

SUPPLEMENTARY INFORMATION

**A Schiff Base Fluorescence Enhancement Probe for Fe(III) and Its Sensing Applications in
Cancer Cells**

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This file includes:

Synthesis of **FeP-1**

Supporting Scheme: Scheme S1

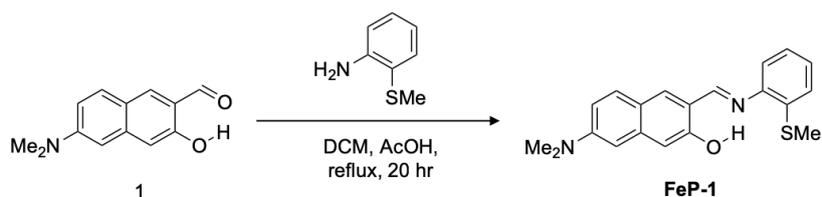
Supporting Figures: Fig. S1 to S10

¹H and ¹³C NMR for **FeP-1**

High resolution mass spectra for **FeP-1**

References and notes

Synthesis of FeP-1.



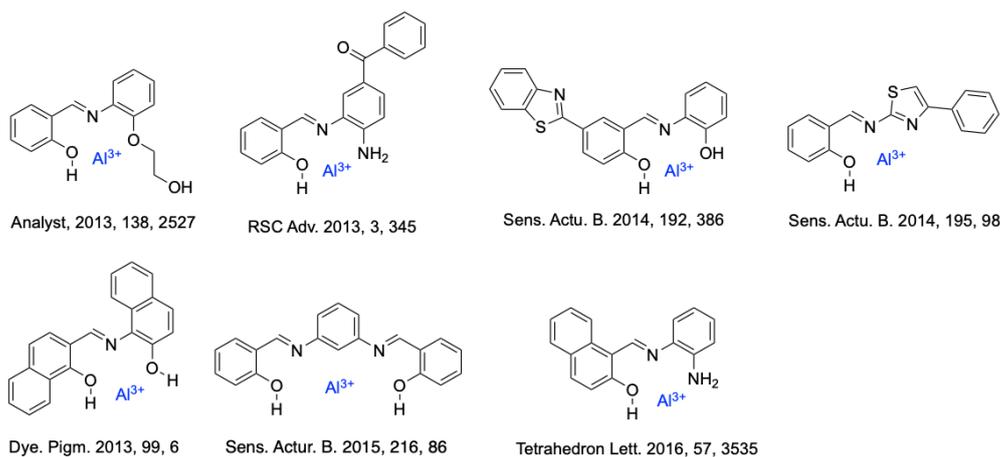
Scheme S1. Synthesis of **FeP-1**: Compound **1** (30 mg, 0.139 mmol), 2-(methylthio)-aniline (26.128 μ L, 0.209 mmol, 1.5 eq), MgSO₄ (83.63 mg, 0.695 mmol, 5 eq), Dichloromethane (DCM, 2 mL), Acetic acid (0.1 mL), reflux for 20 hr, 69% yield.

Synthesis of compound 1. We synthesized the compound 1 by following the reported method by our group. [Ref: 1. Kim, I.; Kim, D.; Sambasivan, S.; Ahn, K. H., Synthesis of π -Extended Coumarins and Evaluation of Their Precursors as Reactive Fluorescent Probes for Mercury Ions. *Asian J. Org. Chem.* **2012**, *1*, 60–64]

