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Supplementary Information

Development of a Multiplex Sandwich Aptamer Microarray for the Detection of $VEGF_{165}$ and Thrombin

Alice Sosic¹, Anna Meneghello², Agnese Antognoli², Erica Cretaio² and Barbara Gatto^{1,*}

- ¹ Dipartimento di Scienze del Farmaco, Universit`à di Padova, via Marzolo 5, 35131 Padova, Italy;
 E-Mail: alice.sosic@studenti.unipd.it
- ² Veneto Nanotech S.C.p.A., Via S. Crispino 106, I -35129 Padova, Italy;
 E-Mails: anna.meneghello@venetonantoech.it (A.M.); agnese.antognoli@venetonanotech.it (A.A.);
 erica.cretaio@venetonanotech.it (E.C.)
- * Author to whom correspondence should be addressed; E-Mail: barbara.gatto@unipd.it; Tel.: +39-049-827-5717; Fax: +39-049-827-5366.

Figure S1. Images of a microarray slide after the incubation of Alexa555-labeled VEGF₁₆₅. Vap7(12T)NH₂ and Vap7-NH₂ were anchored on the glass slide as capture layer on the left and on the right, respectively. The green fluorescence (due to Alexa555 fluorophore) represents the bound labeled protein.

Capture layer	Vap7	Vap7 NH ₂	
	(12T)NH ₂		
Chamber			
Protein	Alexa555-labeled VEGF 165		

Sub-array	1		2	2	3 4			
Capture layer	Vap7 (12T)NH ₂	Random DNA-NH ₂	Vap7 (12T)NH ₂	Random DNA-NH ₂	Vap7 (12T)NH ₂	Random DNA-NH ₂	Vap7 (12T)NH ₂	Random DNA-NH ₂
Protein	VEGF ₁₆₅ ((0,1µM)	Alexa555-VEGF ₁₆₅ (0,1µM) pre-complexed with VEa5-Cy5(0,5µM)		VEGF ₁₆₅ (0,1µM)		Alexa555-VEGF ₁₆₅ (0,1µM)	
Detection layer	vEa5-Cy5	exed with (0,5μM)			VEa5-Cy5(0,5μM)		VEa5-Cy5(0,5μM)	
Procedure		On	ie-step			Tw	o-steps	

Table S1. Schematic representation of the SAM protocol for VEGF detection.

Figure S2. Images of the microarray slide for the Sandwich Aptamer Microarray (SAM) for VEGF. In each subarray, Vap7(12T)NH₂ was printed on the left while the negative control (Random DNA-NH₂) was printed on the right. Red fluorescence represents binding by VEa5-Cy5. In chambers 2 and 4, in which Alexa 555-labeled VEGF₁₆₅ (500 nM) was incubated, the fluorescence of the yellow spots indicates the simultaneous co-localization of the protein (green) and of the detection aptamer (red).

	Two-steps protocol			One-step protocol				
	4		3		2		1	
	Random	Vap7	Random	Vap7	Random	Vap7	Random	Vap7
	DNA-NH ₂	(12T)NH ₂						
Red fluorescence (Cy5)								
Green and red fluorescence merge (Alexa 555 + Cy5)						•••		

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