## **Supplementary Information**

## **Table of Contents**

Table S1	Cell cycle distribution of MDA-MB-231 cells treated with the active						
	ascidian extracts was determined by flow cytometry through DNA						
	quantification of cells stained with propidium iodide						
Figure S1	Morphology analysis of MDA-MB-231 cells treated with the indicated						
	ascidian extracts (1 µge/µL) after 24 h seeding						
<b>S1</b>	Experimental data for the bis-TFA salt of eusynstyelamide B (1)						
Figure S2	HPLC UV chromatogram of the active Didemnum candidum extract						
	(ascidian code: 114) selected for bioassay-guided fractionation						
Figure S3	<sup>1</sup> H NMR spectrum of the <i>bis</i> -TFA salt of eusynstyelamide B (1) in						
	DMSO- $d_6$						
Figure S4	$^{13}$ C NMR spectrum of the <i>bis</i> -TFA salt of eusynstyelamide B (1) in						
	DMSO- $d_6$						
Figure S5	COSY spectrum of the <i>bis</i> -TFA salt of eusynstyelamide (1) in DMSO- <i>d</i> <sub>6</sub>						
Figure S6	HSQC spectrum of the <i>bis</i> -TFA salt of eusynstyelamide (1) in DMSO- <i>d</i> <sub>6</sub>						
Figure S7	HMBC spectrum of the <i>bis</i> -TFA salt of eusynstyelamide (1) in DMSO- $d_6$						
Figure S8	Dose response curves for the <i>bis</i> -TFA salt of eusynstyelamide B (1, EB) in						
	the breast cancer cell line MDA-MB-231 at 72 h post-treatment						

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Extract	Sub-G1		G0/G1		S		G2/M	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
DMSO	6.4	1.7	43.1	3.2	52.0	0.9	5.0	2.3
15	15.2 *	2.2	18.3 *	1.6	67.6 *	2.3	14.1	3.0
17	8.4	0.6	1.4 *	0.2	62.4	0.6	36.2 *	0.3
29	5.8	1.5	18.7 *	15.7	69.4 *	11.9	11.9	9.5
38	11.1	3.4	19.3 *	2.6	72.2 *	4.1	8.5	2.3
43	5.0	1.7	30.3	4.1	60.5	1.2	9.2	3.0
44	4.4	2.5	33.5	15.6	55.7	6.3	10.8	9.3
53	4.5	0.6	41.1	2.4	52.7	3.6	6.2	1.5
61	8.0	1.6	22.4 *	10.4	71.4 *	6.0	6.2	4.4
63	17.2 *	6.6	1.3 *	0.8	84.4 *	9.5	14.4	2.0
71	3.4	1.0	27.7 *	1.7	66.4 *	0.5	5.9	1.3
75	6.4	4.9	4.3 *	2.4	95.7 *	2.4	0.0	0.0
81	14.2 *	4.7	3.8 *	1.0	75.1 *	2.0	21.1 *	1.0
83	6.8	1.6	16.7 *	6.3	58.5	2.9	24.9 *	9.2
85	6.5	1.9	39.0	0.6	56.0	2.7	5.7	1.4
92	5.3	1.5	31.7	2.8	63.3	4.4	5.0	4.7
102	2.1	5.4	28.9 *	0.5	64.9	1.1	6.3	0.6
106	8.6	1.2	51.4	0.8	44.3	1.0	4.3	1.1
114	16.4 *	2.7	5.5 *	0.1	69.6 *	1.1	25.0 *	0.8
117	9.7	3.5	54.0	1.6	39.8	2.7	6.1	1.4
128	5.2	1.7	31.6	3.4	58.6	1.2	9.8	2.3
133	7.2	4.2	43.9	7.4	49.8	5.4	6.3	2.1

**Table S1.** Cell cycle distribution of MDA-MB-231 cells treated with the active ascidian extracts was determined by flow cytometry through DNA quantification of cells stained with propidium iodide. The results are presented as mean  $\pm$  SD of triplicates. Significant samples compared to the control are marked with an asterisk \* (p < 0.05).

**Figure S1.** Morphology analysis of MDA-MB-231 cells treated with the indicated ascidian extracts (200 µge) after 24 h seeding. As controls, cells were treated with complete medium, DMSO (0.1%), or doxorubicin (Dox, 10 µM). The morphology of the cells after treatment was categorized in five groups (A–F). (A) Cells not fully attached; (B) Cells with typical signs of early stages of cell death; (C) Cells well attached to substrate and in contact with other cells; (D) Flat and enlarged cells; (E) High number of cells detached from substrate. The images were obtained with an OlympusIX70 microscope using a  $10 \times$  objective. Scale bar = 100 µm.