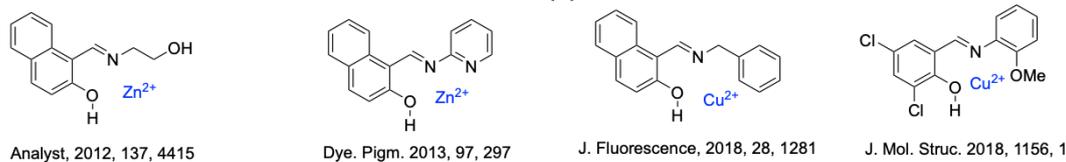
Synthesis of FeP-1. Compound **1** (30 mg, 0.139 mmol), 2-(methylthio)-aniline (26.128 μ L, 0.209 mmol) and DCM (2 mL), magnesium sulfate (83.63 mg, 0.695 mmol), and acetic acid (0.1 mL) were combined at room temperature (25 °C). The reaction was held at reflux temperature overnight. The reaction mixture was washed with brine, and dried over MgSO₄. All the volatile components were removed in vacuo, and the crude residue was purified by column chromatography (*n*-hexane/ethyl acetate = 8:2) to give **FeP-1** (32.3 mg, 69%). ¹H NMR (400 MHz, DMSO-d₆): δ 12.55 (s, 1H), 8.97 (s, 1H), 7.73 (s, 1H), 7.70 (s, 1H), 7.44 (s, 1H), 7.42 (s, 1H), 7.31 (s, 1H), 7.30 (s, 1H), 7.07 (s, 1H), 7.05 (s, 1H), 6.98 (s, 1H), 3.05 (t, 6H), 2.47 (t, 3H); ¹³C NMR (400 MHz, DMSO-d₆): δ 14.82, 40.44, 103.75, 109.00, 113.74, 116.96, 118.07, 120.89, 124.92, 125.25, 127.12, 129.83, 134.39, 135.06, 138.61, 145.70, 150.34, 157.40, 161.44. HRMS (*m/z*): [M+Na]⁺ calcd for C₂₀H₂₀N₂OS, 336.1206; found, 336.1206.

Supporting Figures

(a)

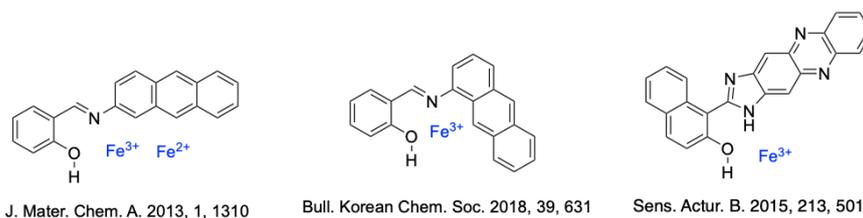


(b)



(c)

(d)



(e)

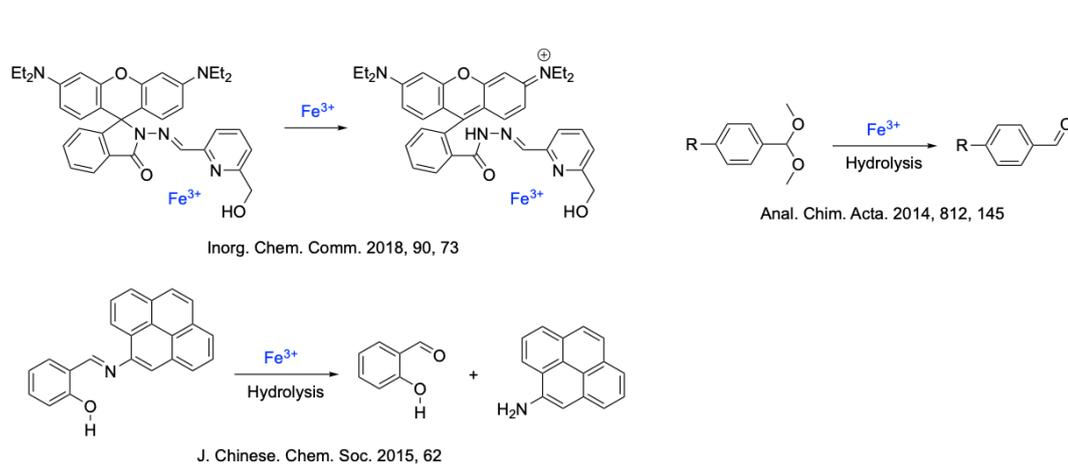


Figure S1. (a–d) Reported Schiff-based fluorescent probes, which have similar structure of FeP-1, for metal ions. (a) Al(III) probes. (b) Zn(II) probes. (c) Cu(II) Probes. (d) Fe(III), Fe(II) probes. (e) Reaction-based fluorescent probes for Fe(III). The references are fully addressed in References and notes.

