–

Supplementary Table S8. Comparative metrical data for cercariae of *Plagiorchis* sp. 10 isolated from different snail hosts (grouped as (1) – *Ampullaceana balthica*, (2) – *A. lagotis*). Data are presented in micrometers as minimum to maximum values, mean in parenthesis, and as a mathematical difference between the two respective mean values (symbol Δ). See Materials and Methods for abbreviations of cercarial morphological parameters.

Snail host	<i>A. balthica</i> (1)	<i>A. lagotis</i> (2)	Δ (1–2)	<i>A. balthica</i> (1)	<i>A. lagotis</i> (2)	Δ (1–2)	<i>A. lagotis</i> (2)
N snails	1	1	1–1	2	1	2–1	1
N cercariae	1	4	1–4	30	19	30–19	19
Fixation	Live	Live	Live	Ethanol	Ethanol	Ethanol	Formalin
ToL	418	353–513 (416)	2	244–353 (319)	287–356 (314)	5	225–316 (266)
BL	282	238–372 (302)	-20	145–227 (194)	171–203 (186)	8	124–165 (147)
BW	138	132–156 (150)	-12	71–100 (87)	75–102 (89)	-2	77–101 (92)
TL	137	87–141 (114)	23	99–150 (124)	104–153 (127)	-3	73–169 (119)
TW	39	39–53 (46)	-7	20–30 (25)	20–28 (25)	0	18–29 (23)
OSL	59	60–69 (65)	-6	35–52 (43)	35–47 (41)	2	36–42 (38)
OSW	66	57–69 (65)	1	36–55 (47)	42–53 (48)	-1	39–45 (42)
VSL	44	43–50 (47)	-3	27–45 (33)	27–34 (31)	2	24–33 (30)
VSW	45	50–58 (53)	-8	32–46 (38)	32–43 (38)	0	31–37 (35)
OSW/VSW	1.5	1.0–1.3 (1.2)	0.3	1.0–1.6 (1.3)	1.1–1.5 (1.3)	0	1.1–1.3 (1.2)
TL/BL (%)	48.6	30.3–48.7 (38.4)	10.2	52.8–92.1 (64.3)	53.6–88.9 (68.7)	-4.4	44.2–134.7 (82.5)
SL	–	28.8–30.3 (29.5)	–	26–29 (27)	27–29 (28)	-1	–
SWantt	–	6.7–6.8 (6.7)	–	–	–	–	–
SWabt	–	4.3–4.5 (4.4)	–	–	–	–	–
SWbt	–	5.1–5.2 (5.1)	–	–	–	–	–
SWantt/SL (%)	–	22.1–23.6 (22.8)	–	–	–	–	–



Supplementary Figure S1. Comparison of *Plagiorchis* cercariae fixed in ethanol and cold formalin used for the measurements. Ethanol-fixed cercariae, **(a)** body view; **(b)** anterior body with detail of stylet; formalin-fixed cercariae **(c)** body view; **(d)** anterior body with detail of stylet. Scale-bars in **(a)** and **(c)**: 50 μm ; Scale-bars in **(b)** and **(d)**: 20 μm .

References to Supplementary Materials

1. Manga-González, Y.; González-Lanza, C.; Kanev, I. *Lymnaea truncatula*, intermediate host of some Plagiorchiidae and Notocotylidae species in Leon, NW Spain. *J. Helminthol.* **1994**, *68*, 135–141. <https://doi.org/10.1017/S0022149X00013663>
2. Faltýnková, A.; Našincová, V.; Kablášková, L. Larval trematodes (Digenea) of the great pond snail, *Lymnaea stagnalis* (L.), (Gastropoda, Pulmonata) in Central Europe: a survey of species and key to their identification. *Parasite* **2007**, *14*, 39–51. <https://doi.org/10.1051/parasite/2007141039>
3. Akimova, L.N.; Shimalov, V.V.; Bychkova, E.I. Species diversity of trematode larvae of gastropods in water bodies of Belarus. *Parazitologiya* **2011**, *45*, 287–305. (in Russian)
4. Našincová, V. Trematode developmental stages in Czech aquatic snails and life–cycles of selected species of the family Omphalometridae and Echinostomatidae. PhD Thesis, Institute of Parasitology, Czechoslovak Academy of Sciences, České Budějovice, 1992. (in Czech)
5. Faltýnková, A. Larval trematodes (Digenea) in molluscs from small water bodies near České Budějovice, Czech Republic. *Acta Parasitol.* **2005**, *50*, 49–55.