## S1. Experimental Data for the bis-TFA Salt of Eusynstyelamide B (1)

*Bis*-TFA salt of eusynstyelamide B (1). Isolated as a stable brown gum (3.5 mg);  $[\alpha]_D^{26} = \pm 0$ (*c* 0.133, MeOH); literature value  $[\alpha]_D^{19} = \pm 0$  (*c* 0.100, MeOH) [1, 2]; CD (MeOH)  $\lambda_{max} (\Delta \epsilon)$  208 (-3.0), 227 (-4.6), 298 (-1.1) nm; literature value CD (MeOH)  $\lambda_{max}$  ( $\Delta\epsilon$ ) 224 (-11) nm [1,2]; <sup>1</sup>H NMR <sup>a</sup> (600 MHz, DMSO-d<sub>6</sub>)  $\delta_{\rm H}$  10.93 (1H, d, J = 2.3, 1-NH), 6.86 (1H, d, J = 2.3, H-2), 7.01 (1H, d, J = 8.5 Hz, H-4), 6.77 (1H, dd, J = 8.5, 1.7 Hz, H-5), 7.45 (1H, d, J = 1.7 Hz, H-7), 3.26 (1H, d, J = 14.5 Hz, H-8a), 2.78 (1H, d, J = 14.5 Hz, H-8b), 6.18 (1H, s, 9-OH), 3.21 (1H, m, H-12a), 2.91 (1H, m, H-12b), 1.48 (2H, m, H-13), 1.40 (2H, tt, J = 6.5, 6.5 Hz, H-14), 3.04 (2H, dt, J = 5.5, 6.5 Hz, H-15), 7.49 (1H, t, J = 5.5 Hz, H-16), 11.15 (1H, d, J = 2.4 Hz, 18-NH), 7.65 (1H, d, J = 2.4 Hz, H-19), 6.81 (1H, d, J = 8.5 Hz, H-21), 6.92 (1H, dd, J = 8.5, 1.7 Hz, H-22), 7.51 (1H, d, J = 1.7 Hz, H-24), 4.02 (1H, s, H-25), 5.42 (1H, s, 26-OH), 7.72 (1H, t, J = 6.0 Hz, H-28), 2.88 (1H, m, H-29a), 2.76 (1H, m, H-29b), 1.12 (2H, m, H-30), 1.12 (2H, m, H-31), 2.87 (2H, m, H-32), 7.37 (1H, t, J = 5.8 Hz, H-33); <sup>13</sup>C NMR (125 MHz, DMSO-d<sub>6</sub>) & 125.6 (C-2), 109.1 (C-3), 126.7 (C-3a), 120.5 (C-4), 120.5 (C-5), 113.6<sup>c</sup> (C-6), 113.3 (C-7), 136.6 (C-7a), 29.0 (C-8), 78.2 (C-9), 175.3 (C-10), 40.2 (C-12), 25.9<sup>d</sup> (C-13), 25.5 (C-14), 40.3 (C-15), 156.7<sup>b</sup> (C-17), 126.7 (C-19), 105.9 (C-20), 127.5 (C-20a), 120.3 (C-21), 120.8 (C-22), 113.5<sup>c</sup> (C-23), 113.5 (C-24), 135.9 (C-24a), 44.0 (C-25), 90.3 (C-26), 168.9 (C-27), 38.4 (C-29), 25.1<sup>d</sup> (C-30), 25.6 (C-31), 40.0 (C-32), 156.6<sup>b</sup> (C-34); (+)-LRESIMS (rel. int.) m/z 787 (30%)  $[M - 2CF_3COO^- + H]^+$ , 789 (100%)  $[M - 2CF_3COO^- + H]^+$ , 791 (30%)  $[M - 2CF_3COO^- + H]^+$ . a Signals for the exchangeable signals 17-NH<sub>3</sub><sup>+</sup>, 17-NH, 34-NH<sub>3</sub><sup>+</sup>, and 34-NH were not assigned; <sup>b-d</sup> Signals are interchangeable.



**Figure S2.** HPLC UV chromatogram of the active *Didemnum candidum* (Extract 114) selected for bioassay-guided fractionation. Every minute of the chromatogram represents one fraction. The yellow shading highlights the active region identified through bioassay-guided fractionation, which corresponds to **1**.







Figure S4. <sup>13</sup>C NMR spectrum of the *bis*-TFA salt of eusynstyelamide B (1) in DMSO-*d*<sub>6</sub>.





Figure S5. COSY spectrum of the bis-TFA salt of eusynstyelamide B (1) in DMSO-d6.

Figure S6. HSQC spectrum of the bis-TFA salt of eusynstyelamide B (1) in DMSO-d6.





Figure S7. HMBC spectrum of the *bis*-TFA salt of eusynstyelamide B (1) in DMSO-*d*<sub>6</sub>.

Figure S8. Dose response curves for eusynstyleamide B (1, EB) in the breast cancer cell line MDA-MB-231 at 72 h post treatment. The data used to calculate the IC<sub>50</sub> of 1 in MDA-MB-231 were acquired in an AlamarBlue<sup>®</sup> assay.



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