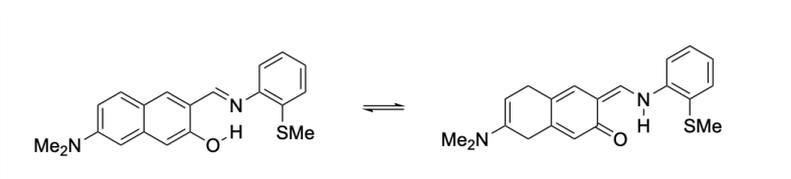


Figure S2. Excited state intramolecular proton transfer (ESIPT) mechanism of **FeP-1**.

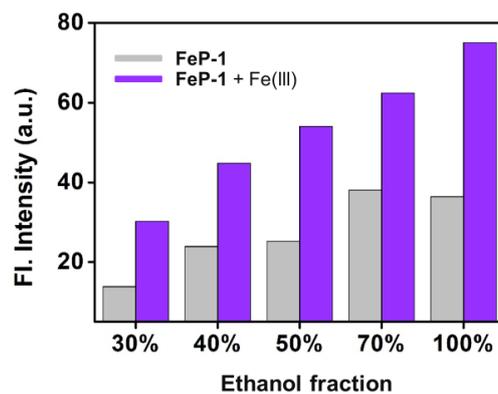


Figure S3. Fluorescence intensity plot (peak height at 550 nm) of **FeP-1** (10 μ M) upon addition of Fe(III) (20 eq) in various ethanol-water solution (30–100% EtOH), measured after mixing (within 1 min) at 25 °C. The emission spectrum was measured under excitation at 407 nm.

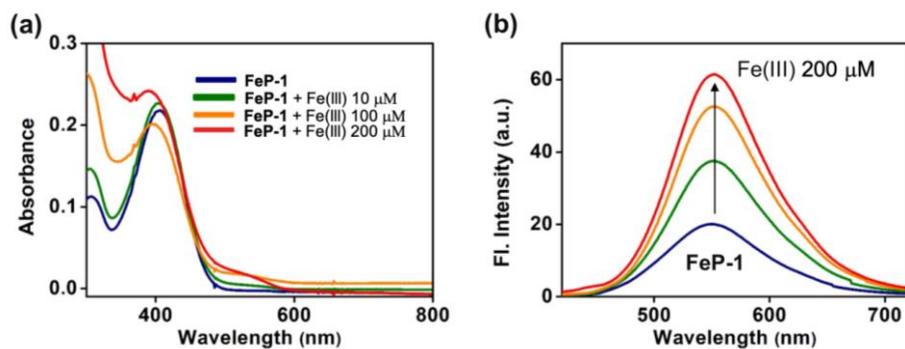


Figure S4. (a) Absorption and (b) Emission spectra of FeP-1 (10 μM) upon addition of Fe(III) (0–200 μM) in ethanol-water solution (EtOH:DI H₂O = 4:6, *v/v*), measured after 1 min at 25 °C. The emission spectrum was measured under excitation at 397 nm.