6. Brown, R.; Soldánová, M.; Barrett, J.; Kostadinova, A. Small-scale to large-scale and back: larval trematodes in *Lymnaea stagnalis* and *Planorbis corneus* in Central Europe. *Parasitol. Res.* **2011**, *108*, 137–150. <https://doi.org/10.1007/s00436-010-2047-z>
7. Soldánová, M.; Faltýnková, A.; Scholz, T.; Kostadinova, A. Parasites in a man-made landscape: contrasting patterns of trematode flow in a fishpond area in Central Europe. *Parasitology* **2011**, *138*, 789–807. <https://doi.org/10.1017/S0031182011000291>
8. Soldánová, M.; Kostadinova, A. Rapid colonisation of *Lymnaea stagnalis* by larval trematodes in eutrophic ponds in central Europe. *Int. J. Parasitol.* **2011**, *41*, 981–990. <https://doi.org/10.1016/j.ijpara.2011.05.005>
9. Soldánová, M.; Kuris, A.M.; Scholz, T.; Lafferty, K.D. The role of spatial and temporal heterogeneity and competition in structuring trematode communities in the great pond snail, *Lymnaea stagnalis* (L.). *J. Parasitol.* **2012**, *98*, 460–471. <https://doi.org/10.1645/ge-2964.1>
10. Zikmundová, J.; Georgieva, S.; Faltýnková, A.; Soldánová, M.; Kostadinova, A. Species diversity of *Plagiorchis* Lühe, 1899 (Digenea: Plagiorchiidae) in lymnaeid snails from freshwater ecosystems in central Europe revealed by molecules and morphology. *Syst. Parasitol.* **2014**, *88*, 37–54. <https://doi.org/10.1007/s11230-014-9481-8>
11. Duan, Y.; Al-Jubury, A.; Kania, P.W.; Buchmann, K. Trematode diversity reflecting the community structure of Danish freshwater systems: molecular clues. *Parasit. Vectors* **2021**, *14*, 43. <https://doi.org/10.1186/s13071-020-04536-x>
12. Niewiadomska, K.; Valtonen, E.T.; Siddall, R. Cercariae from *Lymnaea stagnalis* in lake Kuuhankavesi (central Finland). *Acta Parasitol.* **1997**, *42*, 132–137.
13. Väyrynen, T.; Siddall, R.; Valtonen, E.T.; Taskinen, J. Patterns of trematode parasitism in lymnaeid snails from northern and central Finland. *Ann. Zool. Fennici* **2000**, *37*, 189–199.
14. Voutilainen, A. Interactive effects of predation risk and parasitism on the circadian rhythm of foraging activity in the great pond snail *Lymnaea stagnalis* (Gastropoda: Lymnaeidae). *Ann. Limnol.-Int. J. Lim.* **2010**, *46*, 217–223. <https://doi.org/10.1051/limn/2010025>
15. Kudlai, O.; Pantoja, C.; O'Dwyer, K.; Jouet, D.; Skírnisson, K.; Faltýnková, A. Diversity of *Plagiorchis* (Trematoda: Digenea) in high latitudes: Species composition and snail host spectrum revealed by integrative taxonomy. *J. Zool. Syst. Evol. Res.* **2021**, *59*, 937–962. <https://doi.org/10.1111/jzs.12469>
16. Odening, K. Notizen über Xiphidiozorkarien (Trematoda: Plagiorchiata, Larvae) aus Brandenburg und Sachsen. *Monatsber. Dtsch. Akad. Wiss. Berlin* **1962**, *4*, 300–311.
17. Bursian-Hartung, G. Untersuchungen über die Cercarienfauna des Dieskauer Teichgebietes bei Halle. *Hercynia-Ökolog. Mitt.* **1965**, *2*, 63–111. <http://dx.doi.org/10.25673/93617>
18. Faltýnková, A.; Haas, W. Larval trematodes in freshwater molluscs from the Elbe to Danube rivers (Southeast Germany): before and today. *Parasitol. Res.* **2006**, *99*, 572–582. <https://doi.org/10.1007/s00436-006-0197-9>
19. Soldánová, M.; Selbach, C.; Sures, B.; Kostadinova, A.; Pérez-del-Olmo, A. Larval trematode communities in *Radix auricularia* and *Lymnaea stagnalis* in a reservoir system of the Ruhr River. *Parasit. Vectors* **2010**, *3*, 56. <https://doi.org/10.1186/1756-3305-3-56>
20. Schwelm, J.; Selbach, C.; Kremers, J.; Sures, B. Rare inventory of trematode diversity in a protected natural reserve. *Sci. Rep.* **2021**, *11*, 22066. <https://doi.org/10.1038/s41598-021-01457-2>
21. Styczyńska-Jurewicz, E. The life cycle of *Plagiorchis elegans* (Rud., 1802) and the revision of the genus *Plagiorchis* Lühe, 1889. *Acta Parasitol. Pol.* **1962**, *10*, 419–445.