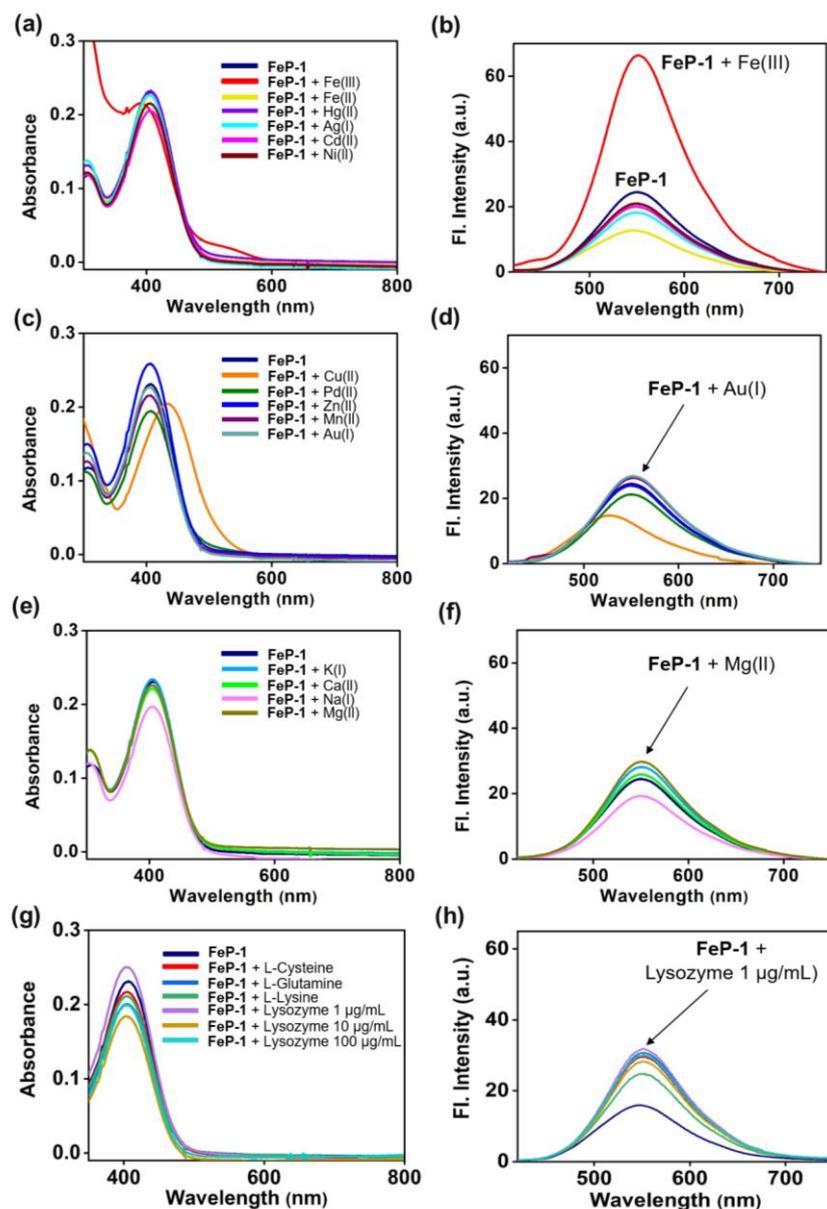


Figure S5. (a, c, e, g) Absorption and (b, d, f, h) Emission spectra of **FeP-1** (10 μM) upon addition of each metal ions (20 eq), amino acid (20 eq), and lysozyme (1–100 $\mu\text{g/mL}$) in ethanol-water solution (EtOH:DI H₂O = 4:6, *v/v*), measured after 1 min at 25 °C. Substrate: FeCl₂, Hg(NO₃)₂, AgCl, CdCl₂, NiCl₂, CuCl₂, PdCl₂, ZnCl₂, MnCl₂, KCl, CaCl₂, NaCl, (C₂H₅)₃PAuCl, MgCl₂, L-Cysteine, L-Glutamine, L-Lysine, Lysozyme 1 $\mu\text{g/mL}$, Lysozyme 10 $\mu\text{g/mL}$, Lysozyme 100 $\mu\text{g/mL}$. The emission spectrum was measured under excitation at 397 nm.

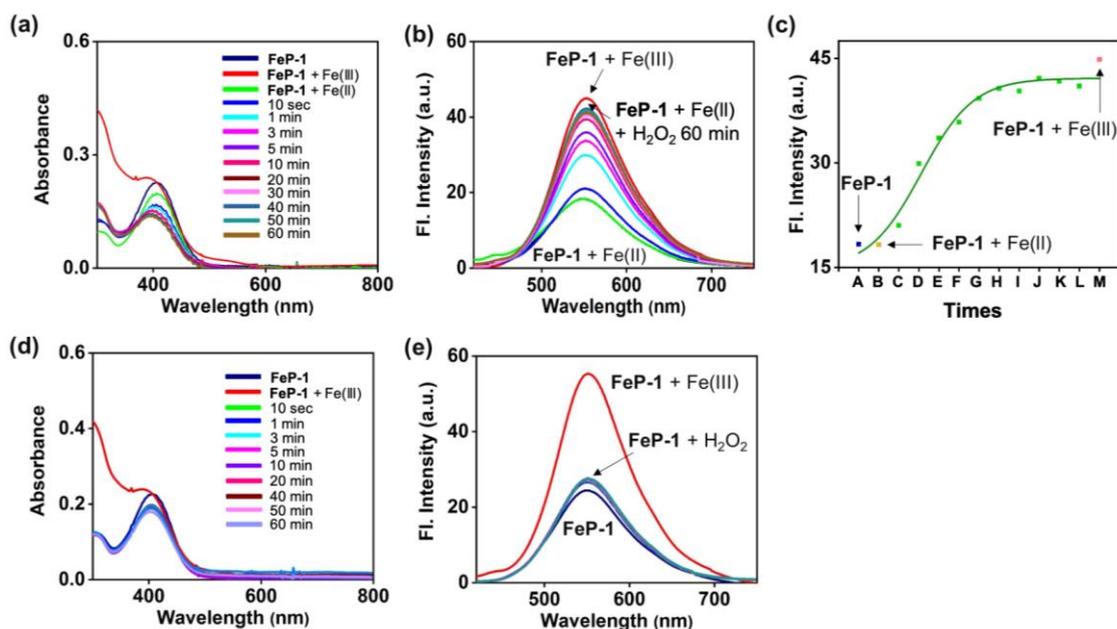


Figure S6. Time-dependent (a) absorption and (b) emission spectra changes of **FeP-1** (10 μM) with **Fe(II)** (20 eq) and **H₂O₂** (20 eq) in ethanol-water solution (EtOH:DI H₂O = 4:6, *v/v*). Control: **FeP-1** (10 μM) with **Fe(III)**. (c) Time-course emission intensity plot at maximum wavelength from panel (b). (A) **FeP-1**, (B) **FeP-1+Fe(II)**, (C) **FeP-1+Fe(II)+Hydrogen peroxide**; 10 s, (D) **FeP-1+Fe(II)+Hydrogen peroxide**; 1 min, (E) **FeP-1+Fe(II)+Hydrogen peroxide**; 3 min, (F) **FeP-1+Fe(II)+Hydrogen peroxide**; 5 min, (G) **FeP-1+Fe(II)+Hydrogen peroxide**; 10 min, (H) **FeP-1+Fe(II)+Hydrogen peroxide**; 20 min, (I) **FeP-1+Fe(II)+Hydrogen peroxide**; 30 min, (J) **FeP-1+Fe(II)+Hydrogen peroxide**; 40 min, (K) **FeP-1+Fe(II)+Hydrogen peroxide**; 50 min, (L) **FeP-1+Fe(II)+Hydrogen peroxide**; 60 min, (M) **FeP-1+Fe(III)**. (d, e) Time-dependent absorption and emission spectra of **FeP-1** (10 μM) with **H₂O₂** (20 eq). Absorption and emission spectrum was measured at 10 s, 1, 3, 5, 10, 20, 30 min after mixing together. Each spectrum was acquired in ethanol-water solution (EtOH:DI H₂O = 4:6, *v/v*) at 25 °C under excitation at 397 nm.