22. Żbikowska, E.; Żbikowski, J. Differences in shell shape of naturally infected *Lymnaea stagnalis* (L.) individuals as the effect of the activity of digenetic trematode larvae. *J. Parasitol.* **2005**, *91*, 1046–1051. <https://doi.org/10.1645/ge-420r1.1>
23. Żbikowska, E.; Kobak, J.; Żbikowski, J.; Kąklewski, J. Infestation of *Lymnaea stagnalis* by digenetic flukes in the Jeziorak Lake. *Parasitol. Res.* **2006**, *99*, 434–439. <https://doi.org/10.1007/s00436-006-0178-z>
24. Żbikowska, E. Digenea species in chosen populations of freshwater snails in northern and central part of Poland. *Wiad. Parazytol.* **2007**, *53*, 301–308.
25. Ginetsinskaya, T.A.; Dobrovolskij, A.A. To the fauna of trematode larvae from freshwater molluscs of the Volga delta. III. Furcocercariae (families Cyathocotylidae) and stylet cercariae (Xiphidiocercariae). *Trudy Astrakhan. Zapov.* **1968**, *11*, 29–95. (in Russian)
26. Zdun, V.I. *Larvae of trematodes in freshwater molluscs of Ukraine*. Ukrainian Academy of Sciences Press: Kiev, Ukraine, 1961. (in Ukrainian)
27. Zhytova, E.P. The first record of parthenitae and cercariae of *Plagiorchis multiglandularis* (Trematoda, Plagiorchiidae) in *Lymnaea stagnalis* in Ukraine. *Vestn. Zool.* **2018**, *52*, 289–294. <https://doi.org/10.2478/vzoo-2018-0030>
28. Selbach, C.; Soldánová, M.; Feld, C.K.; Kostadinova, A.; Sures, B. Hidden parasite diversity in a European freshwater system. *Sci. Rep.* **2020**, *10*, 2694. <https://doi.org/10.1038/s41598-020-59548-5>
29. Stenko, R.P. Role of some species of freshwater molluscs of the Crimea in the biology of trematodes. *Zool. Zhurnal* **1978**, *57*, 658–663. (in Russian)
30. Žďárská Z. The developmental cycle of the trematode *Plagiorchis laricola* (Skrjabin, 1924). *Acta Soc. Zool. Bohem.* **1966**, *20*, 179–184. (in German)
31. Samnaliev, P.; Dimitrov, V.; Bušta, J. Argentophilic integumentary structures of *Plagiorchis laricola* Skrjabin, 1924 cercariae. *Folia Parasitol.* **1983**, *30*, 329–334.
32. Vojtek J. The present situation of the research into the stages of development of trematodes in Czechoslovakia. *Scripta Fac. Sci. Nat. Univ. Purk. Brun.* **1989**, *19*, 339–352.
33. Mutafova, T. Karyological studies on some species of the families Echinostomatidae and Plagiorchiidae and aspects of chromosome evolution in trematodes. *Syst. Parasitol.* **1994**, *28*, 229–238. <https://doi.org/10.1007/bf00009520>
34. Bušta, J. Chaetotaxy of the cercaria of *Plagiorchis maculosus* (Rudolphi, 1802) (Trematoda, Plagiorchiidae). *Folia Parasitol.* **1987**, *34*, 219–224.
35. Khan, D. Studies on larval trematodes infecting freshwater snails in London (UK) and some adjoining areas. Xiphidiocercariae. *Z. Parasitenkd.* **1962**, *21*, 71–87. <https://doi.org/10.1007/bf00260178>
36. Frolova, E.N. *Trematode larvae from molluscs of the lakes in South Karelia*; Nauka: Leningrad, Russia, 1975. (in Russian)
37. Zhytova, O.P. Cercaria of the trematode *Plagiorchis mutationis* (Trematoda, Plagiorchiidae) from the pond snails, *Lymanea stagnalis*, in Ukraine. *Vestn. Zool.* **2010**, *44*, 29–32. <https://doi.org/10.2478/v10058-010-0022-x>
38. Zhytova, E.P.; Romanchuk, L.D.; Gural'ska, S.V.; Andreieva, O.Y.; Shvets, M.V. Circulation pathways of trematodes of freshwater gastropod mollusks in forest biocenoses of the Ukrainian Polissia. *Vestn. Zool.* **2019**, *53*, 13–22. <https://doi.org/10.2478/vzoo-2019-0002>
39. Bušta, J.; Našincová, V. Record of *Plagiorchis neomidis* Brendow, 1970 (Trematoda: Plagiorchiidae) in Czechoslovakia and studies on its life cycle. *Folia Parasitol.* **1986**, *33*, 123–129.