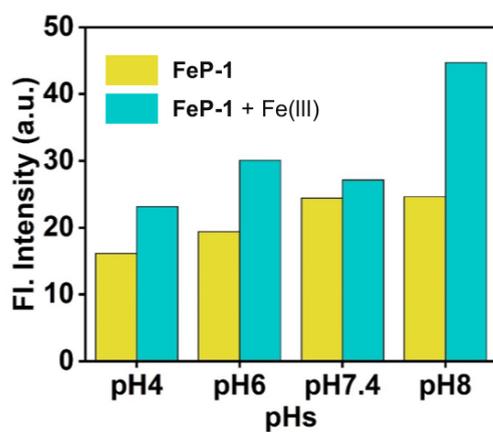


Figure S7. Emission intensity plot (peak height at 485 nm) of FeP-1 (10 μ M) upon addition of Fe(III) (20 eq) in various pHs (pH 4, 6, 7.4, 8), measured after 10 min at 25 $^{\circ}$ C. The emission spectra were measured under excitation at 397 nm.

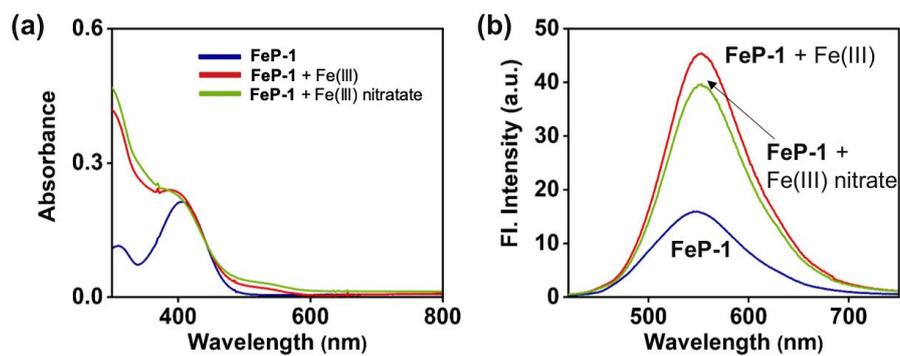


Figure S8. (a, b) Absorption and emission spectra of **FeP-1** (10 μM) upon addition of Fe(III) (20 eq), Fe(III) nitrate (20 eq) in ethanol-water solution (EtOH:DI H₂O = 4:6, *v/v*) after 1 min at 25 $^{\circ}\text{C}$.

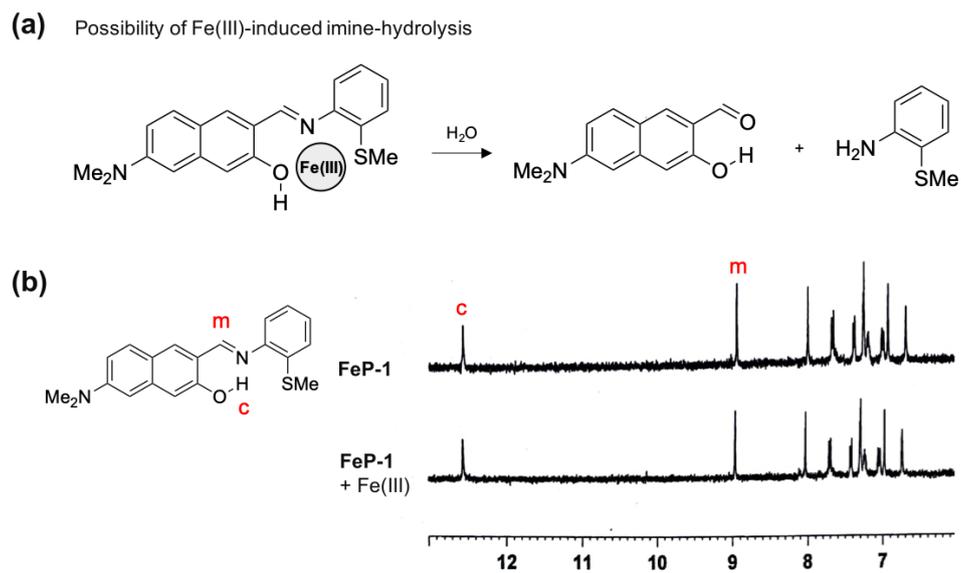


Figure S9. (a) Predicted hydrolysis pathway of **FeP-1**+Fe(III) in aqueous media. (b) ^1H NMR spectra for **FeP-1** (upper, 0.1 mg/mL) and its reaction product (bottom) with Fe(III) (1 mg/mL), 1 h incubation in ethanol-water solution (EtOH:DI H₂O = 4:6, *v/v*) at 25 °C, followed by extraction using dichloromethane. NMR solvent was DMSO-*d*₆.