40. Brendow, V. Ein Beitrag zur Trematodenfauna der Soricidae im Raume Giessen sowie im Naturpark Hoher Vogelsberg. Teil I. *Z. Parasitenkd.* **1970**, *33*, 282–313. <https://doi.org/10.1007/bf00331467>
41. Soldánová, M.; Georgieva, S.; Roháčová, J.; Knudsen, R.; Kuhn, J.A.; Henriksen, E.H.; Siwertsson, A.; Shaw, J.C.; Kuris, A.M.; Amundsen, P.A.; Scholz, T.; Lafferty, K.D.; Kostadinova, A. Molecular analyses reveal high species diversity of trematodes in a sub-Arctic lake. *Int. J. Parasitol.* **2017**, *47*, 327–345. <https://doi.org/10.1016/j.ijpara.2016.12.008>
42. Huguenin, A.; Depaquit, J.; Villena, I.; Ferté, H. MALDI-TOF mass spectrometry: a new tool for rapid identification of cercariae (Trematoda, Digenea). *Parasite* **2019**, *26*, 11. <https://doi.org/10.1051/parasite/2019011>
43. Bock, D. The life cycle of *Plagiorchis* spec. 1, a species of the *Plagiorchis elegans* group (Trematoda, Plagiorchiidae). *Z. Parasitenkd.* **1984**, *70*, 359–373. <https://doi.org/10.1007/bf00927822>
44. Bock, D. Xiphidiocercariae (Trematoda, Larvae) from the «Obedska Bara» nature reserve near Belgrade, Yugoslavia. *Proc. on the Fauna of SR Serbia, Serbian Acad. Of Sci. and Arts, Belgrade* **1985**, *3*, 21–69.
45. Radev, V.; Hrusanov, D.; Georgieva, K.; Bankov, I. *Galba truncatula* (Müller, 1774). A factor for spreading of fasciolosis. *C. R. Acad. Bulg. Sci.* **2018**, *61*, 1033–1036.
46. Allgöwer, R. Zur Trematodenfauna einiger Freiburger Baggerseen, mit besonderer Berücksichtigung des Erregers der Zerkariendermatitis beim Menschen. *Mitt. Bad. Landesv. Naturk. Natursch.* **1990**, *15*, 59–79.

47. Eklun-Natey, D.T.; Al-Khudry, M.; Gauthey, D.; Wüest, J.; Vaucher, C.; Huggel, H. Contribution to the study of the cercariae in the Lake of Geneva. II. Simple tail cercariae. *Rev. Suisse Zool.* **1985**, *92*, 927–938. (in French)
48. Arabuli, L.; Murvanidze, L.; Faltýnková, A.; Mumladze, L. Checklist of digeneans (Platyhelminthes, Trematoda, Digenea) of Georgia. *Biodivers. Data J.* **2024**, *12*, e110204. <https://doi.org/10.3897/BDJ.12.e110204>
49. Aksenova, O.V.; Bolotov, I.N.; Gofarov, M.Y.; Kondakov, A.V.; Vinarski, M.V.; Bepalaya, Y.V.; Kolosova, Y.S.; Palatov, D.M.; Sokolova, S.E.; Spitsyn, V.M.; Tomilova, A.A.; Travina, O.V.; Vikhrev, I.V. Species richness, molecular taxonomy and biogeography of the Radicine pond snails (Gastropoda: Lymnaeidae) in the Old World. *Sci. Rep.* **2018**, *8*, 11199. <https://doi.org/10.1038/s41598-018-29451-1>
50. Horsák, M.; Juříčková, L.; Beran, L.; Čejka, T.; Dvořák, L. Annotated list of mollusc species recorded outdoors in the Czech and Slovak Republics. *Malacol. Bohemoslov.* **2010**, *1*, 1–37. <https://doi.org/10.5817/mab2010-9-s1-v2> (in Czech)
51. Bažant, J. Waterbirds on lake Most 2013–2015. *Sbor. Obl. Muz. Most., ř. př.* **2015**, *37*, 61–80. [in Czech]
52. Bažant, J. Birds of wetland „Bettyňka“ near Vtelno (northwestern Bohemia, Most). *Sbor. Obl. Muz. Most., ř. př.* **2017**, *38*, 92–99. (in Czech)
53. Bažant, J. Birds of wetland „Za pilou“ (Most district, Northwestern Bohemia). *Sbor. Obl. Muz. Most., ř. př.* **2017**, *38*, 100–107. (in Czech)
54. Bažant, J. Interesting ornithological observations in the Most district (Northwestern Bohemia). *Sbor. Obl. Muz. Most., ř. př.* **2018**, *39*, 143–153. (in Czech)
55. Bažant, J. Birds of the sandpit in Polerady village (Most county, Northwestern Bohemia). *Sbor. Obl. Muz. Most., ř. př.* **2020**, *40*, 122–134. (in Czech)
56. Neruda, M.; Filipová, L.; Ambrožová, J.Ř.; Machová, I.; Kubát, K.; Holec, M.; Holcová, D. Ecological research of former brown-coal quarry – the Most Lake in the Czech Republic. *J. Life Sci.* **2014**, *8*, 841–847.
57. Holcová, D.; Holec, M. Bird communities of the Ležáky lake: present and future importance of this ornithology interesting locality. *Stud. Oecol.* **2012**, *4*, 62–72. (in Czech)
58. Česká společnost ornitologická. AVIF database. Available online: <https://avif.birds.cz/> (accessed on 22 January 2024). (in Czech)
59. The Cornell Lab of Ornithology. Birds of the World. Available online: <https://birdsoftheworld.org/bow/home> (accessed on 22 January 2024).
60. BirdLife International. Available online: <https://www.birdlife.org/> (accessed on 22 January 2024).
61. Pozorování ptáků. Available online: <https://pozorovaniptaku.cz/> (accessed on 22 January 2024). (in Czech)
62. Gibson, D.I.; Bray, R.A.; Harris, E.A. (Compilers). Host-Parasite Database of the Natural History Museum. London, 2005. Available online: <https://www.nhm.ac.uk/research-curation/scientific-resources/taxonomy-systematics/host-parasites/index.html> (accessed on 22 January 2024).
63. Sitko, J.; Faltýnková, A.; Scholz, T. *Checklist of the Trematodes (Digenea) of birds of the Czech and Slovak Republics*. Academia, Prague, Czech Republic, 2006, ISBN 802-001-428-4.
64. Rząd, I.; Dzika, E.; Krupa, R. *Actitis hypoleucos* (L., 1758) – new Polish host of the trematodes: *Plagiorchis nanus* (Rudolphi, 1802) and *Leucochloridium perturbatum* Pojmanska, 1969. *Wiad. Parazytol.* **2011**, *57*, 37–41.
65. Okulewicz, A.; Sitko, J. Parasitic helminthes – probable cause of death of birds. *Helminthologia*, **2012**, *49*, 241–246. <https://doi.org/10.2478/s11687-012-0045-7>
66. Sitko, J.; Heneberg, P. Host specificity and seasonality of helminth component communities in central European grebes (Podicipediformes) and loons (Gaviiformes). *Parasitol. Int.* **2015**, *64*, 377–388. <https://doi.org/10.1016/j.parint.2015.05.012>
67. Komorová, P.; Sitko, J.; Špakulová, M.; Hurníková, Z.; Salamatin, R.; Chovancová, G. New data on helminth fauna of birds of prey (Falconiformes, Accipitriformes, Strigiformes) in the Slovak Republic. *Helminthologia* **2017**, *54*, 314–321. <https://doi.org/10.1515/helm-2017-0038>
68. Kirillov, A.A.; Kirillova, N.Y.; Spiridonov, S.N. Trematodes of land birds from the Republic of Mordovia with a checklist of avian trematodes of the Middle Volga Region (European Russia). *Diversity* **2023**, *15*, 330, 1–19. <https://doi.org/10.3390/d15030330>
69. Kirillova, N.Y.; Ruchin, A.B.; Kirillov, A.A.; Chikhlyayev, I.V.; Alpeev, M.A. Overview of helminths in land vertebrates from the Mordovia Nature Reserve, European Russia. *Nat. Env. & Poll. Tech.* **2023**, *22*, 1667–1690. <https://doi.org/10.46488/NEPT.2023.v22i04.001>