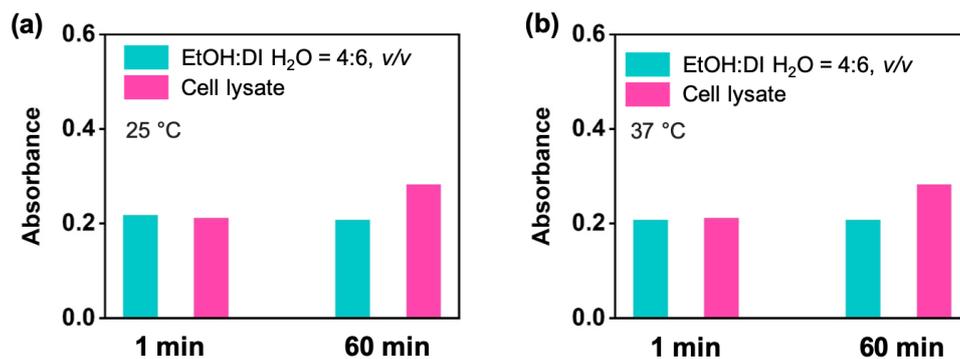
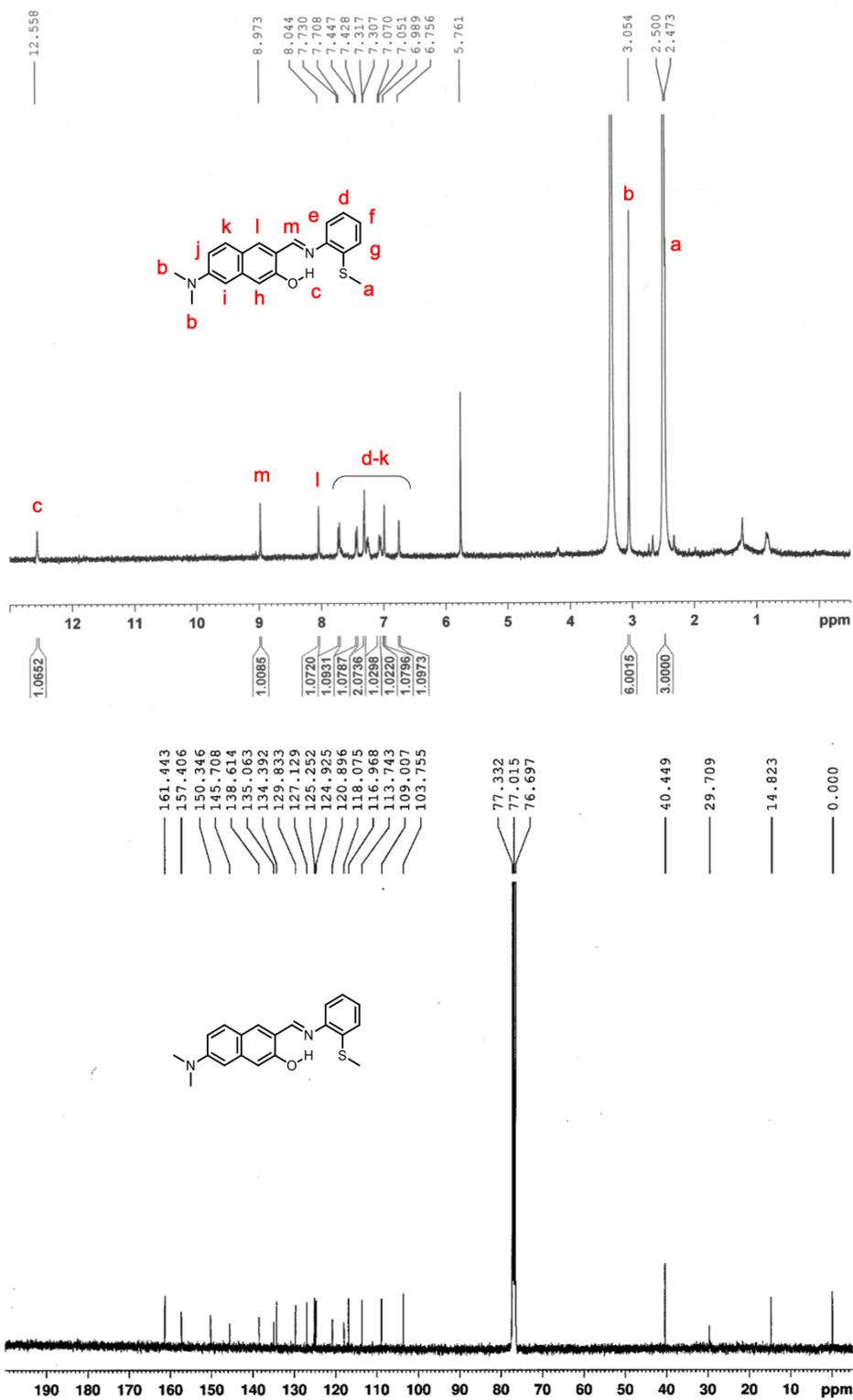
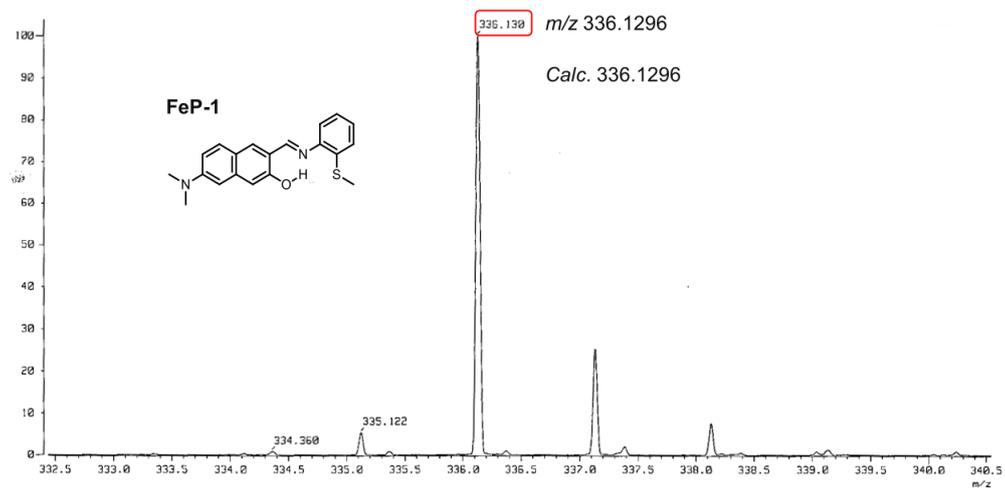


Figure S10. Absorption intensity of **FeP-1** (10 μM) in ethanol-water solution (EtOH:DI H₂O = 4:6, *v/v*) and HeLa cell lysate solution with incubation at (a) 25 °C and (b) 37 °C. The absorption intensity at 407 nm was plotted at 1 min and 60 min after adding the **FeP-1** to each solution. [Cell lysate solution] Lysis buffer (100 μL) was added to HeLa cells (5×10^5) in a 100 mm dish. After tapping the dish with hands, the resulting mixture was centrifuged at 14,000 rpm for 10 min, and the supernatant was collected. DMEM media (2 mL, 10% fetal bovine serum and 1% penicillin-streptomycin, w/o phenol red) was add to the supernatant.

^1H and ^{13}C NMR of FeP-1



High-resolution mass spectra for FeP-1



References and Notes

Figure S1(a)

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Figure S1(b)

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Figure S1(c)

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Figure S1(d)

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Figure S1(e)

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